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Neuropharmacology of N,N-Dimethyltryptamine

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Abstract

N,N-Dimethyltryptamine (DMT) is an indole alkaloid widely found in plants and animals. It is best known for producing brief and intense psychedelic effects when ingested. Increasing evidence suggests that endogenous DMT plays important roles for a number of processes in the periphery and central nervous system, and may act as a neurotransmitter. This paper reviews the current literature of both the recreational use of DMT and its potential roles as an endogenous neurotransmitter. Pharmacokinetics, mechanisms of action in the periphery and central nervous system, clinical uses and adverse effects are also reviewed. DMT appears to have limited neurotoxicity and other adverse effects except for intense cardiovascular effects when administered intravenously in large doses. Because of its role in nervous system signaling, DMT may be a useful experimental tool in exploring how brain works, and may also be a useful clinical tool for treatment of anxiety and psychosis.

Keywords

N,N-Dimethyltryptamine; hallucinogen; neurotransmitter; psychiatric disorder

1. Introduction

N,N-dimethyltryptamine (DMT) is an indole alkaloid widely found in nature. It is an endogenous compound in animals (Saavedra and Axelrod, 1972; Christian et al., 1977, Hollister, 1977) and in a wide variety of plants found around the globe. Major plant genera containing DMT include Phalaris, Delosperma, Acacia, Desmodium, Mimosa, Virola, and Psychotria, but DMT has been found even in apparently innocuous sources, such as leaves of citrus plants (Servillo et al., 2012), and in the leaves, seeds, and inner bark of mimosa tenuiflora, which has become a source of livestock poisoning (Gaujac et al., 2012).

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DMT has become of interest because when ingested, it causes brief, episodic visual hallucinations at high concentrations (Stoff et al. 1977; Strassman and Qualls, 1994; Strassman et al., 1994, Shulgin and Shulgin, 1997). DMT is one of the major psychoactive compounds found in various shamanistic compounds (e.g., ayahuasca, hoasca, yagé) used in South America for centuries (Pochettino et al., 1999) and has, more recently found its way into Europe and North America as a recreational drug (Tupper, 2008).

1.1. Recreational use of DMT

Most hallucinogens such as lysergic acid diethylamide (LSD) and 2,5-dimethoxy-4methylamphetamine (DOM) cause sensory distortion, depersonalization at high doses, and at least one (N,N-Diisopropyltryptamine, DiPT) causes auditory distortions, whereas some compounds such as DMT (found in ayahuasca), psilocybin (mushrooms) or mescaline (peyote) cause episodic visual effects. In the late 1990s, Rick Strassman conducted the first human research with hallucinogens in 20 years, examining the physiological effects and selfreports from people receiving DMT in carefully controlled settings (Strassman et al., 1994; 1996). A book describing these results was published in the popular press (Strassman 2001). Strassman concluded that DMT is a powerful tool for self-discovery and understanding consciousness, which may have helped to drive interest in recreational use of DMT and related tryptamine hallucinogens. In recent years, recreational use of DMT has been increasing; for example, Cakic et al., (2010) reported that 31% of recreational DMT users endorse psychotherapeutic benefits as the main reason for consumption. Similar to ayahuasca, recreational users have made similar concoctions referred to as pharmahuasca. These are of capsules containing free-base DMT and some monoamine oxidase inhibitors (MAOI) such as synthetic harmaline (Ott, 1999) or Syrian Rue (rich in beta-carbolines; Brierley and Davidson, 2012).

It is unclear what proportion of users of hallucinogenic tryptamines have adverse events serious enough for hospitalization, but it seems that the synthetic hallucinogenic compounds, such as 25I-NBOMe may be more dangerous than the plant-derived compounds (Hill et al., 2013; Lowe et al., 2015). Databases derived from Poison Control and Emergency Department visits (via the Drug Abuse Warning Network) only sparing differentiate between hallucinogenic compounds taken and lack adequate records of DMT-specific cases. Street drugs mostly contain powdered DMT, whereas ayahuasca also contains harmine-related compounds, which limit toxic effects (Lanaro et al., 2015). However, aside from the acute cardiovascular effects there have been no consistent reports of toxic effects of long-term use of DMT in the literature. In fact, there has been a report that DMT is neuroprotective (Frecska, 2008). Without more data on the recreational use of this class of compounds, it is not possible to conclude whether the synthetic hallucinogens are indeed more toxic or whether the social context may contribute to the effects.

It is likely that most adverse effects of hallucinogens are psychological effects, such as intense fear, paranoia, anxiety, grief, and depression, that can result in putting the user or others in physical harm or danger (Carbonaro et al., submitted). Anecdotal reports describe psychologically challenging experiences with DMT and other psychedelic compounds. The rates of occurrence for these effects have not been properly accounted for. However, in the

case of psilocybin, about 30% of laboratory experiences include psychologically challenging experiences (Carbonaro et al, submitted). Even though DMT may not produce physical toxicity, severe psychological adverse effects can occur.

1.2. Endogenous roles of DMT

Although widespread biological presence of DMT is acknowledged, the biological function of DMT remains a mystery. DMT is found in low concentrations in brain tissue (Saavedra and Axelrod, 1972; Christian et al., 1977; Hollister, 1977). DMT concentrations can be localized and elevated in certain instances, for example, DMT production increases in rodent brain under stress (Barker et al., 1981). Formerly, endogenous DMT was thought to exist at concentrations too low to produce pharmacological effects, but two discoveries changed that. First, trace amine-associated receptors (TAAR) are activated by DMT and other molecules (Bunzow et al., 2001) and second, DMT can be locally sequestered in neurotransmitter storage vesicles at pharmacologically relevant concentrations, thereby being able to active other pharmacological receptors, e.g. serotonin (Nagai et al., 2007; Cozzi et al., 2009). These findings suggest that DMT may have a role in normal physiological and/or psychopathology. What that role may be has not yet been established.

Although the serotonin system has been thought to be the main contributor to the psychedelic effects of DMT, other behavioral effects have been observed which do not involve the serotonin or other monoaminergic systems; such as jerking, retropulsion, and tremors (Deliganis et al., 1991; Jenner et al., 1980). In addition, molecular effects of DMT have been identified that are not mediated by serotonin receptors. For example, DMT-enhanced phosphatidylinositol production is not blocked by 5-HT_{2A} receptor antagonists (i.e., ketanserin; Deliganis et al., 1991). More recent hypotheses for molecular roles of endogenous DMT have developed over the last decade, and include the potential involvement of TAAR (mentioned above) and sigma-1 receptors. Interactions of both TAAR and sigma-1 receptors will be discussed in detail in subsequent sections.

There has been a great deal of speculation about the role of DMT in naturally occurring altered states of consciousness, such as psychosis, dreams, creativity, imagination, religious and/or spiritual phenomena, and near-death experiences (Callaway, 1988, Strassman 2001). Additionally, DMT may play a role in waking reality (Wallach, 2009). Waking reality is created in a similar way to altered states except that the normal state correlates with event in the "physical" world. Thus, waking reality can be thought of as a tightly regulated psychedelic experience and altered states arise when this regulation is loosened in some fashion. This model predicts that the sensory-altering effects of administered psychedelics are a result of the compound acting directly via neuropharmacological mechanisms in regions of the CNS involved in sensory perception. More simply, DMT may potentially act as a neurotransmitter to exert a signaling function in regions of the CNS, which are involved in sensory perception (Wallach, 2009).

Other theories propose that DMT may be important in psychiatric disorders. Data from early studies of DMT suggested that DMT may be a schizotoxin, and various authors hypothesized that DMT was a key factor in causing schizophrenia (Osmond and Smythies, 1952; Gillin et al., 1976, reviewed by Szara 2007). This hypothesis is no longer accepted,

but it is still thought that DMT may play a role in psychotic symptoms (Daumann et al., 2010; Warren et al., 2012). Similarly, DMT was thought to be neurotoxic, but more recent research suggests that DMT may actually be neuroprotective (Frecska et al., 2013).

More recently, Jacob and Presti (2005) proposed that endogenous DMT may have an anxiolytic role based on the reported subjective effects of DMT administered in low doses, which would result comparable concentrations and biological actions to those of endogenous DMT. Sensory alterations commonly described by people taking DMT occur only when relatively high concentrations of DMT are administered. These high concentrations are similar to those observed in the synapse when endogenous DMT is released (review, Wallach, 2009).

The putative roles of DMT will be explored in more detail in subsequent sections of this review. The review will begin by addressing the basic mechanisms of action of DMT, both pharmacokinetic and pharmacodynamic. It will then examine evidence regarding the neuropharmacological effects of DMT, from both behavioral studies of the exogenous effects of DMT, and from molecular studies of sites of action of endogenous DMT. Next, the review will turn to the use of DMT both as a model for various disorders and the use of DMT to treat some of these disorders. The review will conclude with the effects of DMT on other organ systems besides the central nervous system.

2. Pharmacokinetics of DMT

Intravenous administration of radio-labeled DMT in rabbits produces entry into the brain within 10 s and excretion via the kidneys, such that no traces of DMT or metabolite was measured in urine 24 h post administration. However, DMT could still be detected at 2 and 7 days (0.1% of initial dose) post administration (Vitale et al., 2011). In the same study, tryptamine was eliminated within 10 min. These findings show that even after complete clearance of a dose of DMT from the blood, DMT is still present in the CNS, and imply that DMT is being produced in the CNS. The subjective effects of intravenous administration of DMT (typical dose 0.1 - 0.4 mg/kg; Strassman et al., 1996) peak at about 5 min and are gone by 30 min. Intramuscular effects of DMT hydrochloride or DMT fumarate (reported dose 0.2-1 mg/kg; Szara, 2007) have a rapid onset within 2-5 min and can last 30 - 60 min, and the effects are generally less intense than intravenous or inhalation routes of administration. The hallucinogenic effects of DMT in the formulation of ayahuasca (0.6 -0.85 mg/kg DMT; Riba et al., 2003) generally appear within 60 min, peak at 90 min and can last for approximately 4 h (Cakic et al., 2010). Typical doses of smoked or inhaled free-base DMT are 40 - 50 mg, although dose may be as high as 100 mg (Shulgin and Shulgin, 1997). The onset of these doses of smoked DMT is rapid, similar to that of i.v. administration, but lasts less than 30 min. Smoked DMT effects are extremely intense (Strassman, 2001; Turner, 1994). Intranasal free-base DMT was inactive (0.07 – 0.28 mg/kg; Turner and Merlis, 1959) as was DMT administered rectally as 1.7 mg/kg of the bioxalate salt (de Smet, 1983).

To establish that DMT acts as a neurotransmitter rather than merely being a by-product of the metabolism of other bioactive molecules, it is necessary to establish that it is synthesized, stored, and released. It is of interest that DMT can pass through three barriers

with the help of three different mechanisms so that it can be compartmentalized and stored with the brain (described in detail below). These three mechanisms may yield high intracellular and vesicular concentrations within neurons (Frecska et al., 2013), which suggests that DMT may have a biological role. Processes for the transport of glucose and amino acids are given similar biological priority, which may suggest that DMT is present in the body for more than its psychedelic effects, such as an adaptive role in biological processes, or a universal role in cellular protective mechanisms.

2.1. Synthesis

Endogenous DMT is synthesized from the essential amino acid tryptophan, which is decarboxylated to tryptamine. Tryptamine is then transmethylated by the enzyme indolethylamine-N-methyltransferase (INMT) (using S-adenosyl methionine as a substrate), which catalyzes the addition of methyl groups resulting in the production of Nmethyltryptamine (NMT) and DMT. NMT can also act as a substrate for INMT-dependent DMT biosynthesis (Barker et al., 1981). INMT is widely expressed in the body, primarily in peripheral tissue such as the lungs, thyroid and adrenal gland. INMT is located in intermediate levels in placenta, skeletal muscle, heart, small intestine, stomach, retina, pancreas, and lymph nodes. It is densely located in the anterior horn of the spinal cord (Mandell and Morgan., 1971; Mavlyutov et al., 2012; Morgan and Mandell, 1969; Thompson et al, 1998, 1999; Wyatt et al., 1973). Within the human brain, highest INMT activity has been found in uncus, medulla, amygdala, frontal cortex (Mendell and Morgan, 1971), and in the fronto-parietal and temporal lobes (Saavedra et al., 1973). Cozzi et al. (2011) has shown INMT is also located in the pineal gland. Based on rodent brain cellular fractionation studies 70% of INMT activity is found in the supernatant and 20% in the synaptosomal fractions, suggesting the enzyme is located in the soma of cells, which are fractured during the homogenization process (Saavedra et al., 1973). The wide distribution of INMT implies a wide distribution for DMT.

The enzymatic activity of INMT is closely regulated by endogenous inhibitors (Lin, 1974; Marzullo et al., 1977). DMT at high concentrations (10⁻⁴ M) yields a 90% inhibition of rabbit lung INMT (Thithapandha, 1972). In addition, the same tissues that contain INMT also contain enzymes that metabolize DMT. Only a small fraction of DMT made intracellularly is actually released into the blood. This process helps explain the inconsistent detection levels assessed in many studies discussed below (Karkkainen et al., 2005; Barker et al., 2012; see endogenous section). DMT production is increased under stress in rodent brain and adrenal gland (Christian et al., 1977; Beaton and Christian, 1977). Whether the stress-induced mechanism for increasing DMT is due to increasing INMT activity, or a decrease in DMT metabolism remains unknown.

2.2. Accumulation and storage

As previously mentioned, it has been hypothesized that high, local concentrations of DMT can occur within neurons (Frecska et al., 2013) and potentially widely produced in peripheral organs, especially in the lungs. Frecska and colleagues (2013) summarized a three-step process by which DMT is accumulated and stored. In step 1, DMT crosses the blood brain barrier by active transport across the endothelial plasma membrane, which is

accomplished via Mg2+ and ATP-dependent uptake (Christian et al., 1977; Cohen and Vogel, 1972; Cozzi et al., 2009; Sangiah et al., 1979, Barker et al., 1982; Sitaram et al., 1987; Takahashi et al., 1985; Yanai et al., 1986). In step 2, uptake of DMT into neuronal cells is accomplished via serotonin uptake transporters (SERT) on neuronal plasma membrane (Berge et al., 1983, Nagai et al., 2007, Whipple et al., 1983). In step 3, facilitated sequestration of DMT into synaptic vesicles from the cytoplasm is accomplished by the neuronal vesicle monoamine transporter 2 (VMAT2; Cozzi et al., 2009). DMT inhibited radiolabeled 5-HT uptake via the serotonin transporter (SERT) and VMAT2 with Ki values of 4 and 93 μ M, respectively (Nagai et al., 2007; Cozzi et al., 2009). DMT that has been taken up and stored within cells via SERT and VMAT2 and exhibit high binding-to-uptake ratios, >11 for SERT and >10 for VMAT2. High binding ratios suggest that there are

The high levels of DMT concentration found in vesicles are needed for various pharmacological actions including activation of sigma-1 receptors and TAARs as described below. Once uptake and storage of DMT has been completed, it can remained stored in vesicles for at least 1 week and can be released under appropriate stimuli (Vitale et al., 2011). Through these three steps, peripheral synthesis of DMT, consumption of DMT-containing plant matter, or systemic administration of DMT can influence central nervous system functions (Frecska et al., 2013).

separate substrate and inhibitor sites for SERT and VMAT2 and further supports that DMT

(and other tryptamines) are substrates for both transporters.

