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## Leukocyte immunoglobulin-like receptor B4 regulates key signalling molecules involved in Fc $\gamma$ RI-mediated clathrin-dependent endocytosis and phagocytosis

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Fc $\gamma$ RI cross-linking on monocytes may trigger clathrin-mediated endocytosis, likely through interaction of multiple intracellular molecules that are controlled by phosphorylation and dephosphorylation events. However, the identity of phospho-proteins and their regulation are unknown. We proposed the leukocyte immunoglobulin-like receptor B4 (LILRB4) that inhibits Fc $\gamma$ RI-mediated cytokine production via Tyr dephosphorylation of multiple kinases, may also regulate endocytosis/phagocytosis through similar mechanisms. Fc $\gamma$ RI and/or LILRB4 were antibody-ligated on THP-1 cells, lysates immunoprecipitated using anti-pTyr antibody and peptides sequenced by mass spectrometry. Mascot Search identified 25 Tyr phosphorylated peptides with high confidence. Ingenuity Pathway Analysis revealed that the most significantly affected pathways were clathrin-mediated endocytosis and Fc-receptor dependent phagocytosis. Tyr phosphorylation of key candidate proteins in these pathways included common  $\gamma$ -chain of the Fc receptors, Syk, clathrin, E3 ubiquitin protein ligase Cbl, hepatocyte growth factor-regulated tyrosine kinase substrate, tripartite motif-containing 21 and heat shock protein 70. Importantly, co-ligation of LILRB4 with Fc $\gamma$ RI caused significant dephosphorylation of these proteins and was associated with suppression of Fc receptor-dependent uptake of antibody-opsonised bacterial particles, indicating that LILRB4. These results suggest that Tyr phosphorylation may be critical in Fc $\gamma$ RI-dependent endocytosis/phagocytosis that may be regulated by LILRB4 by triggering dephosphorylation of key signalling proteins.

Fc receptors (FcRs) are key molecules for recognition and elimination of foreign antigens through induction of multiple inflammatory mediators and antigen presentation<sup>1</sup>. Fc $\gamma$ RI (CD64) expressed on mono-myeloid cells<sup>2,3</sup> is a high affinity receptor for monomeric IgG. Cross-linking of Fc $\gamma$ RI by immune complexes initiates cellular responses and internalization of the receptor/ligand(s)<sup>1,4</sup>. The phospho-signalling mechanisms in Fc $\gamma$ RI-mediated activation leading to cytokine release and/or induction of the oxidative burst are well recognised<sup>5</sup>. In contrast, events following Fc $\gamma$ RI receptor internalization after cross-linking of immune complexes are less well defined, although two distinct pathways occurring simultaneously, or alternatively, are implicated: clathrin-mediated endocytosis and clathrin independent phagocytosis<sup>6–9</sup>. Clathrin-mediated endocytosis is associated with internalization of small particles (<0.2  $\mu$ m in diameter) and soluble aggregated molecules into cells via clathrin-coated pits that are formed by multiple accessory and adaptor proteins including dynamin, adaptor protein-2 (AP2) and epsin<sup>10</sup>. These contain ubiquitin-interacting motifs (UIM) that bind ubiquitin ligases, including Cbl, that cause receptor ubiquitination<sup>10–12</sup>. Heat shock cognate protein (HSC) 70 is a constitutive member of the heat shock protein family key in disassembly of the clathrin coat<sup>13,14</sup>. It facilitates fusion of the disassembled clathrin pit with early endosome where receptors are sorted for recycling, directly degraded by Cbl, or delivered to lysosomes<sup>10</sup>.

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Cbl-ubiquitinated proteins may also be recognized by hepatocyte growth factor-regulated tyrosine kinase substrate (HGS or HRS) and sorted from early endosomes to late endosomes for endo-lysosomal degradation<sup>15,16</sup>. Although not reported as part of the clathrin-mediated endocytosis cascade, tripartite motif-containing protein 21 (TRIM21), also known as E3 ubiquitin-protein ligase, was recently described as an intracellular Fc receptor that recognises cytoplasmic antibodies or immune complexes that escape from endosomes and promotes their proteasomal degradation<sup>17</sup>. Thus control of these processes by phosphorylation and dephosphorylation events may regulate the fate of endocytosed particles and their receptors. However, data describing the Tyr phosphorylation state of these molecules following Fc $\gamma$ RI cross-linking is very limited. Moreover, mechanisms regulating dephosphorylation of these molecules, and their functional consequences, are unknown.

Excessive activation of Fc $\gamma$ RI can induce unregulated inflammation leading to host tissue damage<sup>18</sup>, thus requires tight regulation. The leukocyte immunoglobulin-like receptor B4 (LILRB4), an ITIM-containing inhibitory receptor on the surface of mono-myeloid cells, is emerging as a key modulator of activation<sup>19,20</sup>. Co-ligation of LILRB4 with Fc $\gamma$ RI on monocytes potently inhibits cytokine production through recruitment of Src homolog-containing phosphatases (SHP-1, SHP-1-like), resulting in Tyr dephosphorylation of a cascade of protein tyrosine kinases<sup>19</sup>. We proposed that LILRB4 may regulate Fc-receptor-dependent endocytosis/phagocytosis by dephosphorylating key signalling molecules, akin to its inhibitory effects on Fc $\gamma$ RI-mediated cytokine production<sup>19,21,22</sup>. Our aim was to identify Tyr phosphorylated proteins following Fc $\gamma$ RI cross-linking on monocytes and determine whether their phosphorylation was modified by LILRB4 ligation.

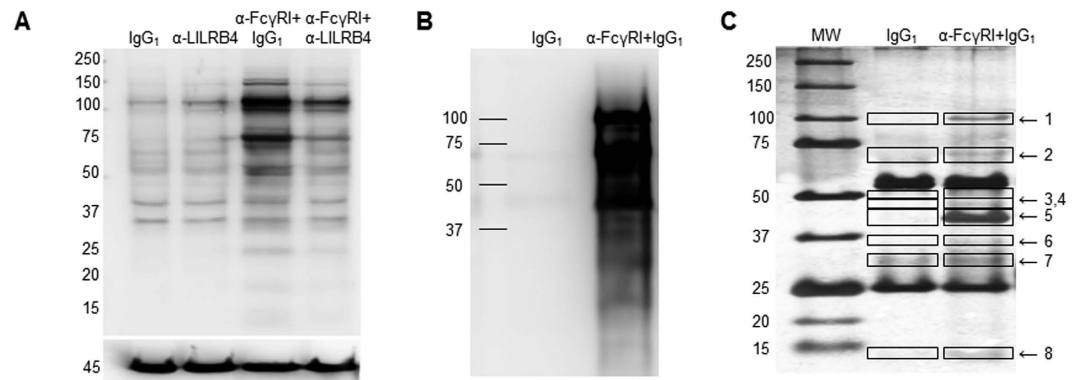
Lysates of Fc $\gamma$ RI-activated THP-1 cells were immunoprecipitated using anti-pTyr antibody and in-gel tryptic digested peptides sequenced by mass spectrometry (LC-MS/MS). Mascot Search output identified 25 high confidence Tyr-phosphorylated proteins and analysis using the Ingenuity Pathway analysis (IPA) revealed that the most significantly affected pathway was clathrin-mediated endocytosis, followed by the Fc receptor-mediated phagocytic pathway. Phosphorylation of 7 key candidate proteins, including the Fc receptor common  $\gamma$ -chain, Syk, clathrin, E3 ubiquitin protein ligase Cbl (a member of Cbl family), HGS, HSP70 and TRIM21 were confirmed by Western blotting and immunoprecipitation. Importantly, co-ligation of LILRB4 with Fc $\gamma$ RI caused significant Tyr dephosphorylation of these proteins, except for HSP70, and significantly suppressed uptake of antibody-opsonised bacterial particles. Unexpectedly, co-ligation of Fc $\gamma$ RI with LILRB4 further increased Tyr phosphorylation of HSP70. Taken together, the results suggest that Tyr phosphorylation may play a critical role in Fc receptor-dependent endocytosis/phagocytosis and that LILRB4 modulates these functions by dephosphorylating key molecules including the upstream Fc receptor common  $\gamma$  chain, Syk and clathrin, as well as downstream molecules including Cbl, HGS and TRIM21.

