

- tremor in Europeans. *Neurobiol Aging* 2013;34(10):2441 e2449-2441 e2411.
5. Hedera P, Davis TL, Phibbs FT, Charles PD, LeDoux MS. FUS in familial essential tremor—the search for common causes is still on. *Parkinsonism Relat Disord* 2013;19(9):818-820.
 6. Labbe C, Soto-Ortolaza AI, Rayaprolu S, et al. Investigating the role of FUS exonic variants in essential tremor. *Parkinsonism Relat Disord* 2013;19(8):755-757.
 7. Zheng W, Deng X, Liang H, et al. Genetic analysis of the fused in sarcoma gene in Chinese Han patients with essential tremor. *Neurobiol Aging* 2013;34(8):2078 e2073-2078 e2074.
 8. Annesi F, De Marco EV, Rocca FE, et al. Association study between the LINGO1 gene and Parkinson's disease in the Italian population. *Parkinsonism Relat Disord* 2011;17(8):638-641.
 9. Radovica I, Inashkina I, Smeltzer L, Vitols E, Jankevics E. Screening of 10 SNPs of LINGO1 gene in patients with essential tremor in the Latvian population. *Parkinsonism Relat Disord* 2012;18(1):93-95.
 10. Wu YW, Rong TY, Li HH, et al. Analysis of Lingo1 variant in sporadic and familial essential tremor among Asians. *Acta Neurol Scand* 2011;124(4):264-268.
 11. Klebe S, Thier S, Lorenz D, et al. LINGO1 is not associated with Parkinson's disease in German patients. *Am J Med Genet B* 2010;153B(6):1173-1178.
 12. Clark LN, Park N, Kisselev S, Rios E, Lee JH, Louis ED. Replication of the LINGO1 gene association with essential tremor in a North American population. *Eur J Hum Genet* 2010;18(7):838-843.
 13. Vilarino-Guell C, Wider C, Ross OA, et al. LINGO1 and LINGO2 variants are associated with essential tremor and Parkinson disease. *Neurogenetics* 2010;11(4):401-408.
 14. Tan EK, Teo YY, Prakash KM, et al. LINGO1 variant increases risk of familial essential tremor. *Neurology* 2009;73(14):1161-1162.
 15. Vilarino-Guell C, Ross OA, Wider C, et al. LINGO1 rs9652490 is associated with essential tremor and Parkinson disease. *Parkinsonism Relat Disord* 2010;16(2):109-111.
 16. Knudsen K, Lorenz D, Deuschl G. A clinical test for the alcohol sensitivity of essential tremor. *Mov Disord* 2011;26(12):2291-2295.
 17. Mostile G, Jankovic J. Alcohol in essential tremor and other movement disorders. *Mov Disord* 2010;25(14):2274-2284.
 18. Wu YR, Foo JN, Tan LC, et al. Identification of a novel risk variant in the FUS gene in essential tremor. *Neurology* 2013;81(6):541-544.
 19. Kuhlentbauer G, Tan EK, Wu YR. Identification of a novel risk variant in the FUS gene in essential tremor. *Neurology* 2014;82(5):461.
 20. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 2009;25(14):1754-1760.
 21. McKenna A, Hanna M, Banks E, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res* 2010;20(9):1297-1303.
 22. Ng SB, Turner EH, Robertson PD, et al. Targeted capture and massively parallel sequencing of 12 human exomes. *Nature* 2009;461(7261):272-276.
 23. Deng H, Gu S, Jankovic J. LINGO1 variants in essential tremor and Parkinson's disease. *Acta Neurol Scand* 2012;125(1):1-7.
 24. Deng H, Le WD, Hunter CB, Mejia N, Xie WJ, Jankovic J. A family with Parkinson disease, essential tremor, bell palsy, and parkin mutations. *Arch Neurol* 2007;64(3):421-424.
 25. Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007;81(3):559-575.
 26. Matise TC, Chen F, Chen W, et al. A second-generation combined linkage physical map of the human genome. *Genome Res* 2007;17(12):1783-1786.
 27. Deng H, Le W, Davidson AL, Xie W, Jankovic J. The LRRK2 I2012T, G2019S and I2020T mutations are not common in patients with essential tremor. *Neurosci Lett* 2006;407(2):97-100.
 28. Deng H, Le WD, Guo Y, Huang MS, Xie WJ, Jankovic J. Extended study of A265G variant of HS1BP3 in essential tremor and Parkinson disease. *Neurology* 2005;65(4):651-652.
 29. Deng H, Le W, Jankovic J. Premutation alleles associated with Parkinson disease and essential tremor. *JAMA* 2004;292(14):1685-1686.
 30. Shatunov A, Sambuughin N, Jankovic J, et al. Genomewide scans in North American families reveal genetic linkage of essential tremor to a region on chromosome 6p23. *Brain* 2006;129(pt 9):2318-2331.
 31. Deng H, Le W, Jankovic J. Genetics of essential tremor. *Brain* 2007;130(pt 6):1456-1464.

