

SCIENTIFIC REPORTS



OPEN

Wnt/ β -catenin signaling pathway activation is required for proliferation of chicken primordial germ cells *in vitro*

Hyung Chul Lee^{1,2}, Sumi Lim^{1,2} & Jae Yong Han^{1,3}

Received: 24 May 2016
 Accepted: 14 September 2016
 Published: 30 September 2016

Here, we investigated the role of the Wnt/ β -catenin signaling pathway in chicken primordial germ cells (PGCs) *in vitro*. We confirmed the expression of Wnt signaling pathway-related genes and the localization of β -catenin in the nucleus, revealing that this pathway is potentially activated in chicken PGCs. Then, using the single-cell pick-up assay, we examined the proliferative capacity of cultured PGCs in response to Wnt ligands, a β -catenin-mediated Wnt signaling activator (6-bromoindirubin-3'-oxime [BIO]) or inhibitor (JW74), in the presence or absence of basic fibroblast growth factor (bFGF). WNT1, WNT3A, and BIO promoted the proliferation of chicken PGCs similarly to bFGF, whereas JW74 inhibited this proliferation. Meanwhile, such treatments in combination with bFGF did not show a synergistic effect. bFGF treatment could not rescue PGC proliferation in the presence of JW74. In addition, we confirmed the translocation of β -catenin into the nucleus by the addition of bFGF after JW74 treatment. These results indicate that there is signaling crosstalk between FGF and Wnt, and that β -catenin acts on PGC proliferation downstream of bFGF. In conclusion, our study suggests that Wnt signaling enhances the proliferation of chicken PGCs via the stabilization of β -catenin and activation of its downstream genes.

Primordial germ cells (PGCs), the precursors of sperm or oocytes, undergo numerous spatially and temporally specific regulatory pathways during early development¹. It has been revealed that these complex regulatory networks in PGCs are governed by several extrinsic factors leading to intracellular pathways². Although the molecular mechanisms underlying signal transduction in PGCs remain poorly defined, leukemia inhibitory factor, basic fibroblast growth factor (bFGF), bone morphogenetic protein 4, and kit ligand are the best studied signaling pathways known to control PGC survival and proliferation³.

Previously, several studies reported a list of PGC specific genes by transcriptome analysis containing Wnt/ β -catenin signaling-related genes^{4,5}. Also, while we were searching for target genes regulated by micro RNA (miRNA) specifically expressed in PGCs based on the previous result⁶, we found that several negative regulators for Wnt signaling pathway, such as Glycogen synthase kinase-3- β (GSK3 β), APC, and AXIN, might be target for those miRNAs, indicating possible role of Wnt pathway in chicken PGCs.

The Wnt pathway is a well-known signaling cascade that is involved in PGC development including specification, migration, and proliferation^{7–10}. The Wnt/ β -catenin pathway, or so-called canonical Wnt signaling, maintains embryonic stem (ES) cells in an undifferentiated state and promotes proliferative capacity in both stem cells and cancer cells^{11,12}. β -catenin is a key mediator of canonical Wnt signaling, and acts as a transcriptional co-activator to control the expression of cell cycle-related genes^{13,14}. The importance of Wnt/ β -catenin signaling in the proliferation of PGCs has been reported in mice and *Drosophila*^{8,15,16}, but not in chickens.

Several inhibitors, such as 6-bromoindirubin-3'-oxime (BIO) and JW74, have been utilized to activate or block such signaling. BIO, an inhibitor of the negative regulator GSK3 β , leads to target gene expression by stabilizing β -catenin from the degradation complex^{11,17}, whereas JW74 stabilizes AXIN, a component of the β -catenin

¹Department of Agricultural Biotechnology, College of Agriculture and Life Sciences, and Research Institute of Agriculture and Life Sciences, Seoul National University, Seoul 08826, Korea. ²Department of Cell and Developmental Biology, University College London, London, United Kingdom. ³Institute for Biomedical Sciences, Shinshu University, Minamiminowa, Nagano 399-4598, Japan. Correspondence and requests for materials should be addressed to J.Y.H. (email: jaehan@snu.ac.kr)

