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Antipsychotic-like effects of M₄ positive allosteric modulators are mediated by CB₂ cannabinoid receptor-dependent inhibition of dopamine release

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Summary

Muscarinic receptors represent a promising therapeutic target for schizophrenia, but the mechanisms underlying the antipsychotic efficacy of muscarinic modulators are not well understood. Here we report that activation of M_4 receptors on striatal spiny projection neurons results in a novel form of dopaminergic regulation resulting in a sustained depression of striatal dopamine release that is observed more than 30 minutes after removal of the muscarinic receptor agonist. Furthermore, both the M_4 -mediated sustained inhibition of dopamine release and the antipsychotic-like efficacy of M_4 activators were found to require intact signaling through CB₂ cannabinoid receptors. These findings highlight a novel mechanism by which striatal cholinergic

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and cannabinoid signaling leads to sustained reductions in dopaminergic transmission and concurrent behavioral effects predictive of antipsychotic efficacy.

Introduction

Currently available treatments for schizophrenia are often effective in ameliorating psychotic symptoms but also induce metabolic, cognitive, and motor side-effects, limiting their therapeutic utility. The seminal finding that the M_1/M_4 -preferring muscarinic acetylcholine receptor (mAChR) agonist xanomeline demonstrated robust antipsychotic efficacy in schizophrenia patients (Shekhar et al., 2008) generated a major interest in developing selective M1 and M4 agonists and understanding the roles of these receptors in forebrain circuits (Foster et al., 2014a). Unfortunately, all known mAChR agonists are non-selective and activate peripheral M_2 and M_3 receptors, leading to adverse effects (Bymaster et al., 2003). By targeting allosteric sights on mAChRs, we and others have succeeded in identifying highly subtype-selective positive allosteric modulators (PAMs) of individual mAChR subtypes that avoid activation of peripheral mAChRs. Interestingly, highly selective M_4 positive allosteric modulators (PAMs) have robust antipsychotic-like effects in multiple rodent models. While the mechanisms by which M₄ PAMs exert their behavioral effects are not entirely clear, these compounds reverse multiple *in vivo* effects of psychomotor stimulants that induce increases in extracellular dopamine (DA; Bubser et al., 2014; Byun et al., 2014; Chan et al., 2008; Leach et al., 2010). These studies raise the possibility that M_4 PAMs may act by inhibiting DA release from midbrain DA neurons. However, the hypothesis that selective M_4 PAMs inhibit DA release has not been directly tested. Furthermore, while M_4 can act as a presynaptic heteroreceptor to inhibit glutamate release in the striatum (Pancani et al., 2014), this receptor is not expressed in DA neurons and is not likely to inhibit DA release by direct actions on DA terminals. We now report that activation of M₄ induces a sustained inhibition of electrically- or optically-induced DA release in midbrain slices that is potentiated by M_4 PAMs and persists for more than 30 minutes after receptor activation. This sustained inhibition of DA release is independent of nicotinic receptor (nAChR) signaling, suggesting that it is not mediated by autoreceptors on cholinergic interneurons. Instead, use of genetically-modified mice revealed that this sustained inhibition of striatal DA release is mediated by M_4 expressed on a subpopulation of striatal spiny projection neurons (SPNs) that express D₁ DA receptors (D₁-SPNs). Surprisingly, both the sustained inhibition of DA release and antipsychotic-like behavioral effects induced by an M_4 PAM were found to require intact endocannabinoid (eCB)mediated CB₂ cannabinoid receptor signaling. Taken together, these studies identify a novel signaling pathway through which activation of M₄ receptors on SPNs dampens dopaminergic signaling and highlights the importance of this pathway to the antipsychoticlike efficacy observed with M₄ PAMs.

Results

mAChR activation induces a sustained inhibition of DA release that is independent of nAChR signaling

We determined the ability of mAChR activation to modulate striatal DA release by bath applying the non-selective mAChR agonist oxotremorine-M (Oxo-M) while monitoring electrically-evoked DA release via fast scan voltammetry (Figure S1A). Consistent with previous studies (Threlfell et al., 2010; Zhang et al., 2002), addition of 10 μ M Oxo-M caused a transient inhibition of DA release that returned to baseline within 10 minutes of removing Oxo-M from the bath (Figure 1A). However, at higher concentrations (30 μ M), Oxo-M induced a sustained inhibition of DA release that persisted long after (>30 minutes) drug washout (Figure 1B).