2.3. Bioavailability of exogenous DMT

DMT is not orally active. This is likely due to rapid degradation by peripheral monoamine oxidase (MAO), the enzyme (located in mitochondria) responsible for catalyzing the oxidative deamination of endogenous biogenic amines (McKenna et al., 1984). For hallucinogenic or psychedelic phenomena to occur, plasma concentration must be between $12-90 \mu g/L$ with an apparent volume of distribution of 36-55 L/kg (Callaway et al., 1999; Riba et al., 2003; Yritia et al., 2002), which roughly corresponds to a plasma concentration of $0.06 - 0.50 \,\mu$ M. In order for DMT to be bioavailable, oral formulations such as ayahuasca contain Banisteriopsis caapi and other beta-carboline harmala alkaloids that act as MAOIs (Schultes, 1972; Shulgin, 1976; Der Marderosian et al., 1968; Agurell et al., 1968). MAO-A inhibitors such as iproniazid prolongs the half-life of DMT in rat brain (Lu et al., 1974), and can extend the time course effects of DMT in drug discrimination from 30 min to 60 min (Gatch, unpublished data). Exogenous DMT formulations containing a reversible MAOI (such as ayahuasca) can result in blood levels up to 1.0 mg/ml or higher (Dos Santos, 2011). On average a 100 mL dose of ayahuasca contains about 24 mg of DMT (Callaway et al., 1996). Interestingly, DMT is itself a short-acting monoamine oxidase inhibitor at high doses (maximum effects at 50 mg/kg), and is selective for MAO-A (Reimann and Schneider, 1993, Smith et al., 1962; Waldmeier and Maitre, 1977). In these studies, DMT decreased serotonin and dopamine deamination in rat striatum concomitantly with rapid onset (15 min). Normalization occurred 2 hours later with an ED_{50} of 25 mg/kg for degradation of both serotonin and dopamine.

2.4. Degradation and elimination

A small fraction of exogenous DMT is excreted in urine as the parent compound (Barker et al., 1981). DMT is primarily metabolized by MAO, although there is evidence that DMT can also be metabolized by peroxidases, leading to a variety of other metabolites. In both human and rodent models, none of the metabolites produced DMT-like effects (Szara, 1956; Szara et al., 1961; Rosenberg et al., 1964; Barker, 1978). From the MAO pathway, several indole compounds are produced: N-methyltryptamine (NMT), 6-hydroxy-DMT (6-OH-DMT), 6-OH-DMT-N-oxide (6-OH-DMT-NO), DMT-N-oxide (DMT-NO), and indole-3-acetic acid (IAA) (Fish et al., 1955; Szara, 1956). The major metabolites of DMT are DMT-NO, and IAA (Fish et al., 1955; Barker et al., 2013). In rabbit liver microsomal fractions pretreated with MAOI iproniazid, five indole compounds were found: DMT, NMT, 6-OH-DMT, 6-OH-DMT-NO, and DMT-NO; however, in rabbit brain microsomal preparation again with iproniazid pretreatment, no 6-hydroxy metabolites were identified (Szara, 1956). These findings suggest that metabolism of DMT is somewhat different in brain and in the periphery.

Formation of IAA was thought to be likely due to oxidase deamination of NMT (Barker et al., 1980), but was later established to be also in part by direct deamination by MAO (Barker et al., 1982). Pretreatment with the MAO-I iproniazid in rat whole brain homogenates inhibited IAA formation by 83%, NMT and DMT-NO formation were inhibited by 90%, suggesting that the increase in behavioral half-life of DMT is due to MAO-inhibition and inhibition of the enzymes responsible for demethylation and N-oxidation as well (Barker et al., 1980).

Erspamer (1955) first recorded IAA as a metabolite of systemically administered DMT in rodent urine, which represented less than 3% of the injected dose of DMT. In human volunteers, 8.3% of the administered dose of DMT was recovered as IAA (Szara, 1956). Around 50% was recovered as IAA but also as DMT-NO (10%) and other MAO-independent compounds (Riba et al., 2012). Neither the Erspamer nor Szara study detected unchanged DMT in urine. Little DMT is found unchanged in the urine of ayahuasca users despite taking it with harmala alkaloids (MAO inhibitors) (McIlhenny et al., 2011). In another study in humans, 0.16% of DMT (injected intramuscularly) was recovered as DMT following a 24-hour urine collection, (Kaplan et al., 1974). In both of these studies, DMT concentration peaked in blood within 10-15 minutes and was essentially undetectable by one hour. Approximately only 1.8% of the injected dose was present in blood at any one time.

Oxidative deamination of DMT by MAO may not be the sole metabolic pathway in humans (Riba et al., 2012). A study by Gomes et al. (2014) suggests that a different metabolic pathway by which DMT can be oxidized by peroxidases may be responsible for increasing cytotoxic activity of peripheral-blood mononuclear cells (Tourino et al., 2013). Metabolites in this pathway include hydroxy-DMT, N,N-dimethyl-N-formyl-kynuramine, and N,N-dimethyl-kynuramine. Barker et al. (1980) suggest other possible metabolites of DMT include 1,2,3,4-tetrahydro-beta-carboline (THBC) and 2-methyl-THBC.

DMT as an endogenous compound can be measured in human body fluids, including blood, urine and cerebral spinal fluid. Levels of endogenous DMT do not appear to be regulated by diet or gut bacteria (Barker et al., 2012). Infrequent and inadequate sampling methods used over time make it difficult to determine specific details pertaining to DMT production in the body. For example, we still do not know if DMT is produced in phasic or diurnal cycles. Measureable concentrations seem to only occur intermittently (Oon et al., 1977), and exact tissue source or sources of DMT is still unclear. It is commonly thought that the adrenal gland and lungs are the most common places for the highest amount of DMT production, since this is where highest levels of INMT have been reported (Thompson and Weinshillboum, 1998; Thompson et al., 1999).

A review by Barker in 2012 assessed 69 studies that reported endogenous DMT detection and quantities reported in urine (29 studies), blood (11 studies), and cerebrospinal fluid (4 studies) from 1955 – 2010 primarily comparing detection levels within healthy controls and schizophrenic patients. DMT in urine was examined in 861 individuals (635 patients), 276 patients and 145 controls were positive for DMT. Throughout the studies, there were inconsistent sampling methods, including various of amounts of urine used in assays, and a range of techniques and analytical approaches were used. Some studies took dietary influences into consideration, but found no associations with endogenous DMT levels. Inconsistent units of measurement were also used across studies. Concentrations in urine range from 0.02 to 42.98 ± -8.6 (SD) ug/24h, and from 0.16 to 19 ng/ml. In blood, data from 417 (300 patients) individuals were examined, 44 patients and 28 controls were positive for DMT. One study was responsible for 137 of the negative samples. Like detection in urine, extraction methods and analytical approaches were highly inconsistent. Testing procedures included discrepancies of samples coming from plasma, serum and/or whole blood, while others had limit detections of 0.2 DMT/ml. Higher concentrations of DMT are extracted from whole blood compared to plasma (Riceberg et al., 1978), but there is no difference in venous and arterial blood (Walker et al., 1979). When concentrations were reported, not just whether it was present or not present, it ranged from 51 pg/ml (HPLC-radioimmunoassay) to 55 ng/ml (direct fluorescence assay of extracts). DMT was detected in cerebrospinal fluid in 4 studies, which tested 136 individuals (82 patients). Of those, 34 patients and 22 controls were positive for DMT. Concentrations ranged from 0.12 to 100 ng/ml. DMT can be detected as an endogenous compound in urine, blood, and cerebrospinal fluid. Even with inconsistent detection methods, DMT does not appear to be related to the onset of schizophrenia, since it seems to be detected more so in healthy controls compared to patients.

3. Pharmacodynamics

3.1. Clinical effects

Oral dosing of DMT via ayahuasca produces both behavioral and neurochemical effects, such as decreases in motor activity (Pic-Taylor et al., 2015), impairment of cognitive function (Alonso et al., 2015; Bouso et al., 2013), sympathomimetic effects, increased prolactin and cortisol levels, and decreased lymphocytes increased natural killer cells (Dos

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Santos, et al., 2011). Doses of ayahuasca 15 or 30-fold higher than commonly used ritual doses increased serotonergic neurotransmission (Pic-Taylor et al., 2015). Long-term use of DMT in ayahuasca produces measurable brain changes. Long-term ayahuasca users show difference in midline brain structures using MRI versus matched controls (Bouso et al., 2015). Interestingly, whereas ayahuasca produced modest impairment of cognitive function in inexperienced users, little or no impairment was observed in experienced users (Bouso et al., 2013).

3.1.1. Tolerance—Several early studies demonstrated that DMT does not produce tolerance. When DMT was administered to squirrel monkeys (2 mg/kg, i.m.) for 36-38 days, it failed to elicit tolerance to the disruption of responding maintained on a fixed-ratio schedule of food reinforcement (Cole and Pieper, 1973). Similarly in cats, Gillin et al. (1973) demonstrated that DMT (3 mg/kg, i.p.) did not produce tolerance when administered 7-15 days twice daily or every 2 or 24 hours to its effects on EEG, pupil dilation, coordination, posture, and other physical signs. To the contrary, an increase in sensitivity to repeated injections were observed. However, following administration of higher doses of DMT and more frequent injections, partial tolerance to DMT in rats occurred with dose ranges of 3.2 – 10 mg/kg every 2 hours for 21 days (Gillin et al., 1973). Cross-tolerance to LSD (0.1 mg/kg) after tolerance to 3.2 mg/kg DMT was established; however, only slight tolerance to LSD was established following 10 mg/kg DMT (Koviac and Domino, 1976).

In humans administered 4 repeated doses of DMT 30 minutes apart, Strassman et al. (1996) observed no tolerance to the subjective effects of intravenous DMT as measured by the Hallucinogen Rating Scale. However, tolerance did develop to change in body temperature and other physiological factors. These findings will be discussed in more detail in the paragraph on cardiovascular effects in section 5. Mild cross-tolerance to DMT was reported in humans made tolerant to LSD (Rosenberg et al., 1964; Jenner et al., 1980). Taken together, these findings suggest that tolerance can develop to the cardiovascular and other peripheral effects of DMT, although little or no tolerance develops to the subjective effects.

3.1.2. Subjective effects of DMT—Because the subjective effects of hallucinogens seem to drive their use rather than effects on the reward/ reinforcement areas of the brain, drug discrimination is often used as an animal model for testing the behavioral effects of hallucinogens. A compound can be tested for its ability to "substitute", that is, produce drug-appropriate responding in test subjects trained to discriminate a psychoactive compound from its vehicle or from other psychoactive compounds. Typically, drug-appropriate responding greater than 80% is considered "full substitution". Conversely, novel compounds can also be trained as discriminative stimuli if they have psychoactive effects, and known compounds can be tested for substitution or antagonism of the novel compound. Asymmetries in cross-substitute for compound A) can indicate that the two compounds may have overlapping, but not identical mechanisms of action (e.g., Grant, 1999). Drug discrimination can be useful in investigating potential mechanisms of action of the trained discriminative stimulus by utilizing selective agonists and antagonists to either mimic or

block the effects. Subsequent paragraphs will examine discrimination studies assessing potential mechanisms of action of DMT.

DMT produced discriminative stimulus effects similar to those of the classic serotonergic hallucinogens DOM and LSD, as DMT fully substituted in DOM-trained rats (Glennon et al. 1983; Glennon 1986) and produced full or near-full (78% drug appropriate responding) substitution in LSD-trained rats and pigeons (Appel et al. 1999; Helsley et al. 1998; Jarbe, 1980). The effects of DMT seem to be mostly hallucinogen-like, as it produced only 50% drug-appropriate responding in 3,4-methylenedioxymethamphetamine (MDMA)-trained rats, and produced a maximum of 37% drug-appropriate responding in methamphetamine-trained rats (Gatch et al., 2009). In rats trained to discriminate between the 5-HT_{2A} antagonist ketanserin and the 5-HT_{2A} agonist 2,5-dimethoxy-4-iodoamphetamine (DOI), DMT produced DOI lever-responding 80% or more of time (Smith et al., 1998), indicating that DMT acted more like a 5-HT_{2A} agonist than a 5-HT_{2A} antagonist.

Despite its very short duration of action, DMT can be trained as a discriminative stimulus (Gatch et al., 2009). A wide range of synthetic phenethylamine hallucinogens fully substitute for DMT, including DOM, DOC, LSD, 2,5-dimethoxy-4-methylphenethylamine (2C-D), 2,5-dimethoxy-4-ethylphenethylamine (2C-E), and 2,5-dimethoxy-4iodophenethylamine (2C-I), whereas 4-chloro-2,5-dimethoxyphenethylamine (2C-C) and 2,5-dimethoxy-4-ethylthiophenethylamine (2C-T-2) produced a maximum of only 75% DMT-appropriate responding (Eshleman et al., 2014). In contrast, other tryptamine hallucinogens produced more equivocal effects, with DiPT and 5-methoxy-diethyl tryptamine (5-MeO-DET) producing full substitution, 4-Hydroxy-N,N-diisopropyl tryptamine (4-OH-DiPT) and 5-methoxy-N-isopropyl-N-methyl tryptamine (5-MeO-IMPT) producing partial substitution, and 5-methoxy- α -methyl tryptamine (5-MeO- α MT) producing little if any DMT-like effects (Gatch et al. 2009; 2011). In addition, although DiPT fully substituted in DMT-trained rats (Gatch et al., 2011), DMT only produced 65 % DAR in DiPT-trained rats (Carbonaro et al., 2013). Taken together, these findings indicate that serotonergic hallucinogens largely produce discriminative stimulus effects similar, but not entirely identical to those of DMT.

3.2. Pharmacological Mechanisms

The mechanisms of action for hallucinogens are currently not well understood. The 5- HT_{2A} receptor is thought to be necessary, but not sufficient for hallucinogenic effects, and 5- HT_{2C} and 5- HT_{1A} receptors may play important roles as well (see review by Nichols, 2004). DMT interacts with a variety of serotonin receptors, but also with ionotropic and metabotropic glutamate receptors, dopamine, acetylcholine, TAAR, and sigma-1 receptors. Current information on the roles of these receptors in mediating the effects of DMT is reviewed in the following paragraphs.

3.2.1. Serotonin—Most studies to date focus on DMT (and most classic psychedelics) as a partial agonist of serotonin (5-HT) receptors, primarily the 1A, 2A, and 2C receptor subtypes, with predominant interest at 5-HT_{2A} receptors. DMT binds 5-HT_{1A}, 5-HT_{1B}, 5-HT_{1D}, 5-HT_{2A}, 5-HT_{2B}, 5-HT_{2C}, 5-HT_{5A}, 5-HT₆ and 5-HT₇ receptors with affinities

ranging from 39 nM to 2.1 μ M (Keiser et al., 2009). The 5-HT_{2A} receptor is thought to be the primary target of classic serotonergic-mediated psychedelic compounds, such as LSD, DOI, psilocin, and mescaline, although 5-HT_{1A} and 5-HT_{2C} receptors may also play some role (Aghajaian and Marek, 1999a, 1999b; review Nichols 2004). DMT has been reported to bind to all three of these receptors in a variety of studies, including the 5-HT_{1A} (Pierce and Peroutka, 1989; McKenna et al., 1990), 5-HT_{2A} (Lyon et al., 1988; Pierce and Peroutka, 1989, McKenna et al., 1990; Smith et al. 1998), and 5-HT_{2C} receptors (Smith et al., 1998; Nagai et al. 2007; Cozzi et al. 2009; Gatch et al., 2011).