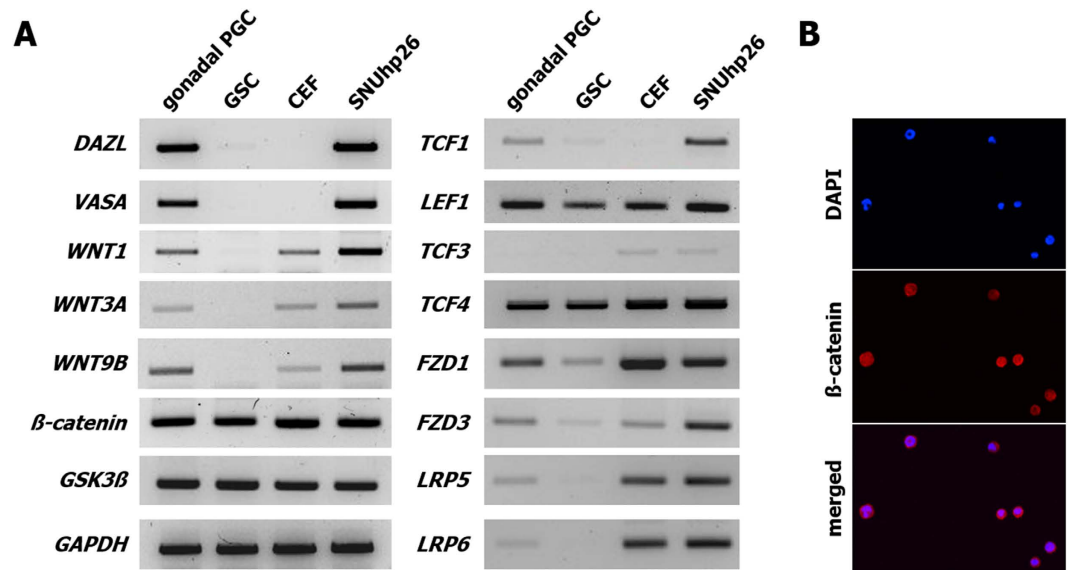


Figure 1. Expression of Wnt signaling pathway-related genes in cultured chicken primordial germ cells (PGCs). (A) The expression of Wnt signaling pathway-related genes including *WNT1*, *WNT3A*, *WNT9B*, *FZD1*, *FZD3*, *LRP5*, *LRP6*, β -catenin, *GSK3 β* , *TCF1*, *LEF1*, *TCF3*, and *TCF4* and germ cell-related genes including *DAZL* and *VASA* was analyzed using reverse transcription polymerase chain reaction (RT-PCR) in gonadal PGCs and gonadal stromal cells (GSCs), which were separated from day 6 embryonic gonads, chicken embryonic fibroblasts (CEFs), and a cultured chicken PGC line, SNUhp26. Most of the Wnt signaling pathway-related genes were upregulated in SNUhp26 compared to those in gonadal PGCs. (B) Immunostaining showed the localization of β -catenin in both the nucleus and cytoplasm in cultured chicken PGCs.

degradation complex, eventually causing β -catenin degradation¹⁸. Accordingly, BIO and JW74 were used in this study to modulate the level of β -catenin in chicken PGCs *in vitro*.

Another growth factor family involved in cell proliferation, migration, and differentiation during embryonic development is bFGF, also known as FGF2^{19,20}. bFGF is a powerful mitogen in PGC culture in mammals²¹ and supports the proliferation of embryonic germ cells²² and PGCs in chickens²³. Choi *et al.*²⁴ and Macdonald *et al.*²⁵ identified bFGF as an essential factor for *in vitro* PGC proliferation^{24,25}. Among several downstream cascades of bFGF, including the PI3K/AKT and MEK/ERK pathways, chicken PGCs mediate MEK/ERK signaling for proliferation²⁴. Recently, Whyte *et al.*²⁶ reported that three different signals, such as TGF beta, FGF, and Insulin are required for PGC proliferation in chicken, suggesting presence of combinatorial action of several signaling pathways²⁶. Meanwhile, numerous recent studies have demonstrated the crosstalk between FGF and Wnt/ β -catenin signaling. For example, β -catenin is activated by FGF signaling through the RAS/ERK or PI3K/AKT cascade, and acts downstream of these pathways in several stem cells or progenitor cells^{27–30}. In addition, FGF can activate Wnt signaling by phosphorylation of the Wnt co-receptor LRP6^{31,32} or inactivation of the Wnt inhibitor Dkk1³³.

In this study, the expression of Wnt/ β -catenin signaling-related genes was examined to determine whether PGCs are capable of activating this signaling pathway. BIO and JW74 were applied to modulate Wnt/ β -catenin signaling in cultured chicken PGCs. Subsequently, an *in vitro* proliferation assay was performed in the presence of a Wnt ligand, BIO, JW74, or bFGF to verify the signaling mechanism underlying PGC proliferation in chickens. Lastly, to verify the crosstalk between bFGF and Wnt/ β -catenin signaling in PGC proliferation, Western blotting and immunocytochemistry were used to examine the translocation of β -catenin into the nucleus, which was stimulated by bFGF after JW74 treatment.

Results

Expression of Wnt signaling-related genes in chicken PGCs. To examine whether chicken PGCs have the ability to process Wnt/ β -catenin signaling, RT-PCR was performed to analyze several components of this pathway. Among the many Wnt ligands and receptors, there was confirmed expression of *WNT1*, *WNT3A*, and *WNT9B*, which are well-known proteins in the canonical Wnt pathway; Frizzled receptors 1 and 3, which interact with *WNT3A*^{34,35}; and *LRP5* and *6*, which are co-receptors of canonical Wnt signaling^{36–38} (Fig. 1A). In light of these findings, Wnt ligands and their receptors were well expressed in both gonadal and cultured PGCs (SNUhp26), and other key factors such as β -catenin, *GSK3 β* , and *TCFs* (except for *TCF3*) were also expressed in gonadal stromal cells and chicken embryonic fibroblasts as well as gonadal PGCs and cultured PGCs, indicating that chicken PGCs might be capable of activating Wnt/ β -catenin signaling (Fig. 1A). In particular, most Wnt signaling pathway-related genes, such as all three Wnt ligands, *FZDs*, *LRPs*, *TCFs*, and *LEF1*, were highly expressed in cultured PGCs compared to those in gonadal PGCs. To determine the status of β -catenin in PGCs, cultured PGCs were subjected to immunostaining (Fig. 1B). The results showed that β -catenin was localized in both the nucleus and cytoplasm, indicating that it is stabilized in cultured PGCs.

