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Inhibition promotes long-term potentiation at cerebellar excitatory synapses

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The ability of the cerebellar cortex to learn from experience ensures the accuracy of movements and reflex adaptation, processes which require long-term plasticity at granule cell (GC) to Purkinje neuron (PN) excitatory synapses. PNs also receive GABAergic inhibitory inputs via GCs activation of interneurons; despite the involvement of inhibition in motor learning, its role in long-term plasticity is poorly characterized. Here we reveal a functional coupling between ionotropic GABA_A receptors and low threshold Ca_v3 calcium channels in PNs that sustains calcium influx and promotes long-term potentiation (LTP) at GC to PN synapses. High frequency stimulation induces LTP at GC to PN synapses and Ca_v3-mediated calcium influx provided that inhibition is intact; LTP is mGluR1, intracellular calcium store and Ca_v3 dependent. LTP is impaired in Ca_v3.1 knockout mice but it is nevertheless recovered by strengthening inhibitory transmission onto PNs; promoting a stronger hyperpolarization via GABA_A receptor activation leads to an enhanced availability of an alternative Purkinje-expressed Ca_v3 isoform compensating for the lack of Ca_v3.1 and restoring LTP. Accordingly, a stronger hyperpolarization also restores Ca_v3-mediated calcium influx in PNs from Ca_v3.1 knockout mice. We conclude that by favoring Ca_v3 channels availability inhibition promotes LTP at cerebellar excitatory synapses.

In everyday life, we all benefit from the fine work performed by the cerebellum which allows us to fine tune our movements during daily actions in response to environmental changes and while executing complicate tasks such as walking or playing the violin. In line with its role in adaptive control of skilled movements and motor learning^{1–4} the cerebellum receives vestibular, sensory and motor information which are conveyed from the entire body to the cerebellar cortex where they converge to Purkinje neurons (PNs). PNs are the sole output of the cerebellar cortex and they receive two main excitatory inputs, parallel fibers (PFs) and climbing fibers (CFs). PFs are the axons of granule cells (GCs) which relay proprioceptive, somatosensory and vestibular information reaching the cerebellum via mossy fibers (MFs) originating from several pre-cerebellar nuclei in the brainstem and spinal cord (Fig. 1a).

Long-term plasticity has been described at GC to PN synapses with long-term depression (LTD) caused by co-activation of PFs and CF⁵ while the sole PFs stimulation leads to long-term potentiation (LTP)^{6–10}. Furthermore, long lasting alterations in the strength of transmission at these excitatory synapses have been proposed as the molecular basis of cerebellar motor learning¹¹. Indeed, mouse models lacking key molecules for LTD and LTP induction also show impairment in adaptation of the vestibulo-ocular reflex (VOR), a well-established model for cerebellum-mediated motor learning^{10,12–15}.

The same bundle of PFs which excites PNs also drives molecular layer interneurons (MLIs: basket and stellate cells, Fig. 1a) which provide feedforward inhibition (FFI) through GABAergic inhibitory synapses on the same PN. PFs-mediated excitation in PNs is therefore balanced by inhibition which ultimately influences the final output of the cerebellum. Indeed, a precise time window for PFs excitatory inputs summation and spike generation in PNs is defined by FFI¹⁶. Sustained PFs activation also promotes repetitive MLIs stimulation and therefore FFI-mediated influence on long-term plasticity is also expected. Accordingly, the block of inhibition promotes LTD¹⁷ and LTP is

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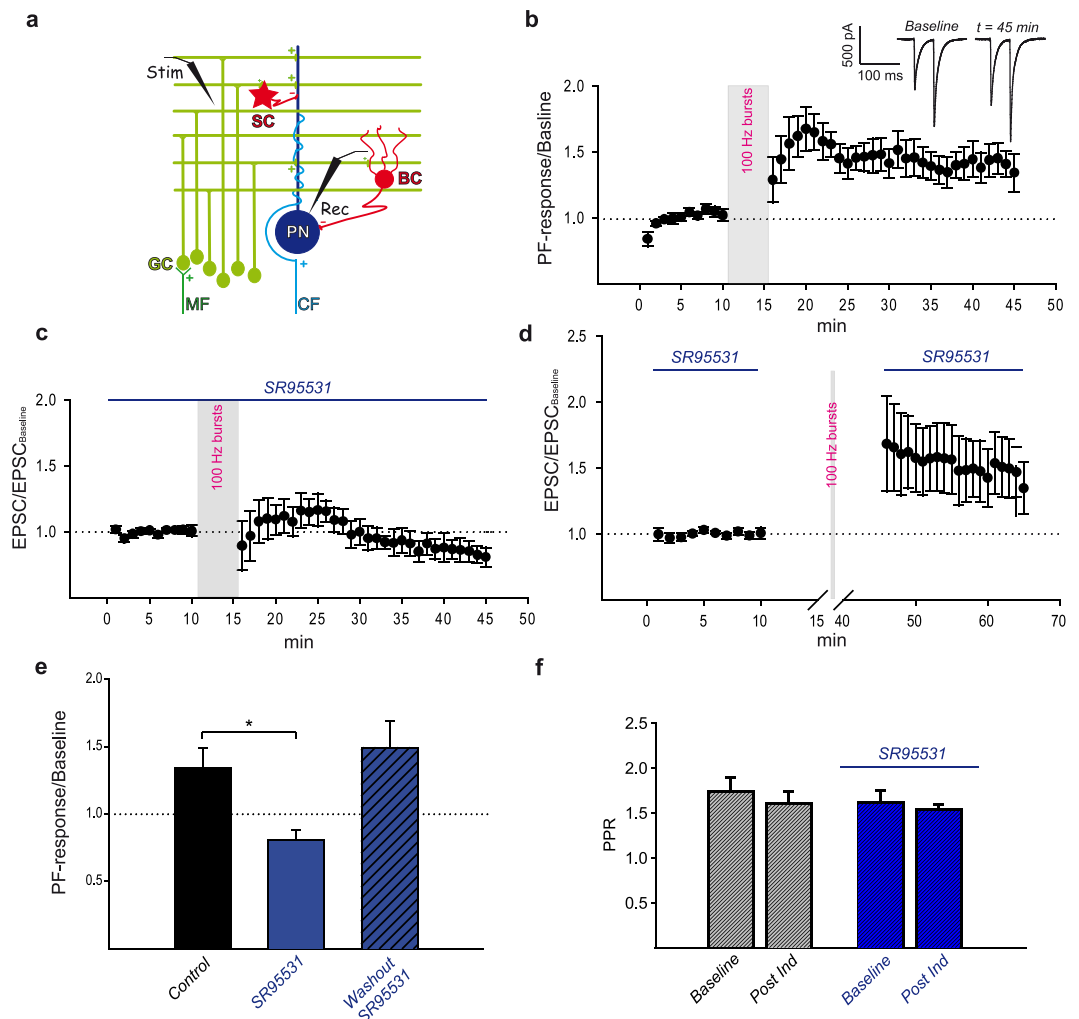


Figure 1. High frequency PFs stimulation induces GABA_A receptor dependent LTP at PF to PN synapses. A schematic representation of the cerebellar microcircuit and experimental setting is shown in panel **a**. Purkinje neurons (PN) receive excitatory inputs from climbing fibers (CF) and mossy fibers (MF) via granule cells (GC) activation. Parallel fibers excitatory synapses (green+) drive PNs and inhibitory (red-) molecular layer interneurons stellate (SC) and basket (BC) cells. Traces elicited by paired-pulses PFs stimulation (Stim, **a**) in a voltage-clamped PN (Rec, **a**) at different time points are showed in the inset of panel **b**. The *Baseline* trace was obtained from averaging of all recordings during baseline while the *t = 45 min* trace is the average of three consecutive PF-mediated responses recorded every 20 seconds at the indicated time point. High frequency PFs stimulation induced a long lasting increase in PNs response (MLI_{dep}-LTP; **b**: mean ± SEM, N = 5, RM ANOVA P < 0.001). Bath application of the GABA_A receptor antagonist SR95531 prevented MLI_{dep}-LTP (**c**, mean ± SEM, N = 5, RM ANOVA P < 0.001). A summary graph for SR95531-mediated effect on MLI_{dep}-LTP is shown in panel **e** (bar represents normalized PF-Rsp at t = 45 min, mean ± SEM, data from panel **b** and **c**), *indicates a statistically significant difference among values (t-test, P = 0.011). Keeping GABAergic transmission intact only during high frequency PFs stimulation was sufficient to induce MLI_{dep}-LTP (**d**). For these experiments, a 10 minutes baseline was established with SR95531 and the antagonist was washed out for at least 15 minutes before the induction protocol was applied; SR95531 was added back to the recording chamber immediately or 15 minutes after high frequency stimulation (**d**, mean ± SEM, N = 7, RM ANOVA P < 0.001; **e**, bar represents normalized PF-Rsp at t = 65 min, mean ± SEM, data from panel **d**). PPRs value (mean ± SEM) for the baseline (t = 10) and the post induction phase (t = 45) with or without bath application of SR95531 are shown in panel **f**.

modulated by activation of the ionotropic GABA_A receptor *in vivo*¹⁸. In addition, mice selectively lacking synaptic GABA_A receptors in PNs show impairment of consolidation of the vestibulo-cerebellar motor learning¹⁹.