Hypothalamic Volume Loss Is Associated With Reduced Melatonin Output in Parkinson's Disease

David P. Breen, MRCP, PhD,^{1*} Cristina Nombela, PhD,¹ Romina Vuono, PhD,¹ P. Simon Jones, MSc,² Kate Fisher, PhD,¹ David J. Burn, FRCP, MD,³ David J. Brooks, MD, DSc,^{4,5} Akhilesh B. Reddy, PhD, MRCP,⁶ James B. Rowe, MRCP, PhD,^{2,7,8} and Roger A. Barker, MRCP, PhD¹

¹John van Geest Centre for Brain Repair, University of Cambridge, Cambridge, UK ²Department of Clinical Neurosciences, University of Cambridge, Cambridge, UK ³Institute of Neuroscience, Newcastle University, Newcastle, UK ⁴Division of Neurology, Imperial College, London, UK ⁵Institute of Clinical Medicine, Aarhus University, Aarhus, Denmark ⁶Institute of Metabolic Science, University of Cambridge, Cambridge, UK ⁷Behavioural and Clinical Neuroscience Institute, University of Cambridge, Cambridge, UK ⁸Medical Research Council Cognition and Brain Sciences Unit, Cambridge, UK

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

*Correspondence to: Dr. David P. Breen, John van Geest Centre for Brain Repair, University of Cambridge, ED Adrian Building, Forvie Site, Robinson Way, Cambridge, CB2 0PY, UK; dpbreen1@gmail.com

Present address for Cristina Nombela: School of Medicine, Universidad Politécnica de Cartagena, Murcia, Spain.

Present address for Kate Fisher: School of Chemistry, University of Edinburgh, Edinburgh UK.

David P. Breen and Cristina Nombela contributed equally to the manuscript.

Funding agencies: The authors acknowledge the study funders: the Big Lottery Fund (C498A738) and Parkinson's UK (J-0802). The research was supported by a National Institute of Health Research Biomedical Research Award (to Addenbrooke's Hospital/University of Cambridge), the Wellcome Trust (103838, 100333/Z/12/Z), and a Raymond and Beverly Sackler Studentship (to D.P.B.).

Relevant conflicts of interests/financial disclosures: The authors report no conflicts of interest.

Received: 23 August 2015; **Revised:** 1 February 2016; **Accepted:** 3 February 2016

Published online 12 March 2016 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.26592

ABSTRACT

Background: Recent studies have suggested that melatonin—a hormone produced by the pineal gland under circadian control—contributes to PD-related sleep dysfunction. We hypothesized that degenerative changes to the neural structures controlling pineal function (especially the suprachiasmatic nuclei of the anterior hypothalamus) may be responsible for reduced melatonin output in these patients. We compared hypothalamic volumes in PD patients with matched controls and determined whether volume loss correlated with reduced melatonin output in the PD group.

Methods: A total of 12 PD patients and 12 matched controls underwent magnetic resonance imaging to determine hypothalamic volume. In addition, PD patients underwent 24-hour blood sampling in a controlled environment to determine serum melatonin concentrations using enzyme-linked immunosorbent assays.

Results: PD patients had significantly reduced hypothalamic gray matter volume when compared with matched controls. Melatonin levels were significantly associated with hypothalamic gray matter volume and disease severity in PD patients.