Previous studies have demonstrated that electrical stimulation of cholinergic afferents in the striatum can increase tonic DA release by activation of β2-containing nicotinic ACh receptors (nAChRs) on DA neuron terminals (Champtiaux et al., 2003). Furthermore, mAChR inhibition of DA release induced by electrical stimulation in striatal slices has been ascribed in large part to activation of mAChR autoreceptors on cholinergic terminals that inhibit acetylcholine (ACh) release and thereby inhibit nAChR-mediated increases in DA release (Threlfell et al., 2010). To assess the effects of mAChR activation on nAChRindependent DA release we pretreated the slices with the β 2-selective nAChR antagonist dihydro- β -erythroidine hydrobromide (DH β E, 1 μ M). Pretreatment with DH β E completely blocked DA release induced by optical stimulation of cholinergic interneurons (Figure S1B) while partially blocking electrically-induced DA transients (Figure S1C). This suggests that this nAChR antagonist can completely inhibit nAChR-mediated increases in DA release as previously observed (Threlfell et al., 2012). Pretreatment with DHBE significantly reduced the Oxo-M induced inhibition of electrically-evoked DA release at early time points (Figure 1C,E). However, the Oxo-M-induced sustained inhibition was not significantly altered by DHBE pretreatment (Figure 1C,F), suggesting that this sustained inhibition of DA release is not mediated by inhibition of nAChR effects. To remove effects of electrical stimulation on cholinergic terminals and selectively activate DA terminals, we utilized mice that selectively express ChR2 in dopamine transporter (DAT)-expressing neurons (DAT-Cre \times ChR2 mice). Optical stimulation in slices from DAT-Cre × ChR2 mice induced robust DA transients (Figure S1D). In these mice, application of 30µM Oxo-M had a blunted effect at early time points (Figure 1D,E), whereas the sustained inhibition at later time points was unchanged (Figure 1D,F). Importantly, inclusion of DH β E had no effect on optically-evoked DA transients or Oxo-M mediated regulation of optically-evoked DA release from DAT-Cre \times ChR2 mice (Figure S1E). Collectively, these findings suggest that while the acute inhibition is largely dependent on nAChR activation, the sustained inhibition of DA release is independent of nAChR signaling.

The sustained inhibition of DA release induced by Oxo-M is mediated by M_4 receptors expressed on D_1 -SPNs

The M_4 receptor is expressed in numerous neuronal populations in the striatum (Figure 2A). In order to determine the role of M_4 in mediating the sustained inhibition of DA release