How inhibition modulates long-term plasticity at PF to PN synapses is nevertheless still unknown. PNs express low threshold voltage-gated T-type calcium channels (Ca_v3) which have been recently linked to calcium signaling, LTP at PF to PN synapses and to some aspects of motor learning^{15,20,21}. Ca_v3 channels are opened by small depolarization of the cell membrane and they generate a transient current which rapidly inactivates²². Recovery from inactivation requires cell membrane re-hyperpolarization and therefore Ca_v3 channels can be influenced by inhibitory inputs as shown in thalamic neurons where GABAergic transmission promotes T-type

channels-mediated low threshold calcium spikes^{23,24}. By investigating the molecular pathway leading to LTP in PNs, we demonstrate a tight cooperation between ionotropic GABA receptors and low threshold voltage-gated T-type calcium channels (Ca_v3) which promotes LTP at PF to PN synapses. We show that potentiation of transmission at PF to PN synapses following high frequency PFs stimulation requires calcium influx via Ca_v3 channels and GABA_A receptor activation. Also, we provide evidence supporting Ca_v3 modulation by inhibitory inputs leading to an increase in channels availability. We then conclude that FFI can control LTP at PF to PN synapses.

Results

LTP at PF to PN synapses is GABA_A receptor-dependent. The role played by inhibition in LTP induction at PF to PN synapses was investigated in acute cerebellar slices. In voltage clamped PNs ($V_h = -60$ mV), molecular layer electrical paired-pulses stimulation (20 Hz) induced PF-mediated fast inward currents with facilitation at the second response (Fig. 1b inset, Baseline). The induced response (PF-Rsp) was stable within 10 minutes from the beginning of the recording (Fig. 1b, average baseline value: -781.31 ± 25.77 pA, mean \pm SEM, $N = 5$). Once baseline was established, PFs were stimulated at high frequency (burst of 15 pulses at 100 Hz repeated every 3 seconds for a period of 5 minutes) while PNs were switched to current clamp mode. This induction protocol was chosen to mimic GCs physiological activity^{25–27} and to ensure reliable PF to MLI transmission whose failure rate has been shown to decrease at high frequency rate²⁸. Train of PFs bursts stimulation caused a long lasting potentiation of the PFs-induced response (Fig. 1b). Higher values were reached at an early phase (Fig. 1b, normalized PF-Rsp_{t20} = 1.68 ± 0.18 , mean \pm SEM, $N = 5$) with a later stable lower level maintained until the end of the recording (steady state) (Fig. 1b inset, $t = 45$ min; Fig. 1b, normalized PF-Rsp_{t45} = 1.34 ± 0.14 , mean \pm SEM, $N = 5$). Since LTP can be expressed at presynaptic and/or postsynaptic site²⁹, the paired-pulse ratio (PPR) was investigated to discriminate between these possibilities. PPRs at steady state showed no statistically significant difference when compared to baseline (Fig. 1f, PPR_{t10} = 1.74 ± 0.15 , PPR_{t45} = 1.6 ± 0.13 , mean \pm SEM; PPR_{t10} vs PPR_{t45}: one-way repeated measures (RM) ANOVA, Tukey's post hoc test, $P > 0.05$, $N = 5$) indicating that a change in the presynaptic probability of release was unlikely.

The role of GABAergic transmission in LTP induction was then pharmacologically investigated by bath application of the selective GABA_A receptor antagonist SR95531 (5 μ M). PNs holding current (I_{holding}) and baseline PF-mediated response decay time constant (τ_{off}) displayed no statistically significant difference in the presence of the antagonist (I_{holding} SR95531: -346.74 ± 44.44 pA, mean \pm SEM, $N = 5$, I_{holding} control: -241.63 ± 55.46 , mean \pm SEM, $P = 0.177$, t test, $N = 5$; τ_{off} SR95531: 7.14 ± 0.8 ms, mean \pm SEM, τ_{off} control: 5.49 ± 2 ms, mean \pm SEM, $P = 0.467$, t -test, $N = 5$) suggesting that, under our experimental conditions, PNs were clamped at a membrane potential ($V_h = -60$ mV) close to the equilibrium potential of GABA_A receptor-mediated currents (calculated $E_{Cl} = -63$ mV) and that the PF-mediated response was mostly mediated by AMPA receptors.

Blocking the GABA_A receptor had a profound effect on LTP (Fig. 1e) which was completely abolished by SR95531 (Fig. 1c, $t = 45$, normalized EPSC = 0.81 ± 0.07 , mean \pm SEM, $N = 5$; Fig. 1e, mean \pm SEM, t -test, $P = 0.011$) with no statistically significant change in PPRs (Fig. 1f, PPR_{t10} = 1.61 ± 0.13 , PPR_{t45} = 1.53 ± 0.06 , mean \pm SEM; PPR_{t10} vs PPR_{t45}: one-way repeated measures (RM) ANOVA, Tukey's post hoc test, $P > 0.05$, $N = 5$).

LTP at PF to PN synapses could therefore be induced by high frequency PFs stimulation provided that the ionotropic GABAergic transmission was intact, supporting a role of MLIs in long-term potentiation at PF to PN synapses; for simplicity we will refer to LTP established under our experimental condition as MLI dependent LTP (MLI_{dep}-LTP) in the later part of the text.

PF to MLI synapses and the MLIs network can also be affected by long-term plasticity that could ultimately influence our recordings^{30–34}. We therefore isolated the effect of high frequency PFs stimulation on EPSCs by applying SR95531 during the baseline and the post induction phase while maintaining inhibition active during the induction protocol. Keeping inhibitory transmission intact while applying PFs bursts stimulation was necessary and sufficient to establish MLI_{dep}-LTP at PF to PN synapses (Fig. 1d) with no statistically significant change in PPRs (PPR_{t10} = 1.78 ± 0.08 , PPR_{t65} = 1.58 ± 0.1 , mean \pm SEM; PPR_{t10} vs, PPR_{t65}, RM ANOVA, Tukey's post hoc test, $P = 0.054$, $N = 7$). Furthermore, EPSC increase was comparable to the potentiation obtained with intact GABAergic transmission (Fig. 1e,d: normalized EPSC_{t65} = 1.49 ± 0.2 ; Fig. 1e,b: normalized PF-response_{t45} = 1.34 ± 0.14 , $P = 0.593$, t -test).

Taken together our data support the requirement of GABA_A receptor activation in MLI_{dep}-LTP; MLI_{dep}-LTP is likely postsynaptic and it is expressed at PF to PN synapses.

Molecular pathway to MLI_{dep}-LTP. Low threshold voltage-gated Ca_v3 channels have recently been linked to postsynaptic LTP at PF to PN synapses¹⁵ and they are influenced by inhibitory inputs as shown in thalamic and deep cerebellar nuclei (DCN) neurons^{23,24,35–37}. We therefore investigated their possible role in MLI_{dep}-LTP by pharmacologically blocking their activation with the highly specific T-type calcium channels antagonist TTA-P2³⁸ (Merck; 500 nM). In the presence of the Ca_v3 channels antagonist, high frequency PFs stimulation failed to induce MLI_{dep}-LTP with the induced responses slightly decreasing below baseline level after the first transient increment (Fig. 2a, $t = 45$, normalized PF-Rsp = 0.9 ± 0.09 mean \pm SEM, $N = 5$); PPRs showed no statistically significant difference from baseline (Fig. 2a inset, PPR_{t10} = 1.5 ± 0.072 , PPR_{t45} = 1.5 ± 0.04 ; mean \pm SEM; PPR_{t10} vs PPR_{t45}, RM ANOVA, Tukey's post hoc test, $P > 0.05$, $N = 5$).

All three Ca_v3 isoforms are found in PNs with weak $Ca_v3.2$ channels staining³⁹ and more pronounced $Ca_v3.1$ and $Ca_v3.3$ channels expression^{39,40}. PFs bursts-induced calcium transients in mature PNs are mostly mediated by $Ca_v3.1$ channels²¹ and $Ca_v3.1$ KO mice showed LTP impairment¹⁵. We therefore investigated whether $Ca_v3.1$ channels play a role in MLI_{dep}-LTP in acute cerebellar slices from $Ca_v3.1$ KO mice⁴¹. Under the same experimental condition which led to MLI_{dep}-LTP in WT mice, $Ca_v3.1$ KO mice showed no MLI_{dep}-LTP (Fig. 2b, $N = 5$).