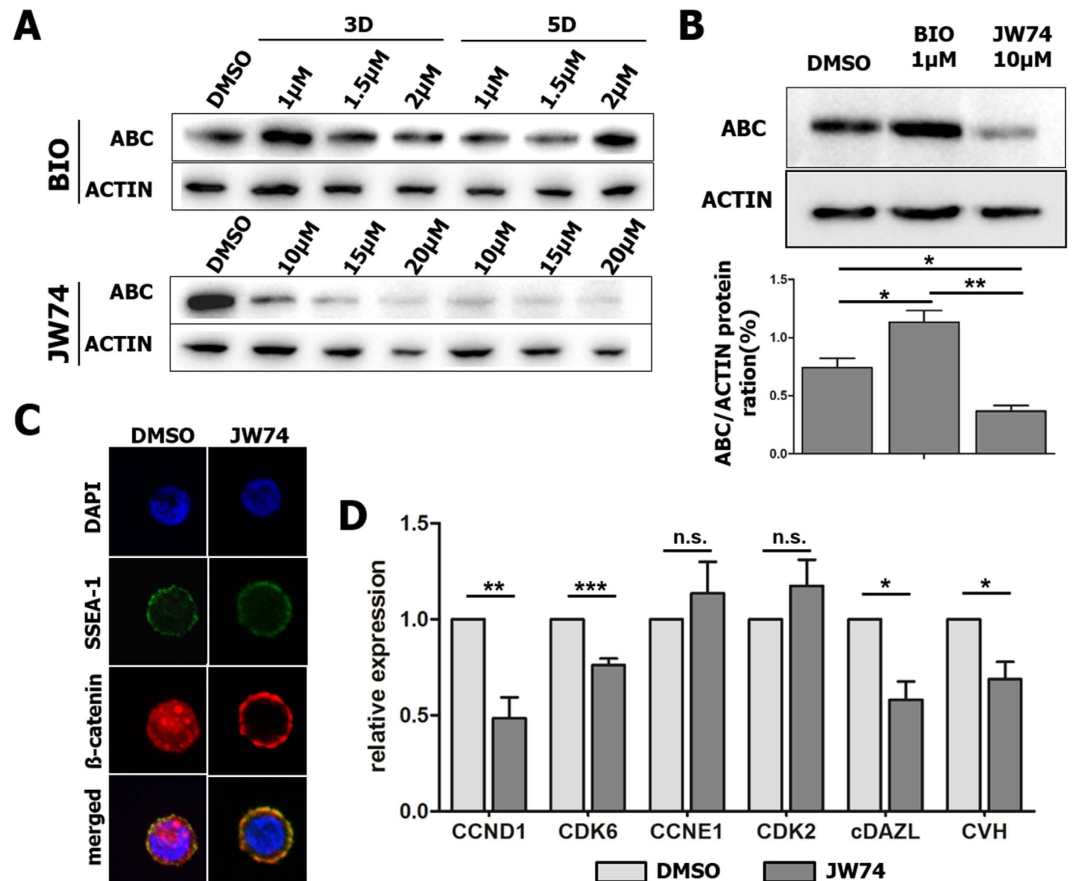


Figure 2. Regulation of β -catenin signaling by 6-bromoindirubin-3'-oxime (BIO) or JW74 treatment for optimization of chicken PGCs (SNUhp26) *in vitro*. (A) The level of active β -catenin (ABC) changed after BIO or JW74 treatment in a dose-dependent manner, as determined by Western blotting. (B) BIO applied at 1 μ M and JW74 at 10 μ M for 3 days showed optimal effects to increase or decrease the β -catenin level. BIO, a GSK3 β inhibitor; JW74, a β -catenin inhibitor. (C) Immunostaining showed that JW74 treatment caused the degradation of β -catenin in both the nucleus and cytoplasm in chicken PGCs. (D) JW74 treatment caused downregulation of cell cycle-related genes essential for entry in G₁ phase (*CCND1* and *CDK6*) and germ cell-specific genes (*cDAZL* and *CVH*), but no effect on cell cycle-related genes regulating progression from G₁ to S phase (*CCNE1* and *CDK2*) as confirmed by qRT-PCR. Values in (B) and (D) represent mean \pm standard error of the mean (SEM) of triplicate analyses. * $P < 0.05$, ** $P < 0.01$, and *** $P < 0.001$. n.s.; not significant.

Verification of the role of Wnt/ β -catenin signaling in chicken PGCs via BIO and JW74. In several types of cells, such as cancer cells and ES cells, BIO and JW74 have been used for modulating β -catenin as an activator and an inhibitor, respectively^{17,18,39}. To test whether β -catenin also works in cultured chicken PGCs, BIO and JW74 were administered at different concentrations for 3 and 5 days (Fig. 2A). Their effects were detected by Western blotting with the antibody for active β -catenin (ABC), which is the non-phosphorylated form (Fig. 2A). Among the various conditions, 1 μ M BIO showed optimal effect of Wnt signaling activation in cultured chicken PGCs when it was applied for 3 rather than 5 days (Fig. 2B). In the case of JW74, the levels of ABC protein gradually decreased in a dose-dependent manner. However, the level of β -actin also decreased when JW74 was applied at a concentration greater than 10 μ M, meaning that it was toxic to chicken PGCs (Fig. 2A). Thus, 1 μ M BIO and 10 μ M JW74 were used for subsequent experiments in this study (Fig. 2B).

To clarify the importance of Wnt/ β -catenin signaling in PGCs, loss-of-function studies were conducted by treating cultured PGCs with JW74. JW74 treatment caused the degradation of β -catenin in both the nucleus and cytoplasm, indicating that Wnt signaling had been inactivated (Fig. 2C). JW74 treatment also caused the downregulation of cell cycle-related genes essential for entry in G₁ phase, such as *CCND1* and *CDK6*, and germ cell-specific genes, such as *cDAZL* and *CVH*, while no effect on cell cycle-related genes regulating progression from G₁ to S phase, such as *CCNE1* and *CDK2* (Fig. 2D).