observed with Oxo-M we utilized tissue specific knock-out mice in combination with the novel M_4 -selective PAM VU0467154. Consistent with its activity as a PAM, application of VU0467154 (3μ M) had no effect on DA release when applied alone (Figure S1F); however, VU0467154 significantly potentiated the inhibition of DA release induced by 10µM Oxo-M at sustained time points (Figure 2B,D) suggesting a key role for M₄ in mediating the sustained inhibition of DA release. However, M_4 is not expressed in DA neurons (Weiner et al., 1990), suggesting that M₄ is not likely to serve as a presynaptic heteroreceptor that directly inhibits DA release in DA terminals. Since this sustained effect of Oxo-M on DA release is independent of nAChR activation, it is also not likely to be mediated by activation of M₄ autoreceptors on cholinergic terminals. The large majority of M₄ in striatum is present on D_1 -SPNs (Ince et al., 1997), raising the possibility that this subpopulation of M_4 could in some way modulate DA release. To assess this, we used mice in which M_4 was selectively deleted from D₁-expressing neurons (D₁-M₄ knockout (KO) mice). Electrically-evoked DA release was similar in magnitude in both D₁-M₄ KOs and control littermates (floxed M₄ receptor mice; M₄ fl/fl; data not shown). However, the acute inhibition observed upon application of 10µM Oxo-M was significantly reduced in the D_1 -M₄ KO mice (28.2 ± 9.1%) acute inhibition of DA release, n=6) compared to littermate controls (72.3 \pm 7.1% % acute inhibition of DA release, n=6; p<0.05, two-tailed Mann-Whitney test), suggesting that M_4 expressed on D_1 neurons may, in part, contribute to the acute inhibition seen with Oxo-M. In the D₁-M₄ KO mice, the acute inhibition was further enhanced by increasing the Oxo-M concentration to $30\mu M$ (Figure 2E), suggesting that M_4 expressed on D₁-SPNs is not the only subpopulation of mAChRs contributing to this acute response. Importantly, VU0467154 had no effect on Oxo-M modulation of DA release in D₁-M₄ KO slices at any of the Oxo-M concentrations examined (Figure 2C-E). Furthermore, the sustained inhibition of DA release observed in littermate control mice after application of Oxo-M (30μ M) in the presence of DH β E was completely absent in D₁-M₄ KO mice (Figure 2F,G). Collectively, these findings suggest that M_4 on D_1 -SPNs contributes to the acute inhibition observed with application of mAChR agonist but is solely responsible for the sustained inhibition of DA signaling observed after mAChR activation.

M_4 -mediated sustained inhibition of DA release requires intact diacylglycerol (DAG) lipase and CB_2 cannabinoid receptor signaling

SPNs do not project to DA terminals, suggesting that in these acute slices M_4 must alter DA release via a form of non-synaptic transmission. One of the best studied forms of non-synaptic communication in the striatum involves eCB signaling and activation of CB₁ receptors and this pathway has been demonstrated to play a key role in M_4 -mediated modulation of striatal synaptic plasticity (Shen et al., 2015). However, the selective CB₁ receptor antagonist AM251 (1µM) had no effect on M_4 modulation of DA release, suggesting that CB₁ is not involved (data not shown). Interestingly, while the CB₂ cannabinoid receptor is not thought to play a major role in regulation of neurotransmitter release, a recent study suggests that DA neurons express the CB₂ receptor (Zhang et al., 2014). Thus, we used a combination of genetic and pharmacological tools to determine whether CB₂ receptors play a role in M_4 effects on DA release. Similar to results in M_4 fl/fl mice, VU0467154 significantly potentiated the sustained inhibition of DA release observed after bath application of 10µM Oxo-M in wild type (WT) mice (Figure 3A,D). Strikingly,

the CB₂ antagonist AM630 (3μ M) had no effect on DA release alone (data not shown), but blocked VU0467154 potentiation of Oxo-M effects (Figure 3B,D). Furthermore, the effects of VU0467154 were absent in CB₂ KO mice (Figure 3C,D) confirming a critical role for the CB₂ receptor in mediating this response.