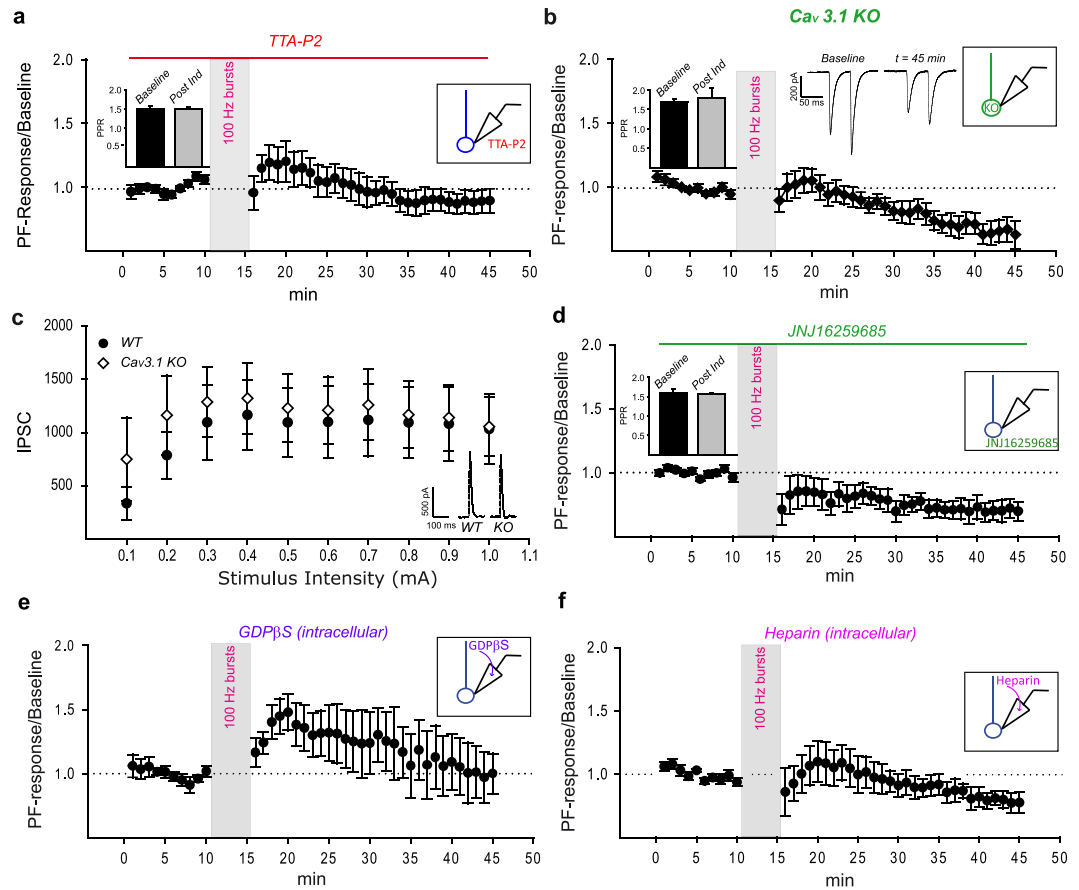


Figure 2. Molecular pathway to MLI_{dep} -LTP. MLI_{dep} -LTP was effectively prevented by the broad spectrum low threshold voltage-gated T-type calcium channels specific antagonist TTA-P2 (**a**, mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$). To identify the Ca_v3 isoform required for the GABA-dependent potentiation, long-term plasticity experiments were performed in $Ca_v3.1$ KO mice. MLI_{dep} -LTP is absent in $Ca_v3.1$ KO mice (**b**, mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$) under the same experimental condition that leads to GABA_A receptor-dependent potentiation of PF to PN transmission in WT mice and traces from a representative experiment are shown in the middle inset. PF-induced inhibitory responses were recorded in PNs from WT and $Ca_v3.1$ KO mice (**c**, inset) and input/output curves obtained (**c**: WT, black circle: $N = 8$; KO, white diamond: $N = 10$, mean \pm SEM). No statistically significant difference was detected among the two curves ($P > 0.05$, t-test at all stimulus intensities). MLI_{dep} -LTP also depends on mGluR1 and intracellular calcium stores; high frequency PFs stimulation failed to induce MLI_{dep} -LTP when activation of the metabotropic glutamate receptor mGluR1 was prevented by bath application of the specific antagonist JNJ16259685 (**d**, mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$). Inclusion of the non-hydrolysable GDP analog GDP β S (**e**, mean \pm SEM, $N = 3$) or heparin (**f**, mean \pm SEM, $N = 6$, RM ANOVA $P = 0.006$) in the intracellular recording solution also impaired MLI_{dep} -LTP. PPRs value (mean \pm SEM) for the baseline ($t = 10$ min) and the post induction phase ($t = 45$ min) under each condition are shown in the left insets in panel **a,b,d**.

Also, PPRs value showed no statistically significant difference to baseline (Fig. 2b inset, $PPR_{t10} = 1.68 \pm 0.1$, $PPR_{t45} = 1.8 \pm 0.26$, mean \pm SEM; PPR_{t10} vs PPR_{t45} , RM ANOVA, Tukey's post hoc test, $P > 0.05$, $N = 5$).

In order to exclude any effect of $Ca_v3.1$ absence on MLI to PN transmission that could potentially interfere with MLI_{dep} -LTP induction, the total PF-induced inhibition was measured in WT and $Ca_v3.1$ KO mice. Input/output curves showed no significant difference in $Ca_v3.1$ KO mice compared to WT animals (Fig. 2c, minimum stimulus intensity: WT = 382.88 ± 152.41 pA, $N = 8$; KO = 748.95 ± 391.01 pA, $N = 10$, $P = 0.379$, t-test) supporting a normal MLI to PN transmission. The absence of MLI_{dep} -LTP is therefore a direct consequence of impaired calcium influx in $Ca_v3.1$ KO PNs dendrites¹⁵ rather than a secondary effect due to altered inhibition.

A functional coupling between mGluR1 receptors and $Ca_v3.1$ channels has been identified in PNs with the activation of the G-coupled receptor leading to the potentiation of calcium influx through T-type calcium channels²¹ and therefore mGluR1 activation may be required for the described long-term potentiation. Indeed, blocking mGluR1 by bath application of the specific antagonist JNJ16259685 ($2 \mu M$) prevented MLI_{dep} -LTP (Fig. 2d, $N = 5$) with no statistically significant change in PPRs when compared to baseline (Fig. 2d inset, $PPR_{t10} = 1.59 \pm 0.09$, $PPR_{t45} = 1.57 \pm 0.03$, mean \pm SEM, $N = 5$; PPR_{t10} vs PPR_{t45} , $P > 0.05$, RM ANOVA, Tukey's test). Strikingly, blocking the receptor also revealed a mGluR1-independent LTD (normalized PF-Rsp_{t45} = 0.7 ± 0.07 , normalized PF-Rsp_{t11} = 1 ± 0.02 , $P = 0.025$ RM ANOVA, Tukey's post hoc test). As expected by MLI_{dep} -LTP impairment

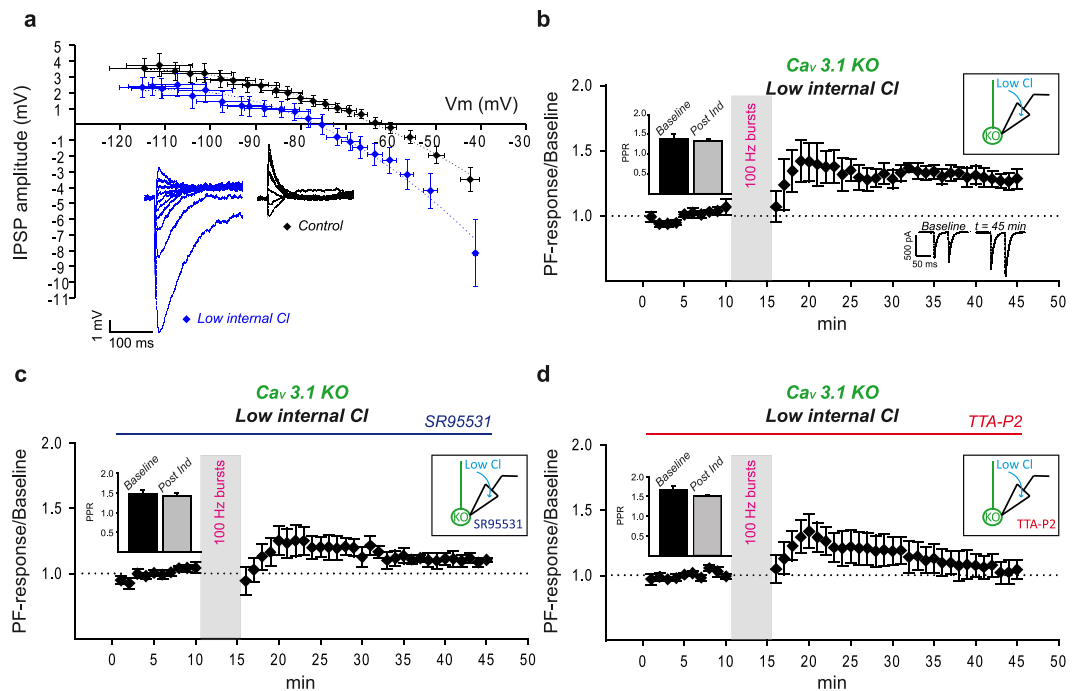


Figure 3. MLI_{dep} -LTP is $Ca_v3.1$ dependent and it relies on $GABA_A$ receptor and T-type calcium channels cooperation. Change in intracellular chloride concentration affects IPSPs/ V_m curves in PNs from $Ca_v3.1$ KO mice. Lowering intracellular chloride caused a 13 mV hyperpolarizing shift of IPSP/ V_m curves (a, blue diamond, $N = 7$, mean \pm SEM) compared to control condition (a, black diamond, $N = 7$, mean \pm SEM). IPSPs recorded at different V_m from two representative PNs are shown in the inset of panel a in control (black) and low chloride (blue) condition. Dot lines represent the polynomial fit of IPSP/ V_m curves obtained under the two experimental conditions. MLI_{dep} -LTP was rescued when the low internal chloride recording solution was used (b, mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$) and traces from one exemplificative experiment are shown in the inset at the bottom of the panel. The rescued MLI_{dep} -LTP was dependent on $GABA_A$ receptor (c: mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$) and T-type calcium channels (d: mean \pm SEM, $N = 5$, RM ANOVA $P < 0.001$) activation. PPRs value (mean \pm SEM) for the baseline ($t = 10$ min) and the post induction phase ($t = 45$ min) under each condition are showed in the left insets in panel c,d.

caused by mGluR1 inactivation in PNs, the inclusion of the non-hydrolysable GDP analog GDP β S (2 mM) in the patch pipette also interfered with long term plasticity (Fig. 2e, $N = 3$). mGluR1 activation induces release of calcium from intracellular stores which could therefore be implicated in MLI_{dep} -LTP. When calcium release from intracellular store via IP3 receptors was prevented by inclusion of heparin (50 μ g/ml) in the patch pipette⁴², MLI_{dep} -LTP was also impaired (Fig. 2f, $N = 6$) suggesting that calcium influx via Ca_v3 channels and internal stores might cooperate for MLI_{dep} -LTP induction.