Proliferation assay of chicken PGCs with Wnt ligand, BIO or JW74. To investigate whether Wnt/ β -catenin is involved in PGC proliferation *in vitro* and related to bFGF signaling in chicken PGCs, a Wnt ligand (WNT1, WNT3A, and WNT9B), BIO or JW74, were applied at different concentrations, after which a cell proliferation assay was performed (Fig. 3). To simplify determination of the cell number, SNUhp26 male chicken PGCs expressing GFP were used. After bFGF starvation for 4 days to remove all of the effects of bFGF signaling, a single PGC was added to a culture under each set of conditions. Then, the number of PGCs was counted on day

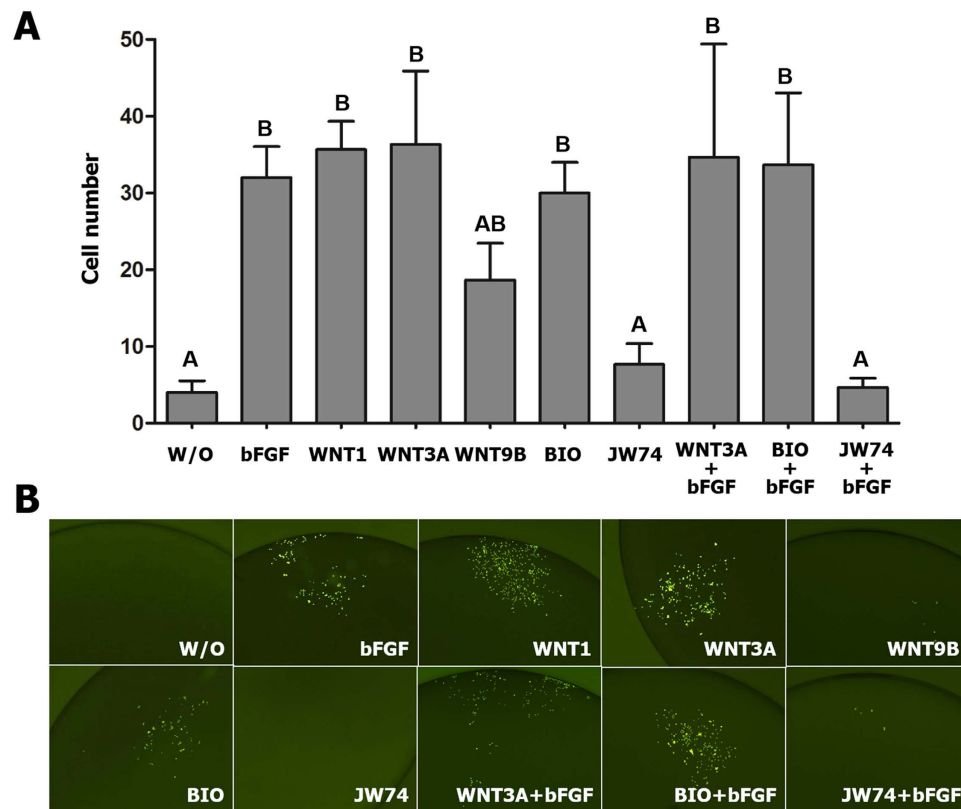


Figure 3. The effects of BIO and JW74 on the proliferation of cultured chicken PGCs (SNUhp26). Proliferation assay was performed with SNUhp26 PGCs, expressing green fluorescent protein (GFP). After bFGF starvation for 4 days, a single PGC was added to each medium containing bFGF, WNT1, WNT3A, WNT9B, BIO, or JW74 with or without bFGF, and cultured for 8 days. (A) The number of PGCs in each treatment was counted after 8 days. Values represent mean \pm SEM of triplicate analyses. Values with different letters are significantly different ($P < 0.05$). (B) The GFP-expressing PGCs after each treatment were observed under a fluorescence microscope.

8 of culture, and GFP expression was observed (Fig. 3A,B). The results showed that, in the experimental sets with bFGF, WNT1, WNT3A, BIO, WNT3A with bFGF, and BIO with bFGF, cell number had significantly increased at day 8 compared to that in culture without a growth factor, in which GFP-expressing PGCs were finally lost after 10 days of culture. In the WNT9B group, an intermediate effect was shown. This indicates that β -catenin as well as bFGF can activate chicken PGCs to proliferate *in vitro* (Fig. 3A,B). On the other hand, JW74-treated PGCs did not grow, as the group without any growth factor showed the same pattern. In the group treated with JW74 supplemented with bFGF, this treatment could not rescue the inhibition of cell growth (Fig. 3A,B).

β -catenin-mediated activation of chicken PGCs through bFGF. Since bFGF is one of the key factors that enable the proliferation of chicken PGCs via MEK/ERK signaling²⁴, we assumed that β -catenin is a downstream gene of bFGF in PGCs. To prove this, we used immunostaining to confirm the translocation of β -catenin into the nucleus from the cytoplasm (Fig. 4). After JW74 treatment (10 μ M) for 2 days, most β -catenin in both the nucleus and cytoplasm had been degraded and was only detected in the membrane (Fig. 4A). When β -catenin had been almost completely cleared from the nucleus and cytoplasm, bFGF was added for 8 h. As a result, the translocation of β -catenin into the nucleus significantly increased (Fig. 4A,B), indicating that bFGF can activate and stabilize β -catenin to promote the proliferation of chicken PGCs.