Conclusion: Melatonin levels are associated with hypothalamic gray matter volume loss and disease severity in PD patients. This provides anatomical and physiological support for an intrinsic sleep and circadian phenotype in PD. © 2016 The Authors. *Movement Disorders* published by Wiley Periodicals, Inc. on behalf of International Parkinson and Movement Disorder Society

Key Words: melatonin, hypothalamus, suprachiasmatic nucleus, Parkinson's, circadian

Sleep disturbances are one of the most common nonmotor complaints in Parkinson's disease (PD) and have been attributed to a variety of factors. Understanding the relative contribution of each is crucial to identify the most effective treatment strategies for individual patients. Some of these relate to the clinically identified features of the disease such as motor impairment, nocturia, pain, or neuropsychiatric symptoms. Dopaminergic and other medications may also exacerbate patients' sleep problems. However, the sleep dysfunction in PD may be a result of neuronal loss in key structures and circuits involved in the regulation of the sleep-wake cycle.

Two recent studies have reported that reduced melatonin output in PD patients is associated with altered sleep architecture, including reduced slow wave and rapid eye movement sleep¹ and excessive daytime sleepiness.² Altered melatonin patterns have also been observed in Huntington's disease³ and Alzheimer's disease,⁴ both of which have prominent sleep and circadian abnormalities. Because melatonin is a hormone

produced by the pineal gland under circadian control, we hypothesized that degenerative changes to the neural structures controlling pineal function (especially the suprachiasmatic nuclei [SCN] of the hypothalamus) may reduce melatonin output and contribute to certain aspects of sleep dysfunction in PD.

This study compared hypothalamic volumes in PD patients with matched controls and determined whether volume loss correlated with reduced melatonin output in the PD group.

Methods**Patients**

A total of 12 PD patients were selected from a previously studied sleep cohort.¹ All of the patients who had also undergone magnetic resonance imaging (MRI) as part of the parallel Incidence of Cognitive Impairment in Cohorts with Longitudinal Evaluation—Parkinson's Disease (ICICLE-PD) study were included in the analysis, in addition to 12 unrelated matched controls from the Medical Research Council Cognition and Brain Sciences Unit healthy volunteer panel. The ICICLE-PD protocol has been published elsewhere.⁵ All participants provided written consent, the study was performed according to the Declaration of Helsinki, and the protocol was approved by the local research ethics committee.

In brief, patients underwent a battery of clinical tests including the Unified Parkinson's Disease Rating Scale (MDS-UPDRS), Addenbrooke's Cognitive Examination (ACE-R), and the Beck Depression Inventory (BDI). Levodopa equivalent dose (LED) was calculated using the conversion factors proposed by Tomlinson and colleagues.⁶ Matching was based on age, gender, years of education, and ACE-R.

Imaging Acquisition and Analysis

MRI data were acquired using a Siemens TIM Trio 3T scanner (Siemens Medical Systems, Erlangen, Germany). Patients underwent T1-weighted magnetisation prepared rapid gradient echo scanning: pulse repetition = 2250 ms, echo time = 2.98 ms, flip angle = 9°, time delay = 900 ms, 256 × 256 mm² field of view, 192 × 1 mm slices). Images were preprocessed according to a pipeline in SPM8 (<http://www.fil.ion.ucl.ac.uk/spm>) run on Matlab 7 (Mathworks, Natick, Massachusetts). T1-weighted images were segmented into gray matter and white matter tissue and registered through the Diffeomorphic Anatomical Registration Through Exponentiated Lie Algebra scheme. The resulting study-specific template was registered to Montreal Neurological Institute space, and individual modulated images were smoothed with an 8 mm, full width at half-maximum Gaussian kernel. A hypothalamic region of interest (dilated by 3 mm) from the