These experiments support the requirement of a postsynaptic molecular cascade for the expression of MLI_{dep} -LTP at PF to PN synapses; we showed that MLI_{dep} -LTP is Ca_v3 channels, mGluR1 and internal calcium stores dependent.

Ionotropic $GABA$ receptors and Ca_v3 channels cooperation is required for MLI_{dep} -LTP. Ca_v3 channels open in response to small depolarization of the cell, quickly inactivate and recovery from inactive state depends on cell membrane re-hyperpolarization after channel opening²². The three Ca_v3 channel isoforms are characterized by different activation curves with $Ca_v3.1$ (half activation V_a : -60 ± 0.9 mV at 37°C) and $Ca_v3.2$ channels (half activation V_a : -51.5 ± 1 mV at 37°C) opening at more depolarized membrane potential compared to $Ca_v3.3$ channels (half activation V_a : -73.5 ± 1.3 mV at 37°C)⁴³ which therefore requires stronger hyperpolarization to de-inactivate. Modulation of $GABA_A$ -mediated transmission could therefore impact Ca_v3 channels availability and affect MLI_{dep} -LTP. We investigated this hypothesis by lowering the chloride concentration in the internal solution for MLI_{dep} -LTP experiments in $Ca_v3.1$ KO mice to obtain a stronger $GABA_A$ receptor-mediated hyperpolarization in PNs as shown by the 13 mV negative shift of the IPSP reversal potential when compared to control condition (control, $N = 7$: -63 mV; low chloride, $N = 7$: -76 mV) (Fig. 3a); a stronger hyperpolarization might enhance $Ca_v3.3$ channels availability and therefore improve Ca_v3 channels-mediated intracellular calcium rise in the $Ca_v3.1$ KO during bursts of PFs stimulation and facilitate MLI_{dep} -LTP induction. Indeed, under these experimental conditions, high frequency PFs stimulation generated a long lasting increase of transmission in $Ca_v3.1$ KO mice (Fig. 3b, $t = 45$, normalized PF-Rsp = 1.29 ± 0.17 , mean \pm SEM, $N = 5$). The recovery of LTP depended on $GABA_A$ activation (KO- MLI_{dep} -LTP) since it was absent in presence of SR95531 (Fig. 3c, normalized EPSC_{t10} = 1.11 ± 0.01 , normalized EPSC_{t45} = 1.05 ± 0.04 , $P > 0.05$ RM ANOVA, Tukey's post hoc test, $N = 5$). More importantly, TTA-P2 bath application (Fig. 3d) also prevented KO- MLI_{dep} -LTP (normalized

PF-Rsp_{t45} = 1.04 ± 0.07, normalized PF-Rsp_{t10} = 0.99 ± 0.02, P > 0.05 RM ANOVA, Tukey's post hoc test, N = 5) indicating that the enhanced availability of an alternative PN-expressed Ca_v3 isoform could compensate for the lack of Ca_v3.1 channels.

Under all experimental condition tested, PPRs showed no difference when compared to baseline value (Fig. 3c inset, control: PPR_{t10} = 1.38 ± 0.13, PPR_{t45} = 1.34 ± 0.04, N = 5; Fig. 3d inset, SR95531: PPR_{t10} = 1.47 ± 0.09, PPR_{t45} = 1.42 ± 0.08, N = 5; Fig. 3e inset, TTA-P2: PPR_{t10} = 1.65 ± 0.1, PPR_{t45} = 1.49 ± 0.04, N = 5; mean ± SEM, PPR_{t10} vs PPR_{t45}, P > 0.05, RM ANOVA, Tukey's test).

Ca_v3 channels-mediated calcium transient is under the control of ionotropic GABA receptors.

The recovery of MLI_{dep}-LTP in Ca_v3.1 KO mice supports a functional coupling between GABA_A receptors and Ca_v3 channels important for LTP at PF to PN synapses. The influence of inhibition on Ca_v3-mediated calcium transient elicited by high frequency PFs stimulation was therefore tested in PNs from WT mice by calcium imaging (Fig. 4). For these experiments, the calcium indicator Oregon Green BAPTA 6F (400 μM) was added to the low chloride internal solution and loaded into the recorded PN via the patch pipette. To prevent calcium release from intracellular stores heparin (50 μg/ml)⁴² was also included in the patch clamp intracellular recording solution. The variation of intracellular calcium concentration was monitored in current clamped PNs (Fig. 4aa–ba) while PFs were stimulated by a single 100 Hz burst (15 pulses). Stimulus intensity was set to induce PFs-mediated responses (Fig. 4ab) similar to those recorded during the LTP induction protocol. The raise in intracellular calcium concentration caused by high frequency PFs stimulation (Fig. 4aa, left panel) was largely Ca_v3 channels-mediated as shown by fluorescence attenuation following TTA-P2 bath application (Fig. 4aa, right panel); the relative change in fluorescence ($\Delta F/F$) was strongly influenced by TTA-P2 with a 64% reduction of its peak value by the antagonist (Fig. 4ac, $\Delta F/F_{\text{Control}} = 1.37 \pm 0.5$, $\Delta F/F_{\text{TTA-P2}} = 0.47 \pm 0.09$, mean ± SEM, P = 0.041, ANOVA, Bonferroni post hoc test, N = 5). Interestingly, the Ca_v3 channels-mediated component of the calcium transient was completely lost when inhibition was blocked. In presence of SR95531 (Fig. 4ba, left panel), the induced increase in intracellular calcium was TTA-P2 insensitive (Fig. 4ba, right panel and Fig. 4bb, N = 5) and therefore mediated by high threshold voltage gated calcium channels.

Plasticity experiments were then conducted with the low chloride intracellular recording solution; when high frequency PFs stimulation was applied at low intracellular chloride concentration (Fig. 4c), MLI_{dep}-LTP was slightly bigger (Fig. 4c, EPSC_{/baseline}^{t65} = 1.63 ± 0.27, mean ± SEM, N = 6) compared to normal chloride (Fig. 1d, EPSC_{/baseline}^{t65} = 1.49 ± 0.2, N = 7) but with no statistically significant difference (normal vs low chloride, t = 65 min, P = 0.678, t-test); PPRs showed no statistically significant difference when compared to baseline value (Fig. 4c inset, PPR_{t10} = 1.66 ± 0.07, PPR_{t65} = 1.59 ± 0.11, mean ± SEM, N = 6, P > 0.05, RM ANOVA, Tukey's test).

These findings demonstrate that ionotropic GABA receptors activation is required for Ca_v3 channels-mediated calcium rise in PNs during high frequency PFs stimulation.

In agreement with the recovery of MLI_{dep}-LTP observed in Ca_v3.1 KO mice, the calcium transient recorded in PNs from KO mice was largely Ca_v3 channels-mediated only when elicited by PFs stimulation at low intracellular chloride concentration (Fig. 4d, *low chloride*); bath application of TTA-P2 strongly reduced calcium influx in low chloride (normalized $\Delta F/F_{\text{MAX}^{\text{Control}}} = 1 \pm 0.27$, normalized $\Delta F/F_{\text{MAX}^{\text{TTAP2}}} = 0.42 \pm 0.13$, mean ± SEM, N = 5, P = 0.017, paired t-test) while no statistically significant difference (normalized $\Delta F/F_{\text{MAX}^{\text{Control}}} = 1 \pm 0.23$, normalized $\Delta F/F_{\text{MAX}^{\text{TTAP2}}} = 0.79 \pm 0.08$, mean ± SEM, N = 5, P = 0.446, paired t-test) was detected in the presence of the antagonist when the regular internal solution was used (Fig. 4d, *normal chloride*).

Taken together these results support the fundamental role of Ca_v3 channels-mediated calcium influx in MLI_{dep}-LTP induction at PF to PN synapses and the requirement of GABA_A receptors activation to ensure the availability of Ca_v3 channels for opening during bursts of PFs activation.

Discussion

In the cerebellar cortex, GCs activation by MFs input leads to excitatory (monosynaptic) and inhibitory (di-synaptic) events in PNs and their interaction in the induction of LTP at PFs excitatory synapses is described in this article. We showed that high frequency PFs stimulation caused LTP at PF to PN synapses only when MLI_s-mediated GABAergic transmission was intact (Fig. 1).

MLI_{dep}-LTP is induced by a postsynaptic mechanism as showed by its recovery in Ca_v3.1 KO mice (Fig. 3b) and its impairment by the intracellular block of GPCRs (Fig. 2e) and IP3 receptors (Fig. 2f); MLI_{dep}-LTP is GABA_A receptors (Fig. 1), Ca_v3 channels (Fig. 2a,b), mGluR1 receptors (Fig. 2d,e) and intracellular calcium store (Fig. 2f) dependent.

