

Pharmacogenetic Associations of Antipsychotic Drug-Related Weight Gain: A Systematic Review and Meta-analysis

Jian-Ping Zhang^{*1-3}, Todd Lencz¹⁻³, Ryan X. Zhang⁴, Masahiro Nitta⁵, Lawrence Maayan⁶, Majnu John^{1,3,7}, Delbert G. Robinson¹⁻³, W. Wolfgang Fleischhacker⁸, Rene S. Kahn⁹, Roel A. Ophoff¹⁰, John M. Kane^{1-3,11}, Anil K. Malhotra^{1-3,12}, and Christoph U. Correll^{1-3,11,12}

¹Division of Psychiatry Research, The Zucker Hillside Hospital, Northwell Health System, Glen Oaks, NY; ²Department of Psychiatry, Hofstra Northwell School of Medicine, Hempstead, NY; ³Center for Psychiatric Neuroscience, The Feinstein Institute for Medical Research, Manhasset, NY; ⁴Department of Psychology and Neuroscience, Duke University, Durham, NY; ⁵Drug Development Division, Sumitomo Dainippon Pharma Co. Ltd, Tokyo, Japan; ⁶Department of Psychiatry, New York University School of Medicine, New York, NY; ⁷Department of Mathematics, Hofstra University, Hempstead, NY; ⁸Department of Psychiatry and Psychotherapy, Medical University Innsbruck, Innsbruck, Austria; ⁹Department of Psychiatry, University Medical Centre Utrecht, Utrecht, The Netherlands; ¹⁰Department of Psychiatry and Behavioral Sciences, University of California, Los Angeles, CA; ¹¹Department of Psychiatry, Albert Einstein College of Medicine, Bronx, NY

¹²Both authors contributed equally to the article.

*To whom correspondence should be addressed; Division of Psychiatry Research, The Zucker Hillside Hospital, Northwell Health System, 75-59 263rd Street, Glen Oaks, NY 11020, US; tel: 718-470-8471, fax: 718-470-1905, e-mail: JZhang1@nshs.edu

Although weight gain is a serious but variable adverse effect of antipsychotics that has genetic underpinnings, a comprehensive meta-analysis of pharmacogenetics of antipsychotic-related weight gain is missing. In this review, random effects meta-analyses were conducted for dominant and recessive models on associations of specific single nucleotide polymorphisms (SNP) with prospectively assessed antipsychotic-related weight or body mass index (BMI) changes (primary outcome), or categorical increases in weight or BMI ($\geq 7\%$; secondary outcome). Published studies, identified via systematic database search (last search: December 31, 2014), plus 3 additional cohorts, including 222 antipsychotic-naïve youth, and 81 and 141 first-episode schizophrenia adults, each with patient-level data at 3 or 4 months treatment, were meta-analyzed. Altogether, 72 articles reporting on 46 non-duplicated samples ($n = 6700$, mean follow-up = 25.1 wk) with 38 SNPs from 20 genes/genomic regions were meta-analyzed (for each meta-analysis, studies = 2–20, $n = 81$ –2082). Eleven SNPs from 8 genes were significantly associated with weight or BMI change, and 4 SNPs from 2 genes were significantly associated with categorical weight or BMI increase. Combined, 13 SNPs from 9 genes (Adrenoceptor Alpha-2A [*ADRA2A*], Adrenoceptor Beta 3 [*ADRB3*], Brain-Derived Neurotrophic Factor [*BDNF*], Dopamine Receptor D2 [*DRD2*], Guanine Nucleotide Binding Protein [*GNB3*], 5-Hydroxytryptamine (Serotonin) Receptor 2C [*HTR2C*], Insulin-induced gene 2 [*INSIG2*], Melanocortin-4 Receptor

[*MC4R*], and Synaptosomal-associated protein, 25kDa [*SNAP25*]) were significantly associated with antipsychotic-related weight gain (P -values $< .05$ –.001). SNPs in *ADRA2A*, *DRD2*, *HTR2C*, and *MC4R* had the largest effect sizes (Hedges' g 's = 0.30–0.80, ORs = 1.47–1.96). Less prior antipsychotic exposure (pediatric or first episode patients) and short follow-up (1–2 mo) were associated with larger effect sizes. Individual antipsychotics did not significantly moderate effect sizes. In conclusion, antipsychotic-related weight gain is polygenic and associated with specific genetic variants, especially in genes coding for antipsychotic pharmacodynamic targets.

Key words: pharmacogenetics/SNP/antipsychotics/weight gain/BMI/meta-analysis

Introduction

Antipsychotics are first-line therapy for schizophrenia-spectrum disorders,¹ and frequently used as monotherapy or combined with mood stabilizers or antidepressants for bipolar disorder² and major depression,^{3,4} respectively. Despite their efficacy, body weight gain and associated metabolic syndrome are prominent side effects, which increase morbidity and mortality in psychiatric patients.^{5–7} Many antipsychotics can cause significant weight gain, especially some second-generation antipsychotics (SGAs), like clozapine, olanzapine, and quetiapine.^{5,8} No

consistent clinical predictors of antipsychotic-induced weight gain have been identified, and the pathophysiology of weight gain remains poorly understood.⁹ Food intake, energy utilization, metabolism, and body weight are regulated by complex interactions among multiple neurotransmitter systems in multiple brain regions, which are pharmacodynamic targets of antipsychotics to some extent.^{5,10} Genetic factors may play an important role because genome-wide association studies found multiple genes associated with obesity in the general population,¹¹ and functions of proteins that are pharmacodynamic antipsychotic targets may be affected by genetic variants.^{12,13}

Since the late 1990s, pharmacogenetic research has attempted to elucidate genetic underpinnings of antipsychotic-related weight gain. The present study aimed to conduct a comprehensive meta-analysis of the associations of genetic variants with antipsychotic-related weight gain. One key methodological issue in the pharmacogenetics of antipsychotic drug response is that most studies used chronic patient samples.¹⁴ Therefore, the present meta-analysis also included data from 3 cohorts of patients with first episode psychosis or minimal prior drug exposure that were largely unpublished, in addition to published studies.

Methods

Literature Search

Two investigators (J-P.Z., R.X.Z., M.N. and/or L.M.) independently conducted an electronic PubMed/Web of Science search (last: December 31, 2014) for pharmacogenetic studies of antipsychotic-related weight change. Combinations of the following key words were used: antipsychotic(s), neuroleptic(s), genetic(s), genomic(s), gene, single nucleotide polymorphism (SNP), polymorphism, weight gain, body mass index (BMI), obesity, and metabolic. We also screened reference lists from identified papers and reviews for additional studies. Inclusion criteria were: (1) humans with mental illness; (2) longitudinal data on body weight or BMI change, or percentage of patients gaining significant weight or BMI within each genotype group after a specified period of antipsychotic treatment; and (3) sufficient data to compute an effect size (ES). Exclusion criteria were: (1) animal or healthy subject studies; (2) cross-sectional studies or longitudinal studies that did not report pre- and post-treatment change in weight or BMI; (3) studies of metabolic syndrome not reporting separate weight or BMI data; (4) studies of SNPs not examined in other studies (“orphan” SNPs); and (5) studies reporting data overlapping with previously published papers (data from the largest report were included). If a SNP was studied in ≥ 2 independent samples, data from the 3 additional cohorts were added whenever possible so that each SNP was meta-analyzed with ≥ 5 samples. One study examined candidate genes in association with antipsychotic-related weight gain¹⁵ using data from the CATIE trial (Clinical Antipsychotic Trials of Intervention Effectiveness), but it

was excluded from the meta-analysis due to its use of different weight gain phenotype. Weight gain was defined in the study as the maximum percent weight change at any time point during the first 18 months of treatment, which was different from all other studies where weight gain was defined as weight change between 2 time points, eg, from baseline to a follow-up time point.

Data Extraction and Outcome Variables

Data were independently extracted by 2 authors (J-P.Z., M.N. and/or L.M.); disagreements were resolved by consensus. For missing information, first and/or last authors were contacted requesting additional/unpublished data.

Primary outcome was change in weight or BMI from baseline until after a specified period of antipsychotic treatment. Whenever ≥ 1 assessment time points were reported, we picked the one closest to 2–3 months (preferring 3 mo) to increase homogeneity. Change in kg or kg/m² were preferred but pooled with percent change in weight or BMI if only these were reported. Secondary outcome was the percentage of patients within each genotype group gaining significant weight or BMI during antipsychotic treatment. Most studies used $\geq 7\%$ body weight gain, but some used $\geq 5\%$ or $\geq 10\%$, which were also pooled.

Additional Cohorts

Three additional cohorts that had published data for at least 1 SNP but for which we obtained patient-level data were also included in the meta-analysis.

1. In the Second-Generation Antipsychotic Treatment Indications, Effectiveness and Tolerability in Youth (SATIETY sample) study, 222 pediatric patients (age = 13.8 ± 3.6 y; male = 57%) with ≤ 7 days of antipsychotic lifetime history initiated clinicians' choice antipsychotics (aripiprazole, olanzapine, quetiapine or risperidone) for the first time and were followed for 3 months.¹⁶
2. In the Zucker Hillside Hospital First Episode Schizophrenia Clinical Trial (ZHH-FE sample), 81 first-episode schizophrenia patients (age = 23.0 ± 4.9 y; male = 75%) were randomized to risperidone or olanzapine and followed for 4 months.^{17,18}
3. In the European First Episode Schizophrenia Trial (EUFEST sample), 141 first-episode schizophrenia patients (age = 25.6 ± 5.2 y; male = 60%) were randomized to amisulpride, haloperidol, olanzapine, quetiapine or ziprasidone and had weight change data at 3 months follow-up.¹⁹

All 3 cohorts were genotyped on approximately 1 million SNPs using the Illumina Omni-1Quad platform, followed by standard quality control procedures (details published previously²⁰). For SNPs included in the meta-analysis, but not genotyped, the SNAP online tool from the Broad Institute was accessed to find proxy SNPs using either

the 1000 Genomes Pilot 1 or HapMap 3 CEU population panel, with parameters set as $r^2 \geq .80$ and distance $\leq 100\text{kb}$.

Statistical Analysis

Outcomes were analyzed separately for each SNP using Comprehensive Meta-Analysis software version 2 (Biostat) whenever ≥ 2 studies contributed data (otherwise data went into the “orphan” SNP category). For continuous and categorical outcomes, Hedges’ g and OR, $\pm 95\%$ CIs, were calculated as the ES measure. For each SNP with sufficient data, both dominant and recessive genetic models were meta-analyzed in association with weight or BMI changes. For selected SNPs in which results from dominant and recessive models on the primary outcome were suggestive of additive effects of the risk allele, a formal test of additive genetic model was conducted. For each study, a linear regression of BMI change on the number of risk alleles (0, 1, 2) was simulated based on summary statistics (mean, SD, n) from each genotype group in R statistical package, and the regression coefficient was converted to Hedges’ g with corresponding SE (supplementary methods). Pooled ESs were computed with a random effects model to accommodate heterogeneity across included studies.²¹ In each meta-analysis, a cohort was included only once. Statistical significance of the pooled ES was set at $\alpha = .05$ without multiple testing correction because each SNP was chosen based on previous research, following a hypothesis-driven approach.

Study heterogeneity was assessed using Q and I^2 statistics, with $I^2 < 25\%$ representing low, $\sim 50\%$ moderate, and $> 75\%$ representing high heterogeneity.²¹ Whenever heterogeneity was present, moderator and meta-regression analyses were conducted to explore moderator effects. Sensitivity analyses were conducted to assess potential influences of any one single study on the pooled ES. Within each meta-analysis, included studies were removed one at a time to check for significant alterations of the pooled ES and associated P -values. Publication bias was assessed with the funnel plot, Egger’s regression test,²² and the “Trim and Fill” method.²³

Based on the Venice guideline of systematically assessing cumulative evidence on genetic association,²⁴ each SNP was assigned a category based on 3 criteria: (1) amount of evidence (A: large-scale evidence, total sample size $n > 1000$; B: moderate amount of evidence, $n = 500\text{--}1000$; C: little evidence, $n < 500$); (2) replication (A: statistically significant overall ES with no/minimal between-study heterogeneity, $I^2 < 25\%$; B: statistically significant overall ES with moderate to large between-study heterogeneity, $I^2 \geq 25\%$; C: insignificant overall ES); and (3) evidence of bias (A: statistically significant overall ES without evidence of bias based on “Trim and Fill” method and Egger’s test; B: evidence of bias without significance level change after adjustment, or insignificant overall ES without evidence

of bias; C: evidence of bias). To generate an overall index of the strength of evidence, categories of A, B, and C were assigned a score of 3, 2, and 1, respectively, and a total score across the 3 categories was calculated. A total score = 8–9 was considered strong evidence supporting the genotype-phenotype association, a score = 6–7 was considered moderate evidence, and a score ≤ 5 was considered minimal evidence.

To explore polygenic effects of SNPs, a polygenic risk score was computed in the SATIETY and EUFEST cohorts using an additive genetic model combining top SNPs that were significantly associated with weight gain from the meta-analysis. We also assessed the percent variance explained by the risk score.