The CB₂-dependence of the sustained reductions in DA release suggests that activation of M₄ on SPNs could facilitate the formation and/or release of an eCB. Production and concurrent physiological effects of eCBs can be stimulated via a multitude of mechanisms at various synapses with some of these mechanisms being Ca²⁺-dependent and others being Ca²⁺-independent (Gladding et al., 2009). While M₄ canonically signals through the Gprotein G_{ai}, there are many examples of G-protein-coupled receptors (GPCRs) that, in addition to coupling with G_{ai}, can activate multiple G-proteins and signaling pathways including intracellular Ca²⁺ mobilization. To evaluate if M₄ in SPNs could promote Ca²⁺ release, single-cell Ca²⁺ mobilization experiments were performed in M₁ KO mice that expressed tdTomato in D1-expressing neurons. These mice allowed us to visualize and patch D₁-SPNs as well as isolate M₄ as the only known mAChR subtype in these cells. Either current injection or activation of Group I metabotropic glutamate (mGlu) receptors produced robust Ca²⁺ transients (Figure S2). In contrast, mAChR activation did not induce Ca²⁺ transients in D₁-SPNs (Figure S2A). To further evaluate a potential role for intracellular Ca²⁺ signaling in the mAChR-induced inhibition of dopamine release, slices were pretreated with either 3uM thapsigargin to deplete intracellular Ca²⁺ stores or 30uM 2-Aminoethoxydiphenyl borate (2-APB) to antagonize the inositol 1,4,5-trisphosphate (IP₃) receptor. Neither of these treatments had any effect on M_4 -mediated sustained inhibitions of DA release (Figure 3F). However, bath application of thapsigargin blocked mGlu-mediated Ca^{2+} transients in D₁-SPNs suggesting that this treatment was effective at depleting intracellular Ca²⁺ stores (Figure S2C,D). These data suggest that the observed M₄-mediated effects on sustained DA release are not likely to be dependent on M₄-induced Ca²⁺ mobilization. Previous examples of Ca²⁺-independent eCB-mediated effects arising from SPNs as well as other brain regions have been demonstrated to occur through DAG lipase which plays a key role in the synthesis of the endocannabinoid 2-arachinonoylglycerol (2-AG; Chevaleyre and Castillo, 2003; Lerner and Kreitzer, 2012). Interestingly, we found that mAChR-induced sustained reductions in DA release were blocked by pretreatment with the DAG lipase inhibitor DO-34 (Figure 3E,F). Collectively, these studies suggest that M₄ expressed on SPNs can lead to CB2 activation through activation of DAG-lipase and formation of the eCB 2-AG.

Efficacy of VU0467154 in rodent models of antipsychotic efficacy are mediated by M_4 in D_1 expressing neurons and are blocked by a CB₂ antagonist

We next evaluated the effects of VU0467154 on amphetamine-induced disruption of prepulse inhibition (PPI; Figure 4A) of the acoustic startle response, a preclinical model of sensory motor gating deficits in schizophrenia patients that is reversed by M₄ PAMs (Byun et al., 2014). Both D₁-M₄ KO mice and control littermates (M₄ fl/fl) had similar baseline acoustic startle responses that were not altered by dosing with amphetamine or VU0467154 alone or in combination (Figure S3). VU0467154 (10mg/kg; intraperitoneal (i.p.)) alone had no effect on PPI compared to vehicle (74.2±3.7% and 72.1±3.8% PPI respectively).

Amphetamine (4 mg/kg; subcutaneous (s.c.)) induced a robust decrease in PPI in both control and D_1 -M₄ KO mice (Figure 4B). In control littermates, this amphetamine disruption was significantly reversed by pretreatment with VU0467154 (Figure 4B left). However, VU0467154 had no effect on amphetamine-disrupted PPI in D_1 -M₄ KO mice (Figure 4B right), highlighting the importance of this M₄ subpopulation in mediating VU0467154 effects on antipsychotic-like behaviors.

In addition, the CB₂ receptor antagonist AM630 (10mg/kg; i.p.) inhibited the ability of VU0467154 (10mg/kg; i.p.) to reverse amphetamine-induced disruption of PPI (Figure 4C) but had no effect in the absence of VU0467154 (27.6 \pm 7% and 26.3 \pm 4.9% PPI in the absence and presence of AM630). We also examined MK801-induced hyperlocomotion, another behavioral paradigm that is used to predict antipsychotic-like efficacy of novel compounds. Consistent with our recent report (Bubser et al., 2014), VU0467154 significantly reversed MK-801-induced ambulation. This effect of VU0467154 was partially blocked by AM630 (Figure S4). Taken together, these findings support the novel concept that activation of M₄ in D₁-SPNs induces eCB release, resulting in a CB₂-mediated sustained inhibition of DA release and that these actions contribute to the antipsychotic-like effects of the M₄ PAM (Figure 4D).

