

Complete Genome Sequences of *Bordetella pertussis* Vaccine Reference Strains 134 and 10536

Michael R. Weigand, Yanhui Peng, Vladimir Loparev, Dhvani Batra, Mark Burroughs, Taccara Johnson, Phalasy Juieng, Lori Rowe, M. Lucia Tondella, Margaret M. Williams

Centers for Disease Control and Prevention, Atlanta, Georgia, USA

Vaccine formulations and vaccination programs against whooping cough (pertussis) vary worldwide. Here, we report the complete genome sequences of two divergent *Bordetella pertussis* reference strains used in the production of pertussis vaccines.

Received 19 July 2016 Accepted 22 July 2016 Published 15 September 2016

Citation Weigand MR, Peng Y, Loparev V, Batra D, Burroughs M, Johnson T, Juieng P, Rowe L, Tondella ML, Williams MM. 2016. Complete genome sequences of *Bordetella pertussis* vaccine reference strains 134 and 10536. *Genome Announc* 4(5):e00979-16. doi:10.1128/genomeA.00979-16.

Copyright © 2016 Weigand et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Michael R. Weigand, mweigand@cdc.gov.

Bordetella pertussis is the primary causative agent of whooping cough (pertussis), a respiratory disease most severe in unvaccinated infants. The introduction of vaccines against pertussis dramatically reduced disease incidence worldwide. However, many countries have recently experienced disease resurgence, in part due to genetic divergence of circulating strains. The resulting antigenic mismatch with vaccine references has led many to conclude that *B. pertussis* is evolving under vaccine-driven selection (1–5). Adaptation of *B. pertussis* is complicated by the varied administration of whole-cell and acellular vaccines between countries and the diversity of reference strains used for vaccine production (6–8). Here, we report the complete genome sequences of two such strains used in manufacturing pertussis vaccines: B202 (Lederle Laboratories, strain 134) and B203 (Sanofi-Pasteur MSD, strain 10536) (9).

Whole-genome shotgun sequencing was performed using a combination of the PacBio RSII (Pacific Biosciences, Menlo Park, CA), Illumina HiSeq/MiSeq (Illumina, San Diego, CA), and Argus (OpGen, Gaithersburg, MD) platforms, as described previously (10). Briefly, genomic DNA libraries were prepared for PacBio sequencing using the SMRTbell template prep kit 1.0 and polymerase binding kit P4, while Illumina libraries were prepared using the NEBNext Ultra library prep kit (New England BioLabs, Ipswich, MA). *De novo* genome assembly of filtered reads was performed using the Hierarchical Genome Assembly Process (HGAP version 3; Pacific Biosciences) at 130× and 144× coverage for B202 and B203, respectively. The resulting consensus sequences were determined with Quiver (version 1), manually checked for circularity, and then reordered to match the start of reference strain Tohama I (accession no. CP010964) (10). To ensure accuracy, assemblies were confirmed by comparison to BamHI and KpnI restriction digestion optical maps using the Argus system (OpGen) with MapSolver (version 2.1.1; OpGen) and further polished by mapping either Illumina HiSeq PE-100 or MiSeq PE-300 reads using CLC Genomics Workbench (version 8.5; CLC bio, Boston, MA). Final assemblies were annotated using

the NCBI automated Prokaryotic Genome Annotation Pipeline (PGAP).

The average G+C content of both B202 and B203 was 67.1%, with genome sizes of 4,128,979 and 4,134,643 bp, respectively. Genome annotation identified 3,645 protein-coding genes in B202 and 3,636 protein-coding genes in B203. Both genomes encoded three rRNA operons and 51 tRNAs.

The assemblies were distinct from genomes of vaccine reference strains Tohama I (GlaxoSmithKline, accession no. CP010964), CS (China, accession no. CP010963), and 137 (Brazil, accession no. CP010323), which have been sequenced previously (10, 11). B202 and B203 were not related, and their genomes differed from that of Tohama I by multiple rearrangements, as well as 186 and 410 single-nucleotide polymorphisms (SNPs), respectively. The genome of B202 was phylogenetically and structurally similar, but not identical, to other strains with the profile *prn1-ptxP1-ptxA2-ptxB2-fimH1*, such as clinical isolate H375 (accession no. CP010961) (10). B203 appeared to be closely related to Brazilian vaccine strain 137, sharing allele profile *prn7-ptxP2-ptxA4-ptxB2-fimH1*, but differed by 13 SNPs and a single ~74-kb inversion flanked by rRNA operon copies.

The availability of these genome sequences provides added resolution to known diversity among references used in vaccine production and will hopefully aid in the research of immune response to clinical infection in vaccinated patients.

Accession number(s). The complete genome sequences have been deposited at DDBJ/EMBL/GenBank under the accession numbers [CP016338](https://accession.cdc.gov/CP016338) and [CP012128](https://accession.cdc.gov/CP012128) for *B. pertussis* B202 and B203, respectively. The versions described in this paper are the first versions.