Discussion

Compared to other somatic cells, primordial germ cells have unique spatial and temporal developmental characteristics¹, and several signaling pathways play important roles in regulating their development². Among them, Wnt signaling has also been shown to be involved in PGC development in several model animals^{16,40,41}, but not yet in chicken. In this study, based on the expression of Wnt signaling-related genes in chicken PGCs, we suggested that the Wnt pathway might play an important role in PGCs in chicken. Most Wnt signaling pathway-related genes were upregulated in cultured PGCs compared to those in gonadal PGCs, indicating that Wnt signaling might be more activated in these specific conditions, *in vitro*, to facilitate proliferation. In addition, β -catenin, an important mediator of Wnt signaling, was stabilized in cultured PGCs considering their localization in both the nucleus and the cytoplasm. These results suggest that chicken PGCs can activate and process the Wnt signaling pathway.

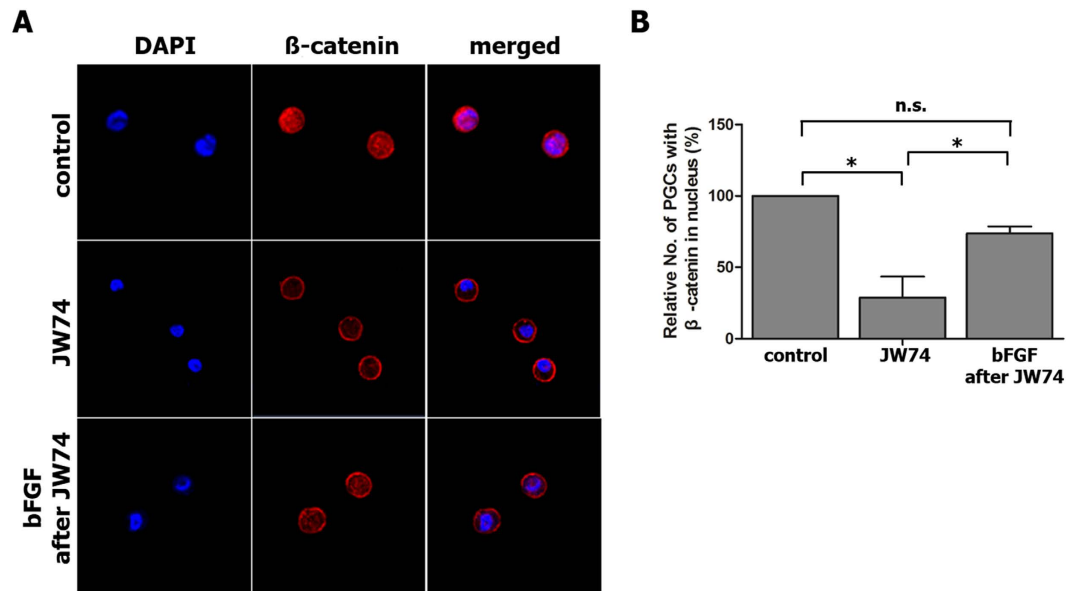


Figure 4. Activation and stabilization of β -catenin by bFGF treatment of chicken PGCs (SNUhp26).

(A) Immunostaining for β -catenin showed that the JW74 treatment caused the degradation of β -catenin in both the nucleus and the cytoplasm in cultured chicken PGCs. When bFGF was added followed by JW74 treatment, β -catenin translocated into the nucleus from the membrane. (B) The relative number of PGCs with β -catenin located in the nucleus was counted after each treatment. * $P < 0.05$. n.s.; not significant.

Aberrant activation of the canonical Wnt/ β -catenin pathway is one of the most common signaling abnormalities in human and mouse cancer^{42,43}, because it activates numerous genes related to cell cycle progression. To investigate the role of the Wnt/ β -catenin pathway in chicken PGCs, we used BIO and JW74, well-known inhibitors of GSK3 β and β -catenin, respectively^{17,18}. We confirmed that both BIO and JW74, respectively, could activate and inactivate β -catenin in cultured PGCs by modulating the level of its active form (ABC). When β -catenin is activated and stabilized in the cytoplasm, it is transported into the nucleus, leading to formation of a transcriptional complex with the TCF transcription factor⁴⁴. Specific gene expression mediated by β -catenin activation generally occurs in a cell-context and cell-dependent manner. Cyclin D1, MYC, and cJun are the best-known target genes that enhance proliferation⁴⁵. In this study, to investigate the role of Wnt/ β -catenin signaling, loss-of-function studies were conducted by applying JW74. The results suggest that inhibition of Wnt/ β -catenin signaling induces cell cycle arrest by downregulation of CCND1 and CDK6⁴⁶, revealing that Wnt/ β -catenin signaling is required for the maintenance of PGC proliferation.

Next, to test whether Wnt/ β -catenin is involved in the proliferation of chicken PGCs, we activated the Wnt signaling pathway in PGCs using Wnt ligands or the activation of a component downstream of it, β -catenin, using BIO. The results of our proliferation assay suggested that the activation of Wnt/ β -catenin signaling induced the proliferation of PGCs *in vitro*, while JW74 treatment prevented this. Also, the proliferative ability induced by the activation of Wnt/ β -catenin signaling was similar as those induced by bFGF, which is known as a major proliferation inducer in chicken PGCs^{23–25}. These results suggest that Wnt/ β -catenin signaling is required for PGC proliferation *in vitro*.