## Results

**Increased global Tyr phosphorylation in THP-1 lysates after surface cross-linking of Fc $\gamma$ RI.** Western blotting with a pan anti-pTyr MAb showed that cross-linking of surface Fc $\gamma$ RI with specific mouse mAb followed by goat anti-mouse secondary antibody in suspension markedly increased Tyr phosphorylation of multiple proteins above those seen when cells were treated with IgG1 alone. The intensity of reactivity was markedly reduced in lysates from cells that had been co-ligated with anti-Fc $\gamma$ RI and anti-LILRB1 (Fig. 1A). Immunoprecipitation using anti-pTyr mAb showed marked enrichment of phosphorylated proteins in lysates of Fc $\gamma$ RI-cross-linked cells compared with those treated with IgG1 alone (Fig. 1B). Silver staining of SDS-PAGE gels loaded with anti-pTyr mAb-precipitated lysates from IgG1 and anti-Fc $\gamma$ RI+IgG1 cross-linked cells showed enrichment of 8 bands that separated at 100, 70, 50, 47, 43, 35, 30 and 14 kDa compared to components separated from IgG1-treated cell immunoprecipitates (Fig. 1C). The 8 bands were excised and peptides sequenced by Nano LC-MS/MS. The Mascot Search output of 3 combined experiments identified 80 hits that comprised 25 Tyr-phosphorylated candidate proteins in peptides sequenced from lysates of anti-Fc $\gamma$ RI+IgG1 cross-linked cells that were not identified in IgG1 control-treated cells (Mowse score > 50,  $p = 0.05$ ; >3 peptide matches) (Table 1).

**Ingenuity Pathway Analysis (IPA) of Tyr phosphorylated proteins generated by Fc $\gamma$ RI cross-linking on THP-1 cells.** To examine potential signalling pathways enriched by antibody cross-linking of Fc $\gamma$ RI, the 80 hits identified by Mascot analysis were imported into the IPA software. Clathrin-mediated endocytosis was predicted as the most enriched signalling pathway ( $p = 2.19 \times 10^{-13}$ ) (Fig. 2). The second most enriched pathway was Fc receptor-mediated phagocytosis in macrophage and monocytes ( $p = 4.11 \times 10^{-13}$ ), a known pathway activated after Fc $\gamma$ RI cross-linking<sup>1,5</sup> (Fig. 2). The key phosphorylated proteins identified in one or both pathways were Fc receptor  $\gamma$  chain, Syk, clathrin, Cbl, HGS, STAM1/2, HSP70, TRIM21, actin, actin-related proteins, actinin-4, tubulin and actin-binding proteins (Fig. 2 and Table 1). This is the first report to show simultaneous Tyr phosphorylation of these proteins following Fc $\gamma$ RI cross-linking. Although not reported to date in monocytes in the context of Fc $\gamma$ RI cross-linking, other Tyr-phosphorylated peptides identified matched proteins involved in cellular activation, migration and differentiation<sup>23–26</sup> (Table 1). These included phospholipase C gamma 2, mitogen-activated protein kinase 9, hematopoietic cell-specific Lyn substrate 1, SH2 domain containing leukocyte protein of 76 kDa, linker for activation of T cell family member 2, Twinfilin-1, docking protein 2, protein tyrosine phosphatase 18, phosphatidylinositol 3, 4, 5-triphosphate 5-phosphatase 1 (SHIP), protein phosphatase 1 gamma, 1-phosphatidylinositol 4, 5-biphosphate phosphodiesterase gamma 2, Toll-like receptor 6, Crk-like protein, Coatomer subunit epsilon and ubiquitin-40S ribosomal protein s27a (Table 1).

**Regulation of Fc $\gamma$ RI-mediated Tyr phosphorylation of Fc receptor  $\gamma$  chain, clathrin, Cbl, HGS, HSP70, TRIM21 and Syk by LILRB4.** Immunoprecipitation of cell lysates using anti-pTyr mAb followed by Western blotting using antibodies against the Fc $\gamma$  chain, clathrin, Cbl, HGS, HSP70 and TRIM21 showed Tyr-phosphorylation of all these proteins in cells activated through Fc $\gamma$ RI cross-linking (Fig. 3A) (supplementary Fig. 1), validating the LC-MS/MS data. Importantly, phosphorylation of all proteins except HSP70 was markedly



**Figure 1. Cross-linking of Fc $\gamma$ RI induced Tyr phosphorylation of multiple proteins that were reduced upon co-ligation with LILRB4.** (A) Representative Western blot using anti-pTyr mAb showing marked increases in multiple Tyr phosphorylated proteins in total cell lysate from THP-1 cells ligated with specific mouse anti-Fc $\gamma$ RI+IgG1 compared with cells treated with IgG1 alone. Tyr phosphorylated proteins in cells following co-ligation with anti-LILRB4 mAb and anti-Fc $\gamma$ RI Ab were markedly reduced and more similar to those in cells treated with anti-LILRB4 mAb alone. The lower panel shows the same membrane stripped and re-probed with anti- $\beta$  actin Ab confirming comparable protein loading (n = 3 experiments using different batches of THP-1 cells). Migration of the molecular weight markers is shown on the left. (B) Immunoprecipitation using anti-pTyr mAb (4G10) followed by Western blotting with biotinylated anti-pTyr mAb-100 mAb showed multiple strongly phosphorylated proteins in lysates from THP-1 cells cross-linked with specific mouse anti-Fc $\gamma$ RI mAb but not control IgG<sub>1</sub> (n = 3 separate experiments). (C) Silver staining of SDS-PAGE gels loaded with anti-pTyr mAb-precipitated lysates from Fc $\gamma$ RI cross-linked cells indicated enrichment of 8 components that separated at approximately 100, 70, 50, 47, 43, 35, 30 and 14 kDa, compared to precipitates from control IgG<sub>1</sub>-treated cells. Bands at 53 and 25 kDa that are evident in each sample likely represented the heavy and light chains of antibodies used for immunoprecipitation and were not sequenced (representative of 3 independent experiments using different batches of THP-1 cells). Mascot Search output of peptides sequenced by LC-MS/MS of the 3 experiments combined identified 80 specific hits of 25 Tyr phosphorylated candidate proteins (Mowse score > 50, p 0.05; >3 peptide matches) (see Table 1).

suppressed upon co-ligation of Fc $\gamma$ RI with LILRB4 (Fig. 3A). Semi-quantitative analysis indicated that LILRB4 significantly reduced Tyr phosphorylation of HGS by an average of 80.2%, Cbl by 51.3%, clathrin by 45.5% and TRIM21 by 36.9% (Fig. 3B; n = 3). In contrast, co-ligation of Fc $\gamma$ RI with LILRB4 significantly enhanced HSP70 phosphorylation by 36.4%, suggesting selective suppressive effects by LILRB4. Similarly, Western blotting of cell lysates from Fc $\gamma$ RI cross-linked cells caused Tyr-phosphorylation of Syk that was markedly reduced upon co-ligation with LILRB4 (Fig. 3E), further validating the LC-MS/MS data and confirming our previous finding<sup>19</sup>. As expected, the brief cross-linking/co-ligation protocols used in this study did not affect the total amounts of any of the above proteins (Fig. 3C,D) (Supplementary Fig. 1).

**LILRB4 suppressed uptake of antibody-opsonised bacterial particles.** For this experiment we used PMA-differentiated THP-1 cells because these have superior phagocytic activity. The mean percentage of these cells that took up antibody-opsonised fluorescent DH5 $\alpha$  *E-coli* particles identified by flow cytometry, was  $32.9 \pm 2.3\%$ . Uptake was significantly reduced to  $3.5 \pm 0.74\%$  when surface Fc $\gamma$ RI was blocked using a specific anti-Fc $\gamma$ RI mAb (blocked by ~90%; p = 0.0005, n = 4; Fig. 4B), but not when cells were treated with an irrelevant negative control mAb ( $36.7 \pm 4.6\%$ ) confirming involvement of Fc $\gamma$ RI-dependent endocytosis/phagocytosis. Ligation of surface LILRB4 using an anti-LILRB4 mAb, followed by goat anti-mouse secondary antibody, significantly suppressed uptake of opsonised DH5 $\alpha$  *E-coli* by up to 55% (p = 0.004, n = 4; Fig. 4B). In contrast, co-ligation of surface MHC-I using anti-MHC-I mAb did not significantly alter uptake of opsonised DH5 $\alpha$  *E-coli* particles when compared with non-ligated cells ( $28.9 \pm 1.7\%$  versus  $32.9 \pm 2.3\%$  Fig. 4B), suggesting specific suppression of endocytosis/phagocytosis by LILRB4.