Discussion

Clinical findings suggest that modulation of mAChR signaling can be efficacious in the cognitive, negative, and psychotic symptoms in schizophrenic and Alzheimer's disease patients (Bodick et al., 1997; Shekhar et al., 2008). Preclinical studies suggest that selective activation of the M_4 receptor may provide a strategy to obtain therapeutic effects while minimizing the side-effects observed with broad spectrum cholinergic/muscarinic compounds (Foster et al., 2014a). However, the mechanism by which M_4 activation exerts antipsychotic efficacy has not been established. Here we make two key findings that further elucidate the mechanism whereby M_4 activation leads to antipsychotic-like effects in preclinical models. First, we report that activation of M_4 in the striatum causes a sustained reduction in striatal DA release. This M₄-mediated reduction in DA release is novel in that it is sustained at time points well after washout of mAChR agonists which is reminiscent of long-term depression (LTD) observed at glutamatergic synapses (Gladding et al., 2009). Furthermore, the inhibition of DA release is mediated by M₄ receptors expressed on D₁-SPNs, which are striatal projection neurons that are postsynaptic targets of dopaminergic projections to the striatum. Interestingly, M_{4} -mediated sustained reductions in DA release are blocked by inhibiting DAG lipase, suggesting that M4 on SPNs modulates DA terminals via the eCB signaling molecule 2-AG. Secondly, we demonstrate that both the VU0467154mediated effects on DA release and the antipsychotic-like behavioral effects require CB₂ receptor signaling. Collectively, these studies highlight a novel interaction between muscarinic and eCB signaling pathways in the striatum leading to reduced dopaminergic signaling and highlight the importance of these pathways in mediating the antipsychotic-like behavioral effects observed with M₄ PAMs.

Previous studies have suggested that mAChR agonist-induced inhibition of striatal DA release is mediated primarily by autoreceptors on cholinergic interneurons (Threlfell et al.,

2010; Zhang et al., 2002), which may include both M_2 and M_4 mAChR subtypes. Here, we find that Oxo-M induced an acute inhibition of DA release that was significantly reduced either in the presence of nAChR antagonist or with direct optical stimulation of DA terminals. While the acute inhibition was largely nAChR-dependent, and accordingly likely to be mediated by autoreceptors, it was not completely abolished under conditions that removed nAChR signaling. Furthermore, the acute inhibition seen with Oxo-M was reduced in D_1 - M_4 KO mice compared to littermate controls. These findings suggest that the acute inhibition observed after mAChR agonist application involves multiple receptor subtypes and populations likely including: M2, M4 (located both pre-synaptically and postsynaptically), and M₅ (Foster et al., 2014b, Shin et al., 2015). Importantly, we also report a novel sustained inhibition of DA release that was observed at time points well after removal of Oxo-M. Unlike the acute inhibition, this sustained inhibition of DA release is independent of nAChR-signaling. Application of the M₄-selective PAM VU0467154 in acute brain slices had no effect on its own, likely owing to the reduced activity of striatal cholinergic interneurons, and corresponding low levels of ACh tone, in ex vivo brain slices compared to in vivo (Shen et al., 2015). However, when VU0467154 was included with a submaximal concentration of agonist, it potentiated the sustained inhibition of DA release. In the striatum the majority of M₄ is expressed on D₁-expressing SPNs and it was found that the VU0467154-mediated effects on DA release and reversal of specific behavioral effects of psychomotor stimulants were dependent on expression of M_4 expressed in D₁-SPNs. These data are consistent with a previous report suggesting that some behavioral effects of the nonselective orthosteric mAChR agonist xanomeline also require activation of M4 receptors on D₁-expressing neurons (Dencker et al., 2011). While these data suggest that M₄ on D₁-SPNs are critically important for mediating antipyschotic-like efficacy, we cannot rule out a role for M_4 expressed on other D₁-expressing populations. Collectively, these data provide exciting new insights into the mechanisms by which activation of M₄ receptors in these neurons can regulate DA-dependent behaviors.