Our proliferation assay confirmed two other findings. First, no synergistic effect was observed upon the treatment of Wnt ligands or BIO in combination with bFGF, suggesting that bFGF and Wnt do not act independently of each other, and that PGCs already reach the threshold of proliferation capacity in the presence of either bFGF or Wnt alone. Second, bFGF treatment could not rescue PGC proliferation in the presence of JW74 (in the absence of Wnt/ β -catenin signaling), indicating that bFGF helps PGCs to proliferate through direct activation of the downstream Wnt/ β -catenin signaling pathway. Previous studies on the crosstalk between β -catenin and bFGF support our results. For example, in the absence of bFGF, β -catenin is only located in the membrane with E-cadherin, whereas it accumulates and is stably translocated into the nucleus to activate downstream genes in the presence of bFGF^{14,27}. In the presence of bFGF, β -catenin helps neural stem cells proliferate when it is overexpressed, while enhancing neuronal differentiation in the absence of bFGF²⁸. Moreover, bFGF increases β -catenin signaling in neural progenitor cells via multiple mechanisms: increasing the level of β -catenin mRNA, nuclear translocation of β -catenin, and phosphorylation of GSK-3 β ⁴⁷. Because the role of β -catenin in the proliferation of chicken PGCs was recently reported⁴⁸, in this study, we focused on evaluating the crosstalk between bFGF and β -catenin in chicken PGCs. First, we applied JW74 for the degradation of β -catenin and then added bFGF to the culture medium. Our results showed the reactivation of β -catenin by bFGF treatment after JW74. These results suggest that bFGF signaling promotes the proliferation of chicken PGCs via the activation of β -catenin and its downstream signaling. Many studies have shown that β -catenin is a target gene of FGFR signaling through RAS/ERK or PI3K/AKT cascades^{27–30}. Therefore, it is worthwhile studying which downstream signals are involved in their crosstalk in PGCs.

In our study, the role of the Wnt/ β -catenin signaling pathway was investigated for the first time in chicken PGCs, and the relationship between Wnt/ β -catenin and bFGF was revealed. We concluded that Wnt/ β -catenin can enhance the proliferation of chicken PGCs, and act as a factor downstream of bFGF signaling. Interestingly, in mouse, Wnt/ β -catenin signaling is suppressed and deleterious for PGC proliferation⁸. Thus, our results suggest that, although many key signaling pathways for PGC development are conserved among animal species, the specific mechanism of regulation could differ. In addition, the recent report suggests the requirement of combinatorial action of different signals on PGC self-renewal²⁶. Thus, a following study could be investigation on epistasis and crosstalk among different signaling pathways, such as TGF beta, FGF, Insulin, and WNT on PGC proliferation. Taken together, this research should contribute to studies on PGC biology and provide a comprehensive understanding of the mechanism behind PGC development from an evo-devo perspective.

Methods

Experimental animals and animal care. The care and experimental use of chickens were approved by the Institute of Laboratory Animal Resources, Seoul National University (SNU-150827-1). The chickens were maintained in accordance with a standard management program at the University Animal Farm, Seoul National University, South Korea. The procedures for animal management, reproduction, and embryo manipulation adhered to the standard operating protocols of our laboratory.

PGC isolation and culture. To conduct gene expression analysis, PGCs from White Leghorn embryonic gonads at day 6 (stage 28) were isolated by magnetic activated cell sorting as previously reported⁴⁹. Anti-stage-specific embryonic antigen (SSEA)-1 antibody-positive cells were regarded as gonadal PGCs, while anti-SSEA-1 antibody-negative cells were regarded as gonadal stromal cells (GSCs). For *in vitro* culture, SNUhp26 male line cultured PGCs⁵⁰, expressing green fluorescent protein (GFP), were maintained with knockout Dulbecco's Modified Eagle Medium (Thermo Fisher–Invitrogen, Carlsbad, CA, USA) supplemented with 20% (vol/vol) fetal bovine serum (Invitrogen), 2% (vol/vol) chicken serum (Sigma-Aldrich, St. Louis, MO, USA), 1× nucleosides (Millipore, Temecula, CA, USA), 2 mM L-glutamine, 1× nonessential amino acids, β -mercaptoethanol, 10 mM sodium pyruvate, and 1× antibiotic–antimycotic (Invitrogen). Human bFGF (Koma Biotech, Seoul, South Korea) was used at 10 ng/mL for PGC self-renewal. Chicken PGCs were cultured in an incubator maintained at 37 °C with an atmosphere of 5% CO₂ and 60–70% relative humidity. The cultured PGCs were subcultured onto mitomycin-inactivated murine embryonic fibroblasts in 5 to 6-day intervals by gentle pipetting without any enzyme treatment.

RT-PCR and qRT-PCR analyses. Total RNA was extracted from each sample with Trizol reagent (Invitrogen), in accordance with the manufacturer's protocol. The RNA quantity was determined by spectrophotometry at 260 nm, and 1 μ g of each RNA sample was reverse-transcribed with the Superscript III First-Strand Synthesis System (Invitrogen). The cDNA was diluted five-fold and quantitatively equalized for PCR amplification. RT-PCR was conducted to examine the change in expression of candidate genes in cultured chicken PGCs using specific primer sets (Table 1). PCR amplification was performed as follows: 94 °C for 5 min, followed by 26–32 cycles at 94 °C for 30 s, 58–62 °C for 30 s, and 72 °C for 30 s. The PCR products were mixed with SYBR[®] Safe (Invitrogen) and loaded for gel electrophoresis. Confirmation of the results was performed using the ChemiDoc XRS System (Bio-Rad Laboratories, Hercules, CA, USA). Downregulation of cell cycle- or germ cell-related genes at the mRNA level was analyzed using quantitative RT-PCR (qRT-PCR). Relative expression was calculated after the threshold cycle had been normalized to that of chicken glyceraldehyde 3-phosphate dehydrogenase.