Results

Literature Search

The literature search produced 586 unduplicated hits, of which 72 reports (see table 1 for details) met inclusion criteria, entering into the meta-analysis (supplementary figure 1). Sample sizes varied from 32 to 481 and most studies included chronic patients (21 studies included first-episode or antipsychotic-naïve patients). Altogether, the 72 reports referred to 46 independent samples, as several published studies reported on different genes or SNPs from the same cohort. After eliminating redundancy, the total sample size from the 46 samples was 6615. Including the 3 additional cohorts ($n = 444$; 289 of which were already published), the total independent sample size in 46 samples was 6770. Most studies were short-term, ranging from 4 weeks to 4 months, but 16 studies (22%) had follow-up at ≥ 1 year (mean follow-up = 25.1 ± 42.0 wk, range = 1 mo to 7 y). Most patients were either Caucasian or Asian, including 30 studies (41.7%) with all Asian patients. The most common antipsychotics were olanzapine (52.8%), risperidone (41.7%), and clozapine (40.3%). Thirty-five studies involved monotherapy with a single antipsychotic, including clozapine (studies = 15), olanzapine (studies = 15), risperidone (studies = 4), and iloperidone (studies = 1). Most studies included schizophrenia patients (94.4% of studies), but some also included patients with various psychiatric diagnoses (table 1).

Included Genes and SNPs

Altogether, 38 SNPs from 20 genes/genomic regions on 15 chromosomes were reported in ≥ 2 independent cohorts, entering into subsequent meta-analyses. Table 2 lists the included SNPs, genes or genomic regions, and other relevant information, as well as proxy SNPs used in the 3 additional cohorts as necessary. The major alleles, minor alleles, and minor allele frequency were based on the 1000 Genome CEU population. Three included SNPs (rs1799732, rs1801028, and rs1051312) did not have proxy SNPs in the 3 additional cohorts, therefore, only

Table 1. Demographic, Illness Treatment and Outcome Information of All 72 Included Studies

Study (First Author, Year)	Genes (SNPs) Included in Meta-Analysis	n	Length (wk)	Age	% Male	% Caucasian	Diagnoses	%FE, Drug-Naïve	APs	Outcome Variables
Basile 2001 ⁷¹	<i>HTR2C</i> (rs6318), <i>ADRB3</i> (rs4994), <i>TNF</i> (rs1800629), <i>HTR2A</i> (rs6313, rs6314)	80	6	33.1 ± 8.4	65	72	SCZ	NR	CLZ	Weight change
Basile 2002 ⁷²	<i>HTR2C</i> (rs3813929)	80	6	NR	65	72	SCZ	NR	CLZ	Weight change
Bishop 2006 ⁷³	<i>GNB3</i> (rs5443)	42	6	36.0 ± 8.7	81	NR	SCZ	50%	OLZ	% Weight change
Brandl 2012 ⁷⁴	<i>LEP</i> (rs7799039)	181	6 to 14	35.9 ± 10.9	65	70	SCZ, SZA	NR	Various APs	% Weight change
Calarge 2009 ⁷⁵	<i>LEPR</i> (rs1137101)	74	2 y	11.7 ± 2.9	91	84	Various diagnoses	NR	RIS	BMI change
Chowdhury 2012 ⁷⁶	<i>MC4R</i> (rs17782313)	224	6 to 14	35.6 ± 10.5	67	70	SCZ, SZA	NR	Various APs	% Weight change
Czerwensky 2013 ⁷⁷	<i>MC4R</i> (rs17782313)	173	4	39.3 ± 14.7	37	NR	Various diagnoses	28%	Various APs	BMI change
Czerwensky 2013 ⁷⁸	<i>MC4R</i> (rs489693)	169	4	39.3 ± 14.7	37	NR	Various diagnoses	28%	Various APs	Weight change
Ellingrod 2005 ⁷⁹	<i>HTR2C</i> (rs3813929)	42	6	NR	81	100	SCZ	NR	OLZ	Weight change >10% Weight gain
Ellingrod 2007 ⁸⁰	<i>LEP</i> (rs7799039)	37	6	37.0 ± 8.4	81	NR	SCZ	NR	OLZ	BMI change
Fernandez 2010 ⁸¹	<i>LEPR</i> (rs1137101)	56	14	39.1 ± 9.0	79	NR	SCZ	NR	CLZ	Weight change, BMI change
Godlewska 2009 ⁸⁶	<i>HTR2C</i> (rs3813929, rs518147)	107	6	29.3 ± 10.0	50	100	SCZ	34	OLZ	% BMI change, ≥10% BMI gain
Herken 2009 ⁸²	<i>PPARG</i> (rs1801282)	95	6	34.4 ± 13.0	52	100	SCZ	NR	OLZ	Weight change
Hoekstra 2010 ⁸³	<i>HTR2C</i> (rs3813929)	32	8	8.7 ± 2.8	88	NR	PDD	NR	RIS	BMI change
Hong 2001 ⁸⁴	<i>HTR2A</i> (rs6313)	93	17	37.1 ± 8.2	65	0 (100% Asian)	SCZ	0	CLZ	Weight change
Hong 2010 ⁸⁵	<i>HTR2C</i> (rs6318)	479	4 y	47.2 ± 13.2	61	0 (100% Asian)	SCZ	NR	CLZ, OLZ, RIS	≥7% weight gain
Houston 2012 ⁸⁶	<i>HTR6</i> (T267C)	205	8	NR	NR	100	Various diagnoses	NR	OLZ	Weight change
Huang 2011 ⁸⁷	<i>ANKKI</i> (rs1800497), <i>DRD2</i> (rs1799978, rs7131056, rs6275, rs2242591)	500	5 y	43.9 ± 9.1	60	0 (100% Asian)	SCZ	NR	CLZ, OLZ, RIS	% Weight change >7% Weight gain
Kang 2008 ⁸⁸	<i>ANKKI</i> (rs1800497), <i>DRD2</i> (rs1079598, rs1801028, rs2242591)	74	>3 mo	47.2 ± 11.6	68	0 (100% Asian)	SCZ	NR	OLZ	Weight change, ≥7% weight gain
	<i>HTR2C</i> (rs3813929, rs518147, rs6318)									
	<i>TNF</i> (rs1800629)									
	<i>LEP</i> (rs7799039)									

Table 1. Continued

Study (First Author, Year)	Genes (SNPs) Included in Meta-Analysis	n	Length (wk)	Age	% Male	% Caucasian	Diagnoses	%FE, Drug-Naive	APs	Outcome Variables									
Kuzman 2008 ⁸⁹ Kuzman 2011 ⁹⁰	<i>HTR2C</i> (rs3813929)	108	4 mo	30.6 ± 11.5	0	100	SCZ	64	OLZ, RIS	≥7% Weight gain									
	<i>MDR1</i> (rs2032582, rs1045642)	101	3 mo	33.5 ± 10.6	0	100	SCZ, SZA, delusional disorder	100	OLZ, RIS	BMI change									
	<i>HTR2C</i> (rs3813929)																		
	<i>MDR1</i> (rs2032582, rs1045642)																		
Laika 2010 ⁹¹	<i>HTR2C</i> (rs3813929)	56	4	41.6 ± 15.9	50	100	Various diagnoses	NR	OLZ	Weight change, BMI change									
Lane 2006 ⁹²	<i>HTR2A</i> (rs6313, rs6314)	123	6	34.0 ± 9.7	55	0 (100% Asian)	SCZ	NR	RIS	Weight change >7% weight gain									
	<i>HTR2C</i> (rs3813929)																		
	<i>HTR6</i> (rs1805054)																		
Le Hellard 2009 ⁹³	<i>DRD2</i> (rs1799732, rs1801028)	160	3 mo	21.9 ± 8.9	61	100	SCZ spectrum disorders	0	Various APs	BMI change									
	<i>ANKKI</i> (rs1800497)																		
	<i>BDNF</i> (rs6265)																		
	<i>INSIG2</i> (rs10490624, rs17047764, rs17587100, rs7566605)																		
	<i>DRD2</i> (rs1799732)																		
Lencz 2010 ⁹⁴	<i>DRD2</i> (rs1799732)	58	16	23.5 ± 4.9	76	28	SCZ, SZA, SZP	100	RIS, OLZ	Weight change									
Lin 2006 ⁹⁵	<i>MDR1</i> (rs1045642)	41	6 wk	35.7 ± 8.8	80	90	SCZ	NR	OLZ	Weight change									
	<i>MC4R</i> (rs489693)																		
Malhotra 2012 ²⁰		139	12	13.4 ± 3.8	58	55.4	Various diagnoses	100	RIS, APZ, QTP	BMI change									
											73	6	33.5 ± 8.3	62	70	SCZ	0	CLZ	Weight change
Miller 2005 ⁹⁶	<i>HTR2C</i> (rs3813929)	41	6 mo	35.6 ± 9.7	63	85	SCZ, SZA, SZP	100	Various APs	Weight change									
											83	26.1	44 ± 10.5	60	100	Various diagnoses	NR	Various APs	Weight change
Monteleone 2010 ⁹⁷	<i>CNR1</i> (rs1049353)	83	26.1	44 ± 10.5	60	100	Various diagnoses	NR	Various APs	Weight change									
Mou 2008 ⁹⁸	<i>LEP</i> (rs7799039)	84	10	24 ± 6.0	65	0 (100% Asian)	SCZ	100	RIS, CPZ	Weight change									
Mueller 2005 ⁹⁹	<i>SNAP25</i> (rs1051312, rs3746544, rs8636)	59	14	40.1 ± 9.5	78	25	SCZ, SZA	0	Various APs	Weight change									
	<i>ANKKI</i> (rs1800497)																		
Mueller 2012 ¹⁰⁰		206	6 or 14	35.7 ± 10.4	68	72 > 62	SCZ, SZA	0	Various APs	BMI change									
Musil 2008 ¹⁰¹	<i>DRD2</i> (rs1799732, rs1799978, rs1079598, rs6275, rs7131056)	162	5	34.2 ± 12.3	57	100	SCZ, SZA	NR	Various APs	Weight change									
	<i>SNAP25</i> (rs1051312, rs3746544, rs8636)																		
Opgen-Rhein 2010 ¹⁰²	<i>HTR2C</i> (rs3813929, rs6318)	128	6	38.6 ± 12.0	63	100	SCZ, SZA	17	Various AP	>7% Weight gain									
	<i>INSIG2</i> (rs17587100, rs10490624, rs17047764, rs7566605)																		
	<i>LEP</i> (rs7799039)																		

Table 1. Continued

Study (First Author, Year)	Genes (SNPs) Included in Meta-Analysis	n	Length (wk)	Age	% Male	% Caucasian (Asian)	Diagnoses	%FE, Drug-Naïve	APs	Outcome Variables
Park 2006 ¹⁰³	<i>ADR42A</i> (rs1800544)	62	Over 1 y	46.5 ± 11.1	71	0 (100% Asian)	SCZ	NR	OLZ	Weight change % Weight change >10% Weight gain
Park 2008 ¹⁰⁴	<i>HTR2C</i> (rs3813929)	79	Over 1 y	46.1 ± 12.1	67	0 (100% Asian)	SCZ	NR	OLZ	Weight change BMI change >7% Weight gain
Park 2009 ¹⁰⁵	<i>GNB3</i> (rs5443)	79	Over 1 y	46.6 ± 11.6	67	0 (100% Asian)	SCZ	NR	OLZ	Weight change % Weight change BMI change >10% Weight gain
Park 2011 ¹⁰⁶	<i>CNR1</i> (rs1049353, rs806368)	78	Over 1 y	46.4 ± 11.6	67	0 (100% Asian)	SCZ	NR	OLZ	Weight change >7% Weight gain
Perez-Iglesias 2010 ¹⁰⁷	<i>LEP</i> (rs7799039) <i>LEPR</i> (rs1137101) <i>FTO</i> (rs9939609)	194	1 y	28.4 ± 8.3	58	94	SCZ spectrum disorders	100	Various APs	Weight change BMI change
Popp 2009 ¹⁰⁸	<i>HTR2C</i> (rs6318)	102	4	37.5 ± 13.7	45	100	SCZ, SZA	NR	Various APs	BMI change
Reynolds 2002 ⁸⁸	<i>HTR2C</i> (rs3813929)	123	6 and 10	26.6 ± 7.7	52	0 (100% Asian)	SCZ	100	Various APs	BMI change >7% Weight gain
Reynolds 2003 ¹⁰⁹	<i>HTR2C</i> (rs3813929)	32	6	NR	66	0 (100% Asian)	SCZ	100	CLZ	BMI change
Reynolds 2012 ¹¹⁰	<i>FTO</i> (rs9939609)	93	1 y	25.5 ± 6.7	74	100	Psychosis	100	Various APs	Weight change BMI change
Ryu 2006 ¹¹¹	<i>LEP</i> (rs7799039)	71	4	30.5 ± 7.6	45	0 (100% Asian)	SCZ	NR	Various APs	BMI change
Ryu 2007 ¹¹²	<i>HTR2C</i> (rs3813929)	84	4	30.1 ± 7.5	46	0 (100% Asian)	SCZ	69	Various APs	BMI change >7% BMI change
Shao 2008 ¹¹³	<i>HTR2C</i> (rs3813929, rs518147)	170	1 y	23.1 ± 5.1	35	0 (100% Asian)	SCZ	100	NR	change >7% Weight gain
Shing 2014 ¹¹⁴	<i>FTO</i> (rs9939609)	218	6–14 wk	NR	66	69	SCZ or SZA	0	Various APs	% Weight change
Sicard 2010 ¹¹⁵	<i>HTR2C</i> (rs518147, rs3813929, rs6318)	205	Average 10 wk	35.9 ± 10.1	69	68	SCZ or SZA	0	Various APs	% Weight change >7% Weight gain
Sickert 2009 ¹¹⁶	<i>ADR42A</i> (rs1800544)	129	10	36.5 ± 9.0	74	50	SCZ or SZA	0	Various APs	Weight change
Song 2014 ¹¹⁷	<i>FTO</i> (rs9939609)	237	6 mo	27.5 ± 7.6	54	0 (100% Asian)	SCZ	100	RIS	Weight change BMI change
Souza 2008 ¹¹⁸	<i>GNB3</i> (rs5443)	208	6 wk, 14 wk	35.9 ± 10.3	68	68	SCZ or SZA	0	Various APs	Weight change