Agonistic properties and affinities for 5-HT_{1A} receptor vary among the classic psychedelics. Interestingly, agonist activity at the 5-HT_{1A} receptor opposes the subjective effect of 5-HT_{2AR} agonists (Araneda and Andrade, 1991; Strassman, 1996; Jacob and Presti, 2005). DMT's affinity for the 5-HT_{1A} receptor is higher compared to 5-methoxy-dimethyl tryptamine (5-MeO-DMT), 6.5 +/- 1.5 nM and 170 +/- 35 nM, respectively (McKenna et al., 1990). A 5-HT_{1A} antagonist significantly increased the reported psychological effects of DMT (Strassman, 1996). DMT, like other tryptamine hallucinogens, but not phenethylamines, inhibits dorsal raphe cell firing. This mechanism is hypothesized to be an underlying basis of psychedelic-like effects (Aghajanian et al., 1970), which may be mediated by stimulation of 5HT_{1A} somatodendritic receptors (Sprouse and Aghajanian, 1987; 1988).

DMT does bind to 5-HT_{1D} receptor (Hamik and Peroutka, 1989; Heuring and Peroutka 1987; Pierce and Peroutka 1989) and 5-HT₃ receptor (Carbonaro, unpublished data), however little has been investigated to follow these results up. Delgado (2005) shows that 5-HT₁ and 5-HT₃ receptors exert anxiolytic effects, which does correspond to some reports of DMT use. DMT is an agonist at 5-HT_{2C} receptors. In drug discrimination, the DMT-like effects were partially blocked by a selective 5-HT_{2C} antagonist, SB242084 (Carbonaro et al., 2015). The 5-HT_{2C} receptor is likely less significant in the psychedelic effects since tolerance develops to the 5-HT_{2C} receptor (Smith, 1998). Little or no tolerance is developed to the subjective effects of DMT in clinical studies (Strassman, 1996).

DMT binds to the 5- HT_{2A} receptor with relative high affinity IC_{50} 75 +/- 1 nM (McKenna 1990), yet other psychedelics that lack visual effects have a higher affinity for the 5- HT_{2A} receptor (5-MeO-DMT; 14+/- 1 nM; McKenna et al., 1990). The 5- HT_{2A} receptor seems to be necessary, but is not sufficient to account for the visual phenomenon common of the classic hallucinogens. Psychedelics and psychedelic-like compounds including MDMA, 5-MeO-DMT, DET (review Wallach 2009), and DiPT (Gatch et al., 2011; Carbonaro et al., 2015) are 5- HT_{2A} receptor agonists. Subjective effects for these compounds are reported to be solely emotional, devoid of visual phenomenon common in other psychedelics such as DMT, except in rare circumstances where individual differences in biology seem to be the regulating factor (Wallach reviewed, 2009).

Head twitch response in rodents is thought to be a 5-HT_{2A} receptor-mediated behavior produced primarily by psychedelics, although it is likely that other receptors play a role in this behavior, including 5-HT_{2C} and glutamatergic receptors. Like other classic psychedelics, DMT does induce this head twitch response in C57Bl/6 mice, which is

blocked by 5-HT_{2A} inverse agonist, MDL100907 (Carbonaro et al., 2015). However, the overall number of head twitches induced by DMT is much smaller compared to most other psychedelic compounds. DMT failed to produce this head twitch response in Swiss Webster mice (Fantegrossi et al., 2006) These discrepancies may be due to the rapid degradation of DMT or other peculiarities specific to DMT.

Functional selectivity on how psychedelic compounds modulate the 5-HT₂ receptor family is not well understood. The 5-HT2 family of receptors are Ga/11 mediated and primarily use the phospholipase C second messenger system pathway (Brown and Tracy, 2013), but also an phospholipase A₂ (Nichols, 2004). Phospholipase C hydrolyzes phosphatidylinositol membrane lipids generating inositol-1,4,5-triphosphate (IP₃) and diacylglycerate. Diacylglycerate remains bound to the membrane and leads downstream activation of protein kinase C and increases the release of calcium from intracellular stores (Conn and Sanders-Bush, 1984; Brown and Tracy, 2013). In particular, protein kinase C can mediate desensitization (Roth et al., 1995) of 5-HT_{2A} receptors during drug exposure. Phospholipase A₂ stimulation can lead to formation of arachidonic acid (Nichols, 2004). DMT stimulates arachidonic acid release (max 5-HT effect = 93%; EC₅₀ = 260 nM) and less inositol phosphate formation (metabolite of IP₃, max 5-HT effect = 39%; EC₅₀ = 269 nM) via the 5-HT_{2A} receptor. Whereas inositol phosphate formation via the 5-HT_{2C} receptor seems to be more efficacious and more potent (max 5-HT effect = 99%; $EC_{50} = 114$ nM; Eshleman et al., 2014). Stimulation of phospholipase A2 does not seem to directly related to the subjective effects of psychedelic compounds (Halberstadt, 2015; Kurrasch-Orbaugh et al., 2003). Other pathways such as the phospholipase D may play a role, but DMT-mediated effects had not been thoroughly investigated. The importance of each second messenger pathway is an important area of future investigation.

DMT, like other classic hallucinogens increase 5-HT levels and/or decrease the turnover of 5-HT (review Nichols, 2004). DMT increases excretion of IAA and 5-hydroxy IAA in humans (Szara, 1956). Other studies have reported an increase in 5-HT and a decrease in 5-hydroxy IAA after DMT administration (Freedman et al., 1970; Randic and Padjen, 1971). DMT seems to have no effect on tryptophan hydroxylase (Andén et al., 1971), but produces a main effect on the rate of 5-HT turnover (Gillin and Wyatt, 1976). DMT inhibited SERT transport and VMAT2, acting as a substrate and not as an uptake blocker.

3.2.2. Glutamate and 5-HT/glutamate interactions—An approach gaining increasing interest within the last decade is to examine interacting roles of serotonin and glutamate in mediating the effects of DMT. Of particular interest are the roles of group II metabotropic glutamate receptors (mGluR2/3), the NMDA receptor, and 5-HT_{2A} receptors in modulating the levels of glutamate in the synapse. These group II glutamate receptors (mGluR2/3) may also be potential target sites for mediating hallucinogenic effects (Gonzalez-Maeso et al. 2007, 2008; Delille et al. 2012; Moreno et al. 2011; Winter et al. 2004).

mGlu2/3 receptor agonists can act presynaptically to suppress glutamate release, although the significance of this effect in mediating the effects of DMT has not been systematically studied. In contrast, mGluR2/3 antagonist increases the amount of glutamate in the synapse, creating a potentiation of hallucinogenic or psychedelic effects (Cartmell et al. 1999;

Forsythe and Barnes- Davies 1997; Ohishi et al., 1994; Shigemoto et al., 1997). The 5-HT_{2A} receptor inverse agonist, MDL100907, fully blocked the discriminative stimulus effects and head twitched produced by DMT, whereas the 5-HT_{2C} receptor antagonist (SB242084) produced little or no effect on the discriminative stimulus effects of DMT. A mGlu2/3 receptor agonist (LY379268) produced modest decreases in the discriminative stimulus effects of DMT, whereas a mGlu2/3 receptor antagonist (LY341495) facilitated the effects of low doses of DMT (Carbonaro et al., 2015). Comparatively, a mGluR2 agonist blocked the discriminative stimulus effects of LSD, whereas a mGlu2 antagonist facilitated the discriminative stimulus effects of LSD (Winter et al., 2004). Further, mGluR2 knockout mice showed little or no head twitch following 2,5-dimethoxy-4-iodo-amphetamine (DOI), and some signaling was disrupted, which may mean that mGlu2 receptors are necessary for hallucinogenic activity (Moreno et al., 2011). Systematic administration of DOI increases glutamate efflux in ventral tegmental area (Pehek et al., 2006).

Electrophysiological studies suggest that stimulation of $5HT_{2A}$ receptors in the medial prefrontal cortex increases pyramidal cell activity and may stimulate corticotegmental glutamatergic projection neurons (Aghajanian and Marek, 1997). A possible explanation for these effects is that mGlu2 receptors co-localize with 5-HT_{2A} receptors to form heteroreceptor complexes (Delille et al. 2012; Gonzalez-Maeso et al. 2007; 2008). It has been suggested that the heteroreceptors induce a psychedelic-specific second messenger cascade (Gonzalez-Maeso et al., 2007; 2008), although this has not been definitively established (Delille et al., 2012).

There has been some evidence that NMDA receptors may also play a role in mediating the effects of DMT. DMT partially blocked the discriminative stimulus effects of phencyclidine, which produces hallucinations through its actions at NMDA receptors (West et al., 2000). In addition, activation of sigma-1 receptor by DMT may lead to potentiation of NMDA receptors (Cozzi et al., 2009).

3.2.3. Dopamine—DMT lacks direct dopaminergic properties, since it did not stimulate dopamine (DA)-sensitive adenylate cyclase (von Hungen et al., 1975). This finding is in agreement with data from a behavioral technique often used to assess direct dopamine agonist effects, which records turning behavior in unilateral nigro-striatal lesioned rats. If a compound stimulates dopamine receptors directly, the animal will rotate toward the intact side (contralateral turning), otherwise if a compound induces dopamine release in the striatum from the nerve terminals of the intact side induces a rotation toward the lesion side (ipsilateral turning). As reviewed by Barker (1981), Pieri et al., (1974) suggest that DMT appears to have no dopamine receptor agonist effects, although higher doses of DMT (10 mg/kg and 20 mg/kg) does produce ipsilateral turning, although were not indicative of a very potent dopamine releasing effect (Trulson et al., 1977; Stern and Dalsass, 1975). Another indication that DMT does not act directly at dopamine receptors is the lack of adenylate cyclase activity in the dorsal striatum of rats (von Hungen et al., 1975).

DMT-induced EEG activation in rabbits can be antagonized by neuroleptics (DA receptor blocking compounds (Moore et al., 1975). This may be due to blockade of downstream sequela. For example, DMT releases dopamine from presynaptic stores (Smith, 1975). This

release of dopamine in combination with the effects of MAO causes an indirect dopaminergic stimulant activity (Waldmeier and Maitre, 1977). DMT (20 mg/ml) caused 42% decrease in concentration of dopamine in rat forebrain (5 - 60 min) while norepinephrine was not affected, no change in the levels of the dopamine metabolite homovanillic acid (HVA, a common marker for dopamine levels) was observed in corpus striatum. This decrease in concentration of dopamine may be caused by a stimulation of the release of dopamine or by inhibition of its synthesis (Haubrich and Wang, 1977). This decrease in dopamine levels is likely not related to change in synthesis, because no change in norepinephrine levels or turnover rate in the diencephalon were observed (Smith, 1977). It appears as if DMT increases central dopamine turnover and enhances striatal dopamine synthesis in rats (Smith, 1977).

Acute and chronic administration of DMT significantly increased endogenous levels of striatal 3-MT (3-methoxytyramine, a dopamine extraneuronal metabolite) (Waldmeier et al., 1976). Dopamine steady state concentrations remained unchanged. Further, DMT increased accumulation of ^{3H}DA and ^{3H}3MT newly formed from ^{3H}DOPA (Waldmeier and Meitre, 1977). 3,4-Dihydroxyphenylacetic acid (DOPAC, a major metabolite of dopamine) more efficiently lowered by DMT rather than HVA (Waldmeier and Meitre, 1977) in the striatum and whole brain. This is distinct from the effects of classic MAOIs, which decrease both DOPAC and HVA (Maitre et al., 1976; Waldmeier et al., 1976). After acute administration striatal dopamine synthesis was increased, yet there was no effect on steady state conditions. Dopamine degradation must be enhanced proportionally and is likely done so extraneuronally, due to the increase in 3-MT (Rech et al., 1971; Smith, 1977). No change in the increase of DA turnover over one month treatment (5 mg/kg, Smith, 1977), with consistent rises in 3-MT is observed.

3.2.4. Acetylcholine—Little investigation has occurred in reference to DMT's effect on acetylcholine. DMT significantly decreases concentration of acetylcholine in corpus striatum, which may be due to a direct release of acetylcholine, thus reducing concentration of striatal acetylcholine (Haubrich and Wang, 1977). Generally, acetylcholine levels in brain are reduced when its rate of release or turnover are increased (Haubrich and Wang, 1977). DMT had no effect on the level of acetylcholine in the cortex.

3.2.5. Trace Amine-Associated Receptors—Trace amine-associated receptors (TAARs) are a more recently discovered class of receptors which may play a role in mediating DMT and other psychedelic drug effects. The rat trace amine-associated receptor – 1 (rTAAR1) is a G protein-coupled receptor with homology with members of the catecholamine receptor family. Trace amines p-tyramine, and p-PEA stimulate cAMP production, are commonly measured to assess activation of the TAAR. DMT binds to the rTAAR-1 with high affinity and acts as an agonist, causing activation of adenylyl cyclase and resultant cAMP accumulation in HEK293 cells transfected with rTAAR1. Other psychedelics such as (+/-)DOI, d-LSD, and 5-MeO-DMT, and non-psychedelics such as R(+)lisuride, (+/-)MDMA and amphetamine also stimulate cAMP production through their effects at rTAAR1. This second-messenger cascade does not seem to be selective for any of these compounds as these effects occurred at approximately 1 μM concentration (no other

concentrations tested). rTAAR1 seems to be located in the intracellular puncta, and not at the plasma membrane *in vitro;* it is not known if this is the case *in vivo* (Bunzow et al., 2001).

Because TAARs were discovered long after research had on DMT (and other psychedelic compounds) had been initiated at the $5HT_{2A}R$, there is a paucity of research on the role of TAAR, which makes it difficult to discern what role this class of receptor may play in mediating the effects of endogenous and exogenously administered DMT. It is unknown whether the typically used $5-HT_{2A}R$ antagonists ketanserin and/or risperidone have any antagonist effects of TAAR as well. This is an area where more research needs to be done to fully understand the importance of TAARs and psychedelic effects.

3.2.6. Sigma-1 receptor—The sigma-1 receptor was once thought to be a subtype of an opioid receptor. It has been implicated to have a role in several neurobiological diseases and conditions such as addiction, depression, amnesia, pain, stroke, and cancer (Collier et al., 2007). It is found widely distributed though out the body including in the CNS, liver, heart, lung, adrenal gland, spleen, and pancreas (Hayashi and Su, 2007; Weissman et al., 1988). They are localized between endoplasmic reticulum and mitochondrion (mitochondria-associated ER membrane; MAM). Sigma-1 receptor agonists signal the receptor to disassociate itself form other endoplasmic reticulum chaperones, which allows the receptor to act as a molecule chaperone to IP₃ receptors. This enhances calcium signaling from the ER to mitochondria, activates TCA cycle and increase ATP production. Sigma-1 receptors can translocate to plasma membrane or sub plasma membrane area when stimulated with higher concentrations of agonists or when sigma-1 receptors are over-expressed. Once sigma-1 receptors translocate to the plasma membrane they can interact with and inhibit several ion channels (reviewed in Cozzi et al., 2009). Sigma-1 receptor activation can also lead to potentiation of NMDA receptors.