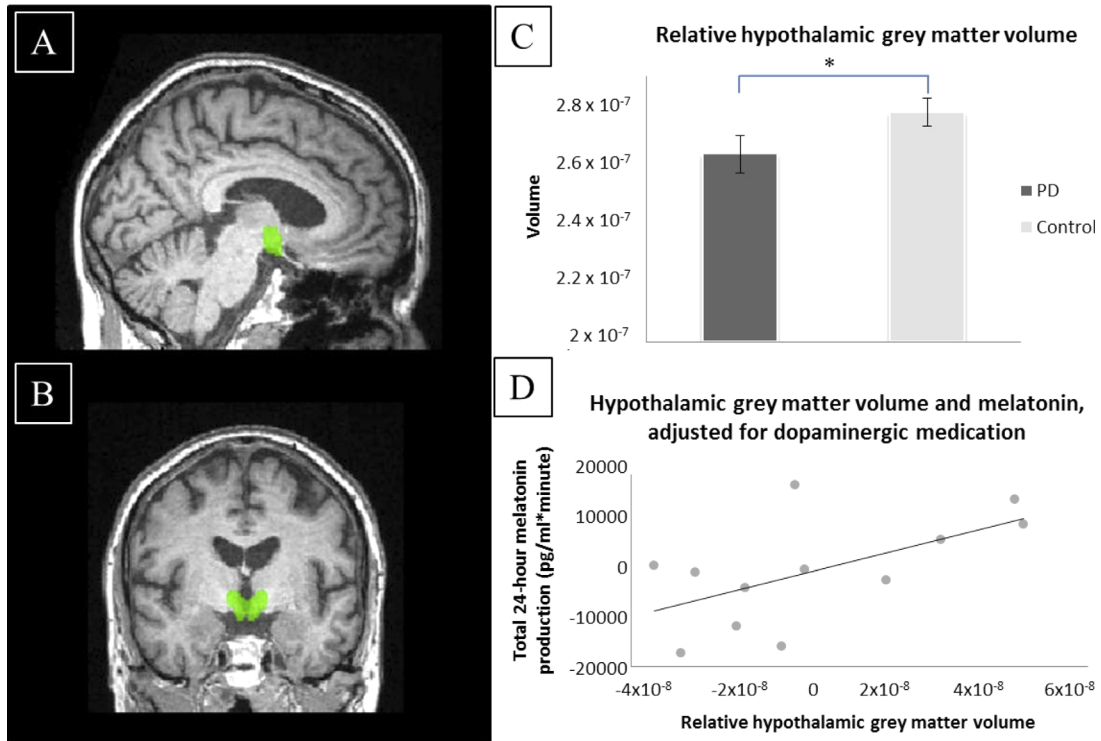


FIG. 1. **A** and **B**: Panels show the region of interest used to calculate the hypothalamic volume for each participant. **C**: Panel is a graphical representation of the significant reduction in relative hypothalamic gray matter volume in PD patients when compared with matched controls (with standard error of the mean error bars). **D**: Panel demonstrates the significant correlation between relative hypothalamic gray matter volume and total 24-hour melatonin output (with both axes showing partial residuals). In both graphs, relative hypothalamic gray matter volume was calculated by dividing gray matter volume by whole brain volume (both measured in voxels). [Color figure can be viewed in the online issue, which is available at wileyonlinelibrary.com.]

WFU Pick Atlas (<http://fmri.wfubmc.edu/software/pickatlas>) was used to obtain an individual hypothalamic volume per participant (Fig. 1A,B). Gray matter volume in the region of interest (measured in voxels) was calculated using the FMRIB Software Library (FSL) tool “fslstats” within FSL version 4.1.7 (www.fmrib.ox.ac.uk). Thereafter, relative hypothalamic gray matter volume was calculated by dividing by whole brain volume (the sum of the gray and white matter segments).

Serum Melatonin Measurement

PD patients were admitted to a single room at the Wellcome Trust Clinical Research Facility at Addenbrooke’s Hospital in Cambridge. A peripheral venous cannula was inserted 30 minutes before the start of sampling at 1:00 PM. During the next 24 hours, participants adhered to their habitual bed times, and blood samples were collected every 90 minutes using a 3-way valve that was attached to a 0.9% sodium chloride infusion to prevent the cannula from clotting. Sampling was performed through a long line to prevent disruption to the patient’s sleep. Participants remained sedentary apart from bathroom visits. Meal times were consistent between participants, and no daytime naps were allowed. Temperature was constant

at 21 °C. Patients were not strictly shielded from external light, but lighting levels were less than 5 lux following lights off. Serum melatonin concentrations were measured using enzyme-linked immunosorbent assays as previously described.¹ Based on hormone