Table 1. Continued

Study (First Author, Year)	Genes (SNPs) Included in Meta-Analysis	n	Length (wk)	Age	% Male	% Caucasian	Diagnoses	%FE, Drug-Naïve	APs	Outcome Variables
Srisawat 2014 ¹⁹	<i>MTHFR</i> (rs1801131, rs1801133)	182	8 wk	26.2 ± 7.4	46	0 (100% Asian)	SCZ	100	Various APs	BMI change
Steaker 2012 ²⁰	<i>PPARG</i> (rs1801282)	72 138	3 mo 4 wk	25.4 ± 6.8 Range ¹⁷⁻³⁸	74 46	100 NR	SCZ Various diagnoses	100 NR	Various APs OLZ	BMI change Weight change
Templeman 2005 ²¹	<i>HTR2C</i> (rs3813929) <i>LEP</i> (rs7799039)	73	6 wk, 3 mo	25.2 ± 0.8	75	100	Psychosis	100	Various APs	BMI change >7% BMI change
Theisen 2004 ²²	<i>HTR2C</i> (rs3813929)	97	12	22.1 ± 7.7	59	100	SCZ spectrum disorders	0	CLZ	BMI change >7% BMI change
Thompson 2010 ²³	<i>HTR2C</i> (rs3813929)	216	4	NR	NR	NR	SCZ	NR	Iloperidone	Weight change
Tiwari 2010 ²⁴	<i>INSIG2</i> (rs17587100, rs7566605, rs10490624, rs17047764)	154	Average 10 wk	35.8 ± 9.8	71	58	SCZ, SZA	0	CLZ, OLZ, RIS, HAL	% Weight change
Tiwari 2010 ²⁵	<i>CNR1</i> (rs1049353, rs806368)	183	Average 10 wk	36.1 ± 10.2	68	64	SCZ, SZA	0	Various APs	% Weight change
Tsai 2002 ²⁶	<i>HTR2C</i> (rs3813929)	80	4 mo	36.7 ± 8.4	65	0 (100% Asian)	SCZ SZA	0	CLZ	BMI change >7% BMI change
Tsai 2003 ²⁷	TNF (rs1800629)	99	4 mo	36.0 ± 8.0	66	0 (100% Asian)	SCZ	0	CLZ	Weight change
Tsai 2004 ²⁸	<i>GNB3</i> (rs5443) <i>ADRB3</i> (rs4994)	87	4 mo	37.0 ± 8.2	64	0 (100% Asian)	SCZ or SZA	0	CLZ	Weight change % Weight change
Tsai 2011 ²⁹	BDNF (rs6265)	481	>2 y	43.9 ± 8.9	60	0 (100% Asian)	SCZ	0	CLZ, OLZ, RIS	% Weight change % BMI change
Ujike 2008 ³⁰	<i>HTR2C</i> (rs3813929, rs6318) <i>HTR2A</i> (rs6213) <i>ADRB3</i> (rs4994) <i>GNB3</i> (rs5443)	164	4 mo	51.8 ± 10.9	62	0 (100% Asian)	SCZ	0	CLZ, OLZ, RIS OLZ	Weight change % Weight change % BMI change
Van Winkel 2010 ³¹	<i>MTHFR</i> (rs1801131, rs1801133)	104	3 mo	31.3 ± 11.7	68	NR	SCZ or SZA	NR	Various APs	Weight change
Wang 2005 ³²	<i>ADRA2A</i> (rs1800544)	93	1 y	38.4 ± 8.1	53	0 (100% Asian)	SCZ	0	CLZ	Weight change >7% Weight gain
Wang 2005 ³³	<i>GNB3</i> (rs5443)	134	1 y	38.5 ± 8.0	60	0 (100% Asian)	SCZ	0	CLZ	Weight change % Weight change
Wang 2010 ³⁴	TNF (rs1800629)	55	8 y	37.2 ± 7.7	49	0 (100% Asian)	SCZ	0	CLZ	Weight change BMI change
Zai 2012 ³⁵	BDNF (rs6265)	257	6 wk	31.8 ± 7.9	76	100	SCZ SZA	0	Various APs	Weight Δ >7% Weight gain
Zhang 2003 ³⁶	<i>ANKKI</i> (rs1800497)	117	10 wk	26.0 ± 8.0	50	0 (100% Asian)	SCZ	100%	Various APs	BMI change Weight change >7% Weight gain

Table 1. Continued

Study (First Author, Year)	Genes (SNPs) Included in Meta-Analysis	n	Length (wk)	Age	% Male	% Caucasian (Asian)	Diagnoses	%FE, Drug-Naïve	APs	Outcome Variables
Zhang 2003 ¹³⁷	LEP (rs7799039)	128	10 wk	26.0 ± 7.0	48	0 (100% Asian)	SCZ	100%	RIS, CPZ	BMI change Weight change >7% Weight gain BMI change
Zhang 2007 ¹³⁸	LEP (rs7799039)	102	Average 7 y	47.2 ± 6.3	66	0 (100% Asian)	SCZ	0	CLZ	BMI change
Zhang 2008 ¹³⁹	BDNF (rs6265)	196	At least 2 y	NR	66	0 (100% Asian)	SCZ	100%	Various APs	BMI change

Notes: AMI, amisulpride; AP, antipsychotic; APZ, aripiprazole; BMI, body mass index; CLZ, clozapine; CPZ, chlorpromazine; FE, first episode; FLU, fluphenazine; HAL, haloperidol; OLZ, olanzapine; QTP, quetiapine; RIS, risperidone; SCZ, schizophrenia; SNP, single nucleotide polymorphism; SLP, sulpiride; SZ, schizoaffective disorder; SZP, schizophreniform disorder; ZIP, ziprasidone; LEP, Leptin; LEPR, Leptin receptor; ADRA2A: Adrenoceptor alpha 2A; ADRB3: Adrenoceptor beta 3; ANKK1: Ankyrin repeat and kinase domain containing 1; BDNF, Brain-derived neurotrophic factor; CNR1, Cannabinoid receptor 1; DRD2, Dopamine receptor D2; FTO, Fat mass and obesity associated; GNB3, Guanine nucleotide binding protein (G protein), beta polypeptide 3; HTR2A, 5-hydroxytryptamine (serotonin) receptor 2A, G protein-coupled; HTR2C, 5-hydroxytryptamine (serotonin) receptor 2C, G protein-coupled; HTR6, 5-hydroxytryptamine (serotonin) receptor 6, G protein-coupled; INSIG2, Insulin-induced gene 2; MC4R, Melanocortin 4 receptor; MDR1 (ABCB1), ATP-binding cassette, sub-family B (MDR/TAP), member 1; MTHFR, Methyltetrahydrofolate reductase; PPARG, Peroxisome proliferator-activated receptor gamma; SNAP25, Synaptosomal-associated protein, 25kDa; TNF, Tumor necrosis factor.

published studies were meta-analyzed for these 3 SNPs. The 5-Hydroxytryptamine (Serotonin) Receptor 2C (*HTR2C*) polymorphism, rs3813929 (−759C/T) was the most studied SNP (published studies = 22). Seven SNPs from Dopamine Receptor D2 (*DRD2*) were included in the meta-analysis, the most in a single gene.

Overview of the Meta-analytic Results

Altogether, 11 SNPs from 8 genes were significantly associated with weight or BMI change, and 4 SNPs from 2 genes were significantly associated with study-defined significant weight gain (table 3). Combined together, 13 SNPs from 9 genes (Adrenoceptor Alpha-2A [*ADRA2A*], Adrenoceptor Beta 3 [*ADRB3*], Brain-Derived Neurotrophic Factor [*BDNF*], *DRD2*, Guanine Nucleotide Binding Protein [*GNB3*], *HTR2C*, Insulin-induced gene 2 [*INSIG2*], Melanocortin-4 Receptor [*MC4R*], and Synaptosomal-associated protein, 25kDa [*SNAP25*]) were significantly associated with antipsychotic-related weight gain (*P*-values: ≤.05–.001). SNPs in *ADRA2A*, *DRD2*, *HTR2C*, and *MC4R* had the largest ES (Hedges' *g*s = 0.30–0.80, ORs = 1.47–1.96). Forest plots for the significant results are included in supplementary figures 1–5. Heterogeneity across studies was not large for most significant SNPs, except for rs489693 (AA vs C carriers, *I*² = 80%), rs1799732 (Ins/Ins vs Del carriers, *I*² = 62.6%), rs3813929 (CC vs T carriers, *I*² = 65.9%), and rs518147 (GG vs C carrier, *I*² = 57.6%). Publication biases existed for several SNPs. However, the direction of publication bias actually underestimated the ES for rs6275, rs7131056, and rs17047764 (ie, the corrected ESs became larger). In contrast, adjusting publication bias eliminated the significance for rs3813929 (secondary outcome, adjusted OR = 1.51, 95% CI = 0.91–2.49). Even after adjusting for potential publication bias, the ES was still significant for rs489693 (AA vs C, adjusted Hedges' *g* = 0.66, 95% CI = 0.09–1.23). Using the modified Venice guideline, 2 *HTR2C* SNPs, rs3813929 and rs518147, achieved a score of 8, 3 SNPs from *ADRA2A*, *DRD2* (rs1799732), and *GNB3* had a score of 7, and 4 SNPs from *DRD2* (rs6275, rs7131056), *INSIG2* (rs17047764), and *MC4R* (rs489693) obtained a score of 6.

Specific Meta-analytic Results

5-Hydroxytryptamine (Serotonin) Receptor 2C Gene. *HTR2C* was the most studied gene, and 3 SNPs were included in the meta-analysis. The most studied SNP, rs3813929, was reported in 22 studies with additional information from the SATIETY and EUFEST sample, totaling 24 studies. The ZHH FE sample did not have any T-allele carrier genotype, and was not included in the analysis. Among these studies, 20 studies reported continuous weight or BMI change data and 18 reported categorical weight gain data. Study duration ranged from 4 weeks to 1 year. The C allele was associated with significantly more weight gain than the T allele. Because

Table 2. List of Genes and SNPs Included in the Meta-analysis

rs#	# Studies	Gene	SNP	Chr	Position	Region	Major Allele	Minor Allele	MAF	Proxy SNP	R ²	D'	Major Allele	Minor Allele	MAF
rs1800544	3	<i>ADRA2A</i>	-1291C/G	10	1111076745		C	G	0.48	rs521674	0.92	1	A	T	0.26
rs4994	3	<i>ADRB3</i>	Trp64Arg	8	37823798	(Missense)	T (Trp)	C (Arg)	0.10						
rs1800497	5	<i>ANKK1</i>	Taq1A (Glu713Lys)	11	112776038	Exon 8	G	A	0.30	rs7118900	0.90	1	G	A	0.18
rs6265	4	<i>BDNF</i>	Val66Met	11	27658369	(Missense)	G	A	0.23						
rs1049353	3	<i>CNR1</i>	1359G/A	6	88143916		G	A	0.14						
rs806368	2	<i>CNR1</i>		6	88140381	3' UTR	T	C	0.28						
rs1799732	3	<i>DRD2</i>	-141C Ins/Del	11	113475529; 113475530	Intron	C	—	0.24	NA					
rs1079598	2	<i>DRD2</i>		11	112801484	Intron 1	T	C	0.21	rs1079594	1	1	A	C	0.13
rs1799978	2	<i>DRD2</i>		11	113475629	Intron	A	G	0.11						
rs1801028	2	<i>DRD2</i>	Ser311Cys	11	112788694	Exon 7	C	G	0.02	NA					
rs2242591	2	<i>DRD2</i>		11	112785131	Downstream	G	A	0.20	rs6278	1	1	C	A	0.13
rs6275	2	<i>DRD2</i>	C939T	11	113412755	Exon (synon)	C	T	0.47						
rs7131056	2	<i>DRD2</i>		11	113459052		C	A	0.48						
rs9939609	3	<i>FTO</i>		16	53820527	Intron	T	A	0.36	rs3751812	1	1	G	T	0.45
rs5443	6	<i>GNB3</i>	C825T	12	6954875	Exon (synon)	C	T	0.48						
rs6313	4	<i>HTR2A</i>	102T/C	13	46895805	Intron	C	T	0.43						
rs6314	2	<i>HTR2A</i>	His452Tyr	13	47409034	Exon	C	T	0.07						
rs3813929	22	<i>HTR2C</i>	-759C/T	X	113818520	Upstream, promoter	C	T	0.12						
rs6318	7	<i>HTR2C</i>	Cys23Ser	X	113871991	Exon 5	G	C	0.17						
rs518147	4	<i>HTR2C</i>	-697G/C	X	113818582	5'UTR	G	C	0.29						
rs1805054	2	<i>HTR6</i>	267T/C	1	19666020		C	T	0.17	rs1977101	1	1	A	G	0.13
rs10490624	3	<i>INSIG2</i>		2	118104916	Intron	A	G	0.09						
rs17047764	3	<i>INSIG2</i>		2	118868582		G	C	0.19	rs3849327	0.94	1	T	C	0.16
rs17587100	3	<i>INSIG2</i>		2	118838410		A	C	0.06	rs17526937	1	1	A	G	0.04
rs7566605	3	<i>INSIG2</i>		2	118078449		G	C	0.30						
rs7799039	12	<i>LEP</i>	-2548A/G	7	128238730		G	A	0.43	rs10487506	1	1	G	A	0.47
rs1137101	4	<i>LEPR</i>	Q223R	1	65592830	(Missense)	A	G	0.41						
rs489693	5	<i>MC4R</i>		18	60215554		C	A	0.33						
rs17782313	2	<i>MC4R</i>		18	60183864		T	C	0.22	rs476828			A	G	0.27
rs1045642	3	<i>MDR1</i>	3435C/T	7	87509329		T, C	A	0.40						
rs2032582	2	<i>MDR1</i>	2677G/T(A)	7	87531302	(Missense)	G	A	0.34						
rs1801131	3	<i>MTHFR</i>	1298A/C	1	11794419	(Missense)	A	C	0.23						
rs1801133	3	<i>MTHFR</i>	677C/T	1	11796321	(Missense)	C	T	0.33						
rs1801282	2	<i>PPARG</i>	Pro12Ala	3	12351626	Intron (Missense)	C	G	0.07						
rs1051312	2	<i>SNAP25</i>	DdelI(T/C)	20	10306440	3' UTR	T	C	0.15	NA					
rs3746544	2	<i>SNAP25</i>	MnlI(T/G)	20	10306436	3' UTR	T	G	0.29						
rs8636	2	<i>SNAP25</i>	TailI(T/C)	20	10307094	3' UTR	C	T	0.27						
rs1800629	4	<i>TNF</i>	G-308A	6	31575254		G	A	0.10						

Note: MAF, minor allele frequency.