Psychedelics and non-psychedelics bind promiscuously to sigma-1 receptors. DMT binds to sigma-1 receptors at low micromolar concentrations. (EC₅₀ = 14 μ M) and appears to have agonist-like effects. DMT inhibits cardiac voltage-activated sodium ion channels at higher concentration (100 M) in HEK293 cells (inhibited 62 +/- 3%), COS-7 cells (inhibited 22 +/- 4%; lower concentration of endogenous sigma-1 receptor), and neonatal mouse cardiac myocytes (inhibited 29 +/- 3%), induces hypermobility in wild-type mice, which is blocked in sigma-1 receptor knock-out mice (Fontanilla et al., 2009). DMT modulated current (patch-clamp) in sigma-receptor-mediated Na⁺ channels, which was reduced by sigma-1 receptor knockdown and by progesterone (Johannessen et al., 2013). In addition, DMT synthesizing enzyme indolethylamine-N-methyltransferase is co-localized with sigma-1 receptor in C-terminals of motor neurons (Mavlyutov et al., 2012), which suggests that there may be adequate levels of endogenous DMT to activate sigma-1 receptors.

The main problem with the theory that DMT is an endogenous sigma-1 receptor agonist is that it requires concentrations in the micromolar range, whereas selective sigma-1R agonists such as (+)-pentazocine have affinities in the nanomolar range (Fontanilla et al., 2009). If DMT is only available in trace amount in humans and is rapidly metabolized (Burchett and Hicks, 2006), how can DMT levels rise enough to account for sigma-1 receptor-mediated effects? One possible explanation for this is the three step process of accumulation and

storage discussed earlier, which includes active transport across the blood brain barrier, and DMT may be a substrate for transporters at the cell surface (SERT; Nagai et al., 2007) and at the neuron level (VMAT2; Cozzi et al., 2009). Supporting the role of sigma-1 receptor is that the SSRI fluvoxamine, has sigma-1 receptor agonist properties with higher affinity than DMT. Fluvoxamine works better with patients suffering from psychotic depression compared to antidepressants without sigma-1 receptor agonist properties (Stahl, 2008). Selective sigma-1 receptor agonists do not cause psychotomimetic effects in animals. At best, sigma-1 receptors may partially mediate the subjective effects of DMT (see review by Su et al., 2009).

Whether or not the sigma-1 receptor plays a significant role in the psychedelic effects of DMT, it may still play an important role in other physiological mechanisms. Sigma-1 receptors agonists are potentially neuroprotective via several mechanisms (see review Frecska et al., 2013). DMT reduced inflammation ostensibly via sigma-1 receptor (Szabo and Rajnavdgyi, 2014), and can induce neuronal plasticity, which is a long-term recuperative process that goes beyond neuroprotection (Ruscher et al., 2011; Tsai et al., 2009; Kourrich et al., 2012). Sigma-1 receptors can regulate cell survival and proliferation (Collina et al., 2013), thus if DMT is an endogenous agonist, this may explain physiological relevance and importance of why DMT has 3-step uptake process.

Regulation of intracellular calcium overload, proapoptotic gene expression via Sigma-1 receptors, can result in neuroprotection during and after ischemia and acidosis. There would be further benefit through sigma-1 receptor dependent plasticity changes (review Frecska et al., 2013; Kourrich et al., 2012; Ruscher et al., 2011; Tsai et al., 2009). Along these lines Frecska colleagues (2013) suggest that DMT may be protective during cardiac arrest, beneficial during perinatal development, immunoregulation, and aid in reducing cancer progression as explained below.

3.2.7. Immediate early gene stimulation—Through second messenger systems, DMT can affect the rate of genetic transcription, such that DMT encodes the transcription factors c-fos (Frankel and Cunningham 2002), egr-1 and egr-2, which are associated with synaptic plasticity (O'Donovan et al., 1999; Gonzalez-Maeso et al., 2007). Increases in expression of brain-derived neurotrophic factor (BDNF) are also observed after DMT administration (Gewirtz et al., 2002). BDNF expression is associated with synaptic plasticity (O'Donovan et al., 1999), cognitive process such as memory (Jones et al., 2001) and attention (DeSteno and Schmauss, 2008), modulation of efficacy and plasticity of synapses (Soule et al., 2006).

3.2.8. Summary—As previously mentioned, DMT interacts with a variety of ionotropic and metabotropic receptors. The subjective effects of large doses of exogenous DMT are most likely mediated primarily by 5- HT_{2A} receptors, with 5- HT_{2C} receptors playing little or no role. mGlu2/3 receptors have significant modulatory effects, and the interaction of serotonergic and glutaminergic receptors may play a central role. DMT does not have direct effects on DA receptors, but indirectly alters the levels of dopamine, with resulting neurochemical and behavioral effects. Similarly, DMT also alters levels of acetylcholine. Finally, DMT may be an endogenous ligand at TAAR and sigma-1 receptors, but at the least, the effects of DMT at these receptors may play important physiological roles.

4. DMT as a model of psychiatric disorders

There has been a revival of interest in clinical uses of hallucinogens. Among the first were a series of controlled clinical studies on DMT (Strassman et al., 1994; 1996). Those studies reported that pure DMT had rapid and extremely strong cardiovascular effects as well as profound psychological effects. The cardiovascular effects preclude the use of pure DMT; however, ayahuasca and other DMT-containing ritual beverages seem to be less toxic while retaining the psychological effects. Based on studies of the health status of ayahuasca users, the use of ayahuasca may be safe and even beneficial (e.g., Barbosa et al., 2012; others from below).

Recently, a series of studies examined the long-term personal and spiritual significance of exposure to psilocybin (Griffiths et al., 2006; 2008; 2011), and others have suggested that psilocybin may be useful for anxiety-related disorders (e.g., Grob et al., 2011; Kometer et al., 2012). Similarly, ayahuasca and similar DMT-containing mixtures have been proposed as treatments for a variety of psychiatric disorders and ayahuasca is mostly well-tolerated. For example, long-term ayahuasca users showed less psychopathology, and better performance on neuropsychological tests compared to matched controls (Bouso et al., 2012) and less substance abuse and fewer psychiatric/psychosocial problems than matched controls (Fábregas et al., 2010). The following paragraphs will summarize evidence for medical applications of ayahuasca for treatment, and the following section will examine research on potential adverse effects of repeated use of ayahuasca.

4.1. Schizophrenia

The classic positive symptoms of schizophrenia include delusions and hallucinogens, so hallucinogenic compounds seem an obvious tool for modeling schizophrenia. Given that hallucinogens produce their effects primarily through activation of the 5-HT_{2A} receptor (review Nichols, 2004), the serotonin system provides an alternative to the dopamine model of schizophrenia. The dopamine model has produced a wide range of treatment medications which are very useful, but do not fully treat the range of symptoms experienced during psychotic episodes and produce substantial adverse effects. Discovery that DMT exists as an endogenous compound led to research focusing on DMT as a model of schizophrenia in the 1960s and 1970s. Reviews of this early research concluded that the data was suggestive but not conclusive (e.g., Gillin et al., 1976). These early studies are not reviewed in the present manuscript.

Subsequent research reported that levels of endogenous DMT increased in schizophrenic patients during psychotic episodes, which declined as their state improved (Checkley et al., 1980; Murrary et al., 1979). However, no changes in DMT levels were observed in rapidly cycling states (manic-depressive) (Checkley et al., 1980). These findings renewed interest in the transmethylation hypothesis, which states that schizophrenia may be due to stress-induced production of psychotomimetic methylated derivatives of catecholamines or indolealkylamines in the brain. DMT seems to fits the bill as it is an indolealkylamine, is an endogenous compound, and is linked to stress reactivity (see reviews by Myin-Germys and van Os, 2007; Grammenos and Barker, 2015).

In addition, DMT was identified as the active ingredient in ayahuasca (Pomilio et al., 1999; Ciprian-Ollivier et al., 1997), which produces effects similar to a psychotic episode, including thought disorders, delusions, and hallucinations (Gouzoulis-Mayfrank, et al., 2005). When given to human subjects, DMT produces complex visual and auditory hallucinations and increases cortisol levels (Strassman 1994; 1996), which supports its possible role as a possible mediator of schizophrenia.

More recent studies have examined the effects of DMT on various experimental models of changes in cognition in schizophrenic patients. Normal subjects are administered DMT and given various cognitive tasks to perform during fMRI scans. DMT slowed reaction time in tests of inhibition of return (Daumann et al., 2008; Gouzoulis-Mayfrank, et al., 2006), decreased alertness (Daumann et al., 2010), but produced less mismatch negativity than did the NMDA glutamate channel blocker ketamine (Heekeren et al., 2008), which commonly serves as a tool for investigating the glutaminergic hypothesis of schizophrenia.

In summary, DMT is still an interesting model of the serotonergic aspects of schizophrenia, but there is no conclusive evidence that endogenous DMT is a primary player. In fact, it has been argued that DMT is anti-anxiety/anti-psychotic via actions at the trace amino acid receptor (TAAR). Jacob and Presti (2005), and others have suggested that the effects of endogenous DMT are mediated via sigma receptor roles (see review by Grammenos and Barker, 2015 or refer to section in this review).

4.2. Depression

Few studies have investigated the effects of DMT-containing compounds on depression. One study investigated the effects of ayahuasca in the forced-swim test, a common animal model of depression. In female Wistar rats, ayahuasca increased swimming, which is considered a sign of potential antidepressant effects (Pic-Taylor et al., 2015). In a human experimental study, long-term ayahuasca users (>10 years) showed reduced ratings of hopelessness while under the influence (Santos et al., 2007). Finally, in an open-label clinical trial in in-patients suffering from depression, ayahuasca produced marked improvement in depressive symptoms with no mania or hypomania for up to up to 21 days after a single dose (Osório et al., 2015). Convergent evidence from three different experimental approaches (animal model, human experimental study, and a clinical trial) provides stronger evidence for potential antidepressant effects of DMT. However, replication of these findings will be necessary to confirm whether DMT-containing compounds will be useful for treatment of depression.

4.3. Anxiety/aggression

It has been proposed that DMT is an endogenous anxiolytic compound through its actions at the trace amino acid receptor (Jacob and Presti, 2005). To date, this hypothesis has generated little interest and DMT has been mostly investigated for its hallucinogenic effects. One early study did examine the effects of DMT in an animal model of anxiety/aggression in which pairs of rats receive shocks while in a test chamber. The shocks produce fighting and anti-anxiety compounds reduce the shock-induced fighting. LSD increased the amount of fighting, whereas DMT suppressed fighting (Walters et al., 1978). However, the effective

doses also produce sedation and reduced locomotor activity, which could also account for the effects.

In a case study of a homeless male with multiple convictions for manslaughter and diagnosed with antisocial disorder, ayahuasca sessions reportedly produced significant moral insights and allowed completion of a rehabilitation program in which the subject had been highly resistant (Frecska et al., 2008). No follow up was conducted, so no data is available on whether incidences of violent behavior decreased. In two larger scale studies, ayahuasca decreased ratings of anxiety in depressive-disorder patients (Osório et al., 2015) and reduced ratings of panic but not state- or trait-anxiety, in long-term users (Santos et al., 2007). Taken together, these findings do not provide support that DMT is useful for treatment of anxiety and/or aggression. It is possible that DMT may be useful in specific settings, similar to the successful use of psilocybin to treat anxiety in cancer patients (Grob et al., 2011; Kometer et al., 2012), but careful experimental research will be necessary before a strong conclusion can be make about DMT's efficacy as an anxiolytic medication.

5. Effects on other organ systems

5.1. Cardiovascular system

Single doses of DMT produced rapid onset of marked sympathomimetic effects including increased heart rate and blood pressure (Strassman et al., 1994). When a 5-HT1A antagonist, pindolol, was co-administered with DMT, the increase in heart rate was diminished whereas the increase in blood pressure was enhanced (Strassman, 1996). Tolerance to the effects of DMT was tested by administration of DMT to human volunteers four times at 30-min intervals. A progressive decrease in heart rate was observed over the four doses, but not in blood pressure (Strassman, et al., 1996). In contrast, two repeated doses of ayahuasca 4-h apart reduced systolic blood pressure and heart rate (Dos Santos et al., 2012). Long-term use of DMT-containing beverages may be of more concern as 14-day exposure to ayahuasca in rats altered the structure of the aorta, leading to a thickening of the walls of the aorta relative to the lumen diameter (Pitol et al., 2015).

5.1.1. Cardiac arrest—DMT has been speculated to aid in extending the survival of brain. A review by Frecska and colleagues (2013), suggests that during physical signals of agony, lungs synthesize large amount of DMT (primarily through the removal of INMT inhibitors) and can release DMT into arterial blood within seconds. Once in blood circulation DMT is safe from degradation as extracellular, circulating MAO enzymes deaminate only primary amines (McEwen and Sober, 1967). DMT is a tertiary amine, thus reaching the brain with minimal degradation. Through the use of active transport mechanisms already discussed for taking DMT from blood into the brain, could potentially keep brain alive longer without the brain having to produce DMT on its own. Exogenous DMT-like psychedelic effects are in essence similar to subjective reports provided after clinical death and near death experiences. Strassman (2001) believes DMT to be very likely involved in the dying process.

5.2. Endocrine system

DMT increased levels of corticotropin, cortisol, prolactin, and growth hormone when administered to human volunteers (Strassman et al., 1994). When DMT was given repeatedly to human volunteers (4 times at 30 min intervals), tolerance to the increases in various endocrine levels was observed, including corticotropin, prolactin and cortisol (Strassman, et al., 1996). Similarly, ayahuasca increased prolactin and cortisol levels in human volunteers (Dos Santos, et al., 2011; 2012), whereas repeated doses resulted in lower levels of GH secretion (Dos Santos et al., 2012).

5.3. Immune system and neurotoxicity

Ayahuasca has been reported to decrease the percentage of CD3 and CD4 lymphocytes, but to increase the number of natural killer cells (Dos Santos et al., 2011). It has been hypothesized that DMT might increase activity of the immune system and could prove useful as a treatment for cancer. Evidence for this hypothesis is equivocal. DMT increased the cytotoxic activity of peripheral blood mononuclear cells (e.g., lymphocytes and monocytes) in the A172 human glioma cell line (Tourino et al., 2013). However, in another study, DMT did not exhibit cytotoxicity of KB or HepG2 carcinoma cells (Gan et al., 2008). In addition, others have proposed that DMT and related compounds are anti-inflammatory and reported that DMT inhibited production of pro-inflammatory compounds IL-1 β , IL-6, IL-8 and TNFa and increased levels of the anti-inflammatory compound IL-10 through actions at the sigma-1 receptor (Szabo et al., 2014).

5.3.1. Immunoregulation—Serotonin plays an important role with immunoregulation (Ahern, 2011; Cloez-Tayarani and Changeux 2007). And on cellular immune functions critical in the elimination of pathogens or cancer cells (Leon-Ponte et al., 2007; O'Connell et al., 2006). It is possible that DMT may also play a role in immunoregulation via its Sigma-1 and 5-HT_{2A} receptor activation. Sigma receptors are also expressed on many cells of the immune system (Gekker et al., 2006). In particular, Dorocq (1995) showed that sigma-1 receptors can reduce pro-inflammatory cytokines and enhance the production of anti-inflammatory cytokine IL-10. DMT through the formulation of ayahuasca increased levels of blood circulating natural killer (NK) cells with concentrations as low as 1 mg DMT/kg body weight (Dos Santos et al., 2011). In vitro DMT administration has shown an increase of secreted interferons (beta and gamma) *in vitro* in NK cell and dendritic cell cultures. Interferons are potent anticancer factors (Caraglia et al., 2009; Gonzalez-Navajas et al., 2012; Szabo et al., 2012; Windbichler et al., 2000). If DMT does increase interferon secretion, it may be beneficial in contributing to or aid in better elimination of malignant and/or infected cells.