TABLE 1. Clinical characteristics of PD patients and controls

| Variable | PD | Controls | P value ^d |
|---------------------------------------|------------|------------|----------------------|
| Number of participants | 12 | 12 | na |
| Gender ratio (male:female) | 6:6 | 6:6 | 1.0 |
| Age (years) | 66.7 (5.5) | 66.3 (5.2) | 0.92 |
| Duration of education (years) | 18.3 (2.9) | 17.2 (2.8) | 0.33 |
| ACE-R | 95.2 (3.1) | 96.1 (3.0) | 0.48 |
| Disease duration (years) ^a | 3.3 (1.1) | na | na |
| LEDD (mg) ^b | 366 (161) | na | na |
| MDS-UPDRS part III ^c | 23.9 (9.0) | na | na |
| BDI | 7.3 (17.8) | 3.3 (3.6) | 0.011* |

*Significant difference at .05 level.
 LEDD, levodopa equivalent daily dose; ACE-R, Addenbrooke’s Cognitive Examination-Revised; MDS-UPDRS, Unified Parkinson’s Disease Rating Scale; BDI, Beck Depression Inventory; na, not applicable.
 Results expressed as mean (SD) unless stated otherwise
^aSignificant difference at .05 level.
^bDisease duration from date of diagnosis.
^cAll but two PD patients were taking dopaminergic medication.
^dBased on MDS-UPDRS assessments performed within the last 6 months.
^eUnpaired t tests performed.

concentrations at each 90-minute time point, total 24-hour melatonin output was defined as the area under the curve (calculated using the trapezoid rule).

Statistical Analysis

All data were approximately normally distributed based on Shapiro-Wilk testing; therefore, unpaired *t* tests were used to compare clinical parameters and volumetric values between patients and controls. Pearson rank correlation testing was used to study the relationship between melatonin output and relative hypothalamic gray matter volume, as well as the relationship between melatonin output and disease severity (adjusted for LED).

Results

Age, gender, duration of education, and ACE-R were not significantly different between PD patients and controls (Table 1). PD patients had a mean disease duration of 3.3 years, mean LED of 366 mg, and mean UPDRS part III score of 23.9. None of the participants were taking hypnotics. The mean duration between melatonin testing and MRI in the PD group was 1.9 months (SD 3.4).

When compared with controls, PD patients had significantly reduced relative hypothalamic gray matter volume (2.56×10^{-7} [SD 2.78×10^{-7}] vs 2.69×10^{-7} [SD 2.07×10^{-7}]; $P = .005$) (Fig. 1C).

Having verified that there were significant differences between patients and controls in terms of hypothalamic volume, we found that melatonin levels were significantly associated with relative hypothalamic gray matter volume in the PD group ($r = .591$, $P = .028$) (Fig. 1D).

Partial correlation between melatonin levels and disease severity, correcting for LED, showed a significant inverse relationship ($r = -.681$, $P = .021$). There was no significant relationship between melatonin output and LED ($r = .180$, $P = .76$).

Discussion

There is increasing evidence from clinical and animal studies that there is circadian dysregulation in a variety of neurodegenerative diseases.⁷ We previously reported significant reductions in melatonin concentration in 30 early-stage PD patients.¹ Videnovic and colleagues also found a significantly diminished amplitude of melatonin secretion in serum samples of 20 PD patients on dopaminergic therapy under modified constant routine conditions.²

There is evidence from neuropathological⁸ and imaging⁹ studies that the hypothalamus is directly affected by PD. The central clock within the hypothalamus, the SCN, is likely to contribute to this volume loss because it has been shown that mice overexpress-

ing alpha-synuclein exhibit a reduced SCN firing rate.¹⁰ This could weaken their ability to communicate neural and hormonal signals from the central clock to the pineal gland, which secretes melatonin into the blood.

This study thus adds to the existing literature by suggesting that hypothalamic volume loss—which we have now shown in this new PD cohort—may be responsible for reduced melatonin output, which has been linked to sleep disturbances in PD.