Table 3. Meta-analytic Results of Associations Between Antipsychotic Drug-Related Weight Gain and Genotype

Gene	rs#	SNP	Genotype Comparison	BMI or Weight Change		BMI or Weight Change >7% or 10%		Category & Score					
				Hedges' g (95% CI)	P	OR (95% CI)	P						
				# Study (Total n)	F	"T&F"	F	"T&F"					
<i>ADRA2A</i>	rs1800544	-1291C/G	CC vs G	-0.22 (-0.39, -0.05)	.01	6 (645)	0% 0	0.50 (0.24, 1.05)	.07	5 (516)	61% +2	B, B, A 7	
			GG vs C	0.30 (0.09, 0.51)	.01	6 (645)	24% 0	1.74 (0.79, 3.85)	.17	5 (516)	58% 0		
			Additive (G)	0.20 (0.06, 0.33)	.004	6 (645)	30% 0						
<i>ADRB3</i>	rs4994	Trp64Arg	Trp/Trp vs Arg	-0.20 (-0.48, 0.09)	.18	6 (680)	54% +1	1.10 (0.44, 2.77)	.83	3 (358)	47% 0	B, B, C 5	
			Arg/Arg vs Trp	0.84 (0.20, 1.47)	.01	2 (235)	0% NA						
			CC vs T	0.05 (-0.09, 0.19)	.51	7 (842)	0% 0	0.97 (0.75, 1.25)	.80	7 (1181)	0% +3		B, C, C 4
<i>BDNF</i>	rs6265	Val66Met (G/A)	TT vs C	-0.09 (-0.44, 0.26)	.61	7 (842)	35% +2	0.93 (0.64, 1.35)	.70	7 (1179)	0% -3	A, C, C 5	
			Additive (C)	0.05 (-0.09, 0.19)	.46	7 (842)	9% -2						
			AA vs G	0.06 (-0.41, 0.53)	.81	7 (1393)	82% +2	0.79 (0.37, 1.68)	.53	5 (609)	0% -1		B, C, C 4
<i>CNR1</i>	rs1049353	1359 G/A	GG vs A	0.13 (-0.07, 0.32)	.21	7 (1393)	61% 0	1.49 (1.02, 2.18)	.04	5 (609)	0% 0	C, C, B 4	
			AA vs G	-0.08 (-0.51, 0.36)	.74	4 (534)	0% 0	1.31 (0.51, 3.39)	.57	5 (522)	0% -3		
			GG vs A	0.04 (-0.14, 0.23)	.64	4 (534)	0% +1	1.04 (0.69, 1.58)	.84	5 (522)	0% +3		
<i>DRD2</i>	rs1799732	-141C Ins/Del	AA vs G	0.36 (-0.07, 0.79)	.10	3 (305)	0% 0	0.84 (0.53, 1.35)	.47	2 (251)	0% NA	C, A, A 7	
			Del/Del vs Ins	-0.44 (-0.86, -0.02)	.04	3 (305)	63% 0	0.92 (0.50, 1.71)	.80	2 (251)	0% NA		
			Ins/Ins vs Del	0.31 (0.07, 0.54)	.01	3 (305)	0% 0	1.94 (0.65, 5.76)	.23	2 (247)	0% NA		
<i>FTO</i>	rs1079598	C939T	Additive (Del)	-0.07 (-0.75, 0.61)	.85	5 (595)	55% 0	0.63 (0.27, 1.46)	.28	2 (247)	46% NA	B, C, C 4	
			CC vs T	0.06 (-0.13, 0.24)	.53	5 (595)	0% +1	2.28 (0.74, 7.07)	.15	4 (485)	0% -1		
			TT vs C	-0.19 (-0.43, 0.06)	.13	4 (486)	0% +1	0.72 (0.45, 1.13)	.15	4 (485)	0% +2		
<i>GNB3</i>	rs1799978	Ser311Cys	AA vs G	0.17 (-0.33, 0.66)	.50	2 (241)	0% NA	0.88 (0.62, 1.26)	.49	5 (940)	7% +3	C, C, C 3	
			CC vs T	-0.17 (-0.91, 0.58)	.66	4 (480)	51% +1						
			Additive (G)	0.10 (-0.11, 0.31)	.34	4 (480)	0% 0	0.84 (0.54, 1.31)	.44	4 (814)	0% -1		
<i>FTO</i>	rs2242591	C939T	GG vs A	0.09 (-0.13, 0.32)	.42	4 (480)	17% 0	1.07 (0.77, 1.48)	.71	4 (814)	0% +1	C, C, C 3	
			CC vs T	-0.35 (-0.54, -0.16)	<.001	4 (482)	0% -1						
			Additive (T)	0.29 (0.04, 0.53)	.02	4 (482)	5% -1	0.79 (0.59, 1.07)	.13	5 (942)	0% -1		
<i>FTO</i>	rs7131056	C825T	TT vs C	0.25 (0.09, 0.41)	.002	4 (482)	0% -1	1.39 (0.86, 2.22)	.18	5 (942)	32% 0	C, A, B 6	
			Additive (T)	0.14 (-0.14, 0.43)	.32	4 (480)	47% -1						
			CC vs A	-0.31 (-0.51, -0.12)	.002	4 (480)	0% -1	1.15 (0.62, 2.14)	.66	5 (939)	68% 0		
<i>FTO</i>	rs9939609	C825T	Additive (A)	0.19 (0.03, 0.34)	.02	4 (480)	0% 0	0.78 (0.52, 1.15)	.20	5 (939)	31% 0	C, A, B 6	
			AA vs T	0.03 (-0.25, 0.31)	.85	6 (790)	49% -1						
			TT vs A	-0.01 (-0.15, 0.12)	.85	7 (1027)	0% 0	0.72 (0.18, 2.92)	.65	3 (364)	66% -1		
<i>GNB3</i>	rs5443	C825T	CC vs T	-0.18 (-0.38, 0.02)	.08	10 (1004)	44% 0	1.52 (0.96, 2.42)	.08	3 (364)	0% +2	A, B, B 7	
			TT vs C	0.28 (0.08, 0.48)	.006	8 (865)	32% 0	0.74 (0.49, 1.13)	.16	4 (443)	0% -2		
			Additive (T)	0.18 (0.04, 0.32)	.01	8 (865)	30% 0	1.20 (0.71, 2.03)	.49	4 (443)	2% +2		

Table 3. Continued

Gene	rs#	SNP	Genotype Comparison	BMI or Weight Change		# Study (Total n)		BMI or Weight Change >7% or 10%		Category & Score				
				Hedges' g (95% CI)	P	"T&F"	P	OR (95% CI)	P		"T&F"			
HTR2A	rs6313	102T/C	CC vs T	-0.12 (-0.28, 0.04)	.16	7 (814)	0% +1	0.79 (0.48, 1.29)	.34	4 (481)	10% +1	B,C,C		
			TT vs C	0.11 (-0.05, 0.27)	.19	7 (814)	0% 0	1.17 (0.71, 1.92)	.54	4 (481)	6% 0	4		
	rs6314	His452Tyr	Additive (T)	0.08 (-0.04, 0.20)	.17	7 (814)	0% -2							
			His/His vs Tyr	-0.05 (-0.42, 0.33)	.81	5 (563)	69% 0	0.91 (0.48, 1.71)	.76	4 (485)	41% 0	4	41% 0	B,C,B
			Tyr/Tyr vs His	0.48 (-0.69, 1.64)	.42	3 (324)	68% 0	1.62 (0.23, 11.38)	.63	2 (246)	32% NA	5	32% NA	5
HTR2C	rs3813929	-759C/T	CC vs T	0.23 (0.04, 0.42)	.02	20 (2082)	66% 0	1.96 (1.19, 3.22)	.009	18 (1738)	67% -4	4	A,B,A	
			GG vs C	0.10 (-0.10, 0.29)	.34	9 (1111)	35% +2	1.47 (1.03, 2.11)	.04	5 (687)	0% -1	5	A,C,C	
			GG vs C	0.18 (0.02, 0.34)	.03	5 (671)	0% 0	1.86 (1.03, 3.35)	.04	5 (659)	58% 0	8	B,A,A	
HTR6	rs1805054	267T/C	CC vs T	-0.03 (-0.33, 0.28)	.87	5 (576)	48% +2	0.92 (0.47, 1.80)	.82	3 (361)	30% 0	4	B,C,C	
			TT vs C	-0.11 (-0.41, 0.20)	.50	3 (338)	0% +2	0.33 (0.04, 3.03)	.33	1 (123)	NA NA	4	NA NA	
			TT vs C	0.07 (-0.19, 0.33)	.61	5 (666)	44% +1	0.96 (0.54, 1.71)	.89	4 (485)	24% +2	4	B,C,C	
INSIG2	rs17587100	Q223R	CC vs G	0.31 (0.00, 0.61)	.05	5 (665)	23% +2	1.55 (0.55, 4.39)	.41	3 (360)	0% -2	6	B,A,C	
			GG vs C	-0.17 (-0.36, 0.02)	.08	5 (665)	10% 0	1.20 (0.81, 1.78)	.37	4 (485)	0% +1	5	B,C,B	
			Additive (C)	0.19 (0.03, 0.35)	.02	5 (665)	10% 0							
			AA vs C	0.05 (-0.22, 0.32)	.73	5 (665)	23% 0	0.65 (0.26, 1.60)	.34	4 (489)	43% +2	5	B,C,C	
			CC vs G	0.06 (-0.19, 0.31)	.64	5 (655)	0% +2	0.86 (0.36, 2.05)	.74	4 (481)	34% -1	4	B,C,C	
LEP	rs7566605	-2548A/G	GG vs C	0.10 (-0.06, 0.27)	.22	5 (655)	11% 0	0.87 (0.59, 1.27)	.46	4 (481)	0% +1	4	B,C,C	
			AA vs G	-0.08 (-0.30, 0.13)	.45	11 (1138)	51% 0	1.24 (0.50, 3.05)	.64	4 (414)	74% +2	5	A,C,C	
			GG vs A	0.16 (-0.01, 0.32)	.06	10 (967)	25% 0	0.73 (0.33, 1.60)	.43	3 (340)	5% +2	5	B,C,C	
			Additive (G)	0.07 (-0.06, 0.19)	.27	7 (763)	0% 0							
			AA vs G	0.03 (-0.14, 0.19)	.74	7 (682)	0% +3							
M4R	rs489693	Q223R	GG vs A	0.09 (-0.11, 0.30)	.38	6 (653)	0% +1							
			AA vs C	0.80 (0.20, 1.41)	.009	6 (583)	80% -1							
			CC vs A	-0.28 (-0.53, -0.03)	.03	6 (583)	52% +1							
			Additive (A)	0.30 (0.04, 0.57)	.03	6 (583)	67% -1							
			CC vs T	0.14 (-0.29, 0.56)	.53	5 (735)	49% +1							
MDR1	rs17782313	3435C/T	TT vs C	-0.25 (-0.52, 0.02)	.07	5 (735)	68% 0							
			Additive (C)	0.19 (-0.05, 0.42)	.12	5 (735)	64% 0							
			CC vs T	-0.02 (-0.22, 0.17)	.81	5 (499)	0% 0	0.87 (0.55, 1.36)	.53	4 (467)	0% +1	3	C,C,C	
			CC vs T	-0.04 (-0.26, 0.18)	.71	5 (499)	0% +2	0.83 (0.36, 1.89)	.65	4 (467)	61% 0	3	C,C,C	
			GG vs T	0.06 (-0.13, 0.25)	.54	4 (462)	0% +1	1.01 (0.53, 1.91)	.98	4 (469)	58% 0	3	C,C,C	
MTHFR	rs2032582	2677G/T(A)	TT vs G	-0.06 (-0.32, 0.20)	.68	4 (462)	0% +1	0.79 (0.44, 1.39)	.41	4 (469)	2% 0	3	B,C,C	
			AA vs C	0.07 (-0.08, 0.22)	.36	6 (707)	0% 0	1.36 (0.86, 2.15)	.19	3 (359)	0% +2	4	B,C,C	
			CC vs A	-0.16 (-0.57, 0.24)	.43	6 (707)	43% -2	0.57 (0.09, 3.77)	.56	3 (359)	77% 0	4	B,C,C	
			CC vs T	0.17 (-0.08, 0.41)	.19	6 (705)	58% 0	0.99 (0.49, 1.99)	.98	3 (357)	49% +1	5	B,C,B	
			TT vs C	-0.01 (-0.24, 0.22)	.92	6 (705)	0% 0	1.16 (0.50, 2.70)	.74	3 (357)	0% -2	5	B,C,B	
PPARG	rs1801282	Pro12Ala	Pro/Pro vs Ala	0.09 (-0.15, 0.33)	.47	5 (622)	23% 0	0.79 (0.49, 1.30)	.35	4 (529)	0% +1	5	B,C,B	