## Discussion

Reversible Tyr phosphorylation of proteins in eukaryotes is critical in regulating intracellular signalling pathways involved in cellular activation, growth, proliferation, differentiation migration and gene transcription<sup>27,28</sup>. Immune-complex mediated activation of Fc $\gamma$ RI on innate immune cells is essential for protection against bacterial infection. Activation of Tyr phosphorylation of selected upstream protein tyrosine kinases and downstream mitogen activated protein kinases result in production of pro-inflammatory cytokines, generation of an oxidative burst, and/or triggering of endocytosis/phagocytosis<sup>1,5</sup>. In this study we immunoprecipitated total Tyr phosphorylated proteins from lysates of THP-1 that had been cross-linked with specific anti-Fc $\gamma$ RI or control IgG mAb and found marked enrichment of phosphorylated proteins in cells activated with the specific mAb. Peptide sequencing by Nano LC-MS/MS identified 80 candidate peptides that were significantly modified, representing 25 Tyr-phosphorylated proteins. Pathway analysis predicted that Ty-phosphorylation of proteins mediating clathrin-mediated endocytosis and Fc receptor-mediated phagocytosis were the most affected. The most

| Band                            | Phosphoproteins detected in anti-Fc $\gamma$ RI cross-linked but not control IgG1 treated THP-1 cells | Score | Peptide matched | Mass |
|---------------------------------|---|-------|-----------------|------|
| 1                               | alpha actinin 1, sarcomeric (F-actin cross-linking protein)   | 5321  | 249             | 102  |
|                                 | alpha actinin 4   | 1927  | 77              | 102  |
|                                 | E3 ubiquitin-protein ligase CBL   | 964   | 71              | 99   |
|                                 | hepatocyte growth factor-regulated tyrosine kinase substrate (HGS)                                    | 231   | 17              | 86   |
|                                 | Toll-like receptor 6  | 116   | 12              | 92   |
|                                 | microtubule-associated protein 2  | 123   | 8               | 199  |
|                                 | ALG-2 interacting protein 1 (hp95/ program cell death 6 interacting protein)                          | 112   | 3               | 96   |
|                                 | phosphatidylinositol 3,4,5-trisphosphate 5-phosphatase 1 (SHIP1)                                      | 98    | 3               | 133  |
| 2                               | protein Tyr kinase Syk  | 951   | 67              | 72   |
|                                 | heat shock cognate 71 kDa protein   | 831   | 46              | 70   |
|                                 | hepatocyte growth factor-regulated tyrosine kinase substrate (HGS)                                    | 209   | 31              | 86   |
|                                 | actinin, alpha 1  | 654   | 22              | 103  |
|                                 | protein SPY75 (hematopoietic cell-specific Lyn substrate 1)   | 289   | 11              | 54   |
|                                 | lymphocyte cytosolic protein 2 (SH2 domain containing leukocyte protein of 76 kDa)                    | 287   | 11              | 60   |
|                                 | E3 ubiquitin-protein ligase CBL (proto-oncogene c-CBL, RING finger protein 55)                        | 146   | 6               | 100  |
|                                 | Fc gamma receptor type I/ Fc fragment of IgG receptor, CD64)  | 76    | 4               | 42   |
| 3                               | ATP-dependent DNA helicase II, 70 kDa subunit (G22P1)   | 64    | 3               | 70   |
|                                 | elongation factor 1 alpha 1   | 402   | 30              | 50   |
|                                 | lymphocyte cytosolic protein 2 (SLP76)  | 293   | 27              | 60   |
|                                 | tripartite motif containing 21 (TRIM21)   | 277   | 21              | 54   |
|                                 | Hematopoietic lineage cell-specific protein   | 261   | 13              | 53   |
|                                 | beta-actin  | 254   | 10              | 42   |
|                                 | tubulin beta chain  | 235   | 10              | 50   |
|                                 | signal transducing adaptor molecule 2B (STAM2)  | 232   | 8               | 58   |
|                                 | coronin, actin binding protein, 1C variant  | 230   | 7               | 53   |
|                                 | alpha actinin (4)   | 227   | 7               | 103  |
|                                 | docking protein 2   | 210   | 6               | 45   |
|                                 | protein Tyr kinase or Syk   | 175   | 7               | 72   |
|                                 | ARP3 actin-related protein 3 homolog  | 154   | 7               | 47   |
|                                 | signal transducing adaptor molecule 1 (STAM1)   | 83    | 4               | 59   |
|                                 | heat shock protein 70 kDa   | 93    | 3               | 70   |
|                                 | E3 ubiquitin-protein ligase CBL (Ring finger protein 55)  | 97    | 5               | 99   |
|                                 | unnamed human protein (IgG receptor Fc region II precursor)   | 78    | 3               | 35   |
| alpha-tubulin                   | 74  | 3     | 50              |      |
| 4                               | ARP3 actin-related protein 3  | 1338  | 61              | 47   |
|                                 | mitogen-activated protein kinase 9  | 103   | 4               | 48   |
|                                 | actin non-muscle 6.2  | 504   | 19              | 41   |
|                                 | alpha actinin 4   | 95    | 3               | 102  |
|                                 | actin 7   | 91    | 3               | 37   |
|                                 | docking protein 2   | 79    | 3               | 45   |
|                                 | signal transducing adaptor molecule 2B (STAM2)  | 75    | 3               | 58   |
| 5                               | POTE ankyrin domain family member E   | 835   | 38              | 35   |
|                                 | actin related protein 2/3 complex subunit 1B (p41-ARC)  | 577   | 35              | 40   |
|                                 | F-actin capping protein alpha-1 subunit   | 480   | 29              | 32   |
|                                 | F-actin capping protein beta subunit (actin filament muscle Z-line)                                   | 208   | 4               | 30   |
|                                 | clathrin light chain (LCB3)   | 170   | 8               | 23   |
|                                 | histone cluster1, H1  | 83    | 8               | 23   |
|                                 | capping protein alpha (actin filament muscle Z-line, alpha 2)   | 141   | 6               | 33   |
|                                 | actin related protein 2/3 complex subunit 2   | 126   | 5               | 34   |
|                                 | EF-hand domain family, member D2  | 100   | 5               | 27   |
|                                 | protein tyrosine kinase (PTK9 or Twinfilin-1)   | 99    | 4               | 40   |
|                                 | protein phosphatase 1 gamma   | 97    | 3               | 37   |
|                                 | protein Tyr kinase (Syk)  | 80    | 4               | 72   |
| E3 ubiquitin-protein ligase CBL | 70  | 3     | 100             |      |

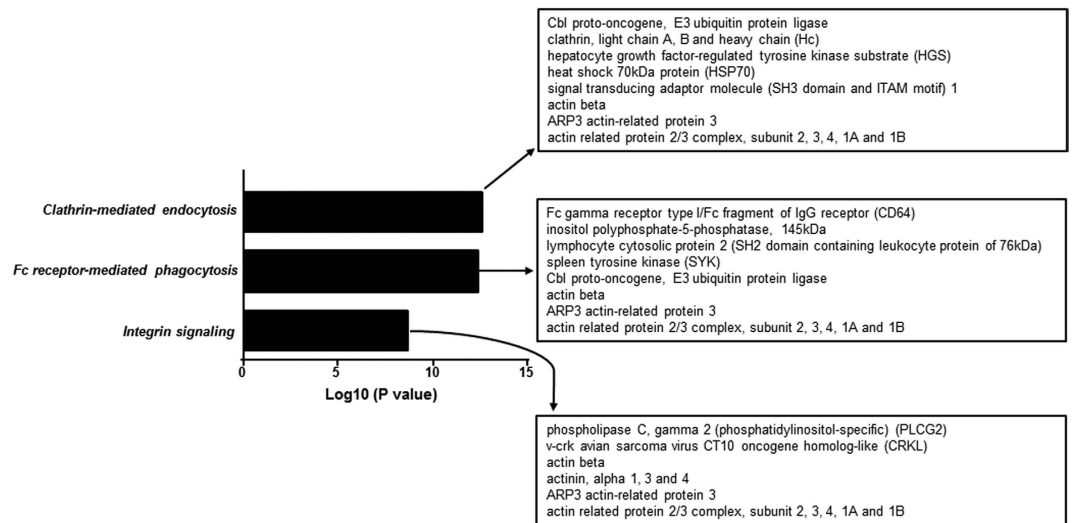
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| Band | Phosphoproteins detected in anti-Fc $\gamma$ RI cross-linked but not control IgG1 treated THP-1 cells | Score | Peptide matched | Mass |
|------|---|-------|-----------------|------|
| 6    | actin related protein 2/3 complex subunit 2   | 515   | 38              | 34   |
|      | actin related protein 2/3 complex subunit 1B (p41-ARC)  | 434   | 17              | 41   |
|      | F-actin capping protein beta subunit (actin filament muscle Z-line, beta)                             | 307   | 15              | 31   |
|      | EF-hand domain family, member D2  | 279   | 10              | 27   |
|      | linker for activation of T cells family member 2  | 133   | 6               | 31   |
|      | F-actin capping protein alpha-1 subunit   | 122   | 5               | 33   |
|      | high affinity immunoglobulin gamma Fc receptor I  | 97    | 5               | 32   |
|      | actin related protein 2/3 complex subunit 4 isoform a   | 96    | 4               | 20   |
|      | Crk-like protein  | 58    | 3               | 33   |
|      | coatamer subunit epsilon  | 56    | 3               | 34   |
| 7    | spectrin beta chain, non-erythrocyte 4  | 221   | 14              | 28   |
|      | EF hand domain containing protein D2  | 311   | 17              | 26   |
|      | clathrin light chain B  | 106   | 13              | 25   |
|      | clathrin light chain A  | 91    | 8               | 27   |
|      | protein tyrosine phosphatase, non-receptor type 18  | 81    | 3               | 50   |
| 8    | alpha actinin 4   | 2158  | 101             | 102  |
|      | actinin, alpha 1  | 1607  | 78              | 103  |
|      | clathrin heavy chain 1 (or KIAA0034)  | 375   | 25              | 191  |
|      | E3 ubiquitin-protein ligase CBL   | 354   | 13              | 100  |
|      | splicing factor proline/glutamine rich  | 288   | 16              | 76   |
|      | Ubiquitin-40S ribosomal protein s27a  | 219   | 17              | 17   |
|      | hepatocyte growth factor-regulated tyrosine kinase substrate  | 186   | 6               | 62   |
|      | high affinity immunoglobulin epsilon receptor subunit gamma   | 179   | 5               | 10   |
|      | protein Tyr kinase (p72 Syk)  | 95    | 4               | 96   |
|      | 1-phosphatidylinositol 4,5-bisphosphate phosphodiesterase gamma-2 (PLCG2)                             | 82    | 3               | 14   |