Ca_v3 channels-dependent LTP has been previously reported to be induced at PF to PN synapses in the presence of the GABA_A receptor antagonist bicuculline¹⁵ in apparent discrepancy with our results. Nevertheless, PNs have been shown to express the ionotropic GABA receptor bicuculline-insensitive ρ subunits^{44,45} suggesting that bicuculline might not be able to completely eliminate inhibition in PNs. Indeed, we were able to record a bicuculline-insensitive IPSC component in PNs following PFs stimulation (Supplementary Information); after bath application of bicuculline (20 μM), 6% of the total IPSQs was still present (Supplementary Figure S1 panel a and panel b) and it was completely blocked by the following application of SR95531 (5 μM). The fact that LTP was successfully induced in the presence of bicuculline suggests that this residual bicuculline-insensitive component of the ionotropic GABA receptors-mediated response was sufficient to mediate Ca_v3 channels recovery from inactivation at least in WT mice. In agreement, MLI_{dep}-LTP was unaffected by bath application of bicuculline (Supplementary Figure S1 panel c and panel d).

The MLI network can be modulated by long-term plasticity and PF to MLI synapses can be potentiated³² at low frequency stimulation (2–8 Hz) while high frequency stimulation induces LTD^{30,31}. Also, high frequency PFs stimulation causes LTP at inhibitory MLI to MLI synapses³⁴ via increase of GABA release³³. High frequency PFs stimulation could therefore reduce inhibitory inputs to PNs via decreased MLI_s excitation and/or by increasing

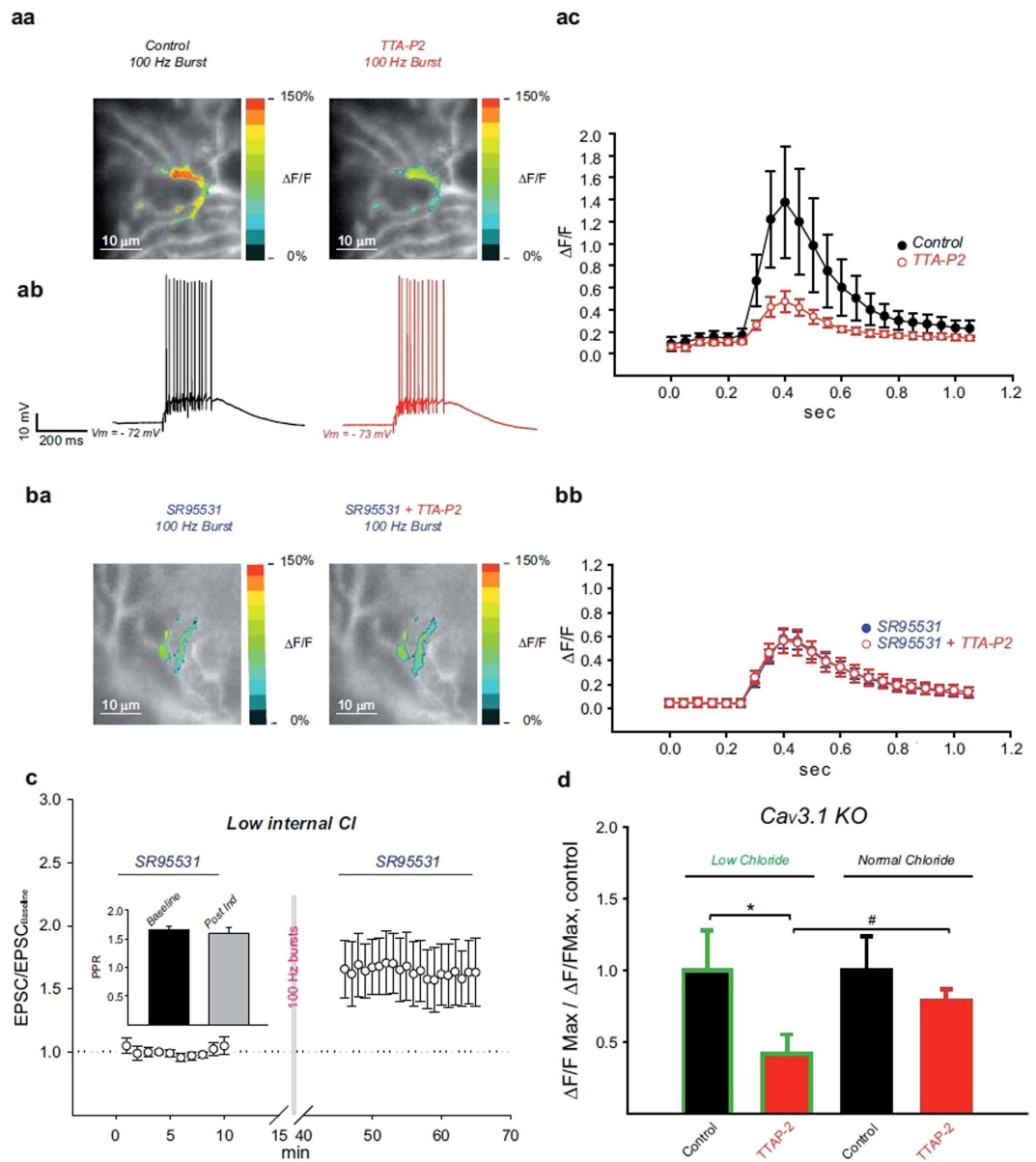


Figure 4. Ca_v3 -mediated calcium influx is controlled by inhibition. High frequency PFs stimulation caused depolarization (**ab**) and calcium increase in PNs (**aa**, left panel; **ba**, left panel); for each representative cell showed, the average $\Delta F/F$ signals was obtained at the peak of the response under the different experimental conditions (pseudocolor) and an it was superimposed on the image of the PN at resting state (grayscale). While TTA-P2 bath application strongly affected the calcium transient recorded in control condition (**aa**, right panel), the antagonist showed no effect in presence of SR95531 (**ba**, right panel). Quantified relative change in fluorescence ($\Delta F/F$) showed a large TTA-P2 sensitive component revealing that calcium influx is mostly mediated by Ca_v3 activation but only in control condition (**ac**, mean \pm SEM, $N = 5$, ANOVA $P < 0.001$). Ca_v3 -mediated calcium influx is lost when inhibition was blocked by bath application of SR95531 (**bb**, mean \pm SEM, $N = 5$, ANOVA $P < 0.001$). High frequency PFs stimulation in low internal chloride induced MLI_{dep}-LTP (**c**, mean \pm SEM, $N = 6$, RM ANOVA $P < 0.001$); PPRs (mean \pm SEM) value for the baseline ($t = 10$ min) and the post-induction phase ($t = 65$ min) are shown in the panel **c** inset. The effect of different intracellular chloride concentrations on the PFs-induced calcium transient was evaluated in PNs from $Ca_v3.1$ KO mice (**d**). The TTA-P2 sensitive component of the calcium transient observed in low chloride (*Low chloride*: control = 1 ± 0.23 , mean \pm SEM, $N = 5$; TTA-P2: 0.42 ± 0.13 , mean \pm SEM, $N = 5$, $P = 0.017$, paired t-test) was lost when the normal chloride internal solution was used (*Normal Chloride*, control: 1 ± 0.23 , mean \pm SEM, $N = 5$; TTA-P2: 0.79 ± 0.08 , mean \pm SEM, $N = 5$, $P = 0.446$, paired t-test). Before cell averaging, the maximal $\Delta F/F$ value obtained before and after TTA-P2 bath application for each cell was normalized to the mean $\Delta F/F$ value obtained in control condition. *statistically significant difference, paired t-test. #statistically significant difference, t-test.

inhibition onto MLIs. The decreased inhibition in PNs could result in indirect potentiation of PFs-mediated responses in these neurons. No significant GABA_A-mediated component in the PFs-mediated responses was observed under our experimental conditions and therefore, if present, indirect potentiation should have had little influence on our recordings. This was confirmed by investigating the specific impact of high frequency PFs stimulation on excitatory transmission by blocking GABAergic inputs before and after the induction protocol. Excitatory transmission at PF to PN showed postsynaptic potentiation to level comparable to what previously observed (Fig. 1d). GABA_A receptor activation is therefore required for MLI_{dep}-LTP at PF to PN synapses only during the induction phase. This and the intracellular pharmacological block of MLI_{dep}-LTP (Fig. 2e,f) strongly argue against indirect potentiation and they support involvement of inhibition in LTP induction at PF to PN synapses.

By investigating the molecular pathway leading to MLI_{dep}-LTP we have provided evidence for a tight cooperation among GABAergic transmission and low threshold voltage-gated calcium channels in PNs (Figs 3 and 4). We propose an active postsynaptic role for GABAergic transmission in MLI_{dep}-LTP induction with inhibition modulating T-type Ca_v3 calcium channels availability (i.e. by favoring de-inactivation). This hypothesis is supported by the recovery of MLI_{dep}-LTP in Ca_v3.1 KO mice by the intracellular modulation of the chloride electrochemical gradient in PNs (Fig. 3b). As shown by KO- MLI_{dep}-LTP dependency on TTA-P2 (Fig. 3d), an alternative Ca_v3 channels isoform is recruited when GABA_A receptor activation favored hyperpolarization toward more negative potentials (Fig. 3a). Accordingly, the TTA-P2-sensitive component of the calcium transient was restored in Ca_v3.1 KO mice when the intracellular chloride was decreased (Fig. 4d). Furthermore, Ca_v3-mediated calcium influx and MLI_{dep}-LTP was detected in PNs from WT mice only when inhibitory GABAergic transmission was intact (Figs 4a,b and 1).