Western blot analysis. Total protein was extracted with the CytoBuster[™] Protein Extraction Reagent (Novagen, Darmstadt, Germany), separated on a 10% polyacrylamide gel, and transferred to a polyvinylidene fluoride membrane (Millipore). The primary antibodies were mouse anti- β -actin (Santa Cruz Biotechnology, Santa Cruz, CA, USA) and rabbit anti-non-phospho (Active) β -catenin (Ser33/37, Thr41) (Cell Signaling, Danvers, MA, USA). Horseradish peroxidase-conjugated anti-mouse IgG and anti-rabbit IgG (Santa Cruz) were used as the secondary antibodies. The blots were treated with the enhanced chemiluminescence substrate solutions and exposed to a ChemiDoc XRS System (Bio-Rad) to detect chemiluminescence.

Proliferation assay by single-cell pick-up. SNUhp26 male line cultured PGCs, expressing GFP, were used for evaluating the rate of proliferation⁵⁰. The SNUhp26 has a full germ cell characteristics and potency that can produce germline chimera and its progeny⁵⁰. A total of 10 culture conditions were prepared in 96-well plates as follows: no growth factor (w/o GF), bFGF (10 ng/mL), WNT1 (100 ng/mL; Abcam, Cambridge, UK), WNT3A (100 ng/mL; Cell Signaling), WNT9B (400 ng/mL; R&D Systems, Minneapolis, MN, USA), BIO (Sigma; B1686), JW74 (Sigma; SML0227), WNT3A with bFGF, BIO with bFGF, and JW74 with bFGF. After 4 days of bFGF starvation, a single PGC was picked up and added to culture under each of the above conditions. The final concentrations of BIO and JW74 were 1 and 10 μ M, respectively. After 8 days of culture, the number of PGCs was counted by fluorescence microscopy.

Immunocytochemistry. Cells were moved from the culture plate and dried on HistoBond[®] adhesive microscope slides (Marienfeld-Superior, Lauda-Königshofen, Germany) for approximately 30 min. Then they were fixed in 100% methanol and prechilled for 3 min at –20 °C. After washing the cells three times for 10 min each in phosphate-buffered saline containing 1% normal goat serum, they were incubated in diluted anti- β -catenin antibody (mouse IgG) or anti-SSEA-1 (mouse IgM) antibody as a primary antibody at 4 °C for 24 h, and then incubated again in diluted Alexa-IgG-568 antibody or fluorescein isothiocyanate (FITC; Santa Cruz)-conjugated goat anti-mouse IgM, respectively, as a secondary antibody for 1 h at room temperature. Samples were then mounted

Gene name	Description	Accession No.	Primer Sequences (5' → 3')	
			Forward	Reverse
<i>cDAZL</i>	deleted in azoospermia — like	NM_204218.1	CTCCTGCACCGCAATTCCAT	TCTTTGCTCCCGAGGAACCC
<i>cVASA</i>	DEAD (Asp — Glu — Ala — Asp) box polypeptide 4	NM_204708.1	CCGCATGCTCGATATGGGT	TCCCTCTTGATAACCGGGG
<i>WNT1</i>	wingless — type MMTV integration site family, member 1	XM_015273182.1	TCTCAGGGCCCCAACATCTT	CGAAGAGCCTCCCGAAATCG
<i>WNT3A</i>	wingless — type MMTV integration site family, member 3A	NM_001171601.1	AGGCCCTTTGTGGTGGGAGT	GAGGGGTGAAGTTGGGCAT
<i>WNT9B</i>	wingless — type MMTV integration site family, member 9B	XM_015299263.1	GCTTTCGCCTTCCACGAGA	CACGTTCTCCAGCAGTTCC
<i>FZD1</i>	frizzled family receptor 1	NM_001030337.1	CTGCTCGTGGGCATCAACA	AATGACGATGGTGGCCGGTA
<i>FZD3</i>	frizzled family receptor 3	NM_001271940.1	GGCTTTTGTACCCCTCCC	AATGTGCCAGGTCACCCAG
<i>LRP5</i>	low density lipoprotein receptor — related protein 5	NM_001012897.1	GCAAGAGCGAGCTCCCAAGA	AGGCCATTGGCTGAAGGAT
<i>LRP6</i>	low density lipoprotein receptor — related protein 6	XM_417286.4	CTGTATGCAAACCGCGTGA	ATGCCAGCCATCAGGTGAC
β — <i>catenin</i>	catenin (cadherin — associated protein), beta 1, 88k	NM_205081.1	ATTTGTGCGCTCCGTCACCT	ACCTTGTTCACGAGTGGGG
<i>GSK3β</i>	glycogen synthase kinase 3	XM_416557.4	TCCATTCCTTTGGGATCTGCC	TACACAGCCCCTGACCACA
<i>TCF1 (TCF7)</i>	transcription factor 7 (T — cell specific, HMG — box)	NM_204547.1	ATGAAAGAGATGCGGGCGAA	CTGCTGCTTTCCCGTGTC
<i>LEF1</i>	lymphoid enhancer — binding factor 1	NM_205013.2	GCGGCTTGTACAGCAAGGGA	GGAGCCTGGGGAAAAGTGCT
<i>TCF3 (TCF7L1)</i>	transcription factor 7 — like 1 (T — cell specific, HMG — box)	XM_003643906.2	GGAATGCTGATGTGGGACCG	CCCAAACCTGTGGGACCGAAA
<i>TCF4 (TCF7L2)</i>	transcription factor 7 — like 2 (T — cell specific, HMG — box)	NM_001206510.2	AAATCCCCATCCGCTAGGA	AGCCGACGTCACCTCTGGGAA
<i>CDK6</i>	cyclin-dependent kinase 6	NM_001007892.2	TGGGGCTGCTTACATTGAC	GCCAGCAGCAAAACGTTTCAG
<i>CCND1</i>	cyclin D1	NM_205381.1	GGTGCTGCAGACCATGCTGA	GCAATTGCAACCGGCTTTTC
<i>CDK2</i>	cyclin-dependent kinase 2	NM_001199857.1	GAGTACCTGTTCCAGCTGC	CCAGATGTCCACAGCAGTCG
<i>CCNE1</i>	cyclin E1	NM_001031358.1	TCGAGTACGCCAGCCACTTA	GAAATGTCGCCTTGCACAGC
<i>GAPDH</i>	glyceraldehyde — 3 — phosphate dehydrogenase	NM_204305.1	GGTGGTGCTAAGCGTGTAT	ACCTCTGCATCTCCACA