**Table 1. Mascot search results of mass spectrometric peptides sequencing of tyrosine phosphorylated proteins following Fc $\gamma$ RI cross linking of THP-1 cells (n = 3).**

prominent phosphorylated proteins included Fc receptor  $\gamma$  chain, Syk, clathrin, Cbl, HGS, STAM1/2, HSP70, TRIM21, actin, actin-related proteins, actinin-4, tubulin and actin binding proteins. This is the first study to demonstrate the simultaneous Tyr phosphorylation of these proteins during Fc $\gamma$ RI-mediated monocyte activation. This is particularly novel for clathrin, Cbl, HGS and HSP70, key molecules involved in the clathrin-mediated endocytosis<sup>10,13,14</sup> and for TRIM21, a ligase recently identified as high affinity intracellular Fc receptor<sup>17,29</sup> which is critical for ubiquitination and degradation of antibody-opsonised viruses<sup>17</sup>. We validated the data generated by LC-MS/MS using a combination of Western blotting and immunoprecipitation and confirmed that these molecules were indeed Tyr phosphorylated following cross-linking of Fc $\gamma$ RI on the surface of THP-1 cells. Moreover, we confirmed our previous finding<sup>19</sup> of Tyr phosphorylation of Syk in response to Fc $\gamma$ RI cross-linking and identified Tyr phosphorylation of the intracellular tyrosine-based activating motifs (ITAMs) of the common  $\gamma$  chain of Fc-receptors that is upstream of Syk, directly linking for the first time, these two critical signalling events. Functionally, Tyr phosphorylation of Syk after Fc $\gamma$ RI cross-linking on monocytes promotes cytokine production<sup>19,21,22</sup> and Syk phosphorylation has been associated with increased phagocytosis of opsonised pathogens<sup>30</sup> and polybeads,<sup>31</sup> and enhanced endocytosis of immune complexes<sup>32</sup>. Here, we found that co-ligation of Fc $\gamma$ RI with LILRB4 that contains intracellular tyrosine-based inhibitory motifs (ITIMs), significantly reduced phosphorylation of the ITAMs of the common  $\gamma$  chain, and of Syk. Suppression of Syk phosphorylation is consistent with our previous report showing that LILRB4, through recruitment of SHP-1-like phosphatase, dephosphorylated Syk and multiple downstream protein tyrosine kinases including Lck, LAT and Erk in THP-1 cells leading to suppressed cytokine production<sup>19</sup>. Importantly, we show here that LILRB4 ligation significantly inhibited Fc $\gamma$ RI-dependent uptake of antibody-opsonised *E. coli* particles. One mechanism may involve dephosphorylation (inactivation) of the common  $\gamma$  chain ITAMs and Syk, similar to its inhibitory effects on Fc $\gamma$ RI-mediated cytokine production<sup>19,21,22</sup>.

Clathrin-mediated endocytosis is an important energy efficient pathway of pathogen/antigen clearance by innate immune cells as an alternate to phagocytosis<sup>7,9,33,9,33</sup>. It is also an important mechanism of endogenous surface ligand/receptor internalisation, ubiquitination and recycling/degradation<sup>7,9,33</sup>. Clathrin-mediated endocytosis may share some upstream signalling molecules with phagocytosis but unlike to phagocytosis, it is associated with internalisation of small particles (<0.2  $\mu$ m in diameter) and soluble aggregated molecules<sup>10</sup>. During endocytosis, receptor-ligand or antibody-antigen complexes are first ubiquitinated and internalised into clathrin-coated pits assembled within AP2, dynamin, epsin and related molecules followed by HSP70-mediated un-coating of the pits and their endosomal sorting<sup>10,13,14</sup>. Receptors and ligands in the endosome are then either ubiquitinated and directly degraded by Cbl<sup>10-12,17</sup> or the ubiquitinated molecules are delivered to lysosome by HGS/STAM1/2 complexes for lysosomal degradation<sup>15,16</sup>. Alternatively, the receptor in the endosomes is rapidly recycled while the ligands undergo endo-lysosomal degradation<sup>10</sup>. Although the involvement of clathrin, Cbl, HSP70, HGS



**Figure 2. Pathway analysis of Tyr-phosphorylated proteins after Fc $\gamma$ RI-cross-linking.** Ingenuity Pathway Analysis of the 80 enriched peptides from the combined 3 experiments showed that clathrin-mediated endocytosis was predicted to be the most enriched signalling pathway ( $p = 2.19 \times 10^{-13}$ ) followed by Fc $\gamma$ RI-mediated phagocytosis and integrin signalling ( $p = 4.11 \times 10^{-13}$  and  $p = 1.88 \times 10^{-9}$ ) respectively.

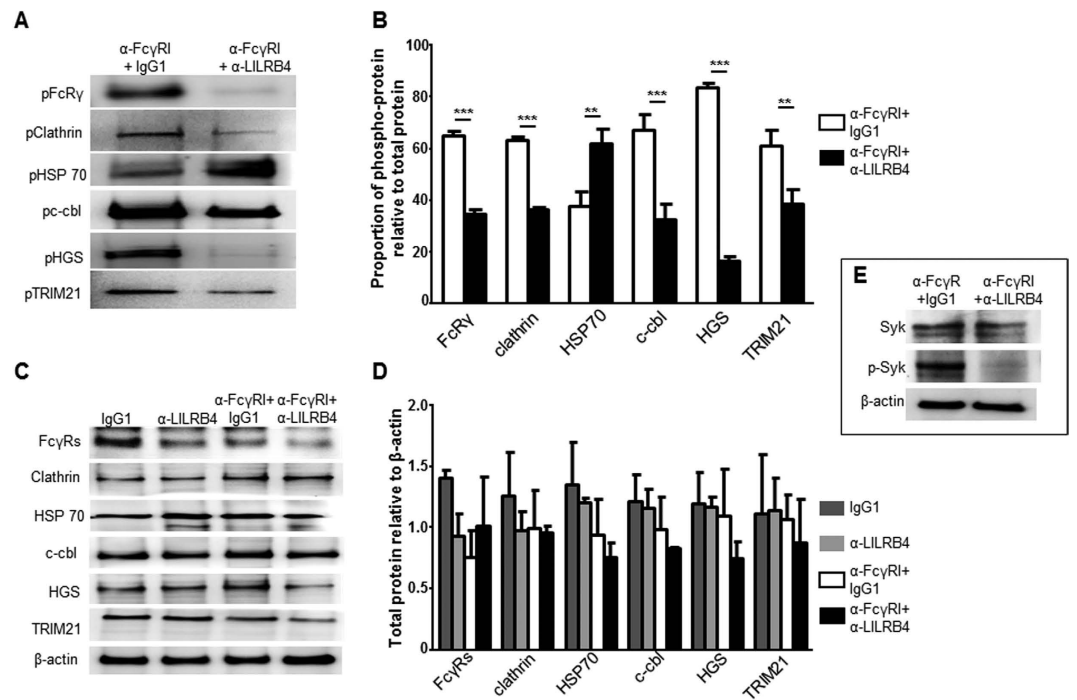
and STAM in this pathway is generally accepted, whether Tyr phosphorylation/dephosphorylation regulate their functions, particularly during Fc-receptors mediated monocyte activation remain unexplored. There is limited evidence that Tyr phosphorylation of clathrin heavy chain promotes bacterial internalisation<sup>34</sup> and that phosphorylation of Cbl is associated with receptor/ligand ubiquitination after receptor clustering in antigen presenting cells, T cells and B cells<sup>35,36</sup>. Tyr phosphorylation of HGS in HeLa cells during epidermal growth factor (EGF)-mediated activation may be involved in intracellular receptor sorting and vesicle formation<sup>37</sup>. C-terminal Tyr phosphorylation of HSP70 is described as a switch that regulates co-chaperon binding in cancer cells and determines whether it facilitates protein folding, or directs proteins for ubiquitin-mediated degradation<sup>38</sup>. These functional observations collectively suggest that Tyr phosphorylation of these particular molecules may play critical roles in Fc receptor-dependent endocytosis of immune complexes. Here we show that co-ligation of LILRB4 with Fc $\gamma$ RI significantly reduced Fc $\gamma$ RI-mediated Tyr phosphorylation (activation) of clathrin, Cbl, HGS and STAM1/2 (Fig. 5). We propose that LILRB4 may inhibit Fc-receptor-dependent endocytosis of antigen-antibody complexes by promoting Tyr dephosphorylation (deactivation) of these key molecules. This proposal is consistent with its reported anti-inflammatory and immunosuppressive properties<sup>19,39–41</sup> and our demonstration that it significantly suppressed Fc $\gamma$ RI-dependent endocytosis/phagocytosis of antibody-opsonised *E. coli* particles (Fig. 4). In contrast to these dephosphorylating events, co-ligation of LILRB4 with Fc $\gamma$ RI significantly enhanced Fc $\gamma$ RI-mediated HSP70 Tyr phosphorylation by 36.4%, indicating selective effects. Whether enhanced HSP70 phosphorylation by LILRB4 regulates ubiquitin-mediated degradation of antibody-opsonised bacterial particles and/or the sorting of the internalised Fc-receptors, requires further investigation.