Table 3. Continued

Gene	rs#	SNP	Genotype Comparison	BMI or Weight Change		BMI or Weight Change >7% or 10%		Category & Score
				Hedges' <i>g</i> (95% CI)	<i>P</i>	Hedges' <i>g</i> (95% CI)	<i>P</i>	
<i>SNAP25</i>	rs1051312	Ddel(T/C)	TT vs C	-0.58 (-0.87, -0.29)	<.001	2 (218)	0% NA	
	rs3746544	MnlI(T/G)	GG vs T	0.03 (-0.27, 0.33)	.86	4 (416)	0% +1	1.40 (0.66, 2.99)
	rs8636	TaiI(T/C)	TT vs G	0.09 (-0.24, 0.43)	.59	4 (416)	63% +1	0.73 (0.46, 1.15)
			CC vs T	0.19 (-0.10, 0.47)	.20	4 (418)	48% 0	0.88 (0.56, 1.38)
			TT vs C	-0.09 (-0.41, 0.23)	.58	4 (418)	0% +2	1.40 (0.67, 2.93)
TNF-alpha	rs1800629	G-308A	AA vs G	0.21 (-0.49, 0.90)	.56	4 (796)	18% 0	0.23 (0.01, 4.53)
			GG vs A	0.15 (-0.08, 0.37)	.20	7 (1090)	44% 0	1.10 (0.65, 1.87)

Note: "T&F": "Trim and Fill" method to assess potential publication bias. 0 = no missing study (no evidence of publication bias); negative value = # missing studies favoring the major or minor allele carriers in the comparison; positive value = # missing studies favoring the homozygotes in the comparison. Bolded results: $P \leq .05$. "Additive": additive genetic model with the risk allele in the parenthesis. "Category & Score": categories and scores of the strength of evidence based on the modified Venice guideline.²⁴

the gene is located in the X chromosome, C hemizyosity in males is equivalent to the CC genotype in females. Because the T allele is relatively rare (frequency = 12%), it was not possible to meta-analyze TT vs C carriers. The pooled ES from 20 studies was small (Hedges' $g = 0.23$, $P = .017$, with significant heterogeneity; table 3; supplementary figure 1). Although meta-regression analysis showed that studies with larger sample sizes had smaller ES ($P = .003$), this finding might be confounded by the fact that larger studies tended to be in chronic patients. Therefore, a series of subgroup analyses was conducted to further dissect the heterogeneity.

When studies were divided into subgroups based on treatment duration, the short-term studies (4–8 wk) produced larger ESs, (Hedges' $g = 0.44$, $P = .004$, studies=12, $n = 1209$). Although there was no evidence of publication bias, the heterogeneity across studies was still high ($I^2 = 76.1\%$). When further classifying samples into chronic vs first-episode patients, the first-episode samples with short-term follow-up produced the largest ES (Hedge's $g = 0.67$, $P = .002$, studies = 6, $n = 589$) compared to chronic samples (Hedge's $g = 0.21$, $P = .27$, studies = 6, $n = 620$), with a significant between-group difference ($Q = 9.74$, $df = 1$, $P = .002$; supplementary figure 2). In contrast, the studies with longer-term duration (3–4 mo, and 6 mo to 1 y) did not produce significant pooled ESs, and there was no difference between first-episode and chronic samples, although the overall trend favored the CC genotype. When mixing studies with different treatment lengths, the pooled ES in first-episode studies (Hedges' $g = 0.47$, $P = .02$, studies = 8, $n = 810$) was significantly higher than in chronic samples (Hedges' $g = 0.14$, $P = .18$, studies=12, $n = 1280$), $Q = 4.75$, $df = 1$, $P = .03$). Results for the categorical outcome variable were similar, but to some extent, less significant. Moderator analyses of race, sex, and specific antipsychotic were not significant.

The above-mentioned results were similar for rs518147 (table 3), but overall ES's were smaller. Due to the small number of included studies, no meta-regression or moderator analysis was conducted. The meta-analytic findings for rs6318 were inconsistent, in that the primary outcome was not significant, but the secondary outcome was significant with evidence of publication bias (table 3).

Dopamine Receptor D2 Gene. Three of seven SNPs in *DRD2* included in the analysis showed significant associations with weight gain (table 3). For rs1799732, an additive model of the risk allele (deletion of C) was identified, ie, each additional risk allele was associated with a 0.31 SD of extra weight gain, $P = .01$. For rs6275, the T-allele was the risk allele because TT homozygotes gained more weight than C carriers (Hedges' $g = 0.29$, $P = .02$) and CC homozygotes gained less weight than T carriers (Hedges' $g = -0.35$, $P < .01$). Both heterogeneity and publication bias were minimal. Similar findings were observed for

rs7131056 (table 3). Due to the small number of studies, moderator analyses were not performed.

ADRA2A Gene and GNB3 Gene. One SNP in *ADRA2A*, rs1800544, was significantly associated with weight gain across 6 studies, with the G allele increasing the risk. Both heterogeneity and publication bias were minimal (table 3; supplementary figure 4). Similarly, 1 SNP in *GNB3*, rs5443, was associated with weight gain across 10 studies. TT homozygotes of this SNP gained significantly more weight than the C carriers (Hedges' $g = 0.26$, $P = .01$; table 3; supplementary figure 5). Heterogeneity was small, and there was no evidence of publication bias. When analyzing 5 short-term studies only (treatment durations ≤ 3 mo), the pooled ES became even larger (Hedges' $g = 0.39$, $P < .001$, studies = 5, $n = 486$).

MC4R Gene and INSIG2 Gene. Two SNPs near *MC4R* were included in the meta-analysis, and one of them (rs489693) was significantly associated with weight gain in 6 studies. AA homozygotes of this SNP gained more weight than the C allele carriers (Hedges' $g = 0.80$, $P = .009$; table 3; supplementary figure 3). Heterogeneity across studies was high, and there was publication bias. However, even after adjusting for potentially missing studies, the association remained significant (Hedges' $g = 0.66$, $P < .05$). The ZHH FE cohort was an extreme outlier. The pooled ES became more significant after dropping this sample (Hedges' $g = 1.05$, $P = 1.9 \times 10^{-7}$). There was no significant moderator variable. Similarly, only 1 of 4 SNPs in *INSIG2*, rs17047764, was significantly associated with weight gain ($P = .048$, table 3). After adjusting for potential publication bias, the pooled ES remained significant.

Polygenic risk scores (PRS) were computed combining 6 top SNPs from *HTR2C*, *DRD2*, *ADRA2A*, *GNB3*, *MC4R*, and *INSIG2*. Number of risk alleles in each SNP (ie, 0, 1, or 2) was multiplied by its pooled ES from the additive model (table 3), and the sum of the 6 products was the PRS. This PRS explained 5.6% of the total variance in weight gain in each of the SATIETY and EUFEST cohorts, $P_s < .01$ (supplementary figure 6).

ADRB3, BDNF, and SNAP25. One SNP from each of these 3 genes was significantly associated with weight gain, but either the sample size was small (studies = 2 for rs4994 in *ADRB3* and rs1051312 in *SNAP25*), or the primary outcome was not significant (rs6265 in *BDNF*; table 3).

Discussion

To our best knowledge, this is the first comprehensive meta-analytic review of pharmacogenetics of antipsychotic-related weight gain that examined quantitatively multiple genetic variants. We investigated 38 SNPs in

20 genes or genetic regions distributed in 15 chromosomes in association with antipsychotic-related weight gain in 6770 patients from 46 non-overlapping samples published in 72 reports and including patient-level data from 3 cohorts providing unpublished data on 33 SNPs that were added to the meta-analysis. We found that 13 SNPs from 9 genes (*ADRA2A*, *ADRB3*, *BDNF*, *DRD2*, *GNB3*, *HTR2C*, *INSIG2*, *MC4R*, and *SNAP25*) were significantly associated with antipsychotic-related weight gain. Among these genes, *HTR2C* was most consistently associated with antipsychotic-related weight gain, and there was moderate evidence supporting the association of *ADRA2A*, *DRD2*, *GNB3*, *MC4R*, and *INSIG2*, based on the modified Venice guidelines. Relationships of other genes (*ADRB3*, *BDNF* and *SNAP25*) with antipsychotic-related weight gain were less consistent. Finally, polygenic scores using 6 SNPs seemed to explain a small proportion of weight gain.

With the widespread use of SGAs, weight gain and related metabolic syndrome have become a significant public health issue.²⁵ Weight gain is especially prominent in young and antipsychotic-naïve patients.^{5,16,26} Extensive effort has been made to understand the pathophysiological mechanisms of drug-related weight gain, and genetic variations seem to play a significant role.^{10,13} The present study aimed at providing a comprehensive and quantitative review of the literature on pharmacogenetics of antipsychotic drug-related weight gain. Previously, meta-analyses of pharmacogenetic association of single genes with antipsychotic-related weight gain have been published, but methodological issues limited their conclusions. First, many previous meta-analyses included cross-sectional studies where obesity or 1-time assessments of weight were correlated with a genetic variant. Body weight is determined by multiple factors including genetic, behavioral and medication effects.⁹ Without longitudinal assessments of weight change during antipsychotic treatment, the alleged genotype-phenotype association may be confounded by other, unmeasured variables. For example, it has been shown in multiple GWAS that *FTO* is associated with obesity in the general populations,²⁷ and most of these studies had 1-time measurements of obesity. However, in longitudinal studies of weight gain, *FTO* was not associated with antipsychotic-related weight gain in 7 studies (table 3). In the present meta-analysis, we included only longitudinal studies. Second, previous meta-analyses tended to pool studies of different treatment durations. ESs of a particular genetic variant on weight gain may be different at different time points. Moreover, antipsychotic-related weight gain is asymptotic and it is unclear when the weight gain begins to plateau, which also depends on the degree of prior antipsychotic-related weight gain. Mixing studies of different treatment durations may bias assessments of the true effect size. In the present review, we preferred time points closest to 2–3 months and we analyzed specific time

points whenever enough studies were available. In addition, previous meta-analyses tended to mix studies with chronic and antipsychotic-naïve or first-episode samples with minimal prior antipsychotic exposure. Studying patients with minimal drug exposure in pharmacogenetics minimizes order effects and increases the signal-to-noise ratio.¹⁴ In the present review, we added unpublished data from 3 first-episode/drug-naïve cohorts, and separated chronic samples from first-episode/antipsychotic-naïve samples whenever possible.

Results for Pharmacodynamic Targets of Antipsychotic Drugs

HTR2C showed the most consistent association with antipsychotic-related weight gain, with multiple SNPs showing this association and in a large number of studies, including both primary (continuous variable) and secondary (categorical) outcomes. Not only is it one of the first studied genes in antipsychotic-related weight gain, it is also one of the most studied genes. The *HTR2C* gene encodes the 2C subtype of serotonin receptor (5-HT_{2C}) and is located on the Chromosome Xq24. Experimental studies demonstrated the relevance of 5-HT_{2C} receptors in regulating appetite and food intake,²⁸ and 5-HT_{2C} agonists, such as dexfenfluramine and lorcaserin, can decrease food intake, resulting in significant weight loss.²⁹ 5-*HTR2C* antagonists, including many antipsychotics, may increase food intake, despite satiety, causing weight gain in animal models.^{30,31} Mice deficient in 5-HT_{2C} develop hyperphagia leading to obesity.^{32,33} The most studied SNP in *HTR2C*, rs3813929, is located in the promoter region of the gene and may play a role in regulating gene expression. Several studies found that the T-allele is associated with increased transcriptional activity of the gene, compared to the C allele.^{34,35} Although the C-allele is the major allele in the population, the T-allele seems to be protective against antipsychotic-related weight gain in the meta-analysis, perhaps by enhancing gene expression of *HTR2C*, which partially counterbalances the 5-HT_{2C} antagonistic antipsychotic effect. This SNP is located in the beginning of a CpG island (83 CpG count, chromosome X:113818520-113819453, UCSC Genome Browser) in the promoter region of *HTR2C*, suggesting that DNA methylation pattern variation may play a role in the SNP effect. Another SNP, rs518147, is only 62bp away from rs3813929. These 2 SNPs may be in high linkage disequilibrium,³⁶ representing probably the same signal. The third SNP in the *HTR2C* gene, rs6318, is located in an exon about 150kb from the first 2 SNPs. It is a missense SNP resulting in cysteine to serine substitution in position 23 of the protein sequence, which may disrupt a disulfide bridge affecting the receptor function.³⁷

DRD2 had a robust association with antipsychotic-related weight gain. Three of seven SNPs included in the meta-analysis showed a significant relationship, although

the number of studies and total sample sizes were not large. Being the main pharmacodynamic target of antipsychotics,³⁸ variations in *DRD2* function are plausible in explaining antipsychotic-related adverse events. At least 2 of the 3 SNPs in *DRD2*, located in Chromosome 11q23.2, rs1799732 (-141C Ins/Del) and rs6275, may be functional polymorphisms. rs1799732 represents a deletion (vs insertion) of cytosine at position -141, located in the 5' promoter region of *DRD2*. In vitro data showed that cell lines transfected with the Del allele were less active in a luciferase reporter assay than cell lines transfected with the Ins allele.³⁹ In vivo data with PET imaging also suggested that this polymorphism may influence D₂ receptor density in the striatum of healthy volunteers unexposed to antipsychotics.⁴⁰ A previous meta-analysis demonstrated that rs1799732 is associated with antipsychotic efficacy.⁴¹ rs6275 (C939T) is a synonymous polymorphism and is in close proximity and high linkage disequilibrium with rs6277 (C957T). Although not resulting in an amino acid sequence change of the D₂ receptor protein, the T-allele of rs6277 is associated with down-regulated D₂ receptors in the striatum⁴² and decreased *DRD2* mRNA stability and half-life.⁴³ Dopaminergic pathways are involved in the brain reward circuitry and modulate motivation, sense of well-being, and feeding behavior.⁴⁴ Many *DRD2* polymorphisms are associated with drug addiction, nicotine consumption, and eating disorders.⁴⁵ In animal studies, D₂ receptor availability in the striatum was significantly lower in obese than lean rats. In human studies, the availability of the striatal dopamine transporter was negatively correlated with BMI in healthy volunteers.⁴⁶ These results led researchers to hypothesize that a hypodopaminergic reward circuitry, which may be caused by the D₂ antagonism from antipsychotic treatment, results in abnormal over-eating and obesity.⁴⁴ It is possible that certain variants of *DRD2*, such as the Del allele of rs1799732 or the T-allele of rs6275 and rs6277, may already produce fewer D₂ receptors, and the subsequent over-eating behavior and weight gain are exacerbated by the use of antipsychotics.