5.3.2. Perinatal INMT Activity—Levels of INMT in the placenta are higher than in adults. It is speculated that activity in fetal lungs compensates for difference. INMT activity in rabbit lung is relatively high in fetus, increases rapidly after birth and peaks at 15 days of age. It then declines to mature levels and remains constant through life (Lin et al., 1974). If INMT levels are paralleled with increased DMT synthesis, it could be possible that DMT-mediated sigma-1 receptor activity induces neuronal plasticity changes that can be expected for newborns. Selective sigma-1 receptor agonists have shown to be protective against

excitotoxic perinatal brain injury (Griesmaier et al., 2012) and ischemic neurodegeneration in neonatal striatum (Yang et al., 2012). Expression of INMT seems to be important for pregnancy success (Nuno-Ayala et al., 2012). Whether DMT, a product of INMT plays any role in these protective and beneficial effects, is unknown.

5.3.3. INMT and Cancer—Down regulation of the expression for the gene responsible for INMT production (*Inmt*) has been associated with cancer (Kopantzev et al., 2008, Larkin et al., 2012). It is believe to be a potential candidate gene in prevention of cancer progression. *Inmt* expression has been associated with a dramatic decrease in recurrence of malignant prostate (Larkin et al., 2012) and lung cancers (Kopantzev et al., 2008). It is possible that the regulating roles of *Inmt* via its product DMT could potentially have a direct tumor suppression effect, but this is role of DMT is unlikely, and highly speculative.

6. Summary and conclusions

DMT is a compound found widely across the plant and animal kingdoms. In mammals, the psychoactive effects produced by DMT seem to be largely mediated by the 5-HT2AR, although the complex subjective effects reported by DMT users are likely modulated by other receptor systems such as the metabotropic glutamate receptors.

The wide use of DMT in the form of ayahuasca for many years has led to a number of studies focusing on adverse health effects or potential benefits of ayahuasca use. There have been few reports of adverse health consequences (see review by Barbosa et al., 2012). Ayahuasca did produce modest impairment of cognitive function in inexperienced users; however, little or no impairment was observed in experienced users (Bouso et al., 2013). Ayahuasca decreased markers of sleep quality and sleep disturbances are common on the night following administration, but the users reported no perception of deterioration of quality (Barbanoj et al., 2008). As mentioned previously, there is little sign of tolerance or dependence to DMT except to the cardiovascular and endocrine effects, which actually could be viewed as the primary adverse effects. Diminution of these effects would preferred by long-term users. The greatest concern appears to the possibility of teratogenicity. Large doses of ayahuasca 50-fold higher than typical ritual doses (approx. 15 mg/kg DMT) were fed to pregnant rats. No lethality was observed, but increased incidence of cleft palate and skeletal malformations was observed in their pups (Gardner et al., 2014).

DMT may be an agent of significant adaptive mechanisms that can also serve as a promising tool in the development of future medical therapies (Frecska et al., 2013). There have been proposals that DMT might be a useful treatment of anxiety, substance abuse, inflammation, or for cancer. Experimental studies have been few and it is premature to conclude that DMT may have clinically relevant uses.

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References

- Aghajanian GK, Foote WE, Sheard MH. Action of psychotogenic drugs on single midbrain raphe neurons. J Pharmacol Exp Ther. 1970; 171:178–187. [PubMed: 5459056]
- Aghajanian GK, Marek GJ. Serotonin induces excitatory postsynaptic potentials in apical dendrites of neocortical pyramidal cells. Neuropharmacol. 1997; 36:589–599.
- Aghajanian GK, Marek GJ. Serotonin, via 5-HT2A receptors, increases EPSCs in layer V pyramidal cells of prefrontal cortex by an asynchronous mode of glutamate release. Brain Res. 1999a; 825:161–171. [PubMed: 10216183]
- Aghajanian GK, Marek GJ. Serotonin and hallucinogens. Neuropsychopharmacol. 1999b; 21:16–23.
- Agurell S, Holmstedt B, Lindgren JE, Schultes RE. Identification of two new p-carboline alkaloids in South American hallucinogenic plants. Biochem Pharmacol. 1968; 17:2487–2488. [PubMed: 5720352]
- Ahern GP. 5-HT and the immune system. Curr Opin Pharmacol. 2011; 11:29–33. [PubMed: 21393060]
- Alonso JF, Romero S, Mañanas MA, Riba J. Serotonergic psychedelics temporarily modify information transfer in humans. Int J Neuropsychopharmacol. 2015; 1-9
- Anden NE, Corrodi H, Fuxe K. Hallucinogenic drugs of the indolealkylamine type and central monoamine neurons. J Pharmacol Exp Ther. 1971; 179:236–249. [PubMed: 5133600]
- Appel JB, West WB, Rolandi WG, Alici T, Pechersky K. Increasing the selectivity of drug discrimination procedures. Pharmacol Biochem Behav. 1999; 64:353–358. [PubMed: 10515312]
- Araneda R, Andrade R. 5-hydroxytryptamine 2 and 5-hydroxy-tryptamine 1A receptors mediate opposing responses on membrane excitability in rat association cortex. Neurosci. 1991; 40:399–412.
- Barbanoj MJ, Riba J, Clos S, Giménez S, Grasa E. Daytime Ayahuasca administration modulates REM and slow-wave sleep in healthy volunteers. Psychopharmacology. 2008; 196(2):315–326. [PubMed: 18030450]
- Barbosa PC, Mizumoto S, Bogenschutz MP, Strassman RJ. Health status of ayahuasca users. Drug Test Anal. 2012; 4:601–609. [PubMed: 22761152]
- Barker SA, Beaton JM, Christian ST, Monti JA, Morris PE. Comparison of the brain levels of N, Ndimethyltryptamine and alpha, alpha, beta, beta-tetradeutero-N, N-dimethyltryptamine following intraperitoneal injection. The in vivo kinetic isotope effect. Biochem Pharmacol. 1982; 31:2513– 2516. [PubMed: 6812592]
- Barker SA, Borjigin J, Lomnicka I, Strassman R. LC/MS/MS analysis of the endogenous dimethyltryptamine hallucinogens, their precursors, and major metabolites in rat pineal gland microdialysate. Biomed Chromatogr. 2013; 27:1690–1700. [PubMed: 23881860]
- Barker SA, McIlhenny EH, Strassman R. A critical review of reports of endogenous psychedelic N, Ndimethyltryptamines in humans: 1955–2010. Drug Test Anal. 2012; 4:617–635. [PubMed: 22371425]
- Barker SA, Monti JA, Christian ST. Metabolism of the hallucinogen *N*,*N*-dimethyltryptamine in rat brain homogenates. Biochem Pharmacol. 1980; 29(7):1049–1057. [PubMed: 6770869]
- Barker SA, Monti JA, Christian ST. N, N-dimethyltryptamine: an endogenous hallucinogen. Int Rev Neurobiol. 1981; 22:83–110. [PubMed: 6792104]
- Beaton JM, Christian ST. Stress induced changes in whole brain indolealkylamine levels in the rat: using gas liquid chromatography-mass spectrometry. Abstr Soc Neurosci. 1977; 4:1322.
- Berge OG, Chacho D, Hole K. Inhibitory effect of 5-methoxy- N,N-dimethyltryptamine on the synaptosomal uptake of 5-hydroxytryptamine. Eur J Pharmacol. 1983; 90:293–296. [PubMed: 6873188]
- Bouso JC, Fábregas JM, Antonijoan RM, Rodríguez-Fornells A, Riba J. Acute effects of ayahuasca on neuropsycho- logical performance: differences in executive function between experienced and occasional users. Psychopharmacology (Berl). 2013; 230:415–424. [PubMed: 23793226]
- Bouso JC, Gonzalez D, Fondevila S, Cutchet M, Fernandez X, Ribeiro Barbosa PC, Alcazar-Corcoles MA, Araujo WS, Barbonoj MJ, Fabregas JM, Riba J. Personality, psychopathology, life attitudes

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and neuropsychological performance among ritual users of ayahuasca: a longitudinal study. PLoS One. 2012; 7:e42421. [PubMed: 22905130]

- Bouso JC, Palhano-Fontes F, Rodríguez-Fornells A, Ribeiro S, Sanches R, Crippa JAS, Hallak JEC, de Araujo DB, Riba J. Long-term use of psychedelic drugs is associated with differences in brain structure and personality in humans. Eur Neuropsychopharmacol. 2015; 15:483–492. [PubMed: 25637267]
- Brierley DI, Davidson C. Developments in harmine pharmacology—implications for ayahuasca use and drug-dependence treatment. Prog Neuropsychopharmacol Biol Psychiatry. 2012; 39(2):263– 72. [PubMed: 22691716]
- Brown K, Tracy D. Lithium: the pharmacodynamic actions of the amazing ion. Ther Adv Psychopharmacol. 2013; 3:163–176. [PubMed: 24167688]
- Bunzow JR, Sonders MS, Arttamangkul S, Harrison LM, Zhang G, Quigley DI, Darland T, Suchland KL, Pasumamula S, Kennedy JL, et al. Amphetamine, 3,4-methylenedioxymethamphetamine, lysergic acid diethylamide, and metabolites of the catecholamine neurotransmitters are agonists of a rat trace amine receptor. Mol Pharmacol. 2001; 60:1181–1188. [PubMed: 11723224]
- Burchett SA, Hicks TP. The mysterious trace amines: protean neuromodulators of synaptic transmission in mammalian brain. Prog Neurobiol. 2006; 79:223–246. [PubMed: 16962229]
- Cakic V, Potkonyak J, Marshall A. Dimethyltryptamine (DMT): subjective effects and patterns of use among Australian recreational users. Drug Alcohol Depend. 2010; 111:30–37. [PubMed: 20570058]
- Callaway JC, McKenna DJ, Grob CS, Brito GS, Raymon LP, Poland RE, Andrade EN, Andrade EO, Mash DC. Pharmacokinetics of hoasca alkaloids in healthy humans. J Ethnopharmacol. 1999; 65:243–256. [PubMed: 10404423]
- Callaway JC. A proposed mechanism for the visions of dream sleep. Med Hypotheses. 1988; 26:119–124. [PubMed: 3412201]
- Callaway JC, Raymon LP, Hearn WL, McKenna DJ, Grob CS, Brito GS, Mash DC. Quantitation of *N*,*N*-dimethyltryptamine and harmala alkaloids in human plasma after oral dosing with ayahuasca. J Anal Toxicol. 1996; 20(6):492–497. [PubMed: 8889686]
- Caraglia M, Marra M, Tagliaferri P, Lamberts SW, Zappavigna S, Misso G, Cavagnini F, Facchini G, Abbruzzese A, Hofland LJ, Vitale G. Emerging strategies to strengthen the anti-tumour activity of type I interferons: overcoming survival pathways. Curr Cancer Drug Targets. 2009; 9:690–704. [PubMed: 19508175]
- Carbonaro TM, Eshleman AJ, Forster MJ, Cheng K, Rice KC, Gatch MB. The role of 5-HT2A, 5-HT2C and mGluR2 receptors in the behavioral effects of tyrptamine hallucinogens *N*,*N*dimethyltryptamine and *N*,*N*-diisopropyltryptamine in rats and mice. Psychopharmacology. 2015; 232:275–284. [PubMed: 24985890]
- Carbonaro TM, Forster MJ, Gatch MB. Discriminative stimulus effects of *N*,*N*-diisopropyltryptamine. Psychopharmacology. 2013; 226(2):241–246. [PubMed: 23070023]
- Cartmell J, Monn JA, Schoepp DD. The metabotropic glutamate 2/3 receptor agonists LY354740 and Ly379268 selectively attenuate phencyclidine versus d-amphetamine motor behaviors in rats. J Pharmacol Exp Ther. 1999; 291:161–170. [PubMed: 10490900]
- Checkley SA, Murray RM, Oon MC, Rodnight R, Birley JL. A longitudinal study of urinary excretion of N,N-dimethyltryptamine in psychotic patients. Br J Psychiatry. 1980; 137:236–239. [PubMed: 6777009]
- Christian ST, Harrison R, Quayle E, Pagel J, Monti J. The in vitro identification of dimethyltryptamine (DMT) in mammalian brain and its characterization as a possible endogenous neuroregulatory agent. Biochem Med. 1977; 18:164–183. [PubMed: 20877]
- Ciprian-Ollivier J, Cetkovich-Bakmas MG. Altered consciousness states and endogenous psychoses: a common molecular pathway? Schizophr Res. 1997; 28:257–265. [PubMed: 9468359]
- Cloez-Tayarani I, Changeux JP. Nicotine and serotonin in immune regulation and inflammatory processes: a perspective. J Leukoc Biol. 2007; 81:599–606. [PubMed: 17108054]
- Cohen I, Vogel WH. Determination and physiological disposition of dimethyltryptamine and diethyltryptamine in rat brain, liver and plasma. Biochem Pharmacol. 1972; 21:1214–1216. [PubMed: 5034205]