TRIM21 is described as an important high affinity intracellular Fc receptor implicated in elimination of antibody-bound intracellular viruses<sup>17</sup>, although little is known about interactions that link extracellular antibody-bound pathogen to intracellular TRIM21. Here we found that surface cross-linking of Fc $\gamma$ RI that uses a method that mimics antibody-antigen complexes, promoted strong phosphorylation of TRIM21. Thus Fc $\gamma$ RI might be the missing link between activation by extracellular antibody-bound pathogens (antigen) and their intracellular Fc receptor (TRIM21) that may also function as a novel downstream pathway in Fc receptor-dependent endocytosis of immune complexes. Interestingly, TRIM21 is Tyr phosphorylated in TLR3 or TLR4-stimulated monocytes and macrophages and is suggested to activate downstream TLR-mediated signalling<sup>42</sup>. The significant dephosphorylation of TRIM21 caused by LILRB4 ligation shown by us, may therefore indicate functional deactivation of this molecule when Fc $\gamma$ RI-induced monocyte activation is moderated by LILRB4.

In conclusion, results presented here suggest that Tyr phosphorylation of the upstream common  $\gamma$ -chain, Syk, and clathrin and the downstream molecules such as TRIM21 may be critical in Fc-receptor (Fc $\gamma$ RI)-dependent endocytosis/phagocytosis of antibody-opsonised particles. Importantly, LILRB4 may regulate this important innate immune function by promoting mechanisms that dephosphorylate these proteins.

## Materials and Methods

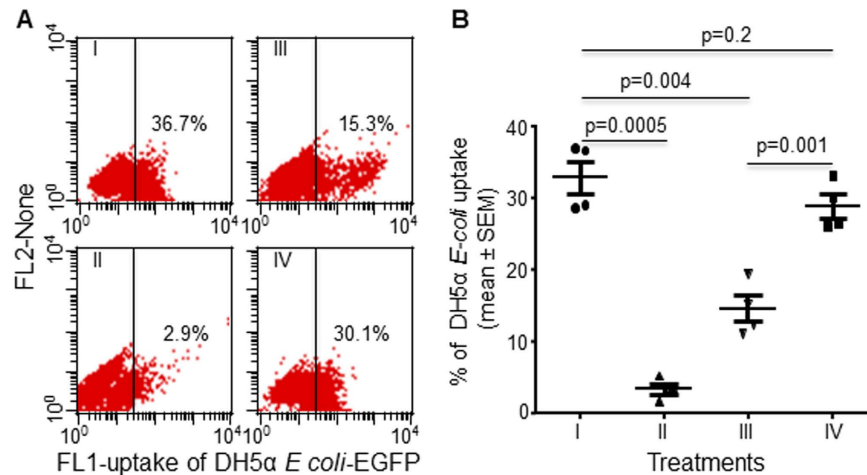
**Cells and antibodies.** Human monocytic leukemic THP-1 cells (ATCC clone TIB-202, Manassas, VA, USA) were cultured in RPMI 1640 supplemented with 2 mM L-glutamine, 10% heat-inactivated fetal bovine serum (FBS), 100 U/ml penicillin, 100  $\mu$ g/ml streptomycin, 1 mM sodium pyruvate, 10 mM HEPES and 0.1%  $\beta$ -mercaptoethanol (all from Life Technologies) and 20 mM sodium bicarbonate (Sigma-Aldrich) at 37 °C with 5% CO<sub>2</sub><sup>19</sup>. The following antibodies were used for flow cytometry and/or cross-linking/co-ligation experiments; anti-LILRB4 (kindly donated by Dr. Luis Borges, Amgen Inc), anti-Fc $\gamma$ RI (R&D System, Minneapolis, MN,



**Figure 3. Co-ligation of LILRB4 with Fc $\gamma$ RI suppressed Tyr phosphorylation of multiple proteins involved in clathrin-mediated endocytosis, and of TRIM21.** (A) Representative immunoprecipitation of THP-1 cell lysates using anti-pTyr mAb (4G10) followed by Western blotting using selected antibodies showed abundant Tyr phosphorylation of Fc $\gamma$ Rs, clathrin, HSP70, Cbl, HGS and TRIM21 in cells co-ligated with anti-Fc $\gamma$ RI+IgG<sub>1</sub> control mAb validating LC-MS/MS data. Importantly, LILRB4 co-ligation with Fc $\gamma$ RI markedly reduced Tyr phosphorylation of all proteins except for HSP70 (n = 3). (B) Summary of densitometry of bands from 3 independent experiments showed significant reduction of Fc $\gamma$ Rs, clathrin, Cbl, HGS and TRIM21 phosphorylation, but not HSP70, in THP-1 cells co-ligated with anti-Fc $\gamma$ RI and anti-LILRB4 mAbs, compared to cells co-ligated with anti-Fc $\gamma$ RI and negative control mAb (n = 3, \*\*p < 0.01; \*\*\*p < 0.001). (C) Representative Western blotting of total cell lysates showed that co-ligation of Fc $\gamma$ RI with LILRB4 did not alter the total amounts of any of the above proteins when compared to co-ligation of Fc $\gamma$ RI+IgG<sub>1</sub> control, ligation of LILRB4 alone or treatment with IgG<sub>1</sub> control alone; the lower panel is the same membrane stripped and re-probed with anti- $\beta$  actin Ab, confirming comparable protein loading. (D) Summary of densitometry analysis of 3 independent experiments showed no significant differences in total Fc $\gamma$ Rs, clathrin, HSP70, Cbl, HGS and TRIM21 in THP-1 cells within the 4 different treatment groups (n = 3). Full image of the Western blots is shown in Supplementary Fig. 1. (E) Western blotting of cell lysates from Fc $\gamma$ RI cross-linked cells showing increased Tyr-phosphorylated Syk that was markedly reduced upon co-ligation with LILRB4, confirming our earlier finding<sup>21</sup> and validating current LC-MS/MS data (Fig. 2) (n = 1).

USA), IgG<sub>1</sub> negative control (Sigma-Aldrich) mouse primary mAbs and F(ab')<sub>2</sub> fragment goat anti-mouse IgG (Fc-specific) secondary Ab (Jackson ImmunoResearch, West Grove, PA, USA). Mouse anti-pTyr mAb (clone 4G10; Upstate Biotechnology, Lake Placid, NY, USA) was used for immunoprecipitation. The following antibodies were used for Western blotting: biotinylated mouse  $\alpha$ -pTyr-100 mAb (Cell Signaling, Danvers, MA, USA), mouse anti-human clathrin (Thermo Fisher Scientific, Waltham, MA, USA), mouse anti-HSP70 (Stressgen/Enzo Life Sciences, Farmingdale, NY, USA), rabbit anti-HGS (Thermo Fisher Scientific), rabbit anti-Cbl (Sigma-Aldrich), rabbit anti-Fc $\gamma$ Rs (Upstate Biotechnology Inc, Lake placid, NY, USA), rabbit anti-Syk (Cell Signaling), rabbit anti-pSyk (Tyr 525/526) (Cell Signaling), mouse anti- $\beta$ -actin (Sigma-Aldrich), and mouse anti-human TRIM21 mAb (R&D System) in-house labelled with biotin using lightning-link<sup>TM</sup> biotin conjugation kit (Innova Biosciences, Babraham, Cambridge, UK), primary antibodies, and HRP-conjugated goat anti-mouse or goat anti-rabbit secondary antibodies, and HRP-conjugated streptavidin (all from Bio-Rad, Gladesville, NSW, Australia). Mouse anti-MHC-I mAb (anti-HLA-ABC) (BD Biosciences, Mountain View, CA, USA) was used as relevant surface binding control Ab in detection of antibody opsonised bacteria uptake upon LILRB4 ligation.