Because SPNs do not make synaptic connections with DA terminals, we hypothesized that M₄ receptors on SPNs must alter DA release via a form of non-synaptic transmission. A recent study demonstrated that M4 activation with M4 PAMs could modulate multiple forms of spike-timing dependent plasticity at corticostriatal synapses via an eCB-dependent mechanism. These effects on plasticity were absent when M₄ was deleted from D₁expressing neurons and required intact CB₁ receptor signaling (Shen et al., 2015). Most previously described eCB actions in the CNS are primarily mediated by CB1. However, we found that a selective antagonist of CB1 receptors did not inhibit the effect of M4 activation on DA release. In contrast, selective blockade or genetic deletion of CB2 receptors completely blocked the ability of M_4 PAMs to inhibit DA release. Furthermore, the M_4 mediated sustained reductions in DA release were completely blocked by a DAG-lipase inhibitor suggesting a key role for 2-AG in mediating these effects. Collectively, these findings suggest that the subpopulation of M₄ on direct pathway SPNs can efficiently promote eCB signaling in the striatum with important physiological implications. In addition to changes in synaptic plasticity induced by CB_1 activation, reduced DA release via CB_2 activation could impact forms of synaptic plasticity that are modulated by D1 and D2 receptor activation. In the future it will be important to fully parse out the effects of M₄

activation on overall striatal function that are mediated by M_4 on SPNs (regulation of DA release through CB_2 activation, activation of CB_1 receptors on glutamatergic and GABAergic terminals) and activation of M_4 in other neuronal populations.

The finding that CB2 was required for M4-mediated modulation of DA release was surprising in the light of previous studies indicating that CB₁ and CB₂ were thought to be primarily expressed in the CNS and immune system, respectively. However, this original dichotomy has been challenged by recent studies that have found CB₂ expressed in numerous brain areas (Mechoulam and Parker, 2013), including data suggesting that DA neurons express CB₂ receptors (Zhang et al., 2014). Furthermore, CB₂ activation has been demonstrated to modulate cocaine-induced locomotion (Xi et al., 2011) and CB₂ function has been linked to schizophrenia in patients and to schizophrenia-related behaviors in rodent models (Ishiguro et al., 2010; Ortega-Alvaro et al., 2011). Furthermore, we demonstrate that CB₂ blockade occluded M₄-mediated behavioral effects in two preclinical behavioral models predictive of antipsychotic efficacy. While activation of M₄ leading to CB₂ activation on DA terminals is the most parsimonious explanation of our results, it is not clear what population(s) of CB₂ mediate these effects. Further studies with tissue-specific CB₂ KO mice will be needed to determine the role of CB2 receptors expressed on various neuronal and glial populations in modulating DA release and DA-dependent behaviors. In addition to providing a novel mechanism whereby M_4 activation can lead to antipsychotic-like efficacy, these studies suggest that CB2-selective agonists could provide an exciting new therapeutic strategy for the treatment of schizophrenia.

The present study is especially important in the context of currently available antipsychotic agents that act by blocking striatal DA receptors. While exerting antipsychotic activity through blockade of midbrain DA receptors, DA receptor antagonists also block DA receptors in the cortex and other brain regions where they induce adverse effects, including impaired cognitive function. In contrast, M_4 PAMs improve cognitive function in animal models (Bubser et al., 2014). These studies provide a mechanism by which M_4 may act locally to reduce DA signaling without blockade of DA receptors throughout the CNS. The combined cellular and behavioral studies reported suggest that this mechanism may be responsible, at least in part, for the antipsychotic-like effects of M_4 PAMs.

Experimental Procedures

Voltammetry data plotted with box and whisker plots was analyzed using one-way analysis of variance (ANOVA) with a Dunnett's or Bonferroni post hoc test where appropriate. Time courses of DA release are depicted as the mean \pm SEM. PPI data are expressed as mean \pm SEM and analyzed using one-way ANOVA with a Dunnett's post hoc test comparing all dosing groups to amphetamine-treated controls. Experimental procedures are described in detail in the Supplemental Experimental Procedures. All animals were handled in accordance with federal guidelines and protocols approved by Vanderbilt University.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Foster et al.