One SNP in *ADRA2A*, rs1800544, was consistently associated with antipsychotic-related weight gain. It is located in the upstream of *ADRA2A*, and may be a binding site for transcription factors (ENCODE, UCSC Genome Browser). Just like 5-HT_{2C} and D₂, the alpha-2A receptor is a pharmacodynamic target of many antipsychotics, especially SGAs, including risperidone, olanzapine, and clozapine.⁴⁷ Interestingly, the adrenergic system also innervates adipose tissue.⁴⁸ Beta-3 adrenergic receptors in the brown adipose tissue are involved in production of heat (thermogenesis or fat burning),⁴⁹ whereas alpha-receptors have an inhibitory effect on lipolysis in the adipose tissue.⁵⁰ The SNP of interest, rs1800544, was associated with body fat accumulation⁵¹ in a large epidemiological study and affected plasma concentrations of glucose, insulin and cortisol in a human experimental

study.⁵² However, it is not clear how the different alleles affect receptor density. Notably, the allele frequency is different between racial groups in that G is the minor allele in CEU (minor allele frequency [MAF] = 27.5%) but the C-allele is the minor allele in Asians (MAF = 27.5%) and Africans (23.7%). To add to the complexity, some of the included studies did not specify whether genotyping was done along the positive or negative DNA strand. In the present meta-analysis, an effort was made to align the correct allele across studies. Nevertheless, caution is warranted when interpreting these findings.

Results for Genes Implicated in Obesity in the General Population

GNB3 also seems to be consistently associated with antipsychotic-related weight gain. In 10 studies ($n = 1004$), the TT homozygotes of rs5443 gained more weight than the C-allele carriers, with minimal heterogeneity across studies and no evidence of publication bias. Heterotrimeric G-proteins are important regulators of intracellular signaling pathways⁵³ and the beta-3 subunit is encoded by the *GNB3* gene. The SNP, rs5443 (C825T), is located on exon 10 of the *GNB3* gene. The T-allele is associated with alternative splicing of *GNB3* transcription, which results in enhanced signal transduction,⁵⁴ and the T-allele carries a higher risk of cardiovascular disease⁵⁵ and obesity⁵⁶ in the general population. Increased signaling by G-proteins stimulates adipogenesis and may lead to obesity.⁵⁶ This SNP seems to be associated with antidepressant efficacy,⁵⁷ but it is unclear whether antipsychotics interact with the G-protein subunit directly. It is possible that the SNP moderates the effect of antipsychotics on weight gain. Further studies are warranted to elucidate underlying mechanisms.

Another gene that has moderate evidence for its association with antipsychotic-related weight gain is *MC4R*, located in Chromosome 18q21. In several genome-wide association studies,^{58–60} a genomic locus near *MC4R* was associated with obesity in the general population. *MC4R* mutations have been linked to extreme early-onset obesity,⁶¹ and *MC4R* knockout mice develop obesity.⁶² The SNP, rs489693, was approaching genome-wide significance in association with antipsychotic-related weight gain in a pediatric antipsychotic-naïve cohort, and this effect was replicated in 3 independent samples.²⁰ The present meta-analysis added 1 unpublished cohort and 1 published study to the 4 samples. Despite heterogeneity and potential publication bias, AA homozygotes gained significantly more weight than C carriers, with a moderate effect size. *MC4R* neurons in the hypothalamic paraventricular nucleus, activated by α -melanocyte stimulating hormone (α -MSH) produced by the pro-opiomelanocortin (POMC)-expressing neurons, can induce decreased food intake and increased energy expenditure.⁶³ POMC neurons are partly controlled by serotonergic signals via

the 2-HT2C receptors.⁶⁴ Studies have shown that deficits in either *MC4R*,⁶¹ *POMC*,⁶⁵ prohormone convertase 1/3 (one of the key enzymes that converts POMC to α -MSH, encoded by the *PCSK1* gene),^{66,67} or 5-HT2C⁶⁸ resulted in obesity or drug-induced weight gain. Therefore, the POMC pathway may be an important mechanism. In addition, *MC4R* seems to interact with multiple other neurotransmitter pathways, including dopamine, leptin, and BDNF, in regulating appetite, eating, and energy homeostasis.⁶⁹

In contrast to *MC4R*, other genes that have been significantly associated with obesity in the general population or that are involved in appetite regulation and energy homeostasis, including *FTO*, *LEP* (Leptin), *LEPR* (Leptin receptor), *BDNF* and *INSIG2*, were not consistently associated with antipsychotic-related weight gain. In the present meta-analysis, genes that are direct pharmacodynamic targets of antipsychotics were more likely significantly associated with weight gain, except for *MC4R* and *GNB3*. Perhaps, genes that are involved in metabolic regulation, energy homeostasis, and appetite control may impact upon the downstream effects of antipsychotic actions, therefore, it may be more difficult to find significant associations with antipsychotic-induced adverse events.

Several limitations of this meta-analysis must be considered. First, many results were heterogeneous across studies, but for most SNPs the number of studies was not large enough to perform moderator or meta-regression analyses. However, we were able to conduct these analyses for a few SNPs with ≥ 8 studies. The most significant finding is that pooled ESs tended to be larger in studies with short-term follow-up and those including patients with minimal prior antipsychotic exposure. This finding was true for both *HTR2C* and *GNB3*. The diminishing gene effects during longer-term antipsychotic treatment and follow-up could be explained by a diminishing weight gain signal overall and/or greater contributions of behavioral and environmental factors, including changes in diet and exercise or antipsychotic adherence. Thus, although we attempted to examine more homogeneous samples in terms of patient populations, prior antipsychotic exposure and follow-up duration, the analyzed studies remained heterogeneous. One source of heterogeneity may come from variability in ancestry background, which may present different allele structures and was not well accounted for in meta-analysis. We have attempted to run subgroup analysis in different racial groups whenever possible, but no significant difference was found. Future studies should always include a 2- to 3-month time point, and a greater focus on antipsychotic-naïve and first episode patients would be helpful. Interestingly, the pooled effect size for rs7700039 (–2548A/G) in *LEP* was only marginally significant in 10 studies, and examining the studies with short-term durations and first-episode patients failed to improve the overall ES, suggesting that perhaps

LEP does not play an important role in antipsychotic-related weight gain. Additionally, we detected a significant publication bias for several results, as is expected in mostly single-gene studies. However, the overall bias seemed conservative, as generally the ES increased when potentially missing studies were imputed. Another issue is that although we attempted to evaluate the strength of evidence on the phenotype-genotype association based on the modified Venice criteria, it needs to be acknowledged that the Venice criteria was not designed for pharmacogenetic studies. Therefore, interpretation of these findings should be cautious. Finally, the antipsychotics used in each study were heterogeneous, which may limit the detection of pharmacogenetic signals. Many studies examined clozapine and olanzapine, which confer the highest weight gain liability and are pharmacologically different from other antipsychotics. The power of detecting a signal may be increased by studying a single agent or agents with similar weight gain properties.²⁰

In summary, the present meta-analysis attempted to overcome the limitations of previous reviews and comprehensively examined all genetic variants deemed relevant in association with antipsychotic-related weight gain. Several genes, including *HTR2C*, *DRD2*, *ADRA2A*, *MC4R* and *GNB3* seem to be consistently associated with antipsychotic-related weight gain. ES were larger in patients with minimal prior antipsychotic exposure and in short-term studies. Despite promising findings, the ES of individual SNPs and genes are too small to fulfill the promise of personalized medicine. Because antipsychotic-related weight gain is likely polygenic and affected by environmental factors,⁹ future studies should carefully consider study design issues^{14,70} and explore combining multiple genetic markers and relevant clinical factors to improve clinical prediction.

Supplementary Material

Supplementary material is available at <http://schizophreniabulletin.oxfordjournals.org>.

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References

1. Kane JM, Correll CU. Pharmacologic treatment of schizophrenia. *Dialogues Clin Neurosci*. 2010;12:345–357.
2. Galling B, Garcia MA, Osuchukwu U, Hagi K, Correll CU. Safety and tolerability of antipsychotic-mood stabilizer co-treatment in the management of acute bipolar disorder: results from a systematic review and exploratory meta-analysis. *Expert Opin Drug Saf*. 2015;14:1181–1199.
3. American Psychiatric Association. *Practice Guideline for the Treatment of Patients With Major Depressive Disorder [Internet]*. Washington, DC: American Psychiatric Association; 2010. www.psychiatryonline.com. Accessed July 15, 2015.
4. Farahani A, Correll CU. Are antipsychotics or antidepressants needed for psychotic depression? A systematic review and meta-analysis of trials comparing antidepressant or antipsychotic monotherapy with combination treatment. *J Clin Psychiatry*. 2012;73:486–496.
5. De Hert M, Detraux J, van Winkel R, Correll CU. Metabolic and cardiovascular adverse effects associated with antipsychotic drugs. *Nat Rev Endocrinol*. 2011;8:114–126.
6. Correll CU, Joffe BI, Rosen LM, Sullivan TB, Joffe RT. Cardiovascular and cerebrovascular risk factors and events associated with second-generation antipsychotic compared to antidepressant use in a non-elderly adult sample: results from a claims-based inception cohort study. *World Psychiatry*. 2015;14:56–63.
7. Correll CU, Detraux J, De Lepeleire J, De Hert M. Effects of antipsychotics, antidepressants and mood stabilizers on risk for physical diseases in people with schizophrenia, depression and bipolar disorder. *World Psychiatry*. 2015;14:119–136.
8. Allison DB, Mentore JL, Heo M, et al. Antipsychotic-induced weight gain: a comprehensive research synthesis. *Am J Psychiatry*. 1999;156:1686–1696.
9. Correll CU, Lencz T, Malhotra AK. Antipsychotic drugs and obesity. *Trends Mol Med*. 2011;17:97–107.