- Cole JM, Pieper WA. The effects of N,N-dimethyltryptamine on operant behavior in squirrel monkeys. Psychopharmacology. 1973; 29:107–112.
- Collier TL, Waterhouse RN, Kassiou M. Imaging sigma receptors: Applications in drug development. Curr Pharm Des. 2007; 13:51–72. [PubMed: 17266588]
- Conn PJ, Sanders-Bush E. Selective 5HT-2 antagonists inhibit serotonin stimulated phosphatidylinositol metabolism in cerebral cortex. Neuropharmacol. 1984; 23:993–996.
- Cozzi NV, Gopalakrishnan A, Anderson LL, Feih JT, Shulgin AT, Daley PF, Ruoho AE. Dimethyltryptamine and other hallucinogenic tryptamines exhibit substrate behavior at the serotonin uptake transporter and the vesicle monoamine transporter. J Neural Transm. 2009; 116:1591–1599. [PubMed: 19756361]
- Cozzi NV, Mavlyutov TA, Thompson MA, Ruoho AE. Indolethylamine-N-methyltransferase expression in primate ner- vous tissue. Abstr Soc Neurosci. 2011; 37:840.9.
- Daumann J, Heekeren K, Neukirch A, Thiel CM, Moller-Hartmann W, Gouzoulis-Mayfrank E. Pharmacological modulation of the neural basis underlying inhibition of return (IOR) in the human 5HT2A agonist and NMDA antagonist model of psychosis. Psychopharmacology. 2008; 200:573– 583. [PubMed: 18649072]
- Daumann J, Wagner D, Heekeren K, Neukirch A, Thiel CM, Gouzoulis- Mayfrank E. Neuronal correlates of visual and auditory alertness in the DMT and ketamine model of psychosis. J Psychopharmacol. 2010; 24:1515–1524. [PubMed: 19304859]
- Delgado M, Caicoya AG, Greciano V, Benhamu B, Lopez-Rodriguez ML, Fernandez-Alfonso MS, Pozo MA, Manzanares J, Fuentes JA. Anxiolytic-like effect of a serotonergic ligand with high affinity for 5-HT1A, 5-HT2A and 5-HT3 receptors. Eur J Pharmacol. 2005; 511:9–19. [PubMed: 15777774]
- Deliganis AV, Pierce PA, Peroutka SJ. Differential interactions of dimethyltryptamine (DMT) with 5-HT1A and 5-HT2 receptors. Biochem Pharmacol. 1991; 41:1739–1744. [PubMed: 1828347]
- Delille HK, Becker JM, Burkhardt S, Bleher B, Terstappen GC, Schmidt M, Meyer AH, Unger L, Marek GJ, Mezler M. Heterocomplex formation of 5-HT2A-mGlu2 and its relevance for cellular signaling cascades. Neuropharmacol. 2012; 62:2184–2191.
- Der Marderosian AH, Pinkley HV, Dobbins MF IV. Native use and occurrence of N,Ndimethyltryptamine in the leaves of Banisteriopsis rusbyana. Am J Pharm. 1968; 140:137–147. [PubMed: 5698438]
- Derocq JM, Bourrie B, Segui M, Le Fur G, Casellas P. In vivo inhibition of endotoxin-induced proinflammatory cytokines production by the sigma ligand SR-31747. J Pharmacol Exp Ther. 1995; 272:224–230. [PubMed: 7815336]
- de Smet PAGM. A multidisciplinary overview of intoxicating enema rituals in the western hemisphere. J Ethnopharmacol. 1983; 9:129–66. [PubMed: 6677814]
- DeSteno DA, Schmauss C. Induction of early growth response gene 2 expression in the forebrain of mice performing an attention-set-shifting task. Neurosci. 2008; 152:417–428.
- Dos Santos RG, Grasa E, Valle M, Ballester MR, Bouso JC, Nomdedeu JF, Homs R, Barbanoj MJ, Riba J. Pharmacology of ayahuasca administered in two repeated doses. Psychopharmacology. 2012; 219(4):1039–1053. [PubMed: 21842159]
- Dos Santos RG, Valle M, Bouso JC, Nomdedeu JF, Rodriguez-Espinosa J, McIlhenny EH, Barker SA, Barbonoj MJ, Riba J. Autonomic, neuroendocrine, and immunological effects of ayahuasca: a comparative study with d-amphetamine. J Clin Psychopharmacol. 2011; 31(6):717–726. [PubMed: 22005052]
- Erspamer V. Observations on the fate of indolalkylamines in the organism. J Physiol. 1955; 127(1): 118–133. [PubMed: 14354632]
- Eshleman AJ, Forster MJ, Wolfrum KM, Johnson RA, Janowsky A, Gatch MB. Behavioral and neurochemical pharmacology of six psychoactive substituted phenethylamines: mouse locomotion, rat drug discrimination and in vitro receptor and transporter binding and function. Psychopharmacology. 2014; 231(5):875–88. [PubMed: 24142203]
- Fábregas JM, González D, Fondevila S, Cutchet M, Fernán- dez X, Barbosa PCR, Alcázar-Córcoles MÁ, Barbanoj MJ, Riba J, Bouso JC. Assessment of addiction severity among ritual users of ayahuasca. Drug Alcohol Depend. 2010; 111:257–261. [PubMed: 20554400]

- Fantegrossi WE, Harrington AW, Kiessel CL, Eckler JR, Rabin RA, Winter JC, Coop A, Rice KC, Woods JH. Hallucinogen-like actions of 5-methoxy-N, N-diisopropyltryptamine in mice and rats. Pharmacol Biochem Behav. 2006; 83:122–129. [PubMed: 16460788]
- Forsythe ID, Barnes-Davies M. Synaptic transmission: well-placed modulators. Curr Biol. 1997; 7:R362–R365. [PubMed: 9197230]
- Fish MS, Johnson NM, Horning EC. Piptadenia Alkaloids Indole bases of P. Peregrina (L.) Benth. and Related Species. J Am Chem Soc. 1955; 77:5892–5895.
- Fontanilla D, Johannessen M, Hajipour AR, Cozzi NV, Jackson MB, Ruoho AE. The hallucinogen N, N-dimethyltryptamine (DMT) is an endogenous sigma-1 receptor regulator. Science. 2009; 323:934–937. [PubMed: 19213917]
- Frankel PS, Cunningham KA. The hallucinogen d-lysergic acid diethylamide (d-LSD) induces the immediate-early gene c- Fos in rat forebrain. Brain Res. 2002; 958:251–260. [PubMed: 12470860]
- Frecska E. Ayahuasca versus violence-a case report. Neuropsychopharmacol Hung. 2008; 10(2):103–106. [PubMed: 18959142]
- Frecska E, Szabo A, Winkelman MJ, Luna LE, McKenna DJ. A possible sigma-1 receptor mediated role of dimethyltryptamine in tissue protection, regeneration, and immunity. J Neural Transm. 2013; 120:1295–1303. [PubMed: 23619992]
- Freedman DX, Gottleib R, Lovell RA. Psychotomimetic drugs and brain 5-hydroxytryptamine metabolism. Biochem Pharmacol. 1970; 19:1181–1188.
- Gan N, Yang X, Th L, He P. Studies on constituents of rootsanel leaves from Desmodium blandum and their cytotoxic activity against growth of several tumor cells. Zhongguo Zhong Yoa Za Zhi. 2008; 33(18):2077–2080.
- Gardner D, Riet-Correa F, Lemos D, Welch K, Pfister J, Panter K. Teratogenic effects of Mimosa tenuiflora in a rat model and possible role of N-methyl- and N,N-dimethyltryptamine. J Agric Food Chem. 2014; 62(30):7398–401. [PubMed: 24689494]
- Gatch MB, Rutledge MA, Carbonaro T, Forster MJ. Comparison of the discriminative stimulus effects of dimethyltryptamine with different classes of psychoactive compounds in rats. Psychopharmacology. 2009; 204:715–724. [PubMed: 19288085]
- Gatch MB, Forster MJ, Janowsky A, Eshleman AJ. Abuse liability profile of three substituted tryptamines. J Pharmacol Exp Ther. 2011; 338:280–289. [PubMed: 21474568]
- Gaujac A, Aquino A, Navickiene S, de Andrade JB. Determination of N,N-dimethyltryptamine in Mimosa tenuiflora inner barks by matrix solid-phase dispersion procedure and GC–MS. J Chromatogr B Analyt Technol Biomed Life Sci. 2012; 881–882:107–10.
- Gekker G, Hu S, Sheng WS, Rock RB, Lokensgard JR, Peterson PK. Cocaine-induced HIV-1 expression in microglia involves sigma-1 receptors and transforming growth factor-beta1. Int Immunopharmacol. 2006; 6:1029–1033. [PubMed: 16644490]
- Gewirtz JC, Chen AC, Terwilliger R, Duman RC, Marek GJ. Modulation of DOI-induced increases in cortical BDNF expression by group II mGlu receptors. Pharmacol Biochem Behav. 2002; 73:317– 326. [PubMed: 12117585]
- Gillin J, Cannon E, Magyar R, Schwartz M, Wyatt R. Failure of N,N-dimethyltryptamine to evoke tolerance in cats. Biol Psychiatry. 1973; 7(3):213–220. [PubMed: 4519415]
- Gillin JC, Kaplan J, Stillman R, Wyatt RJ. The psychedelic model of schizophrenia: the case of N,Ndimethyltryptamine. Am J Psychiatry. 1976a; 133:203–208. [PubMed: 1062171]
- Gillin JC, Wyatt RJ. Evidence for and against the involvement of N,N-dimethyl-tryptamine (DMT) and 5-methoxy-N,N-dimethyltryptamine (5-MeO-DMT) in schizophrenia. Psychopharmacol Bull. 1976b; 12(4):12–23. [PubMed: 1067623]
- Glennon RA. Discriminative stimulus properties of phenylisopropylamine derivatives. Drug Alcohol Depend. 1986; 17:119–134. [PubMed: 2874967]
- Glennon RA, Young R, Jacyno JM, Slusher M, Rosecrans JA. DOM-stimulus generalization to LSD and other hallucinogenic indolealkylamines. Eur J Pharmacol. 1983; 86:453–459. [PubMed: 6572591]
- Gomes MM, Coimbra JB, Clara RO, Dorr FA, Moreno ACR, Chagas JR, Tufik S, Pinto E Jr, Catalani LH, Campa A. Biosynthesis of N,N-dimethyltryptamine (DMT) in a melanoma cell line and its metabolization by peroxidases. Biochem Pharmacol. 2014; 88:393–401. [PubMed: 24508833]

- Gonzalez-Maeso J, Ang RL, Yuen T, Chan P, Weisstaub NV, Lopez-Gimenez JF, Zhou M, Okawa Y, Callado LF, Milligan G, Gingrich JA, Filizola M, Meana JJ, Sealfon SC. Identification of a serotonin/glutamate receptor complex implicated in psychosis. Nature. 2008; 452:93–97. [PubMed: 18297054]
- Gonzalez-Maeso J, Weisstaub NV, Zhou M, Chan P, Ivic L, Ang R, Lira A, Bradley-Moore M, Ge Y, Zhou Q, Sealfon SC, Gingrich JA. Hallucinogens recruit specific cortical 5-HT(2A) receptormediated signaling pathways to affect behavior. Neuron. 2007; 53:439–452. [PubMed: 17270739]
- Gonzalez-Navajas JM, Lee J, David M, Raz E. Immunomodulatory functions of type I interferons. Nat Rev Immunol. 2012; 12:125–135. [PubMed: 22222875]
- Gouzoulis-Mayfrank E, Heekeren K, Neukirch A, Stoll M, Stock C, Dauman J, Obradovic M, Kovar KA. Inhibition of return in the human 5HT2A agonist and NMDA antagonist model of psychosis. Neuropsychopharmacology. 2006; 31:431–441. [PubMed: 16123739]
- Gouzoulis-Mayfrank E, Heekeren K, Neukirch A, Stoll M, Stock C, Obradovic M, Kovar K. Psychological effects of (S)-ketamine and N,N-dimethyltryptamine (DMT): a double-blind, crossover study in healthy volunteers. Pharmacopsychiatry. 2005; 38(6):301–311. [PubMed: 16342002]
- Grammenos D, Barker SA. On the transmethylation hypothesis: stress, N,N-dimethyltryptamine, and positive symptoms of psychosis. J Neural Transm. 2015; 122:733–739. [PubMed: 25362533]
- Grant KA. Strategies for understanding the pharmacological effects of ethanol with drug discrimination procedures. Pharmacol Biochem Behav. 1999; 64:261–267. [PubMed: 10515301]
- Griesmaier E, Posod A, Gross M, Neubauer V, Wegleiter K, Hermann M, Urbanek M, Keller M, Kiechl-Kohlendorfer U. Neuroprotective effects of the sigma-1 receptor ligand PRE- 084 against excitotoxic perinatal brain injury in newborn mice. Exp Neurol. 2012; 237:388–395. [PubMed: 22771763]
- Griffiths RR, Johnson MW, Richards WA, Richards BD, McCann U, Jesse R. Psilocybin occasioned mystical-type experiences: immediate and persisting dose-related effects. Psychopharmacology. 2011; 218(4):649–665. [PubMed: 21674151]
- Griffiths RR, Richards WA, Johnson MW, McCann U, Jesse R. Mystical-type experiences occasioned by psilocybin mediate the attribution of personal meaning and spiritual significance 14 months later. J Psychopharmacol. 2008; 22(6):621–632. [PubMed: 18593735]
- Griffiths RR, Richards WA, McCann U, Jesse R. Psilocybin can occasion mystical-type experiences having substantial and sustained personal meaning and spiritual significance. Psychopharmacology. 2006; 187(3):268–283. [PubMed: 16826400]
- Grob CS, Danforth AL, Chopra GS, Hagerty M, McKay CR, Halberstadt AL, Greer GR. Pilot study of psilocybin treatment for anxiety in patients with advanced-stage cancer. Arch Gen Psychiatry. 2011; 68(1):71–78. [PubMed: 20819978]
- Halberstadt AL. Recent advances in the neuropsychopharmacology of serotonergic hallucinogens. Behav Brain Res. 2015; 277:99–120. [PubMed: 25036425]
- Hamik A, Peroutka SJ. 1-(m-chlorophenyl) piperazine (mCPP) interactions with neurotransmitter receptors in the human brain. Biol Psychiatry. 1989; 25:569–575. [PubMed: 2537663]
- Haubrich D, Wang P. N,N-dimethyltryptamine lowers rat brain acetylcholine and dopamine. Brain Res. 1977; 131(1):158–161. [PubMed: 267501]
- Hayashi T, Su TP. Sigma-1 receptor chaperones at the ER- mitochondrion interface regulate Ca signaling and cell survival. Cell. 2007; 131:596–610. [PubMed: 17981125]
- Heekeren K, Daumann J, Neukirch A, Stock C, Kawohl W, Norra C, Waberski TD, Gouzoulis-Mayfrank E. Mismatch negativity generation in the human 5HT2A agonist and NMDA antagonist model of psychosis. Psychopharmacology. 2008; 199:77–88. [PubMed: 18488201]
- Helsley S, Fiorella D, Rabin RA, Winter JC. A comparison of N,N-dimethylamine, harmaline, and selected congeners in rats trained with LSD as a discriminative stimulus. Prog Neuropsychopharmacol Biol Psychiatry. 1998; 22:649–663. [PubMed: 9682278]
- Heuring RE, Peroutka SJ. Characterization of a novel 3H-5-hydroxytryptamine binding site subtype in bovine brain membranes. J Neurosci. 1987; 7:894–903. [PubMed: 2951504]
- Hill SL, Doris T, Gurung S, Katebe S, Lomas A, Dunn M, Blain P, Thomas SH. Severe clinical toxicity associated with analytically confirmed recreational use of 25I-NBOMe: case series. Clin Toxicol (Phila). 2013; 51:487–92. [PubMed: 23731373]