The major limitation of this study is the relatively small number of patients, which precluded the use of linear regression and adjustment of confounders. Furthermore, patients were not strictly shielded from external light during the melatonin sampling period, which may have influenced the results. Although we lacked serum melatonin measurements in the control group, the critical test for our hypothesis was the correlation between hypothalamic volume and melatonin levels in PD patients. It is not yet possible to perform dedicated imaging of the SCN within the hypothalamus using 3Tesla MRI; therefore, ultra-high field imaging or clinico-pathological studies will be required to allow more thorough dissection of the relative role of the different hypothalamic nuclei to this deficit.

In summary, we have shown that melatonin levels are associated with hypothalamic gray matter volume loss and disease severity in PD patients. This provides anatomical and physiological support for an intrinsic sleep and circadian phenotype in PD and the fact that this is related to the disease itself rather than being an indirect consequence of other symptoms or treatments. ■

Acknowledgments: We would like to thank Dr. Saber Sami for his assistance with imaging acquisition and Dr. Noham Wolpe for his technical support. We would also like to thank the staff at the Wellcome Trust Clinical Research Facility in Addenbrooke's Hospital, Cambridge, for performing the melatonin blood sampling.

References

1. Breen DP, Vuono R, Nawarathna U, et al. Sleep and circadian rhythm regulation in early Parkinson disease. *JAMA Neurol* 2014; 71:589-595.
2. Videnovic A, Noble C, Reid KJ, et al. Circadian melatonin rhythm and excessive daytime sleepiness in Parkinson disease. *JAMA Neurol* 2014;71:463-466.
3. Kakkilola E, Silajdzic E, Nambron R, et al. Plasma melatonin is reduced in Huntington's disease. *Mov Disord* 2014;29:1511-1515.
4. Mishima K, Tozawa T, Satoh K, et al. Melatonin secretion rhythm disorders in patients with senile dementia of Alzheimer's type with disturbed sleep-waking. *Biol Psychiatry* 1999;45:417-421.
5. Nombela C, Rowe JB, Winder-Rhodes SE, et al. Genetic impact on cognition and brain function in newly diagnosed Parkinson's disease: ICICLE-PD study. *Brain* 2014;137:2743-2758.
6. Tomlinson CL, Stowe R, Patel S, et al. Systematic review of levodopa dose equivalency reporting in Parkinson's disease. *Mov Disord* 2010;25:2649-2653.
7. Videnovic A, Lazar AS, Barker RA, et al. 'The clocks that time us'—circadian rhythms in neurodegenerative disorders. *Nat Rev Neurol* 2014;10:683-693.
8. Langston JW, Forno LS. The hypothalamus in Parkinson disease. *Ann Neurol* 1978;3:129-133.

9. Politis M, Piccini P, Pavese N, et al. Evidence of dopamine dysfunction in the hypothalamus of patients with Parkinson's disease: an *in vivo* 11C-raclorpride PET study. *Exp Neurol* 2008;214:112-116.
10. Kudo T, Loh DH, Truong D, et al. Circadian dysfunction in a mouse model of Parkinson's disease. *Exp Neurol* 2011;232:66-75.

GBA Mutations Are Associated With Earlier Onset and Male Sex in Dementia With Lewy Bodies

Ana Gámez-Valero, MSc,^{1,2} Patricia Prada-Dacasa, MSc,^{1,3} Cristina Santos, PhD,³ Cristina Adame-Castillo, BSc,¹ Jaume Campdelacreu, MD,⁴ Ramón Reñé, MD,⁴ Jordi Gascón-Bayarri, MD,⁴ Lourdes Ispuerto, MD,⁵ Ramiro Álvarez, MD,⁵ Aurelio Ariza, MD,¹ and Katrin Beyer, PhD^{1*}

¹Department of Pathology, Hospital Universitari and Health Sciences Research Institute Germans Trias i Pujol, Universitat Autònoma de Barcelona, Barcelona, Spain ²IVECAT Group, Health Sciences Research Institute Germans Trias i Pujol, Badalona, Spain ³Unitat d'Antropologia Biològica, Departament de Biologia Animal, Biologia Vegetal i Ecologia, Universitat Autònoma de Barcelona, Barcelona, Spain ⁴Department of Neurology, Hospital Universitari de Bellvitge, L'Hospitalet de Llobregat, Spain ⁵Department of Neurology, Hospital Universitari Germans Trias i Pujol, Badalona, Barcelona, Spain