10. Correll CU, Malhotra AK. Pharmacogenetics of antipsychotic-induced weight gain. *Psychopharmacology (Berl)*. 2004;174:477–489.
11. Fall T, Ingelsson E. Genome-wide association studies of obesity and metabolic syndrome. *Mol Cell Endocrinol*. 2014;382:740–757.
12. Zhang J-P, Malhotra AK. Pharmacogenetics and antipsychotics: therapeutic efficacy and side effects prediction. *Expert Opin Metabol Toxicol*. 2011;7:9–37.
13. Lett TA, Wallace TJ, Chowdhury NI, Tiwari AK, Kennedy JL, Müller DJ. Pharmacogenetics of antipsychotic-induced weight gain: review and clinical implications. *Mol Psychiatry*. 2012;17:242–266.
14. Malhotra AK, Zhang JP, Lencz T. Pharmacogenetics in psychiatry: translating research into clinical practice. *Mol Psychiatry*. 2012;17:760–769.
15. Need AC, Keefe RS, Ge D, Grossman I, Dickson S, McEvoy JP, Goldstein DB. Pharmacogenetics of antipsychotic response in the CATIE trial: a candidate gene analysis. *Eur J Hum Genet*. 2009;17:946–957.
16. Correll CU, Manu P, Olshanskiy V, Napolitano B, Kane JM, Malhotra AK. Cardiometabolic risk of second-generation antipsychotic medications during first-time use in children and adolescents. *JAMA*. 2009;302:1765–1773.
17. Robinson DG, Woerner MG, Napolitano B, et al. Randomized comparison of olanzapine versus risperidone for the treatment of first-episode schizophrenia: 4-month outcomes. *Am J Psychiatry*. 2006;163:2096–2102.
18. Lencz T, Robinson DG, Xu K, et al. DRD2 promoter region variation as a predictor of sustained response to antipsychotic medication in first-episode schizophrenia patients. *Am J Psychiatry*. 2006;163:529–531.
19. Kahn RS, Fleischhacker WW, Boter H, et al. Effectiveness of antipsychotic drugs in first-episode schizophrenia and schizophreniform disorder: an open randomised clinical trial. *Lancet*. 2008;371:1085–1097.
20. Malhotra AK, Correll CU, Chowdhury NI, et al. Association between common variants near the Melanocortin 4 Receptor Gene and severe antipsychotic drug-induced weight gain. *Arch Gen Psychiatry*. 2012;69:904–912.
21. Borenstein M, Hedges LV, Higgins JPT, Rothstein HR. *Introduction to Meta-Analysis*. Chichester, UK: John Wiley & Sons, Ltd; 2009.
22. Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. 1997;315:629–634.
23. Duval SJ, Tweedie RL. A non-parametric “trim and fill” method of assessing publication bias in meta-analysis. *J Am Stat Assoc*. 2000;95:89–98.
24. Ioannidis JP, Boffetta P, Little J, et al. Assessment of cumulative evidence on genetic associations: interim guidelines. *Int J Epidemiol*. 2008;37:120–132.
25. DE Hert M, Correll CU, Bobes J, et al. Physical illness in patients with severe mental disorders. I. Prevalence, impact of medications and disparities in health care. *World Psychiatry*. 2011;10:52–77.
26. Fleischhacker WW, Siu CO, Bodén R, Pappadopulos E, Karayal ON, Kahn RS; EUFEST Study Group. Metabolic risk factors in first-episode schizophrenia: baseline prevalence and course analysed from the European First-Episode Schizophrenia Trial. *Int J Neuropsychopharmacol*. 2013;16:987–995.
27. Fall T, Ingelsson E. Genome-wide association studies of obesity and metabolic syndrome. *Mol Cell Endocrinol*. 2014;382:740–757.
28. Vickers SP, Easton N, Webster LJ, et al. Oral administration of the 5-HT_{2C} receptor agonist, mCPP, reduces body weight gain in rats over 28 days as a result of maintained hypophagia. *Psychopharmacology (Berl)*. 2003;167:274–280.
29. Smith SR, Weissman NJ, Anderson CM, et al. Multicenter, placebo-controlled trial of lorcaserin for weight management. *N Engl J Med*. 2010;363:245–256.
30. Simansky KJ. Serotonergic control of the organization of feeding and satiety. *Behav Brain Res*. 1996;73:37–42.
31. Bonhaus DW, Weinhardt KK, Taylor M, et al. RS-102221: a novel high affinity and selective, 5-HT_{2C} receptor antagonist. *Neuropharmacology*. 1997;36:621–629.
32. Tecott LH, Sun LM, Akana SF, et al. Eating disorder and epilepsy in mice lacking 5-HT_{2c} serotonin receptors. *Nature*. 1995;374:542–546.
33. Nonogaki K, Strack AM, Dallman MF, Tecott LH. Leptin-independent hyperphagia and type 2 diabetes in mice with a mutated serotonin 5-HT_{2C} receptor gene. *Nature Med*. 1998;4:1152–1156.
34. Yuan X, Yamada K, Ishiyama-Shigemoto S, Koyama W, Nonaka K. Identification of polymorphic loci in the promoter region of the serotonin 5-HT_{2C} receptor gene and their association with obesity and type II diabetes. *Diabetologia*. 2000;43:373–376.
35. Buckland PR, Hoogendoorn B, Guy CA, Smith SK, Coleman SL, O'Donovan MC. Low gene expression conferred by association of an allele of the 5-HT_{2C} receptor gene with antipsychotic-induced weight gain. *Am J Psychiatry*. 2005;162:613–615.
36. Godlewska BR, Olajossy-Hilkesberger L, Ciwoniuk M, et al. Olanzapine-induced weight gain is associated with the -759C/T and -697G/C polymorphisms of the HTR_{2C} gene. *Pharmacogenomics J*. 2009;9:234–241.
37. Drago A, Serretti A. Focus on HTR_{2C}: a possible suggestion for genetic studies of complex disorders. *Am J Med Genet B Neuropsychiatr Genet*. 2009;150B:601–637.
38. Kapur S, Mamo D. Half a century of antipsychotics and still a central role for dopamine D₂ receptors. *Prog Neuropsychopharmacol Biol Psychiatry*. 2003;27:1081–1090.
39. Arinami T, Gao M, Hamaguchi H, Toru M. A functional polymorphism in the promoter region of the dopamine D₂ receptor gene is associated with schizophrenia. *Hum Mol Genet*. 1997;6:577–582.
40. Ritchie T, Noble EP. Association of seven polymorphisms of the D₂ dopamine receptor gene with brain receptor-binding characteristics. *Neurochemical Res*. 2003;28:73–82.
41. Zhang JP, Lencz T, Malhotra AK. D₂ receptor genetic variation and clinical response to antipsychotic drug treatment: a meta-analysis. *Am J Psychiatry*. 2010;167:763–772.
42. Hirvonen MM, Laakso A, Nägren K, Rinne JO, Pohjalainen T, Hietala J. C957T polymorphism of dopamine D₂ receptor gene affects striatal DRD₂ in vivo availability by changing the receptor affinity. *Synapse*. 2009;63:907–912.
43. Duan J, Wainwright MS, Cameron JM, et al. Synonymous mutations in the human dopamine receptor D₂ (DRD₂) affect mRNA stability and synthesis of the receptor. *Hum Mol Genet*. 2003;12:205–216.
44. Blum K, Thanos PK, Gold MS. Dopamine and glucose, obesity, and reward deficiency syndrome. *Front Psychol*. 2014;5:919.
45. Hamdi A, Porter J, Prasad C. Decreased striatal D₂ dopamine receptors in obese Zucker rats: changes during aging. *Brain Res*. 1992;589:338–340.

46. Chen PS, Yang YK, Yeh TL, et al. Correlation between body mass index and striatal dopamine transporter availability in healthy volunteers--a SPECT study. *NeuroImage*. 2008;40:275–279.
47. Miyamoto S, Duncan GE, Marx CE, Lieberman JA. Treatments for schizophrenia: a critical review of pharmacology and mechanisms of action of antipsychotic drugs. *Mol Psychiatry*. 2005;10:79–104.
48. Snitker S, Macdonald I, Ravussin E, Astrup A. The sympathetic nervous system and obesity: role in aetiology and treatment. *Obes Rev*. 2000;1:5–15.
49. Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. *Physiol Rev*. 2004;84:277–359.
50. van Baak MA. The peripheral sympathetic nervous system in human obesity. *Obes Rev*. 2001;2:3–14.
51. Garenc C, Perusse L, Chagnon YC, et al. The alpha 2-adrenergic receptor gene and body fat content and distribution: the HERITAGE Family Study. *Mol Med*. 2002;8:88–94.
52. Rosmond R, Bouchard C, Bjorntorp P. A C-1291G polymorphism in the alpha2A-adrenergic receptor gene (ADRA2A) promoter is associated with cortisol escape from dexamethasone and elevated glucose levels. *J Inter Med*. 2002;251:252–257.
53. Kozasa T, Hajicek N, Chow CR, Suzuki N. Signalling mechanisms of RhoGTPase regulation by the heterotrimeric G proteins G12 and G13. *J Biochem*. 2011;150:357–369.
54. Siffert W, Roszkopf D, Siffert G, et al. Association of a human G-protein beta3 subunit variant with hypertension. *Nat Genet*. 1998;18:45–48.
55. Zethelius B, Berglund L, Sundstrom J, et al. Use of multiple biomarkers to improve the prediction of death from cardiovascular causes. *N Engl J Med*. 2008;358:2107–2116.
56. Siffert W. G protein polymorphisms in hypertension, atherosclerosis, and diabetes. *Annu Rev Med*. 2005;56:17–28.
57. Hu Q, Zhang SY, Liu F, et al. Influence of GNB3 C825T polymorphism on the efficacy of antidepressants in the treatment of major depressive disorder: a meta-analysis. *J Affect Disord*. 2014;172C:103–109.
58. Chambers JC, Elliott P, Zabaneh D, et al. Common genetic variation near MC4R is associated with waist circumference and insulin resistance. *Nat Genet*. 2008;40:716–718.
59. Loos RJ, Lindgren CM, Li S, et al. Common variants near MC4R are associated with fat mass, weight and risk of obesity. *Nat Genet*. 2008;40:768–775.
60. Zobel DP, Andreasen CH, Grarup N, et al. Variants near MC4R are associated with obesity and influence obesity-related quantitative traits in a population of middle-aged people: studies of 14,940 Danes. *Diabetes*. 2009;58:757–764.
61. Farooqi IS, O'Rahilly S. Monogenic obesity in humans. *Annu Rev Med*. 2005;56:443–458.
62. Huszar D, Lynch CA, Fairchild-Huntress V, et al. Targeted disruption of the melanocortin-4 receptor results in obesity in mice. *Cell*. 1997;88:131–141.
63. Kim JD, Leyva S, Diano S. Hormonal regulation of the hypothalamic melanocortin system. *Front Physiol*. 2014;5:480.
64. Yeo GS, Heisler LK. Unraveling the brain regulation of appetite: lessons from genetics. *Nature Neurosci*. 2012;15:1343–1349.
65. Krude H, Gruters A. Implications of proopiomelanocortin (POMC) mutations in humans: the POMC deficiency syndrome. *Trends Endocrinol Metab*. 2000;11:15–22.
66. Benzinou M, Creemers JW, Choquet H, et al. Common non-synonymous variants in PCSK1 confer risk of obesity. *Nat Genet*. 2008;40:943–945.
67. Kilpelainen TO, Bingham SA, Khaw KT, Wareham NJ, Loos RJ. Association of variants in the PCSK1 gene with obesity in the EPIC-Norfolk study. *Hum Mol Genet*. 2009;18:3496–3501.
68. Reynolds GP, Zhang ZJ, Zhang XB. Association of antipsychotic drug-induced weight gain with a 5-HT2C receptor gene polymorphism. *Lancet*. 2002;359:2086–2087.
69. Harrold JA, Williams G. Melanocortin-4 receptors, beta-MSH and leptin: key elements in the satiety pathway. *Peptides*. 2006;27:365–371.
70. Hamilton SP. The Promise of Psychiatric Pharmacogenomics. *Biol Psychiatry*. 2015;77:29–35.
71. Basile VS, Masellis M, McIntyre RS, Meltzer HY, Lieberman JA, Kennedy JL. Genetic dissection of atypical antipsychotic-induced weight gain: novel preliminary data on the pharmacogenetic puzzle. *J Clin Psychiatry*. 2001;62(suppl 23):45–66.
72. Basile VS, Masellis M, De Luca V, Meltzer HY, Kennedy JL. 759C/T genetic variation of 5HT(2C) receptor and clozapine-induced weight gain. *Lancet*. 2002;360:1790–1791.
73. Bishop JR, Ellingrod VL, Moline J, Miller D. Pilot study of the G-protein beta3 subunit gene (C825T) polymorphism and clinical response to olanzapine or olanzapine-related weight gain in persons with schizophrenia. *Med Sci Monit*. 2006;12:BR47–BR50.
74. Brandl EJ, Frydrychowicz C, Tiwari AK, et al. Association study of polymorphisms in leptin and leptin receptor genes with antipsychotic-induced body weight gain. *Prog Neuropsychopharmacol Biol Psychiatry*. 2012;38:134–141.
75. Calarge CA, Ellingrod VL, Zimmerman B, Acion L, Sivitz WI, Schlechte JA. Leptin gene -2548G/A variants predict risperidone-associated weight gain in children and adolescents. *Psychiatr Genet*. 2009;19:320–327.
76. Chowdhury NI, Tiwari AK, Souza RP, et al. Genetic association study between antipsychotic-induced weight gain and the melanocortin-4 receptor gene. *Pharmacogenomics J*. 2013;13:272–279.
77. Czerwensky F, Leucht S, Steimer W. Association of the common MC4R rs17782313 polymorphism with antipsychotic-related weight gain. *J Clin Psychopharmacol*. 2013;33:74–79.
78. Czerwensky F, Leucht S, Steimer W. MC4R rs489693: a clinical risk factor for second generation antipsychotic-related weight gain? *Int J Neuropsychopharmacol*. 2013;16:2103–2109.
79. Ellingrod VL, Perry PJ, Ringold JC, et al. Weight gain associated with the -759C/T polymorphism of the 5HT2C receptor and olanzapine. *Am J Med Genet B Neuropsychiatr Genet*. 2005;134B:76–78.
80. Ellingrod VL, Bishop JR, Moline J, Lin YC, Miller DD. Leptin and leptin receptor gene polymorphisms and increases in body mass index (BMI) from olanzapine treatment in persons with schizophrenia. *Psychopharmacol Bull*. 2007;40:57–62.
81. Fernandez E, Carrizo E, Fernandez V, et al. Polymorphisms of the LEP- and LEPR genes, metabolic profile after prolonged clozapine administration and response to the antidiabetic metformin. *Schizophrenia Res*. 2010;121:213–217.
82. Herken H, Erdal M, Aydin N, et al. The association of olanzapine-induced weight gain with peroxisome proliferator-activated receptor-gamma2 Pro12Ala polymorphism in patients with schizophrenia. *DNA Cell Biol*. 2009;28:515–519.
83. Hoekstra PJ, Troost PW, Lahuis BE, et al. Risperidone-induced weight gain in referred children with autism spectrum