MLIs-mediated postsynaptic response causes membrane potential re-hyperpolarization in depolarized PNs both in the dendritic compartment and at the soma^{46,47}. Since high frequency PFs stimulation is able to bring PNs membrane voltage to firing threshold (Fig. 4ab), the recovery of MLI_{dep}-LTP (Fig. 3b) and Ca_v3 channels-mediated calcium transient (Fig. 4d) in Ca_v3.1 KO mice suggests that activation of GABA_A receptors favor re-hyperpolarization toward a level suitable for Ca_v3 channels de-inactivation. Ca_v3 channels inactivation has been shown to be effectively removed by GABA_A-mediated IPSPs in DCN neurons^{35–37}; the IPSP reversal potential in these neurons (−75 mV)³⁷ is close the one obtained under our experimental condition in low chloride (Fig. 3a) supporting an effective Ca_v3 channels de-inactivation also in our experiments.

Ca_v3.3 channels are highly expressed in PNs and they require a stronger hyperpolarization to recovery from inactivation; based on its expression pattern and biophysical characteristics, Ca_v3.3 channels are therefore the isoform most likely to be involved in the rescue of MLI_{dep}-LTP in Ca_v3.1 KO mice although a role of Ca_v3.2 channels cannot be excluded at this time⁴⁸. The reversal potential of GABAergic currents (E_{GABA}: −85/−87 mV) measured in mature PNs^{49,50} predict a strong hyperpolarization in PNs caused by GABA_A receptors activation suggesting that Ca_v3.3 channels might be also recruited under physiological conditions and it might also participate with Ca_v3.1 in MLI_{dep}-LTP induction. Ca_v3.3 channels sole requirement is nevertheless unlikely. Cerebellar long-term plasticity is a calcium-dependent mechanism⁷ and PFs-mediated increment of calcium in PNs spines is mostly mediated by Ca_v3.1 channels even though in Ca_v3.1 KO mice a residual T-type dependent influx is still present²¹. Furthermore, Ca_v3.1 channels-mediated calcium influx is potentiated by mGluR1 activation and MLI_{dep}-LTP dependency on this metabotropic glutamate receptor (Fig. 2d,e) further supports the requirement for the Ca_v3.1 channel isoform in MLI_{dep}-LTP. Interestingly, the impairment exhibited by Ca_v3.1 KO mice in long term VOR phase-reversal training seems less severe when compared to the one of WT mice systemically injected with TTA-P2¹⁵ suggesting that Ca_v3.3 channels might also participate in cerebellar-mediated motor learning in Ca_v3.1 KO mice and slightly attenuate their phenotype.

When MLI_{dep}-LTP was compromised, decrease efficiency in PFs transmission was also revealed unmasking a pathway leading to depression at PF to PN synapses. Interestingly, this long term depression was still present when mGluR1 was inactivated (Fig. 2d). At PF to PN synapses, mGluR1 plays a central role in LTD^{51–55} but long term depression is nevertheless also reliably induced by nitric oxide (NO) uncaging when coupled to PNs depolarization⁵⁶ suggesting that this PF-released anterograde messenger⁵⁷ together with the depolarization-induced intracellular calcium raise in PNs might be sufficient to permanently decrease the synaptic transmission strength at these synapses. Indeed a NO synthase (NOS) dependent LTD has been previously described at PF to PN synapses⁵⁸; this LTD requires NMDA receptors activation⁵⁸ and it is mGluR1 independent⁵⁹. NO is likely to be released under our experimental conditions and therefore it might play a role in the depression observed whenever MLI_{dep}-LTP was impaired.

A simple model describing the first events leading to MLI_{dep}-LTP at PF to PN synapses can be proposed (Fig. 5): high frequency PFs stimulation activates AMPA receptors and causes PN dendrites depolarization counterbalanced by the GABA_A-induced hyperpolarization elicited by MLIs; membrane depolarization triggers T-type calcium channels activation and their availability for opening is controlled by inhibition. In order to induce MLI_{dep}-LTP at PF to PN synapses, Ca_v3 channels require mGluR1 activation that potentiates Ca_v3.1 channels-mediated calcium influx via a PLC independent pathway²¹. Via the G_q/PLC pathway, mGluR1 activation also leads to the release of calcium from the intracellular stores via activation of IP3 receptors; intracellular released calcium also seems to be required for MLI_{dep}-LTP.

While high frequency stimulation of PFs induced a mGluR1-dependent LTP *in vivo*¹⁸, low frequency (1 Hz) stimulation of PFs in the molecular layer of cerebellar slices induced a long lasting enhancement of transmission at PF to PN synapses which was unaffected by the pharmacological inactivation of this metabotropic receptor⁹. Together with our results, these findings support the existence of two distinct pathways leading to LTP which are differently engaged by PFs activity. Low frequency stimulation induces a mGluR1 independent potentiation which is unaffected by GABAergic ionotropic transmission impairment¹⁹ but it depends on NO release⁶ and PP2B activation¹⁰. When PFs are stimulated at high frequency, long term potentiation requires MLIs activation

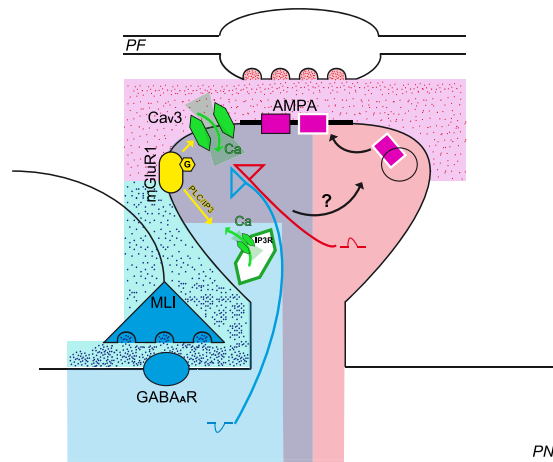


Figure 5. MLI_{dep}-LTP model. Schematic representation of the parallel fiber (PF) to Purkinje neuron (PN) synapse with a PF varicosity drawn in the upper part and the corresponding PN spine shown below. Molecular layer interneuron inhibitory synapse (MLI) is also shown in the scheme. Following high frequency parallel fibers (PF) stimulation, GABA_A-mediated hyperpolarization (blue shadow) limits AMPA-induced depolarization (pink shadow) to a range suitable for Ca_v3 activation (light violet shadow). AMPA-mediated depolarization (red arrow) activates Ca_v3 calcium channels independently on their availability regulated by inhibition (blue arrow) and Ca_v3-mediated calcium influx is enhanced by mGluR1 activation (yellow arrow). mGluR1 activation also leads to calcium release from intracellular stores via IP3 receptors. The described molecular steps initiate the cascade that leads to MLI_{dep}-LTP and downstream events are still to be determined.

and it is dependent on ionotropic GABA receptors and mGluR1 activation which cooperate to ensure a reliable activation of low threshold voltage-gated calcium channels. We therefore describe here a complementary pathway regulating PF to PN synaptic efficacy in a context of high frequency GCs inputs. Since GCs inputs recorded *in vivo* range from few to several hundred hertz²⁷, both mechanisms might coexist in order to control plasticity in different conditions or they might underlie specific pathways expressed in different groups of GC to PN synapses as suggested by the discovery that zebrin band specific physiological mechanisms could regulate cerebellar information processing^{60,61}.

Suggestions on the possible role played by MLI_{dep}-LTP in cerebellar physiology come from previous findings obtained *in vivo*. When PFs in the cat forelimb movements-related C3 zone are electrically stimulated with the same paradigm used in this paper, a bidirectional change in PFs receptive field of PNs is induced depending on the co-activation of CF input to the recorded cell. While co-activation leads to depression, an enlargement of PFs receptive field is observed when PFs are the sole excitatory pathway stimulated. Following PFs stimulation, PNs in the C3 zone are driven by cutaneous stimulation from several parts of the body in agreement with what could be expected following awakening of silent connection between PFs and PNs⁶². In attempt to mimic protocols performed *in vivo*, LTP has been established in this study without pharmacological perturbation of synaptic transmission and intracellular signal transductions. Thus, our work might provide a detailed description of the initial molecular events which lead to the PFs receptor field enlargement observed *in vivo*. Consequently, it is tempting to speculate that the described MLI-dependent LTP could be the result of the summation of newly awakened PF to PN synapses rather than the sole increased membrane expression of AMPA receptors at activated synapses.

Methods

Ethical approval. All animal procedures were performed in accordance with the University of Strasbourg animal care committee's regulations and they were approved by the Ethical Committee of the University of Strasbourg (A67-2018-38).

Electrophysiology. Patch clamp experiments were conducted on acute coronal slices from cerebellum of adult C57BL/6 male mice (P27-P46) and age-matched Ca_v3.1 knockout (KO) male mice from homozygous breeding. Mice were anesthetized by exposure to isoflurane, decapitated and the cerebellum dissected in ice cold bubbled (95% O₂/5% CO₂) aCSF containing (in mM): NaCl 120, KCl 3, NaHCO₃ 26, NaH₂PO₄ 1.25, CaCl₂ 2, MgCl₂ 1 and glucose 16. 300 μm thick slices were obtained with a vibratome (Microm 650 V; Thermo Scientific Microm, Waltham, Massachusetts) in ice cold slicing medium containing (in mM): KGlucuronate 130, KCl 14.6, EGTA 2, HEPES 20, Glucose 25, pH 7.3 and supplemented with D-APV 50 μM and minocycline 50 nM. Recovery for 1–5 seconds at 35 °C was allowed in bubbled transfer buffer containing (in mM): sucrose 230, KCl 2.5, NaHCO₃ 26, NaH₂PO₄ 1.25, CaCl₂ 0.8, MgCl₂ 8, glucose 25 and supplemented with D-APV 50 μM and minocycline 50 nM. Slices were then transferred to a holding chamber containing bubbled aCSF and kept at 35 °C for at least 40 minutes before they were moved to room temperature for the remaining experimental time.