**Immunoprecipitation of tyrosine-phosphorylated proteins after Fc $\gamma$ RI cross-linking and identification of phosphorylated peptides by mass spectrometry.** A total  $2 \times 10^7$  THP-1 cells in 50  $\mu$ l cross-linking buffer (CLB; RPMI supplemented with 10 mM HEPES, 1 mM MgCl<sub>2</sub>, 0.1 mM CaCl<sub>2</sub> and 0.1% bovine serum albumin CLB) were incubated with 5  $\mu$ g/ml IgG<sub>1</sub> control, anti-Fc $\gamma$ RI, anti-LILRB4 or anti-Fc $\gamma$ RI+anti-LILRB4 mAbs for 15 min at RT. Cells were washed in 1 ml CLB then resuspended in 100  $\mu$ l CLB, then cross-linked with 15  $\mu$ g/ml goat anti-mouse IgG (Fc-specific) secondary antibody for 90 sec at RT. Cell activation was stopped by adding cold PBS, cells harvested by centrifugation at 4  $^{\circ}$ C then lysed with cold Western lysis buffer



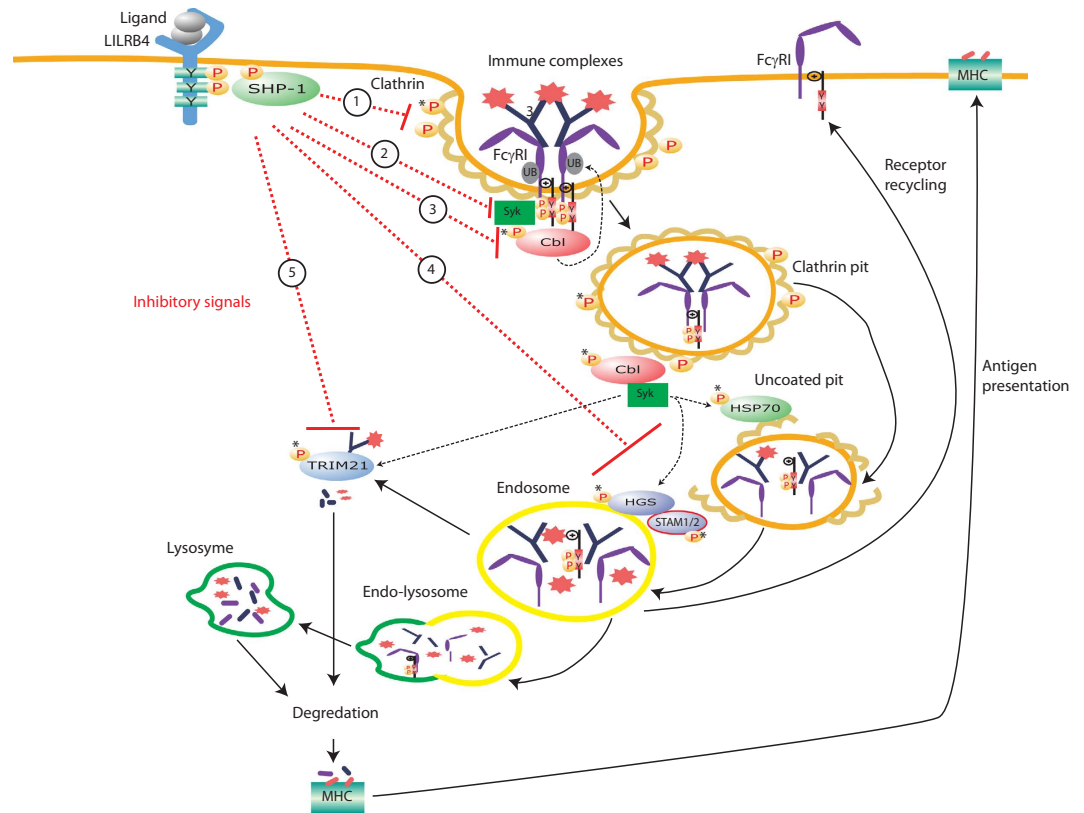
**Figure 4. Ligation of LILRB4 significantly suppressed Fc receptor-dependent uptake of antibody-opsinized *E. coli* by differentiated THP-1 cells.** (A) Representative dot plot showing uptake of Ab opsonized EGFP-expressing DH5 $\alpha$  *E-coli* particles by 36.7% of PMA-differentiated THP-1 cells (I) that was reduced by 90% when surface Fc $\gamma$ RI was blocked by pre-incubating cells with 20  $\mu$ g/ml anti-Fc $\gamma$ RI mAb (II), but not by cells pre-incubated with negative control IgG $_1$  mAb, indicating Fc-receptor dependent endocytosis/phagocytosis ( $n = 4$ ). Ligation of LILRB4 with mouse anti-LILRB4 mAb followed by goat anti-mouse secondary Ab reduced uptake of Ab opsonized EGFP-expressing DH5 $\alpha$  *E-coli* particles by >50% (III) when compared with non-ligated cells (I). Ligation of control mouse anti-MHC-I mAb (IV) or negative control mouse mAb had little effect on uptake, confirming specific LILRB4-mediated suppression. (B) Summary analysis of 4 independent experiments presenting mean percentages ( $\pm$ SEM) showing that numbers of PMA-differentiated THP-1 cells that took up Ab-opsinized EGFP-expressing DH5 $\alpha$  *E-coli* particles were significantly less if cells were pre-incubated with anti-Fc $\gamma$ RI mAb ( $p = 0.0005$ ), and markedly less following Ab ligation of surface LILRB4 ( $p = 0.004$ ), but not Ab ligation of MHC-I ( $p = 0.2$ ).

containing 150 mM NaCl, 50 mM Tris-HCl (pH 8.0), 5 mM EDTA and 1% NP-40, freshly-made protease inhibitors (2 mg/ml; Roche Applied Science) and 10  $\mu$ M sodium pervanadate (Sigma-Aldrich). After vortexing for 1 min, samples were incubated on ice for 30 min then supernatants collected by centrifugation at 20,000 $\times$ g for 10 min at 4  $^{\circ}$ C. Tyr-phosphorylated proteins were immunoprecipitated using 5  $\mu$ g/ml anti-pTyr mAb (clone 4G10) at overnight 4  $^{\circ}$ C then incubated with goat anti-mouse secondary antibody conjugated to Sepharose beads (10  $\mu$ g/ml; Zymed Laboratories Inc., San Francisco, CA, USA) for 2 hrs at 4  $^{\circ}$ C. Bead-bound proteins were washed once with 1 ml cold dilution buffer (0.1% Triton X-100 in TSA buffer pH 8.0; 0.01 M Tris buffer, 0.14 M NaCl, 0.025% Na $_2$ S $_2$ O $_8$ ), two washes with TSA and a single wash with 50 mM Tris buffer pH 6.9. Beads were then resuspended in Tricine gel loading buffer containing 10 mM dithiothreitol, heated for 5 min at 100  $^{\circ}$ C and supernatants resolved in 10% Tris-Tricine SDS-PAGE gels under reducing conditions then silver-stained. Specific silver-stained bands were excised and Tyr-phosphorylated proteins were identified by Nano Liquid Chromatography tandem Mass Spectrometry (Nano LC-MS/MS) as described<sup>43,44</sup>. Bands excised from lanes loaded with immunoprecipitates of irrelevant IgG $_1$ -cross-linked THP-1 cells were used as negative controls. Peak lists of MS/MS data were generated using Mascot Daemon/extract\_msn (Matrix Science, London, England, Thermo) were interrogated using Mascot version 2.1 (<http://www.matrixscience.com>) and searched against *Homo sapiens* proteins in the Swissprot protein database (version 80). Precursor tolerances were 4.0 ppm and product ion tolerances were  $\pm 0.4$  Da and acceptable cut-off scores for individual MS/MS spectra were set to 20. Specific phosphorylated peptides identified in Fc $\gamma$ RI cross-linked cells, but not in cells treated with control IgG $_1$  from 3 independent experiments, were combined and uploaded onto Ingenuity Pathway Analysis software version 24718999, used to predict the most significantly-enriched pathways (IPA $^{\circ}$ ; [www.qiagen.com/ingenuity](http://www.qiagen.com/ingenuity), QIAGEN, Redwood City, CA). Alternatively, proteins were transferred onto PVDF membranes (0.2  $\mu$ m pore size; Millipore, Bayswater, VIC, Australia) for Western blotting using 1  $\mu$ g/ml biotinylated  $\alpha$ -pTyr-100 mAb.