- Hollister, LE. Some general thoughts about endogenous psychotogens. In: Usdin, E.; Hamburg, DA.; Barchas, JD., editors. Neuroregulators and psychiatric disorders. Oxford University Press; New York: 1977. p. 550-556.
- Jacob MS, Presti DE. Endogenous psychoactive tryptamines reconsidered: an anxiolytic role for dimethyltryptamine. Med Hypotheses. 2005; 64(5):930–937. [PubMed: 15780487]
- Jarbe TUC. LSD-25 as a discriminative stimulus for response selection by pigeons. Pharmacol Biochem Behav. 1980; 13:549–554. [PubMed: 6107936]
- Jenner P, Marsden CD, Thanki CM. Behavioural changes induced by N, N-dimethyltryptamine in rodents. Br J Pharmacol. 1980; 69:69–80. [PubMed: 6769527]
- Johannessen M, Fontanilla D, Mavlyutov T, Ruoho AE, Jackson MB. Antagonist action of progesterone at σ-receptors in the modulation of voltage-gated sodium channels. Am J Physiol Cell Physiol. 2013; 305(9):C997.
- Jones MW, Errington ML, French PJ, Fine A, Bliss TV, Garel S, Charnay P, Bozon B, Laroche S, Davis S. A requirement for the immediate early gene Zif268 in the expression of late LTP and long-term memories. Nat Neurosci. 2001; 4:289–296. [PubMed: 11224546]
- Kaplan J, Mandel LR, Stillman R, Walker RW, Heuval WJA, Gillin JC, Wyatt RJ. Blood and urine levels of N,N-dimethyltryptamine following administration of psychoactive dosages to human subjects. Psychopharmacology. 1974; 38:239–245.
- Karkkainen J, Forsstrom T, Tornaeus J, Wahala K, Kiuru P, Honkanen A, Stenman UH, Turpeinen U, Hesso A. Potentially hallucinogenic 5-hydroxytryptamine receptor ligands bufotenine and dimethyltryptamine in blood and tissues. Scand J Clin Lab Invest. 2005; 65:189–199. [PubMed: 16095048]
- Keiser MJ, Setola V, Irwin JJ, Laggner C, Abbas AI, Hufeisen SJ, Jensen NH, Kuijer MB, Matos RC, Tran TB, et al. Predicting new molecular targets for known drugs. Nature. 2009; 462:175–81. [PubMed: 19881490]
- Kometer M, Schmidt A, Bachmann R, Studerus E, Seifritz E, Vollenweider FX. Psilocybin biases facial recognition, goal-directed behavior, and mood state toward positive relative to negative emotions through different serotonergic subreceptors. Biol Psychiatry. 2012; 72(11):898–906. [PubMed: 22578254]
- Kopantzev EP, Monastyrskaya GS, Vinogradova TV, Zinovyeva MV, Kostina MB, Filyukova OB, Tonevitsky AG, Sukhikh GT, Sverdlov ED. Differences in gene expression levels between early and later stages of human lung development are opposite to those between normal lung tissue and non-small lung cell carcinoma. Lung Cancer. 2008; 62:23–34. [PubMed: 18394749]
- Kovacic B, Domino EF. Tolerance and limited cross-tolerance to the effects of N,Ndimethyltryptamine (DMT) and lysergic acid diethylamide-25 (LSD) on food-rewarded bar pressing in the rat. J Pharmacol Exp Ther. 1976; 197:495–501. [PubMed: 1064726]
- Kurrasch-Orbaugh DM, Watts VJ, Barker EL, Nichols DE. Serotonin 5-hydroxytryptamine2A receptor-coupled phospholipase C and phospholipase A2 signaling pathways have different receptor reserves. J Pharmacol Exp Ther. 2003; 304:229–237. [PubMed: 12490596]
- Kourrich S, Su TP, Fujimoto M, Bonci A. The sigma-1 receptor: roles in neuronal plasticity and disease. Trends Neurosci. 2012; 35:762–771. [PubMed: 23102998]
- Lanaro R, Calemi DB, Togni LR, Costa JL, Yonamine M, Cazenave Sde O, Linardi A. Ritualistic Use of Ayahuasca versus Street Use of Similar Substances Seized by the Police: A Key Factor Involved in the Potential for Intoxications and Overdose? J Psychoactive Drugs. 2015; 47(2): 132–9. [PubMed: 25950593]
- Larkin SE, Holmes S, Cree IA, Walker T, Basketter V, Bickers B, Harris S, Garbis SD, Townsend PA, Aukim-Hastie C. Identification of markers of prostate cancer progression using candidate gene expression. Br J Cancer. 2012; 106:157–165. [PubMed: 22075945]
- Leon-Ponte M, Ahern GP, O'Connell PJ. Serotonin provides an accessory signal to enhance T-cell activation by signaling through the 5-HT7 receptor. Blood. 2007; 109:3139–3146. [PubMed: 17158224]
- Lin RL, Sargeant S, Narasimhachari N. Indolethylamine-N-methyltransferase in developing rabbit lung. Dev Psychobiol. 1974; 7:475–481. [PubMed: 4426474]

- Lu LJ, Wilson AE, Moore RH, Domino EF. Correlation between brain, N,N-dimethyltryptamine (DMT) levels and bar pressing behaviour in rats: effect of MAO inhibition. Pharmacologist. 1974; 16:237.
- Lowe LM, Peterson BL, Couper FJ. A case review of the first analytically confirmed 25I-NBOMerelated seath in Washington State. J Anal Toxicol. 2015; 39:668–671. [PubMed: 26378143]
- Lyon RA, Titeler M, Seggel MR, Glennon RA. Indolealkylamine analogs share 5-HT2 binding characteristics with phenylalkylamine hallucinogens. Eur J Pharmacol. 1988; 145:291–297. [PubMed: 3350047]
- Maitre, L.; Delini-Stula, A.; Waldmeier, PC. Ciba Found Syrup on "Monoamine oxidase and its inhibition". Amsterdam-Oxford-New York: Elsevier; 1976. Relations between the degree of monoamine oxidase inhibition and some psychopharmacological responses to monoamine oxidase inhibitors in rats; p. 247-267.
- Mandell AJ, Morgan M. Indole(ethyl)amine N-methyltransferase in human brain. Nat New Biol. 1971; 230:85–87. [PubMed: 5279043]
- Marzullo G, Rosengarten H, Friedhoff AJ. A peptide-like inhibitor of N-methyltransferase in rabbit brain. Life Sci. 1977; 20:775–783. [PubMed: 15714759]
- Mavlyutov TA, Epstein ML, Liu P, Verbny YI, Ziskind-Conhaim L, Ruoho AE. Development of the sigma-1 receptor in C-terminals of motoneurons and colocalization with the N, N0dimethyltryptamine forming enzyme, indole-N-methyl transferase. Neuroscience. 2012; 206:60– 68. [PubMed: 22265729]
- McEwen CM Jr, Sober AJ. Rabbit serum monoamine oxidase. II. Determinants of substrate specificity. J Biol Chem. 1967; 242:3068–3078. [PubMed: 6027789]
- McIlhenny EH, Riba J, Barbanoj MJ, Strassman R, Barker SA. Methodology for and the determination of the major constituents and metabolites of the Amazonian botanical medicine aya- huasca in human urine. Biomed Chromatogr. 2011; 25(9):970–984. [PubMed: 21058415]
- McKenna DJ, Repke DB, Lo L, Peroutka SJ. Differential interactions of indolealkylamines with 5hydroxytryptamine receptor subtypes. Neuropharmacology. 1990; 29(3):193–198. [PubMed: 2139186]
- McKenna DJ, Towers GH, Abbott F. Monoamine oxidase inhibitors in South American hallucinogenic plants: Tryptamine and beta-carboline constituents of ayahuasca. J Ethnopharmacol. 1984; 10:195–223. [PubMed: 6587171]
- Moore RH, Demetriou SK, Domino EF. Effects of iproniazid, chlorpromazine and methiothepin on DMT-induced changes in body temperature, pupillary dilatation, blood pressure and EEG in the rabbit. Arch Int Pharmacodyn. 1975; 213:64–72. [PubMed: 1057382]
- Moreno JL, Holloway T, Albizu L, Sealfon SC, González-Maeso J. Metabotropic glutamate mGlu2 receptor is necessary for the pharmacological and behavioral effects induced by hallucinogenic 5-HT2A receptor agonists. Neurosci Lett. 2011; 493:76–79. [PubMed: 21276828]
- Morgan M, MandellL J. Indole(ethyl)amine N-methyltransferase in the brain. Science. 1969; 165:492– 493. [PubMed: 5793241]
- Murray RM, Oon MC, Rodnight R, Birley JL, Smith A. Increased excretion of dimethyltryptamine and certain features of psychosis: a possible association. Arch Gen Psychiatry. 1979; 36(6):644–9. [PubMed: 286576]
- Myin-Germeys I, van Os J. Stress-reactivity in psychosis: evidence for an affective pathway to psychosis. Clin Psychol Rev. 2007; 27(4):409–424. [PubMed: 17222489]
- Nagai F, Nonaka R, Satoh K, Kamimura H. The effects of non- medically used psychoactive drugs on monoamine neurotransmission in rat brain. Eur J Pharmacol. 2007; 559:132–137. [PubMed: 17223101]
- Nichols DE. Hallucinogens. Pharmacol Ther. 2004; 101:131-181. [PubMed: 14761703]
- Nuno-Ayala M, Guillen N, Arnal C, Lou-Bonafonte JM, de Martino A, Garcia-de-Jalon JA, Gascon S, Osaba L, Osada J, Navarro MA. Cystathionine b-synthase deficiency causes infertil- ity by impairing decidualization and gene expression networks in uterus implantation sites. Physiol Genomics. 2012; 44:702–716. [PubMed: 22617046]

- O'Connell PJ, Wang X, Leon-Ponte M, Griffiths C, Pingle SC, Ahern GP. A novel form of immune signaling revealed by transmission of the inflammatory mediator serotonin between dendritic cells and T cells. Blood. 2006; 107:1010–1017. [PubMed: 16223770]
- O'Donovan KJ, Tourtellotte WG, Millbrandt J, Baraban JM. The EGR family of transcriptionregulatory factors: progress at the interface of molecular and systems neuroscience. TrendsNeurosci. 1999; 22:167–173.
- Ohishi H, Ogawa-Meguro R, Shigemoto R, Kaneko T, Nakanishi S, Mizuno N. Immunohistochemical localization of metabotro- pic glutamate receptors, mGluR2 and mGluR3, in rat cerebellar cortex. Neuron. 1994; 13:55–66. [PubMed: 8043281]
- Oon MC, Murray RM, Rodnight R, Murphy MP, Birley JL. Factors affecting the urinary excretion of endogenous formed dimethyltryptamine in normal humans subjects. Psychopharmacology. 1977; 54:171–175. [PubMed: 22091]
- Osmond H, Smythies J. Schizophrenia: a new approach. J Ment Sci. 1952; 98:309–315. [PubMed: 14917992]
- Osório FL, Sanches RF, Macedo LR, Santos RG, Maia-de-Oliveira JP, Wichert-Ana L, de Araujo DB, Riba J, Crippa JA, Hallak JE. Antidepressant effects of a single dose of ayahuasca in patients with recurrent depression: a preliminary report. Rev Bras Psiquiatr. 2015; 37:13–20. [PubMed: 25806551]
- Ott J. Pharmahuasca: human pharmacology of oral DMT plus harmine. J Psychoactive Drugs. 1999; 31(2):171–7. [PubMed: 10438001]
- Pehek E, Nocjar C, Roth B, Byrd T, Mabrouk O. Evidence for the preferential involvement of 5-HT2A serotonin receptors in stress- and drug-induced dopamine release in the rat medial prefrontal cortex. Neuropsychopharmacology. 2006; 31:265–277. [PubMed: 15999145]
- Pic-Taylor A, Gueiros da Mottab L, Alves de Moraisa J, Juniora JM, de Fátima W, Santosb A, Camposc LA, Mortaric Marcia Renata, von Zubend Marcus Vinicius, Caldasb Eloisa Dutra. Behavioural and neurotoxic effects of ayahuasca infusion (Banisteriopsis caapi and Psychotria viridis) in female Wistar rat. Behav Proc. 2015; 118:102–110.
- Pierce PA, Peroutak SJ. Hallucinogenic drug interaction with neurotransmitter receptor binding sites in human cortex. Psychopharmacol. 1989; 97:188–122.
- Pieri L, Pieri M, Haefely W. LSD as an agonist of dopamine receptors in the striatum. Nature. 1974; 252:586–588. [PubMed: 4431521]
- Pitol DL, Siéssere S, dos Santos RG, Rosa MLNM, Hallak JEC, Scalize PH, Pereira BF, Iyomasa MM, Semprini M, Riba J, Regalo SCH. Ayahuasca Alters Structural Parameters of the Rat Aorta. J Cardiovasc Pharmacol. 2015; 66:58–62. [PubMed: 25714595]
- Pochettino ML, Cortella AR, Ruiz M. Hallucinogenic snuff from northwestern Argentina: microscopical identification of Anadenanthera colubrina var Cebil (fabaceae) in powdered archaeological material. Econ Bot. 1999; 53:127–132.
- Pomilio A, Vitale A, Ciprian-Ollivier J, Cetkovich-Bakmas M, Gomez R, Vazquez G. Ayahoasca: an experimental psychosis that mirrors the transmethylation hypothesis of schizophrenia. J Ethnopharmacol. 1999; 65(1):29–51. [PubMed: 10350367]
- Randic M, Padjen A. Effect of N,N-dimethyltryptamine and D- lysergic acid diethylamide on the release of 5-hydroxyindoles in rat forebrain. Nature. 1971; 230:532–533. [PubMed: 4927758]
- Rech, R.; Inore, K. An Introduction to Psychopharmacology. Raven Press; New York: 1971. p. 100
- Reimann W, Schneider F. The serotonin receptor agonist 5-methoxy-N,N-dimethyltryptamine facilitates noradrenaline release from rat spinal cord slices and inhibits monoamine oxidase activity. Gen Pharmacol. 1993; 24:449–453. [PubMed: 8482527]
- Riba J, McIlhenny EH, Calle M, Bouso JC, Barker SA. Metabolism and disposition of N,Ndimethyltryptamine and harmala alkaloids after oral administration of ayahuasca. Drug Tes Anal. 2012; 4:610.
- Riba J, Valle M, Urbano G, Yritia M, Morte A, Barbanoj MJ. Human pharmacology of ayahuasca: Subjective and cardiovascular effects, monoamine metabolite secretion, and pharmacokinetics. J Pharmacol Exp Ther. 2003; 306:73–83. [PubMed: 12660312]