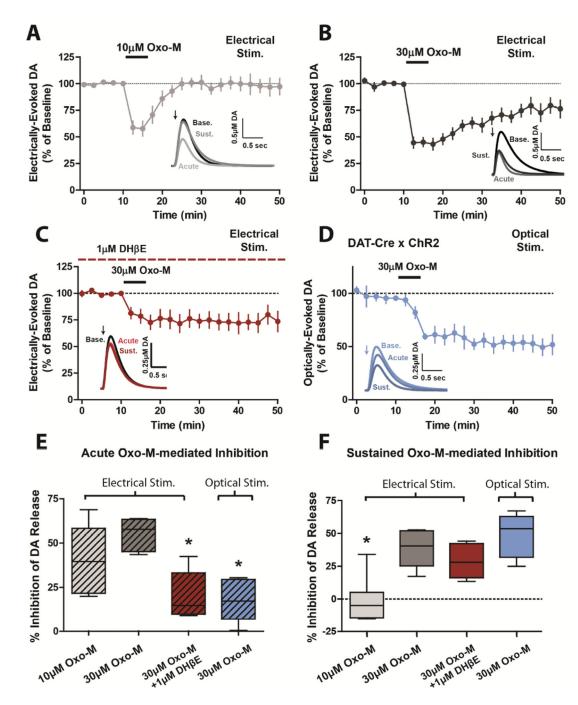


Figure 1.

mAChR activation induces a sustained reduction in DA release that is independent of nAChR signaling. Time-courses of Oxo-M-induced inhibition of DA release in the presence or absence of the nAChR antagonist DH β E using electrically-evoked (**A**–**C**) or optically-evoked (**D**) DA release paradigms. (**E**,**F**) Box plot summaries depicting the % inhibition of DA release observed under different conditions at acute (15 min) or sustained (30 min) time points (*n*=5–7; * Significant difference from 30µM Oxo-M, p<0.05; one-way ANOVA with a *post hoc* Dunnett's test).

Foster et al.

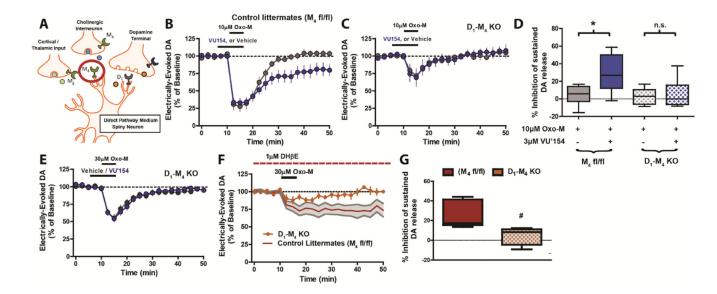


Figure 2.

 M_4 expressed on spiny projection neurons mediate the muscarinic-induced sustained suppression of DA release. (A) Cartoon depicting the anatomical location of M_4 in the striatum. (**B**,**C**) Time-courses and (**D**) box plot summaries of DA release following bath application of 10µM Oxo-M +/– 3µM VU0467154 (VU'154) in D₁-M₄ KO mice and control littermates (M₄ fl/fl; *n*=5–6; * Significant difference from 10µM Oxo-M, p<0.05; one-way ANOVA with a *post hoc* Bonferroni test). (**E**) Time-course showing the effect of 30µM Oxo-M +/– 3µM VU0467154 in D₁-M₄ KO mice. Time-course (**F**) and box plot summaries (**G**) depicting the effects of 30µM Oxo-M following pretreatment with DHβE (*n*=5, # Significant difference from control littermate; p<0.05; two-tailed Mann-Whitney test).