#### Validation of enriched Tyr phosphorylated proteins by immunoprecipitation and Western blotting and regulation by LILRB4.

Six proteins including the common  $\gamma$  chain of the Fc receptor, clathrin, Cbl, HGS, TRIM21 and HSP70 had multiple Tyr phosphorylated peptides with high Mascot scores following Fc $\gamma$ RI cross-linking. Four of these are reported to be involved in clathrin-mediated receptor endocytosis, although this was the first demonstration of their simultaneous phosphorylation. Hence, results were validated, and their regulation by LILRB4 examined using a combination of immunoprecipitation and Western blotting. In brief,  $2 \times 10^7$  THP-1 cells were activated via Fc $\gamma$ RI cross-linking with or without LILRB4 co-ligation [21], lysates immunoprecipitated using anti-pTyr mAb (4G10) followed by serial Western blots using antibodies against FcR common  $\gamma$  chain, clathrin, Cbl, HGS, TRIM21 or HSP70. In separate experiments, Western blotting using anti-pTyr (4G10) was performed using 20  $\mu$ g total lysates to detect global protein phosphorylation; membranes were re-probed with 1  $\mu$ g/ml mouse anti- $\beta$ -actin mAb to confirm equal protein loading. For detection of Syk and





**Figure 5. Schematic diagram suggesting possible roles of Tyr phosphorylation of key molecules involved in clathrin-mediated endocytosis of  $Fc\gamma RI$  and ligands, and their regulation by LILRB4.** Cross-linking of  $Fc\gamma RI$  by immune-complexes causes Tyr phosphorylation of the ITAMs of its common  $\gamma$  chain and binding of pSyk transduces activating signals. This simultaneously initiates phosphorylation of clathrin that causes lateral diffusion of receptor-ligand complexes to clathrin-coated pits, membrane invagination and generation of clathrin-coated vesicles, and/or initiates phosphorylation of Cbl that may directly ubiquitinate the receptor. Phosphorylated Cbl triggers phosphorylation of HSP70 that facilitates un-coating of the vesicles, a precondition for vesicles to fuse with early endosomes and release ligands. The released receptors are transported to either the late endosome and/or lysosome for proteosomal and/or lysosomal degradation or are recycled to the cell surface. The immune complexes in the endosome are either directly degraded by Cbl, or delivered to the lysosome by phosphorylated HGS-STAM 1/2 complex for final degradation. During transfer, immune complexes that escape the endosome are recognised by phosphorylated TRIM21 for proteasomal degradation. Co-ligation of  $Fc\gamma RI$  with LILRB4 may recruit phosphatases such as SHP-1 to its ITIMs that subsequently dephosphorylate (deactivate) the key molecules including clathrin (1),  $Fc\gamma RI$  and Syk (2), Cbl (3), HGS and STAM 1/2 (4) and TRIM21 (5). These effects may reduce cellular activation and/or suppress receptor/ligand endocytosis. \*New Tyr phosphorylated and dephosphorylated proteins identified in this study.

p-Syk proteins, 20  $\mu g$  of total lysates were serially Western blotted using rabbit anti-pSyk Ab followed by rabbit anti-Syk Ab and mouse anti- $\beta$ -actin mAb.

#### Detection of uptake of antibody opsonised bacteria particles by differentiated THP-1 cells and modulation by LILRB4.

To determine uptake of antibody-opsonised bacteria by phorbol 12-myristate 13-acetate (PMA) differentiated THP-1 cells a modified Fc-receptor dependent endocytosis/phagocytosis assay was developed. In brief, desired numbers of THP-1 cells were cultured in RPMI complete medium containing 100 ng/ml PMA (Sigma-Aldrich) at 37°C in 5% CO<sub>2</sub> for 3 days. Differentiation was confirmed by assessing morphological changes; larger, partially adherent cells, vesicular with ruffled edges and non-replicating. Enhanced green fluorescent protein (EGFP) expressing DH5 $\alpha$  *E. coli* (Addgene, Cambridge, MA, USA) (10<sup>7</sup> cells/ml PBS) were killed by freeze-thawing twice and pellets incubated with 5  $\mu g$ /ml goat anti-DH5 $\alpha$  *E. coli* in PBS (Abcam 25823, Melbourne, VIC, Australia) for 2 hrs at 37°C. The opsonised particles were added to 2  $\times$  10<sup>5</sup> PMA-differentiated THP-1 cells in 400  $\mu l$  CLB, at an estimated bacteria to cell ratio of 10:1, and incubated at 37°C for 4 hrs. Cells were then washed twice in 1.5 ml cold PBS containing 0.05% NaN<sub>3</sub> and 1% bovine serum albumin, and re-suspended in 0.5 ml of 1% paraformaldehyde in PBS. The percentage of cells that took up bacterial particles was determined by flow cytometry. To confirm Fc-dependent uptake,  $Fc\gamma RI$  (the primary Fc receptor expressed on THP-1 cells<sup>19</sup>) function was blocked by pre-incubating cells with 20  $\mu g$ /ml anti- $Fc\gamma RI$  mAb for 15 min at RT followed by a single CLB wash prior to addition of opsonised bacteria particles; 20  $\mu g$ /ml irrelevant mouse

negative control IgG<sub>1</sub> mAb (Sigma-Aldrich) was used as a control. A total of  $2 \times 10^4$  events were acquired using BD FACSCalibur™, and data analysed using Cell Quest software (BD Biosciences, Mountain View, CA, USA).

To assess the effect of LILRB4 ligation on the uptake of the opsonised DH5 $\alpha$  *E. coli* particles,  $2 \times 10^5$  differentiated THP-1 cells were resuspended in 100  $\mu$ l CLB and incubated with 10  $\mu$ g/ml anti-LILRB4 mAb for 20 min at RT, followed by ligation using 15  $\mu$ g/ml goat anti-mouse IgG<sub>1</sub> (Fc-specific) secondary Ab at RT for 10 min, and a single CLB wash prior to addition of the *E. coli* particles. Mouse anti-MHC-I mAb (anti-HLA-ABC) was used as a relevant surface-binding control Ab.

**Statistical analysis.** Fisher's exact test was used to determine the most significantly enriched pathways as predicted by Ingenuity Pathway Analysis software. Western blots were semi-quantified by densitometry using ImageJ software (<http://rsbweb.nih.gov/ij>) and compared using one-way ANOVA with Dunnett post-test for multiple comparisons. The mean percentages of differentiated THP-1 cells that took up bacterial particles without LILRB4 ligation were compared with cells pre-ligated with anti-LILRB4, or treated with control anti-MHC-I mAb using two-tailed unpaired t-test. P values < 0.05 were considered statistically significant.