Abstract

Background: Parkinson's disease (PD) and dementia with Lewy bodies (DLB) are Lewy body diseases characterized by similar pathological features. Several studies have shown a relation between alterations in the glucocerebrosidase gene (*GBA*) and the development of LB diseases. Here, we explored the role of *GBA* mutations in Spanish DLB patients.

Methods: *GBA* mRNA sequences were analyzed in a neuropathological (50 DLB, 43 PD, and 34 control brains) and in a clinical cohort (47 DLB patients and 131 unaffected individuals).

Results: Sixteen *GBA* mutation carriers were identified, 5 of which were brains with pure DLB. The most common mutation, E326K, was strongly associated with pure

DLB and PD with dementia. *GBA* mutations were overrepresented in men and associated with earlier DLB onset.

Conclusions: *GBA* mutations are also an important risk factor for DLB development in the Spanish population, are associated with earlier disease onset, and are more prevalent in men. © 2015 International Parkinson and Movement Disorder Society

Key Words: *GBA* mutations, Parkinson's disease, dementia with Lewy bodies

Both Parkinson's disease (PD) and dementia with Lewy bodies (DLB) are characterized neuropathologically by the presence of α -synuclein-immunopositive Lewy bodies (LBs) in the brain.¹ Their morphological substrate is the same, and although the clinical course of DLB and PD with dementia (PDD) differs, it is difficult to detect neuropathological differences between the 2 conditions. When LB pathology represents the main neuropathological finding, cases are classified as pure LB pathology, and when concomitant Alzheimer's disease (AD) pathology is present as common LB pathology.² About 20%-50% of PD patients develop dementia 10 to 15 years following PD diagnosis, and its incidence increases with age and disease duration.^{3,4}

The lysosomal enzyme glucocerebrosidase (GCase) is responsible for the breakdown of glucocerebroside into glucose and ceramide.⁵ Mutations in the GCase gene *GBA* cause GCase deficiency,⁶ leading to lysosomal glucocerebroside accumulation, resulting in Gaucher's disease (GD).⁵ Because PD is more frequent in first-degree relatives with GD,⁷⁻⁹ *GBA* has been studied in PD and later also in DLB patients, showing a strong association between *GBA* mutations and synucleinopathies.¹⁰⁻¹⁶ Two extensive multicenter studies confirmed the strong association between *GBA* mutations and PD as well as DLB, with odds ratios of 5.43 and 8.28, respectively.^{17,18}

The reduction of GCase levels in PD is associated with an increase in abnormal α -synuclein accumulation.¹⁶ Two models that aim to explain the role of *GBA* mutations for LB disease development are a gain-of-function model and a loss-of-function model. Supporting the first hypothesis, *GBA* mutants were found to enhance α -synuclein accumulation.¹⁹ In the loss-of-function model, lysosomal glucocerebroside accumulation directly promoted the formation of α -synuclein fibrils, which inhibit GCase activity, creating a bidirectional pathogenic loop.²⁰

Although several studies have shown the association between *GBA* mutations and LB diseases in diverse populations, only 1 work evaluating the role of *GBA* mutations in PD has been carried out in the Spanish population.²¹ Here, we have characterized *GBA*

*Correspondence to: Katrin Beyer, PhD, Department of Pathology, Hospital Universitari Germans Trias i Pujol, 08916 Badalona, Barcelona, Spain, E-mail: katrinbeyer@hotmail.com

Funding agencies: This work was supported by Spain's Ministry of Health FIS grant PI12/1702 and the Marató TV3 grant 201405/10.

Relevant conflicts of interests/financial disclosures: The authors report no conflict of interest regarding the contents of the article, and no financial conflict is declared.

Received: 30 November 2015; **Revised:** 3 February 2016; **Accepted:** 7 February 2016

Published online 29 March 2016 in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/mds.26593