- Riceberg LJ, Vunakis HV. Determination of N,N-dimethylindolealkylamines in plasma, blood and urine extracts by radioimmunoassay and high pressure liquid chromatography. J Pharmacol Exp Ther. 1978; 206:158–166. [PubMed: 275476]
- Rosenberg DE, Isbell H, Miner EJ, Logan CR. The effect of N,N-dimethyltryptamine in human subjects tolerant to lysergic acid diethylamide. Psychopharmacology. 1964; 5:217–227.
- Roth BL, Palvimaki EP, Berry S, Khan N, Sachs N, Uluer A, Choudhary MS. 5-Hydroxytryptamine2A (5-HT2A) receptor desensitization can occur without down-regulation. J Pharmacol Exp Ther. 1995; 275:1638–1646. [PubMed: 8531139]
- Ruscher K, Shamloo M, Rickhag M, Ladunga I, Soriano L, Gisselsson L, Toresson H, Ruslim-Litrus L, Oksenberg D, Urfer R, Johansson BB, Nikolich K, Wieloch T. The sigma-1 receptor enhances brain plasticity and functional recovery after experimental stroke. Brain. 2011; 134:732–746. [PubMed: 21278085]
- Saavedra JM, Axelrod J. Psychotomimetic N-methylated tryptamines: formation in brain in vivo and in vitro. Science. 1972; 175(4028):1365–1366. [PubMed: 5059565]
- Saavedra JM, Coyle JT, Axelrod J. The distribution and properties of the nonspecific Nmethyltransferase in brain. J Neurochem. 1973; 20:743–752. [PubMed: 4703789]
- Sangiah S, Gomez MV, Domino EF. Accumulation of N,N-dimethyltryptamine in rat brain cortical slices. Biol Psychiatry. 1979; 14:925–936. [PubMed: 41604]
- Santos RG, Landeira-Fernandez J, Strassman RJ, Motta V, Cruz AP. Effects of ayahuasca on psychometric measures of anxiety, panic-like and hopelessness in Santo Daime members. J Ethnopharmacol. 2007; 112:507–513. [PubMed: 17532158]
- Schultes RE. Ethnotoxicological significance of additives to New World hallucinogens. Plant Sci Bull. 1972; 18:34–41.
- Servillo L, Giovane A, Balestrieri ML, Cautela D, Castaldo D. N-methylated tryptamine derivatives in Citrus genus plants: Identification of N,N,N-trimethyltryptamine in bergamot. J Agric Food Chem. 2012; 60:9512–9518. [PubMed: 22957740]
- Shigemoto R, Kinoshita A, Wada E, Nomura S, Ohishi H, Takada M, Flor PJ, Neki A, Abe T, Nakanishi S, Mizuno N. Differential presynaptic localization of metabotropic glutamate receptor subtypes in the rat hippocampus. J Neurosci. 1997; 17:7503–7522. [PubMed: 9295396]
- Shulgin, AT. Psychotomimetic agents. In: Gordon, Maxwell, editor. Psychopharmacological Agents, Vol IV. Vol. Chap. 4. Academic Press; New York, London: 1976.
- Shulgin, A.; Shulgin, A. Tihkal: the continuation. Joy, D., editor. US: Transform Press; 1997.
- Sitaram BR, Lockett L, Talomsin R, Blackman GL, McLeod WR. In vivo metabolism of 5-methoxy-N, N-dimethyltryptamine and N, N-dimethyltryptamine in the rat. Biochem Pharmacol. 1987; 36:1509–1512. [PubMed: 3472526]
- Soulé J, Messaoudi E, Bramham CR. Brain-derived neurotrophic factor and control of synaptic consolidation in the adultbrain. Biochem Soc Trans. 2006; 34:600–604. [PubMed: 16856871]
- Smith RL, Canton H, Barrett RJ, Sanders-Bush E. Agonist properties of N, N-dimethyltryptamine at serotonin 5–HT2A and 5–HT2C receptors. Pharmacol Biochem Behav. 1998; 61:323–330. [PubMed: 9768567]
- Smith TE, Weissbach H, Udenfriend S. Studies on the mechanism of action of monoamine oxidase: metabolism of N,N-dimethyltryptamine and N,N-dimethyltryptamine-N-oxide. Biochemistry. 1962; 1:137–143. [PubMed: 13914428]
- Smith TL. Some neurochemical effects of N,N-dimethyltryptamine and their possible relation to acute schizophrenia. Thesis, Reno, Nev USA 1975; Dissertation Abst Intern. 1975; B 35:4374 B–4375 B.
- Smith TL. Increased synthesis of striatal DA by N,N-DMT. Life Sci. 1977; 21:1597–1602. [PubMed: 271755]
- Sprouse JS, Aghajanian GK. Electrophysiological responses of serotoninergic dorsal raphe neurons to 5-HT1A and 5-HT1B agonists. Synapse. 1987; 1(1):3–9. [PubMed: 3505364]
- Sprouse JS, Aghajanian GK. Responses of hippocampal pyramidal cells to putative serotonin 5-HT1A and 5-HT1B agonists: a comparative study with dorsal raphe neurons. Neuropharmacology. 1988; 27(7):707–715. [PubMed: 2901680]

- Stahl SM. The sigma enigma: can sigma receptors provide a novel target for disorders of mood and cognition? J Clin Psychiatry. 2008; 69:1673–1674. [PubMed: 19200426]
- Stern WC, Dalsass M. DMT-induced turning in unilateral nigral lesioned rats: evidence for central dopaminergic activation. Abstr Soc Neurosci Sixth Ann Meeting. 1976; 2:880.
- Stoff DM, Moja EA, Gillin JC, Wyatt RJ. Dose response and time course effects of N, Ndimethyltryptamine on disruption of rat shuttlebox avoidance. Biol Psychiatry. 1977; 12:339– 346. [PubMed: 266950]
- Strassman RJ. Human psychopharmacology of N,N-dimethyltryptamine. Behavioural Brain Research. 1996; 73:121–124. [PubMed: 8788488]
- Strassman, RJ. A doctor's revolutionary research into the biology of near-death and mystical experiences. Park Street Press; Rochester: 2001. DMT: the spirit molecule.
- Strassman RJ, Qualls CR. Dose-response study of N,N-dimethyltryptamine in humans: I. Neuroendocrine, autonomic, and cardiovascular effects. Arch Gen Psychiatry. 1994; 51:85–97. [PubMed: 8297216]
- Strassman RJ, Qualls CR, Berg LM. Differential tolerance to biological and subjective effects of four closely spaced doses of N,N-dimethyltryptamine in humans. Biol Psychiatry. 1996; 39(9):784– 795. [PubMed: 8731519]
- Strassman RJ, Qualls CR, Uhlenhuth EH, Kellner R. Dose-response study of N,N-dimethyltryptamine in humans. I. Neuroendocrine, autonomic, and cardiovascular effects. Arch Gen Psychiatry. 1994; 51(2):85–97. [PubMed: 8297216]
- Strassman RJ, Qualls CR, Uhlenhuth EH, Kellner R. Dose-response study of N,N-dimethyltryptamine in humans. II. Subjective effects and preliminary results of a new rating scale. Arch Gen Psychiatry. 1994; 51(2):98–108. [PubMed: 8297217]
- Su TP, Hayashi T, Vaupel DB. When the endogenous hallucinogenic trace amine N,Ndimethyltryptamine meets the sigma-1 receptor. Sci Signal. 2009; 2:pe12. [PubMed: 19278957]
- Szabo A, Osman RM, Bacskai I, Kumar BV, Agod Z, Lanyi A, Gogolak P, Rajnavolgyi E. Temporally designed treatment of melanoma cells by ATRA and polyI: C results in enhanced chemokine and IFNb secretion controlled differently by TLR3 and MDA5. Melanoma Res. 2012; 22:351–361. [PubMed: 22797253]
- Szabo A, Rajnavolgyi E. Finding a fairy in the forest: ELF4, a novel and critical element of type I interferon responses. Cell Mol Immunol. 2014; 11(3):218–220. [PubMed: 24658434]
- Szára S. Dimethyltryptamine: its metabolism in man; the relation to its psychotic effect to the serotonin metabolism. Experientia. 1956; 12:441–442. [PubMed: 13384414]
- Szára S. Hallucinogenic effects and metabolism of tryptamine derivatives in man. Fed Proc. 1961; 20:885–888.
- Szára S. DMT at fifty. Neuropsychopharmacologia Hungarica. 2007; IX/4:201–205. [PubMed: 18510265]
- Takahashi T, Takahashi K, Ido T, Yanai K, Iwata R, Ishiwata K, Nozoe S. 11C-labeling of indolealkylamine alkaloids and the comparative study of their tissue distributions. Int J Appl Radiat Isot. 1985; 36:965–969. [PubMed: 3866749]
- Thompson MA, Moon E, Kim UJ, Xu J, Siciliano MJ, Weinshilboum RM. Human indolethylamine Nmethyltransferase: cDNA cloning and expression, gene cloning, and chromosomal localization. Genomics. 1999; 61:285–297. [PubMed: 10552930]
- Thompson MA, Weinshilboum RM. Rabbit lung indolethylamine N-methyltransferase: cDNA and gene cloning and characterization. J Bio Chem. 1998; 273:34502–34510. [PubMed: 9852119]
- Tithaphandha A. Substrate specificity and heterogeneity of N-methyltransferases. Biochem Biophys Res Commun. 1972; 47(2):301–308. [PubMed: 4662391]
- Tourino MC, de Oliveira EM, Belle LP, Knebel FH, Albuquerque RC, Dorr FA, Okada SS, Migliorini S, Soares IS, Campa A. Tryptamine and dimethyltryptamine inhibit indoleamine 2,3 dioxygenase and increase the tumor-reactive effect of peripheral blood mononuclear cells. Cell Biochem Funct. 2013; 31(5):361–4. [PubMed: 23754498]
- Trulson ME, Ross CA, Jacobs BL. Lack of tolerance to the depression of raphe unit activity by lysergic acid diethylamide. Neuropharmacology. 1977; 16:771–774. [PubMed: 593537]

- Tsai SY, Hayashi T, Harvey BK, Wang Y, Wu WW, Shen RF, Zhang Y, Becker KG, Hoffer BJ, Su TP. Sigma-1 receptors regulate hippocampal dendritic spine formation via a free radical-sensitive mechanism involving Rac1×GTP pathway. Proc Natl Acad Sci. 2009; 106:22468–72243. [PubMed: 20018732]
- Tupper KW. The globalization of ayahuasca: harm reduction or benefit maximization? Int J Drug Policy. 2008; 19(4):297–303. [PubMed: 18638702]
- Turner, DM. The essential psychedelic handbook. Panther Press; San Francisco: 1994.
- Turner WJ, Merlis S. Effects of some indolealkylamines in man. AMA Arch Neurol Psychiat. 1959; 81:121–129. [PubMed: 13605329]
- Vitale AA, Pomilio AB, Canellas CO, Vitale MG, Putz EM, Ciprian-Ollivier J. In vivo long-term kinetics of radiolabeled N, N-dimethyltryptamine and tryptamine. J Nucl Med. 2011; 52:970– 977. [PubMed: 21622895]
- von Hungen K, Roberts S, Hill DF. Interactions between lysergic acid diethylamide and dopaminesensitive adenylate cyclase systems in rat brain. Brain Res. 1975; 94:57–66. [PubMed: 238721]
- Waldmeier PC, Delini-Stula A, Maitre L. Preferential deamination of dopamine by an A-type monoamine oxidase in rat brain. Arch Pharmacol. 1976; 292:9–14.
- Waldmeier PC, Maitre L. Comparison of short and longlasting effects of pargyline on cerebral dopamine metabolism. Arch Pharmacol. 1976; 294:133–140.
- Waldmeier PC, Maitre L. Neurochemical investigations of the interactions of N,N-DMT with the dopaminergic system in rat brain. Psychopharmacology. 1977; 52:137–144. [PubMed: 407597]
- Walker RW, Mandel LR, Kleinman JE, Gillin JC, Wyatt RJ, Vandenheuvel WJ. Improved selective ion monitoring mass-spectrometric assay for deterruination of N,N-dimethyltryptamine in human blood utilizing capillary column gas chromatography. J Chromatogr Biomed Appl. 1979; 162:539–546.
- Wallach JV. Endogenous hallucinogens as ligands of the trace amine receptors: a possible role in sensory perception. Med Hypotheses. 2009; 72:91–94. [PubMed: 18805646]
- Walters JK, Sheard MH, Davis M. Effects of N,N-dimethyltryptamine (DMT) and 5-methoxy-N,Ndimethyltryptamine (5-MeODMT) on shock elicited fighting in rats. Pharmacol Biochem Behav. 1978; 9(1):87–90. [PubMed: 279938]
- Warren JM, Dham-Nayyar P, Alexander J. Recreational use of naturally occurring dimethyltryptamine contributing to psychosis? Aust N Z J Psychiatry. 2013; 47(4):398–399. [PubMed: 23047957]
- Weissman AD, Su TP, Hedreen JC, London ED. Sigma receptors in post-mortem human brains. J Pharmacol Exp Ther. 1988; 247:29–33. [PubMed: 2845055]
- West WB, Lou A, Pechersky K, Chachich ME, Appel JB. Antagonism of a PCP drug discrimination by hallucinogens and related drugs. Neuropsychopharmacology. 2000; 22:618–625. [PubMed: 10788761]
- Whipple MR, Reinecke MG, Gage FH. Inhibition of synaptosomal neurotransmitter uptake by hallucinogens. J Neurochem. 1983; 40:1185–1188. [PubMed: 6131933]
- Windbichler GH, Hausmaninger H, Stummvoll W, Graf AH, Kainz C, Lahodny J, Denison U, Muller-Holzner E, Marth C. Interferon-gamma in the first-line therapy of ovarian cancer: a randomized phase III trial. Br J Cancer. 2000; 82:1138–1144. [PubMed: 10735496]
- Winter JC, Eckler JR, Rabin RA. Serotonergic/glutamatergic interactions: the effects of mGlu2/3 receptor ligands in rats trained with LSD and PCP as discriminative stimuli. Psychopharmacol. 2004; 172:233–240.
- Wyatt RJ, Saavedra JM, Axelrod J. A dimethyltryptamine forming enzyme in human blood. Am J Psychiatry. 1973; 130:754–760. [PubMed: 4514540]
- Yanai K, Ido T, Ishiwata K, Hatazawa J, Takahashi T, Iwata R, Matsuzawa T. In vivo kinetics and displacement study of a carbon-11-labeled hallucinogen, N, N [11C]dimethyltryptamine. Eur J Nucl Med. 1986; 12:141–146. [PubMed: 3489620]
- Yang ZJ, Carter EL, Torbey MT, Martin LJ, Koehler RC. Sigma receptor ligand 4-phenyl-1-(4phenylbutyl)-piperidine modulates neuronal nitric oxide synthase/postsynaptic density-95 coupling mechanisms and protects against neonatal ischemic degeneration of striatal neurons. Exp Neurol. 2010; 221:166–174. [PubMed: 19883643]

Yritia M, Riba J, Ortuño J, Ramirez A, Castillo A, Alfaro Y, de la Torre R, Barbanoj MJ. Determination of N,N-dimethyltryptamine and beta-carboline alkaloids in human plasma following oral administration of Ayahuasca. J Chromatogr B Analyt Technol Biomed Life Sci. 2002; 779:271–281.

Abbreviations

DMT-NO	DMT-N-oxide
HVA	homovanillic acid
IAA	indole-3-acetic acid
INMT	indolethylamine-N-methyltransferase
IP ₃	inositol-1,4,5-triphosphate
LSD	lysergic acid diethylamide
MAOI	monoamine oxidase inhibitor
ΜΑΟ	monoamine oxidase
DMT	N,N-Dimethyltryptamine
DiPT	N,N-Diisopropyltryptamine
NMT	N-methyltryptamine
SERT	serotonin transporter
TAAR	trace amine-associated receptors
VMAT2	vesicle monoamine transporter 2
ТНВС	1,2,3,4-tetrahydro-beta-carboline
DOI	2,5-dimethoxy-4-iodoamphetamine
DOM	2,5-dimethoxy-4-methylamphetamine
2С-Е	2,5-dimethoxy-4-ethylphenethylamine
2С-D	2,5-dimethoxy-4-methylphenethylamine
2C-I	2,5-Dimethoxy-4-iodophenethylamine
2C-T-2	2,5-Dimethoxy-4-ethylthiophenethylamine
3-MT	3-methoxytyramine
DOPAC	3,4-Dihydroxyphenylacetic acid
MDMA	3,4-methylenedioxymethamphetamine
2C-C	4-chloro-2,5-dimethoxyphenethylamine

4-OH-DiPT	4-Hydroxy-N,N-diisopropyl tryptamine
5-MeO-DET	5-Methoxy-diethyl tryptamine
5-MeO-IMPT	5-Methoxy-N-isopropyl-N-methyl tryptamine
5-MeO-aMT	5-Methoxy-a-methyl tryptamine
5-MeO-DMT	5-Methoxy-dimethyl tryptamine
6-OH-DMT	6-hydroxy-DMT

6-OH-DMT-NO 6-OH-DMT-N-oxide

•	N, N-Dimethyltryptamine (DMT) is an endogenous compound in both plants and animals.
•	Subjective effects of DMT are mediated by several neurotransmitter systems.
•	DMT may act as a neurotransmitter.
•	DMT has limited neurotoxicity and may have therapeutic significance.