- disorders is associated with a common polymorphism in the 5-hydroxytryptamine 2C receptor gene. *J Child Adolesc Psychopharmacol.* 2010;20:473–477.
84. Hong CJ, Lin CH, Yu YW, Yang KH, Tsai SJ. Genetic variants of the serotonin system and weight change during clozapine treatment. *Pharmacogenetics.* 2001;11:265–268.
 85. Hong CJ, Liou YJ, Bai YM, Chen TT, Wang YC, Tsai SJ. Dopamine receptor D2 gene is associated with weight gain in schizophrenic patients under long-term atypical antipsychotic treatment. *Pharmacogenet Genomics.* 2010;20:359–366.
 86. Houston JP, Kohler J, Bishop JR, et al. Pharmacogenomic associations with weight gain in olanzapine treatment of patients without schizophrenia. *J Clin Psychiatry.* 2012;73:1077–1086.
 87. Huang HH, Wang YC, Wu CL, et al. TNF-alpha -308 G>A polymorphism and weight gain in patients with schizophrenia under long-term clozapine, risperidone or olanzapine treatment. *Neurosci Lett.* 2011;504:277–280.
 88. Kang SG, Lee HJ, Park YM, et al. Possible association between the -2548A/G polymorphism of the leptin gene and olanzapine-induced weight gain. *Prog Neuropsychopharmacol Biol Psychiatry.* 2008;32:160–163.
 89. Kuzman MR, Medved V, Bozina N, Hotujac L, Sain I, Bilusic H. The influence of 5-HT(2C) and MDR1 genetic polymorphisms on antipsychotic-induced weight gain in female schizophrenic patients. *Psychiatry Res.* 2008;160:308–315.
 90. Kuzman MR, Medved V, Bozina N, Grubišić J, Jovanović N, Sertić J. Association study of MDR1 and 5-HT2C genetic polymorphisms and antipsychotic-induced metabolic disturbances in female patients with schizophrenia. *Pharmacogenomics J.* 2011;11:35–44.
 91. Laika B, Leucht S, Heres S, Schneider H, Steimer W. Pharmacogenetics and olanzapine treatment: CYP1A2*1F and serotonergic polymorphisms influence therapeutic outcome. *Pharmacogenomics J.* 2010;10:20–29.
 92. Lane HY, Liu YC, Huang CL, et al. Risperidone-related weight gain: genetic and nongenetic predictors. *J Clin Psychopharmacol.* 2006;26:128–134.
 93. Le Hellard S, Theisen FM, Haberhausen M, et al. Association between the insulin-induced gene 2 (INSIG2) and weight gain in a German sample of antipsychotic-treated schizophrenic patients: perturbation of SREBP-controlled lipogenesis in drug-related metabolic adverse effects? *Mol Psychiatry.* 2009;14:308–317.
 94. Lencz T, Robinson DG, Napolitano B, et al. DRD2 promoter region variation predicts antipsychotic-induced weight gain in first episode schizophrenia. *Pharmacogenet Genomics.* 2010;20:569–572.
 95. Lin YC, Ellingrod VL, Bishop JR, Miller DD. The relationship between P-glycoprotein (PGP) polymorphisms and response to olanzapine treatment in schizophrenia. *Ther Drug Monit.* 2006;28:668–672.
 96. Miller DD, Ellingrod VL, Holman TL, Buckley PF, Arndt S. Clozapine-induced weight gain associated with the 5HT2C receptor -759C/T polymorphism. *Am J Med Genet B Neuropsychiatr Genet.* 2005;133B:97–100.
 97. Monteleone P, Milano W, Petrella C, Canestrelli B, Maj M. Endocannabinoid Pro129Thr FAAH functional polymorphism but not 1359G/A CNR1 polymorphism is associated with antipsychotic-induced weight gain. *J Clin Psychopharmacol.* 2010;30:441–445.
 98. Mou XD, Zhang ZJ, Zhang XR, Shi JB, Sun J. [-2548G/A functional polymorphism in the promoter region of leptin gene and antipsychotic agent-induced weight gain in schizophrenic patients: a study of nuclear family-based association]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* 2008;33:316–320.
 99. Müller DJ, Klempan TA, De Luca V, et al. The SNAP-25 gene may be associated with clinical response and weight gain in antipsychotic treatment of schizophrenia. *Neurosci Lett.* 2005;379:81–89.
 100. Müller DJ, Zai CC, Sicard M, et al. Systematic analysis of dopamine receptor genes (DRD1-DRD5) in antipsychotic-induced weight gain. *Pharmacogenomics J.* 2012;12:156–164.
 101. Musil R, Spellmann I, Riedel M, et al. SNAP-25 gene polymorphisms and weight gain in schizophrenic patients. *J Psychiatr Res.* 2008;42:963–970.
 102. Opgen-Rhein C, Brandl EJ, Müller DJ, et al. Association of HTR2C, but not LEP or INSIG2, genes with antipsychotic-induced weight gain in a German sample. *Pharmacogenomics.* 2010;11:773–780.
 103. Park YM, Chung YC, Lee SH, et al. Weight gain associated with the alpha2a-adrenergic receptor -1,291 C/G polymorphism and olanzapine treatment. *Am J Med Genet B Neuropsychiatr Genet.* 2006;141B:394–397.
 104. Park YM, Cho JH, Kang SG, et al. Lack of association between the -759C/T polymorphism of the 5-HT2C receptor gene and olanzapine-induced weight gain among Korean schizophrenic patients. *J Clin Pharm Ther.* 2008;33:55–60.
 105. Park YM, Chung YC, Lee SH, et al. G-protein beta3 subunit gene 825C/T polymorphism is not associated with olanzapine-induced weight gain in Korean schizophrenic patients. *Psychiatry Investig.* 2009;6:39–43.
 106. Park YM, Choi JE, Kang SG, et al. Cannabinoid type 1 receptor gene polymorphisms are not associated with olanzapine-induced weight gain. *Hum Psychopharmacol.* 2011;26:332–337.
 107. Perez-Iglesias R, Mata I, Amado JA, et al. Effect of FTO, SH2B1, LEP, and LEPR polymorphisms on weight gain associated with antipsychotic treatment. *J Clin Psychopharmacol.* 2010;30:661–666.
 108. Popp J, Leucht S, Heres S, Steimer W. DRD4 48bp VNTR but not 5-HT 2C Cys23Ser receptor polymorphism is related to antipsychotic-induced weight gain. *Pharmacogenomics J.* 2009;9:71–77.
 109. Reynolds GP, Zhang Z, Zhang X. Polymorphism of the promoter region of the serotonin 5-HT(2C) receptor gene and clozapine-induced weight gain. *Am J Psychiatry.* 2003;160:677–679.
 110. Reynolds GP, Yevtushenko OO, Gordon S, Arranz B, San L, Cooper SJ. The obesity risk gene FTO influences body mass in chronic schizophrenia but not initial antipsychotic drug-induced weight gain in first-episode patients. *Int J Neuropsychopharmacol.* 2013;16:1421–1425.
 111. Ryu S, Jang WS, Cho EY, Kim SK, Lee D, Hong KS. Association study of -2548A/g polymorphism of leptin gene with antipsychotics-induced weight gain. *Korean J Psychopharmacol.* 2006;17:423–428.
 112. Ryu S, Cho EY, Park T, et al. -759 C/T polymorphism of 5-HT2C receptor gene and early phase weight gain associated with antipsychotic drug treatment. *Prog Neuropsychopharmacol Biol Psychiatry.* 2007;31:673–677.
 113. Shao P, Zhao JP, Chen JD, Wu RR, He YQ. [Association of HTR2C-759C/T and -697G/C polymorphisms with antipsychotic agent-induced weight gain]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban.* 2008;33:312–315.
 114. Shing EC, Tiwari AK, Brandl EJ, et al. Fat mass- and obesity-associated (FTO) gene and antipsychotic-induced

- weight gain: an association study. *Neuropsychobiology*. 2014;69:59–63.
115. Sicard MN, Zai CC, Tiwari AK, et al. Polymorphisms of the HTR2C gene and antipsychotic-induced weight gain: an update and meta-analysis. *Pharmacogenomics*. 2010;11:1561–1571.
 116. Sickert L, Müller DJ, Tiwari AK, et al. Association of the alpha 2A adrenergic receptor -1291C/G polymorphism and antipsychotic-induced weight gain in European-Americans. *Pharmacogenomics*. 2009;10:1169–1176.
 117. Song X, Pang L, Feng Y, et al. Fat-mass and obesity-associated gene polymorphisms and weight gain after risperidone treatment in first episode schizophrenia. *Behav Brain Funct*. 2014;10:35.
 118. Souza RP, De Luca V, Muscettola G, et al. Association of antipsychotic induced weight gain and body mass index with GNB3 gene: a meta-analysis. *Prog Neuropsychopharmacol Biol Psychiatry*. 2008;32:1848–1853.
 119. Srisawat U, Reynolds GP, Zhang ZJ, et al. Methyltetrahydrofolate reductase (MTHFR) 677C/T polymorphism is associated with antipsychotic-induced weight gain in first-episode schizophrenia. *Int J Neuropsychopharmacol*. 2014;17:485–490.
 120. Staeker J, Leucht S, Steimer W. Peroxisome proliferator-activated receptor gamma (PPARG) Pro12Ala: lack of association with weight gain in psychiatric inpatients treated with olanzapine or clozapine. *Mol Diagn Ther*. 2012;16:93–98.
 121. Templeman LA, Reynolds GP, Arranz B, San L. Polymorphisms of the 5-HT2C receptor and leptin genes are associated with antipsychotic drug-induced weight gain in Caucasian subjects with a first-episode psychosis. *Pharmacogenet Genomics*. 2005;15:195–200.
 122. Theisen FM, Hinney A, Bromel T, et al. Lack of association between the -759C/T polymorphism of the 5-HT2C receptor gene and clozapine-induced weight gain among German schizophrenic individuals. *Psychiatr Genet*. 2004;14:139–142.
 123. Thompson A, Lavedan C, Volpi S. Absence of weight gain association with the HTR2C -759C/T polymorphism in patients with schizophrenia treated with iloperidone. *Psychiatry Res*. 2010;175:271–273.
 124. Tiwari AK, Zai CC, Meltzer HY, Lieberman JA, Müller DJ, Kennedy JL. Association study of polymorphisms in insulin induced gene 2 (INSIG2) with antipsychotic-induced weight gain in European and African-American schizophrenia patients. *Hum Psychopharmacol*. 2010;25:253–259.
 125. Tiwari AK, Zai CC, Likhodi O, et al. A common polymorphism in the cannabinoid receptor 1 (CNR1) gene is associated with antipsychotic-induced weight gain in Schizophrenia. *Neuropsychopharmacology*. 2010;35:1315–1324.
 126. Tsai SJ, Hong CJ, Yu YW, Lin CH. -759C/T genetic variation of 5HT(2C) receptor and clozapine-induced weight gain. *Lancet*. 2002;360:1790.
 127. Tsai SJ, Hong CJ, Yu YW, Lin CH, Liu LL. No association of tumor necrosis factor alpha gene polymorphisms with schizophrenia or response to clozapine. *Schizophr Res*. 2003;65:27–32.
 128. Tsai SJ, Yu YW, Lin CH, Wang YC, Chen JY, Hong CJ. Association study of adrenergic beta3 receptor (Trp64Arg) and G-protein beta3 subunit gene (C825T) polymorphisms and weight change during clozapine treatment. *Neuropsychobiology*. 2004;50:37–40.
 129. Tsai A, Liou YJ, Hong CJ, Wu CL, Tsai SJ, Bai YM. Association study of brain-derived neurotrophic factor gene polymorphisms and body weight change in schizophrenic patients under long-term atypical antipsychotic treatment. *Neuromolecular Med*. 2011;13:328–333.
 130. Ujike H, Nomura A, Morita Y, et al. Multiple genetic factors in olanzapine-induced weight gain in schizophrenia patients: a cohort study. *J Clin Psychiatry*. 2008;69:1416–1422.
 131. van Winkel R, Moons T, Peerbooms O, et al. MTHFR genotype and differential evolution of metabolic parameters after initiation of a second generation antipsychotic: an observational study. *Int Clin Psychopharmacol*. 2010;25:270–276.
 132. Wang YC, Bai YM, Chen JY, Lin CC, Lai IC, Liou YJ. Polymorphism of the adrenergic receptor alpha 2a -1291C>G genetic variation and clozapine-induced weight gain. *J Neural Transm (Vienna)*. 2005;112:1463–1468.
 133. Wang YC, Bai YM, Chen JY, Lin CC, Lai IC, Liou YJ. C825T polymorphism in the human G protein beta3 subunit gene is associated with long-term clozapine treatment-induced body weight change in the Chinese population. *Pharmacogenet Genomics*. 2005;15:743–748.
 134. Wang YC, Bai YM, Chen JY, Lin CC, Lai IC, Liou YJ. Genetic association between TNF-alpha -308 G>A polymorphism and longitudinal weight change during clozapine treatment. *Hum Psychopharmacol*. 2010;25:303–309.
 135. Zai GC, Zai CC, Chowdhury NI, et al. The role of brain-derived neurotrophic factor (BDNF) gene variants in antipsychotic response and antipsychotic-induced weight gain. *Prog Neuropsychopharmacol Biol Psychiatry*. 2012;39:96–101.
 136. Zhang ZJ, Yao ZJ, Zhang XB, et al. No association of antipsychotic agent-induced weight gain with a DA receptor gene polymorphism and therapeutic response. *Acta Pharmacol Sinica*. 2003;24:235–240.
 137. Zhang ZJ, Yao ZJ, Mou XD, et al. [Association of -2548G/A functional polymorphism in the promoter region of leptin gene with antipsychotic agent-induced weight gain]. *Zhonghua Yi Xue Za Zhi*. 2003;83:2119–2123.
 138. Zhang XY, Tan YL, Zhou DF, et al. Association of clozapine-induced weight gain with a polymorphism in the leptin promoter region in patients with chronic schizophrenia in a Chinese population. *J Clin Psychopharmacol*. 2007;27:246–251.
 139. Zhang XY, Zhou DF, Wu GY, et al. BDNF levels and genotype are associated with antipsychotic-induced weight gain in patients with chronic schizophrenia. *Neuropsychopharmacology*. 2008;33:2200–2205.