ACKNOWLEDGMENTS

We thank Pam Cassidy for technical assistance with bacterial culture and Bruce Meade (U.S. Food and Drug Administration) for supplying strains.

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the Centers for Disease Control and Prevention.

FUNDING INFORMATION

This work was supported by internal funds.

REFERENCES

1. Bart MJ, Harris SR, Advani A, Arakawa Y, Bottero D, Bouchez V, Cassiday PK, Chiang CS, Dalby T, Fry NK, Gaillard ME, van Gent M, Guiso N, Hallander HO, Harvill ET, He Q, van der Heide HG, Heuvelman K, Hozbor DF, Kamachi K, Karataev GI, Lan R, Lutynska A, Maharjan RP, Mertsola J, Miyamura T, Octavia S, Preston A, Quail MA, Sintchenko V, Stefanelli P, Tondella ML, Tsang RS, Xu Y, Yao SM, Zhang S, Parkhill J, Mooi FR. 2014. Global population structure and evolution of *Bordetella pertussis* and their relationship with vaccination. *mBio* 5:e01074-14. <http://dx.doi.org/10.1128/mBio.01074-14>.
2. Octavia S, Maharjan RP, Sintchenko V, Stevenson G, Reeves PR, Gilbert GL, Lan R. 2011. Insight into evolution of *Bordetella pertussis* from comparative genomic analysis: evidence of vaccine-driven selection. *Mol Biol Evol* 28:707–715. <http://dx.doi.org/10.1093/molbev/msq245>.
3. Sealey KL, Harris SR, Fry NK, Hurst LD, Gorringer AR, Parkhill J, Preston A. 2015. Genomic analysis of isolates from the United Kingdom 2012 pertussis outbreak reveals that vaccine antigen genes are unusually fast evolving. *J Infect Dis* 212:294–301. <http://dx.doi.org/10.1093/infdis/jiu665>.
4. van Gent M, Bart MJ, van der Heide HG, Heuvelman KJ, Mooi FR. 2012. Small mutations in *Bordetella pertussis* are associated with selective sweeps. *PLoS One* 7:e46407. <http://dx.doi.org/10.1371/journal.pone.0046407>.
5. Xu Y, Liu B, Grondahl-Yli-Hannuksela K, Tan Y, Feng L, Kallonen T, Wang L, Peng D, He Q, Wang L, Zhang S. 2015. Whole-genome sequencing reveals the effect of vaccination on the evolution of *Bordetella pertussis*. *Sci Rep* 5:12888.
6. Van Amersfoort SC, Schouls LM, van der Heide HG, Advani A, Hallander HO, Bondeson K, von König CH, Riffelmann M, Vahrenholz C, Guiso N, Caro V, Njamkepo E, He Q, Mertsola J, Mooi FR. 2005. Analysis of *Bordetella pertussis* populations in European countries with different vaccination policies. *J Clin Microbiol* 43:2837–2843. <http://dx.doi.org/10.1128/JCM.43.6.2837-2843.2005>.
7. Kallonen T, Grøndahl-Yli-Hannuksela K, Elomaa A, Lutyńska A, Fry NK, Mertsola J, He Q. 2011. Differences in the genomic content of *Bordetella pertussis* isolates before and after introduction of pertussis vaccines in four European countries. *Infect Genet Evol* 11:2034–2042. <http://dx.doi.org/10.1016/j.meegid.2011.09.012>.
8. Kilgore PE, Salim AM, Zervos MJ, Schmitt HJ. 2016. Pertussis: microbiology, disease, treatment, and prevention. *Clin Microbiol Rev* 29:449–486. <http://dx.doi.org/10.1128/CMR.00083-15>.
9. Litt DJ, Neal SE, Fry NK. 2009. Changes in genetic diversity of the *Bordetella pertussis* population in the United Kingdom between 1920 and 2006 reflect vaccination coverage and emergence of a single dominant clonal type. *J Clin Microbiol* 47:680–688. <http://dx.doi.org/10.1128/JCM.01838-08>.
10. Bowden KE, Weigand MR, Peng Y, Cassiday PK, Sammons S, Knipe K, Rowe LA, Loparev V, Sheth M, Weening K, Tondella ML, Williams MM. 2016. Genome structural diversity among 31 *Bordetella pertussis* isolates from two recent U.S. whooping cough statewide epidemics. *mSphere* 1:e00036-16. <http://dx.doi.org/10.1128/mSphere.00036-16>.
11. Akamatsu MA, Nishiyama MY, Jr, Morone M, Oliveira UC, Bezerra MF, Sakauchi MA, Raw I, Junqueira de Azevedo IL, Kitajima JP, Carvalho E, Ho PL. 2015. Whole-genome sequence of a *Bordetella pertussis* Brazilian vaccine strain. *Genome Announc* 3(1):e01570-14. <http://dx.doi.org/10.1128/genomeA.01570-14>.