For patch clamp recordings slices were moved to a recording chamber at 34 °C and continuously perfused with bubbled aCSF eventually supplemented with antagonists as stated in the main text. Borosilicate glass pipettes were pulled using a vertical puller (Narishige PC-10; Narishige, Tokyo, Japan) to a final resistance of 4–4.5 MΩ and filled with the following internal solution (in mM): KGlucuronate 130, KCl 10, MgCl₂ 1, HEPES 10, Na₂ATP

4, NaGTP 0.4, sucrose 16, pH 7.3. For experiments in low internal chloride concentration the following internal solution was used (in mM): KGlucuronate 136, KCl 4, MgCl₂ 1, HEPES 10, Na₂ATP 4, NaGTP 0.4, sucrose 16, pH 7.3. Once whole-cell configuration was established, a period of at least 20 minutes was waited before the start of the experiment. PNs were clamped at -60 mV and PFs-induced responses elicited by electrical stimulation delivered by a patch pipette positioned in the molecular layer distant from the recorded cell. The stimulation pipette was filled with the following solution (in mM): NaCl 120, KCl 3, HEPES 10, NaH₂PO₄ 1.25, CaCl₂ 2, MgCl₂ 1, glucose 10, pH 7.3. The PF-induced response was monitored over time by a test protocol of paired stimulation pulses (20 Hz) applied every 20 seconds. Three consecutive induced responses were averaged to obtain a mean trace of the evoked response per minute of recording. The induction protocol was applied in current-clamp mode with cells held at -68 mV.

Data were collected with a MultiClamp 700B (Molecular Devices, Sunnyvale, California), filtered at 2 kHz and digitized at 20 kHz. Data from each cell were normalized to the mean baseline value before cell averaging. Data are expressed as mean \pm SEM.

A small hyperpolarize step (-10 mV) was applied before each stimulation to follow series resistance during the experiment. Series resistance was compensated by 70–80% and cells were discarded if significant changes were detected.

For input/output curves, cerebellar coronal acute slices from adult C57BL/6 and Ca_v3.1 KO mice were prepared as previously described and the following intracellular recording solution was used in whole cell patch clamp experiments (in mM): cesium methanesulfonate 135, NaCl 6, MgCl₂ 1, HEPES 10, MgATP 4, Na₂GTP 0.4, EGTA 1.5, QX314Cl 5, pH 7.3. The glutamatergic transmission was kept intact during the recordings to allow the full recruitment of all MLIs contributing to the GABA-mediated inhibitory response; to minimize the EPSCs contribution to the induced response, the recorded PNs were clamped at -10 mV and the PF-mediated inhibitory response was elicited by molecular layer electrical stimulation; the stimulation electrode was pulled with a vertical puller (Narishige PC-10) to a final 5 M Ω resistance when filled with the following solution (in mM): NaCl 120, KCl 3, HEPES 10, NaH₂PO₄ 1.25, CaCl₂ 2, MgCl₂ 1, glucose 10, pH 7.3. Input/output curves were obtained by progressively increasing the strength of stimulation by 0.1 mA increments. At each intensity, PFs-mediated responses were recorded for three consecutive stimuli (one every 10 seconds) and traces averaged before analysis.

IPSPs were recorded in current clamp mode in presence of NBQX 5 μ M. For IPSP/V_m curves, PNs were held at -40 mV and the membrane potential progressively hyperpolarized by 22 steps with a 200 pA increment.

Calcium imaging. 300 μ m sagittal slices were prepared from adult (4–5 weeks) C57BL/6 and Ca_v3.1 KO male mice as previously described and calcium imaging experiments were performed at room temperature. The calcium indicator Oregon Green 488 BAPTA 6F (K_D = \sim 3 μ M, Molecular Probes) and heparin were added to the low chloride or regular internal solution to a final concentration of 400 μ M and 50 μ g/ml respectively and they were loaded into PNs via the patch clamp pipette. After whole cell establishment, the calcium indicator was allowed to diffuse for at least 20 minutes before starting the experiment. PNs were held at a potential close to -70 mV in current clamp configuration and PF stimulation was achieved by electrical stimulation via a glass stimulation pipette placed in the molecular layer. A single 100 Hz burst stimulation (15 pulses) was applied while calcium images were acquired every 50 ms by a sCMOS CCD camera (Xyla 5.5, Andor Technology Ltd, UK) with a 20 ms exposure time. Stimulation was repeated at long interval (at least 3 minutes between trials) to avoid plasticity in the recorded cell and collected images analyzed by using ImageJ⁶³. Relative change in fluorescence ($\Delta F/F$) was quantified in ROIs including the entire dendritic tree area in which PF-induced increase in calcium concentration was detected. Average $\Delta F/F$ value for each cell was obtained from 5 consecutive imaging sections before and after TTA-P2 500 nM bath application with or without SR95531 5 μ M.

Statistics. For plasticity experiments, PF-induced responses and paired-pulse ratios obtained at different time point were compared by one-way repeated measure (RM) ANOVA followed by Tukey's post hoc test.

References

- Ito, M. *The cerebellum and neural control*. Raven Press (1984).
- McCormick, D. A. & Thompson, R. F. Cerebellum: essential involvement in the classically conditioned eyelid response. *Science* **223**(4633), 296–299 (1984).
- Thach, W. T., Goodkin, H. P. & Keating, J. G. The cerebellum and the adaptive coordination of movement. *Annu Rev Neurosci* **15**, 403–442 (1992).
- Boyden, E. S., Katoh, A. & Raymond, J. L. Cerebellum-dependent learning: the role of multiple plasticity mechanisms. *Annu Rev Neurosci* **27**, 581–609 (2004).
- Ito, M., Sakurai, M. & Tongroach, P. Climbing fibre induced depression of both mossy fibre responsiveness and glutamate sensitivity of cerebellar Purkinje cells. *J Physiol* **324**, 113–134 (1982).
- Lev-Ram, V., Wong, S. T., Storm, D. R. & Tsien, R. Y. A new form of cerebellar long-term potentiation is postsynaptic and depends on nitric oxide but not cAMP. *Proc Natl Acad Sci USA* **99**(12), 8389–8393 (2002).
- Coesmans, M., Weber, J. T., De Zeeuw, C. I. & Hansel, C. Bidirectional parallel fiber plasticity in the cerebellum under climbing fiber control. *Neuron* **44**(4), 691–700 (2004).
- Belmeguenai, A. & Hansel, C. A role for protein phosphatases 1, 2A, and 2B in cerebellar long-term potentiation. *J Neurosci* **25**(46), 10768–10772 (2005).
- Belmeguenai, A. *et al.* Alcohol impairs long-term depression at the cerebellar parallel fiber–Purkinje cell synapse. *J Neurophysiol* **100**(6), 3167–3174 (2008).
- Schonewille, M. *et al.* Purkinje cell-specific knockout of the protein phosphatase PP2B impairs potentiation and cerebellar motor learning. *Neuron* **67**(4), 618–628 (2010).
- Marr, D. A theory of cerebellar cortex. *J Physiol* **202**(2), 437–470 (1969).
- De Zeeuw, C. I. *et al.* Expression of a protein kinase C inhibitor in Purkinje cells blocks cerebellar LTD and adaptation of the vestibulo-ocular reflex. *Neuron* **20**(3), 495–508 (1998).
- Hansel, C. *et al.* α CaMKII Is essential for cerebellar LTD and motor learning. *Neuron* **51**(6), 835–843 (2006).
- Boyden, E. S. *et al.* Selective engagement of plasticity mechanisms for motor memory storage. *Neuron* **5**(6), 823–834 (2006).