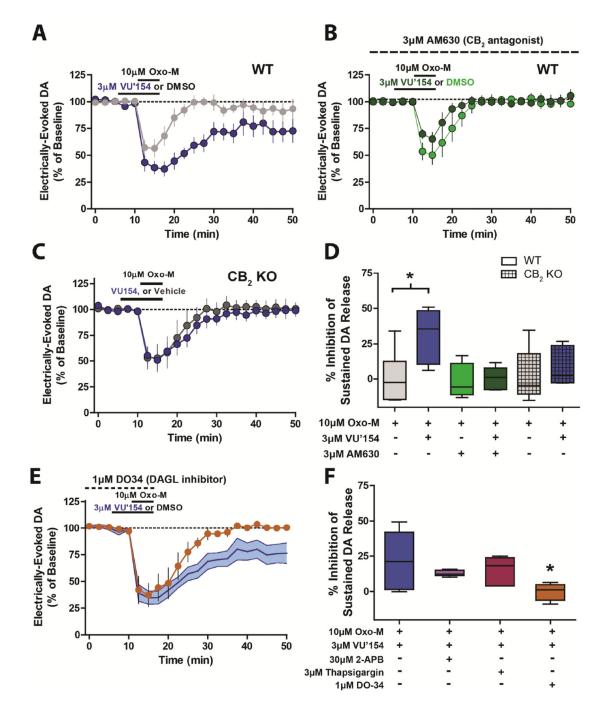


Figure 3.

VU0467154-mediated effects on DA release are CB₂-dependent. (**A,B,C**) Time-courses and (**D**) box plot summaries showing that the effects of VU0467154 (VU'154) on DA release are blocked by pretreatment with the CB₂ antagonist AM630 (3μ M) and absent in CB₂ KO mice (*n*=5–6; * Significant difference from 10 μ M Oxo-M, p<0.05; one-way ANOVA with a *post hoc* Bonferroni test). (**E**) Time-course and (**F**) box plot summaries showing that VU'154-mediated effects on sustained dopamine release are blocked by pretreating with DAG-Lipase inhibitor DO34 (1 μ M) but unaffected by treatment with 2-APB or thapsigargin (*n*=5–6; *

Significant difference from VU'154 + Oxo-M, p<0.05; one-way ANOVA with a *post hoc* Dunnett's test).

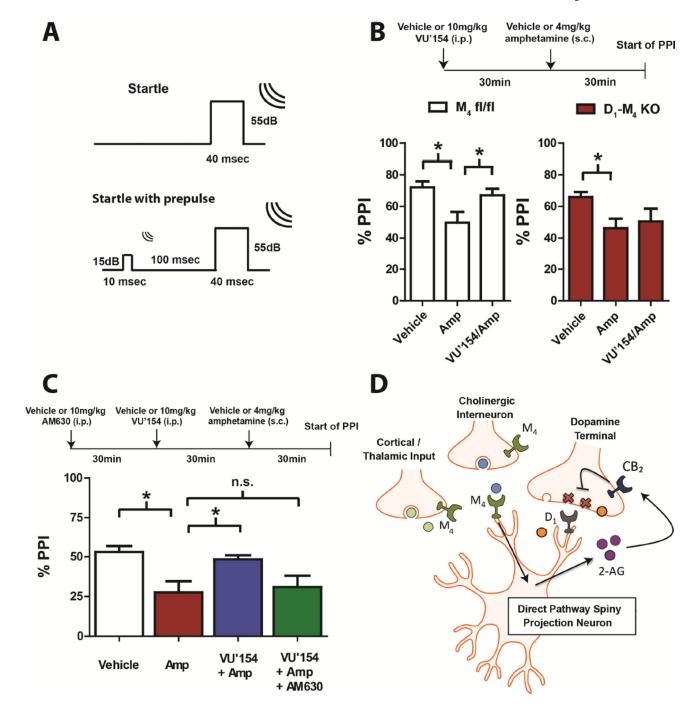


Figure 4.

VU0467154 reverses disrupted prepulse inhibition (PPI) via a mechanism requiring activation of M_4 on D_1 -expressing neurons and subsequent CB_2 receptor activation. (A) Experimental paradigm for monitoring PPI (B) Averaged data depicting % PPI observed in the absence or presence of VU0467154 (VU'154; 10mg/kg), amphetamine (Amp; 4mg/kg) or vehicle in D_1 - M_4 KO mice and control littermates (M_4 fl/fl; *n*=11–21; * Significant difference from Amp, p<0.05). (C) Averaged data depicting the % PPI observed in WT mice dosed with VU'154, AMP, and/or 10mg/kg AM630 (*n*=11–21; * Significant difference from

Amp, p<0.05; one-way ANOVA with a *post hoc* Dunnett's test). (**D**) Cartoon depicting mechanistic model for how striatal M_4 receptors mediate sustained reductions in DA release and antipsychotic-like efficacy.