## References

- Nimmerjahn, F. & Ravetch, J. V. Fc $\gamma$  receptors as regulators of immune responses. *Nature reviews. Immunology* **8**, 34–47, doi: 10.1038/nri2206 (2008).
- Fridman, W. H. Fc receptors and immunoglobulin binding factors. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* **5**, 2684–2690 (1991).
- Smith, K. G. & Clatworthy, M. R. Fc $\gamma$ RIIB in autoimmunity and infection: evolutionary and therapeutic implications. *Nature reviews. Immunology* **10**, 328–343, doi: 10.1038/nri2762 (2010).
- Davis, W., Harrison, P. T., Hutchinson, M. J. & Allen, J. M. Two distinct regions of FC gamma RI initiate separate signalling pathways involved in endocytosis and phagocytosis. *The EMBO journal* **14**, 432–441 (1995).
- Debets, J. M., Van der Linden, C. J., Dieteren, I. E., Leeuwenberg, J. F. & Buurman, W. A. Fc-receptor cross-linking induces rapid secretion of tumor necrosis factor (cachectin) by human peripheral blood monocytes. *Journal of immunology* **141**, 1197–1201 (1988).
- Molfetta, R. *et al.* Regulation of fc receptor endocytic trafficking by ubiquitination. *Frontiers in immunology* **5**, 449, doi: 10.3389/fimmu.2014.00449 (2014).
- Huang, Z. Y. *et al.* Differential kinase requirements in human and mouse Fc-gamma receptor phagocytosis and endocytosis. *Journal of leukocyte biology* **80**, 1553–1562, doi: 10.1189/jlb.0106019 (2006).
- Norman, J. C. & Allen, J. M. Endocytosis of Fc $\gamma$ RI is regulated by two distinct signalling pathways. *FEBS letters* **484**, 179–183 (2000).
- Veiga, E. & Cossart, P. The role of clathrin-dependent endocytosis in bacterial internalization. *Trends in cell biology* **16**, 499–504, doi: 10.1016/j.tcb.2006.08.005 (2006).
- Le Roy, C. & Wrana, J. L. Clathrin- and non-clathrin-mediated endocytic regulation of cell signalling. *Nature reviews. Molecular cell biology* **6**, 112–126, doi: 10.1038/nrm1571 (2005).
- Meier, O. & Greber, U. F. Adenovirus endocytosis. *The journal of gene medicine* **6** Suppl 1, S152–163, doi: 10.1002/jgm.553 (2004).
- Doherty, G. J. & McMahon, H. T. Mechanisms of endocytosis. *Annual review of biochemistry* **78**, 857–902, doi: 10.1146/annurev.biochem.78.081307.110540 (2009).
- DeLuca-Flaherty, C., McKay, D. B., Parham, P. & Hill, B. L. Uncoating protein (hsc70) binds a conformationally labile domain of clathrin light chain LCa to stimulate ATP hydrolysis. *Cell* **62**, 875–887 (1990).
- Newmyer, S. L. & Schmid, S. L. Dominant-interfering Hsc70 mutants disrupt multiple stages of the clathrin-coated vesicle cycle *in vivo*. *The Journal of cell biology* **152**, 607–620 (2001).
- Hirano, S. *et al.* Double-sided ubiquitin binding of Hrs-UIM in endosomal protein sorting. *Nature structural & molecular biology* **13**, 272–277, doi: 10.1038/nsmb1051 (2006).
- Raiborg, C. *et al.* Hrs sorts ubiquitinated proteins into clathrin-coated microdomains of early endosomes. *Nature cell biology* **4**, 394–398, doi: 10.1038/ncb791 (2002).
- McEwan, W. A. *et al.* Intracellular antibody-bound pathogens stimulate immune signaling via the Fc receptor TRIM21. *Nature immunology* **14**, 327–336, doi: 10.1038/ni.2548 (2013).
- Mancardi, D. A. *et al.* The high-affinity human IgG receptor Fc $\gamma$ RI (CD64) promotes IgG-mediated inflammation, anaphylaxis, and antitumor immunotherapy. *Blood* **121**, 1563–1573, doi: 10.1182/blood-2012-07-442541 (2013).
- Lu, H. K. *et al.* Leukocyte Ig-like receptor B4 (LILRB4) is a potent inhibitor of Fc $\gamma$ RI-mediated monocyte activation via dephosphorylation of multiple kinases. *The Journal of biological chemistry* **284**, 34839–34848, doi: 10.1074/jbc.M109.035683 (2009).
- Cella, M. *et al.* A novel inhibitory receptor (ILT3) expressed on monocytes, macrophages, and dendritic cells involved in antigen processing. *The Journal of experimental medicine* **185**, 1743–1751 (1997).
- Debets, J., Van de Winkel, J., Ceuppens, J., Dieteren, I. & Buurman, W. Cross-linking of both Fc gamma RI and Fc gamma RII induces secretion of tumor necrosis factor by human monocytes, requiring high affinity Fc-Fc gamma R interactions. Functional activation of Fc gamma RII by treatment with proteases or neuraminidase. *The Journal of Immunology* **144**, 1304–1310 (1990).
- Krutmann, J. *et al.* Cross-linking Fc receptors on monocytes triggers IL-6 production. Role in anti-CD3-induced T cell activation. *The Journal of Immunology* **145**, 1337–1342 (1990).
- Zhang, W., Sloan-Lancaster, J., Kitchen, J., Tribble, R. P. & Samelson, L. E. LAT: the ZAP-70 tyrosine kinase substrate that links T cell receptor to cellular activation. *Cell* **92**, 83–92 (1998).
- Vivanco, I. & Sawyers, C. L. The phosphatidylinositol 3-Kinase AKT pathway in human cancer. *Nature reviews. Cancer* **2**, 489–501, doi: 10.1038/nrc839 (2002).
- Akira, S., Takeda, K. & Kaisho, T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nature immunology* **2**, 675–680, doi: 10.1038/90609 (2001).
- Nakamura, K., Malykhin, A. & Coggeshall, K. M. The Src homology 2 domain-containing inositol 5-phosphatase negatively regulates Fc $\gamma$  receptor-mediated phagocytosis through immunoreceptor tyrosine-based activation motif-bearing phagocytic receptors. *Blood* **100**, 3374–3382, doi: 10.1182/blood-2002-03-0787 (2002).
- Schemarova, I. V. The role of tyrosine phosphorylation in regulation of signal transduction pathways in unicellular eukaryotes. *Current issues in molecular biology* **8**, 27–49 (2006).
- Hunter, T. The Croonian Lecture 1997. The phosphorylation of proteins on tyrosine: its role in cell growth and disease. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* **353**, 583–605, doi: 10.1098/rstb.1998.0228 (1998).
- Mallery, D. L. *et al.* Antibodies mediate intracellular immunity through tripartite motif-containing 21 (TRIM21). *Proceedings of the National Academy of Sciences of the United States of America* **107**, 19985–19990, doi: 10.1073/pnas.1014074107 (2010).

30. Shi, Y. *et al.* Protein-tyrosine kinase Syk is required for pathogen engulfment in complement-mediated phagocytosis. *Blood* **107**, 4554–4562 (2006).
31. Majeed, M., Cavegion, E., Lowell, C. A. & Berton, G. Role of Src kinases and Syk in Fc $\gamma$  receptor-mediated phagocytosis and phagosome-lysosome fusion. *Journal of leukocyte biology* **70**, 801–811 (2001).
32. Lau, C. *et al.* Syk associates with clathrin and mediates phosphatidylinositol 3-kinase activation during human rhinovirus internalization. *Journal of immunology* **180**, 870–880 (2008).
33. Rejman, J., Oberle, V., Zuhorn, I. S. & Hoekstra, D. Size-dependent internalization of particles via the pathways of clathrin- and caveolae-mediated endocytosis. *The Biochemical journal* **377**, 159–169, doi: 10.1042/BJ20031253 (2004).
34. Bonazzi, M. *et al.* Clathrin phosphorylation is required for actin recruitment at sites of bacterial adhesion and internalization. *The Journal of cell biology* **195**, 525–536, doi: 10.1083/jcb.201105152 (2011).
35. Tanaka, S., Neff, L., Baron, R. & Levy, J. B. Tyrosine phosphorylation and translocation of the c-cbl protein after activation of tyrosine kinase signaling pathways. *The Journal of biological chemistry* **270**, 14347–14351 (1995).
36. Feshchenko, E. A., Langdon, W. Y. & Tsygankov, A. Y. Fyn, Yes, and Syk phosphorylation sites in c-Cbl map to the same tyrosine residues that become phosphorylated in activated T cells. *The Journal of biological chemistry* **273**, 8323–8331 (1998).
37. Urbe, S. *et al.* The UIM domain of Hrs couples receptor sorting to vesicle formation. *Journal of cell science* **116**, 4169–4179, doi: 10.1242/jcs.00723 (2003).
38. Muller, P. *et al.* C-terminal phosphorylation of Hsp70 and Hsp90 regulates alternate binding to co-chaperones CHIP and HOP to determine cellular protein folding/degradation balances. *Oncogene* **32**, 3101–3110, doi: 10.1038/ncr.2012.314 (2013).
39. Katz, H. R. Inhibition of inflammatory responses by leukocyte Ig-like receptors. *Advances in immunology* **91**, 251–272, doi: 10.1016/S0065-2776(06)91007-4 (2006).
40. Ravetch, J. V. & Lanier, L. L. Immune inhibitory receptors. *Science* **290**, 84–89 (2000).
41. Brown, D. P. *et al.* The inhibitory receptor LILRB4 (ILT3) modulates antigen presenting cell phenotype and, along with LILRB2 (ILT4), is upregulated in response to Salmonella infection. *BMC immunology* **10**, 56, doi: 10.1186/1471-2172-10-56 (2009).
42. Stacey, K. B., Breen, E. & Jefferies, C. A. Tyrosine phosphorylation of the E3 ubiquitin ligase TRIM21 positively regulates interaction with IRF3 and hence TRIM21 activity. *PloS one* **7**, e34041, doi: 10.1371/journal.pone.0034041 (2012).
43. Park, M. *et al.* Hypersensitivity Pneumonitis Caused by House Cricket, *Acheta domesticus*. *Journal of Clinical & Cellular Immunology* **5**, doi: 10.4172/2155-9899.1000248 (2014).
44. Lee, T. H. *et al.* Glycosylation in a mammalian expression system is critical for the production of functionally active leukocyte immunoglobulin-like receptor A3 protein. *The Journal of biological chemistry* **288**, 32873–32885, doi: 10.1074/jbc.M113.478578 (2013).

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## Author Contributions

M.P. and N.T. designed experiments and prepared the manuscript. M.P. did the majority of the experiments and M.J.R. performed mass spectrometry and IPA analysis. N.T., P.S.T., C.L.G. and K.B. provided advice for the experiments and preparation of the manuscript. N.T. directed the research.

## Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

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