15. Ly, R. *et al.* T-type channel blockade impairs long-term potentiation at the parallel fiber–Purkinje cell synapse and cerebellar learning. *Proc Natl Acad Sci USA* **110**(50), 20302–20307 (2013).
16. Mittmann, W., Koch, U. & Häusser, M. Feed-forward inhibition shapes the spike output of cerebellar Purkinje cells. *J Physiol* **563** (Pt 2), 369–378 (2005).
17. Schreurs, B. G. & Alkon, D. L. Rabbit cerebellar slice analysis of long-term depression and its role in classical conditioning. *Brain Res* **631**(2), 235–240 (1993).
18. Wang, X., Chen, G., Gao, W. & Ebner, T. Long-term potentiation of the responses to parallel fiber stimulation in mouse cerebellar cortex *in vivo*. *Neuroscience* **162**(3), 713–722 (2009).
19. Wulff, P. *et al.* Synaptic inhibition of Purkinje cells mediates consolidation of vestibulo-cerebellar motor learning. *Nat Neurosci* **12**(8), 1042–1049 (2009).
20. Isope, P. & Murphy, T. H. Low threshold calcium currents in rat cerebellar Purkinje cell dendritic spines are mediated by T-type calcium channels. *J Physiol* **562** (Pt 1), 257–269 (2005).
21. Hildebrand, M. E. *et al.* Functional coupling between mGluR1 and Cav3.1 T-type calcium channels contributes to parallel fiber-induced fast calcium signaling within Purkinje cell dendritic spines. *J Neurosci* **29**(31), 9668–9682 (2009).
22. Perez-Reyes, E. Molecular physiology of low-voltage-activated t-type calcium channels. *Physiol Rev* **83**(1), 117–161 (2003).
23. Thomson, A. M. Inhibitory postsynaptic potentials evoked in thalamic neurons by stimulation of the reticularis nucleus evoke slow spikes in isolated rat brain slices–I. *Neuroscience* **25**(2), 491–502 (1988).
24. Suzuki, S. & Rogawski, M. A. T-type calcium channels mediate the transition between tonic and phasic firing in thalamic neurons. *Proc Natl Acad Sci USA* **86**(18), 7228–7232 (1989).
25. Chadderton, P., Margrie, T. W. & Häusser, M. Integration of quanta in cerebellar granule cells during sensory processing. *Nature* **428**(6985), 856–860 (2004).
26. Jorntell, H. & Ekerot, C. F. Properties of somatosensory synaptic integration in cerebellar granule cells *in vivo*. *J Neurosci* **26**(45), 11786–11797 (2006).
27. Arenz, A., Silver, R. A., Schaefer, A. T. & Margrie, T. W. The contribution of single synapses to sensory representation *in vivo*. *Science* **321**(5891), 977–980 (2008).
28. Rancillac, A. & Barbara, J. G. Frequency-dependent recruitment of inhibition mediated by stellate cells in the rat cerebellar cortex. *J Neurosci Res* **80**(3), 414–423 (2005).
29. Gao, Z., van Beugen, B. J. & De Zeeuw, C. I. Distributed synergistic plasticity and cerebellar learning. *Nat Rev Neurosci* **13**(9), 619–635 (2012).
30. Liu, S. Q. & Cull-Candy, S. G. Synaptic activity at calcium-permeable AMPA receptors induces a switch in receptor subtype. *Nature* **405**(6785), 454–458 (2000).
31. Soler-Llavina, G. J. & Sabatini, B. L. Synapse-specific plasticity and compartmentalized signaling in cerebellar stellate cells. *Nat Neurosci* **9**(6), 798–806 (2006).
32. Rancillac, A. & Crepel, F. Synapses between parallel fibres and stellate cells express long-term changes in synaptic efficacy in rat cerebellum. *J Physiol* **554** (Pt 3), 707–720 (2004).
33. Liu, S. J. & Lachamp, P. The activation of excitatory glutamate receptors evokes a long-lasting increase in the release of GABA from cerebellar stellate cells. *J Neurosci* **26**(36), 9332–9339 (2006).
34. Lachamp, P. M., Liu, Y. & Liu, S. J. Glutamatergic modulation of cerebellar interneuron activity is mediated by an enhancement of GABA release and requires protein kinase A/RIM1 α signaling. *J Neurosci* **29**(2), 381–392 (2009).
35. Engbers, J. D. *et al.* Distinct roles for I(T) and I(H) in controlling the frequency and timing of rebound spike responses. *J Physiol* **589** (Pt 22), 5391–5413 (2011).
36. Llinas, R. & Muhlethaler, M. Electrophysiology of guinea-pig cerebellar nuclear cells in the *in vitro* brain stem-cerebellar preparation. *J Physiol* **404**, 241–258 (1988).
37. Aizenman, C. D. & Linden, D. J. Regulation of the rebound depolarization and spontaneous firing patterns of deep nuclear neurons in slices of rat cerebellum. *J Neurophysiol* **82**(4), 1697–1709 (1999).
38. Dreyfus, F. M. *et al.* Selective T-type calcium channel block in thalamic neurons reveals channel redundancy and physiological impact of I(T) window. *J Neurosci* **30**(1), 99–109 (2010).
39. Molineux, M. L. *et al.* Specific T-type calcium channel isoforms are associated with distinct burst phenotypes in deep cerebellar nuclear neurons. *Proc Natl Acad Sci USA* **103**(14), 5555–5560 (2006).
40. Talley, E. M. *et al.* Differential distribution of three members of a gene family encoding low voltage-activated (T-type) calcium channels. *J Neurosci* **19**(6), 1895–1911 (1999).
41. Petrenko, A. B., Tsujita, M., Kohno, T., Sakimura, K. & Baba, H. Mutation of $\alpha 1G$ T-type calcium channels in mice does not change anesthetic requirements for loss of the righting reflex and minimum alveolar concentration but delays the onset of anesthetic induction. *Anesthesiology* **106**(6), 1177–1185 (2007).
42. Finch, E. A. & Augustine, G. J. Local calcium signalling by inositol-1,4,5-trisphosphate in Purkinje cell dendrites. *Nature* **396**(6713), 753–756 (1998).
43. Iftinca, M. *et al.* Temperature dependence of T-type calcium channel gating. *Neuroscience* **142**(4), 1031–1042 (2006).
44. Boue-Grabot, E. *et al.* Expression of GABA receptor ρ subunits in rat brain. *J Neurochem* **70**(3), 899–907 (1998).
45. Rozzo, A. *et al.* Expression and dendritic mRNA localization of GABAC receptor $\rho 1$ and $\rho 2$ subunits in developing rat brain and spinal cord. *Eur J Neurosci* **15**(11), 1747–1758 (2002).
46. Midtgaard, J. Stellate cell inhibition of Purkinje cells in the turtle cerebellum *in vitro*. *J Physiol* **457**, 355–367 (1992).
47. Oldfield, C. S., Marty, A. & Stell, B. M. Interneurons of the cerebellar cortex toggle Purkinje cells between up and down states. *Proc Natl Acad Sci USA* **107**(29), 13153–13158 (2010).
48. Engbers, J. D. *et al.* Intermediate conductance calcium-activated potassium channels modulate summation of parallel fiber input in cerebellar Purkinje cells. *Proc Natl Acad Sci USA* **109**(7), 2601–2606 (2012).
49. Eilers, J., Plant, T. D., Marandi, N. & Konnerth, A. GABA-mediated Ca $^{2+}$ signalling in developing rat cerebellar Purkinje neurones. *J Physiol* **536** (Pt 2), 429–437 (2001).
50. Chavas, J. & Marty, A. Coexistence of excitatory and inhibitory GABA synapses in the cerebellar interneuron network. *J Neurosci* **23**(6), 2019–2031 (2003).
51. Aiba, A. *et al.* Deficient cerebellar long-term depression and impaired motor learning in mGluR1 mutant mice. *Cell* **79**(2), 377–388 (1994).
52. Conquet, F. *et al.* Motor deficit and impairment of synaptic plasticity in mice lacking mGluR1. *Nature* **372**(6503), 237–243 (1994).
53. Hartell, N. A. Induction of cerebellar long-term depression requires activation of glutamate metabotropic receptors. *Neuroreport* **5**(8), 913–916 (1994).
54. Shigemoto, R., Abe, T., Nomura, S., Nakanishi, S. & Hirano, T. Antibodies inactivating mGluR1 metabotropic glutamate receptor block long-term depression in cultured Purkinje cells. *Neuron* **12**(6), 1245–1255 (1994).
55. Ichise, T. *et al.* mGluR1 in cerebellar Purkinje cells essential for long-term depression, synapse elimination, and motor coordination. *Science* **288**(5472), 1832–1835 (2000).
56. Lev-Ram, V., Makings, L. R., Keitz, P. F., Kao, J. P. & Tsien, R. Y. Long-term depression in cerebellar Purkinje neurons results from coincidence of nitric oxide and depolarization-induced Ca $^{2+}$ transients. *Neuron* **15**(2), 407–415 (1995).

57. Shibuki, K. & Kimura, S. Dynamic properties of nitric oxide release from parallel fibres in rat cerebellar slices. *J Physiol* **498** (Pt 2), 443–452 (1997).
58. Casado, M., Isope, P. & Ascher, P. Involvement of presynaptic N-methyl-D-aspartate receptors in cerebellar long-term depression. *Neuron* **33**(1), 123–130 (2002).
59. Bidoret, C., Ayon, A., Barbour, B. & Casado, M. Presynaptic NR2A-containing NMDA receptors implement a high-pass filter synaptic plasticity rule. *Proc Natl Acad Sci USA* **106**(33), 14126–14131 (2009).
60. Zhou, H. *et al.* Cerebellar modules operate at different frequencies. *Elife* **3**, e02536 (2014).
61. Cerminara, N. L., Lang, E. J., Sillitoe, R. V. & Apps, R., Redefining the cerebellar cortex as an assembly of non-uniform Purkinje cell microcircuits. *Nat Rev Neurosci* **16**(2), 79–93 (2015).
62. Jorntell, H. & Ekerot, C. F., Reciprocal bidirectional plasticity of parallel fiber receptive fields in cerebellar Purkinje cells and their afferent interneurons. *Neuron* **34**(5), 797–806 (2002).
63. Rasband, W. S. *ImageJ*, U. S. National Institutes of Health, Bethesda, Maryland, USA (1997–2014).

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Author Contributions

F.B. designed research, performed research, analyzed data wrote the paper. K.D. performed research. S.R. provided technical assistance. K.S. and M.K. provided $Ca_v3.1$ KO mice. B.P. wrote the paper. P.I. designed research and wrote the paper.

Additional Information

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