Table 1. Primers used for RT-PCR.

with Prolong Gold antifade reagent with 4',6-diamidino-2-phenylindole (Invitrogen) and visualized using fluorescence microscopy.

Statistical analysis. Statistical analysis was performed using SAS version 9.4 software (SAS Institute, Cary, NC, USA). When a significant main effect was detected by analysis of variance using SAS, the least-squares analysis was used to compare the different treatments. Differences between control and treatment groups were deemed to be significant when $P < 0.05$.

References

- Han, J. Y. Germ cells and transgenesis in chickens. *Comp Immunol Microbiol Infect Dis* **32**, 61–80 (2009).
- De Felici, M., Farini, D. & Dolci, S. In or out stemness: comparing growth factor signalling in mouse embryonic stem cells and primordial germ cells. *Curr Stem Cell Res Ther* **4**, 87–97 (2009).
- De Felici, M. & Farini, D. The control of cell cycle in mouse primordial germ cells: old and new players. *Curr Pharm Des* **18**, 233–244 (2012).
- Jean, C. *et al.* Transcriptome analysis of chicken ES, blastodermal and germ cells reveals that chick ES cells are equivalent to mouse ES cells rather than EpiSC. *Stem Cell Res* **14**, 54–67 (2015).
- Rengaraj, D. *et al.* Gene pathways and cell cycle-related genes in cultured avian primordial germ cells. *Poult Sci* **91**, 3167–3177 (2012).
- Lee, S. I. *et al.* MicroRNA-mediated posttranscriptional regulation is required for maintaining undifferentiated properties of blastoderm and primordial germ cells in chickens. *Proc Natl Acad Sci USA* **108**, 10426–10431 (2011).
- Chawengsaksophak, K. *et al.* Loss of Wnt5a disrupts primordial germ cell migration and male sexual development in mice. *Biol Reprod* **86**, 1–12 (2012).
- Kimura, T. *et al.* The stabilization of beta-catenin leads to impaired primordial germ cell development via aberrant cell cycle progression. *Dev Biol* **300**, 545–553 (2006).
- Laird, D. J., Altshuler-Keylin, S., Kissner, M. D., Zhou, X. & Anderson, K. V. Ror2 enhances polarity and directional migration of primordial germ cells. *PLoS Genet* **7**, e1002428 (2011).
- Ohinata, Y. *et al.* A signaling principle for the specification of the germ cell lineage in mice. *Cell* **137**, 571–584 (2009).
- Sato, N., Meijer, L., Skaltsounis, L., Greengard, P. & Brivanlou, A. H. Maintenance of pluripotency in human and mouse embryonic stem cells through activation of Wnt signaling by a pharmacological GSK-3-specific inhibitor. *Nat Med* **10**, 55–63 (2004).
- Polakis, P. Wnt signaling in cancer. *Cold Spring Harb Perspect Biol* **4**, a008052 (2012).
- Fu, Y. *et al.* beta-catenin as a potential key target for tumor suppression. *Int J Cancer* **129**, 1541–1551 (2011).
- Valenta, T., Hausmann, G. & Basler, K. The many faces and functions of beta-catenin. *EMBO J* **31**, 2714–2736 (2012).
- Bialecka, M. *et al.* Cdx2 contributes to the expansion of the early primordial germ cell population in the mouse. *Dev Biol* **371**, 227–234 (2012).
- Sato, T., Ueda, S. & Niki, Y. Wingless signaling initiates mitosis of primordial germ cells during development in Drosophila. *Mech Dev* **125**, 498–507 (2008).
- Tseng, A. S., Engel, F. B. & Keating, M. T. The GSK-3 inhibitor BIO promotes proliferation in mammalian cardiomyocytes. *Chem Biol* **13**, 957–963 (2006).
- Waalder, J. *et al.* Novel synthetic antagonists of canonical Wnt signaling inhibit colorectal cancer cell growth. *Cancer Res* **71**, 197–205 (2011).
- Delaune, E., Lemaire, P. & Kodjabachian, L. Neural induction in Xenopus requires early FGF signalling in addition to BMP inhibition. *Development* **132**, 299–310 (2005).
- Feldman, B., Poueymirou, W., Papaioannou, V. E., DeChiara, T. M. & Goldfarb, M. Requirement of FGF-4 for postimplantation mouse development. *Science* **267**, 246–249 (1995).

21. Resnick, J. L., Ortiz, M., Keller, J. R. & Donovan, P. J. Role of fibroblast growth factors and their receptors in mouse primordial germ cell growth. *Biol Reprod* **59**, 1224–1229 (1998).
22. Park, T. S. & Han, J. Y. Derivation and characterization of pluripotent embryonic germ cells in chicken. *Mol Reprod Dev* **56**, 475–482 (2000).
23. van de Lavoie, M. C. *et al.* Germline transmission of genetically modified primordial germ cells. *Nature* **441**, 766–769 (2006).
24. Choi, J. W. *et al.* Basic fibroblast growth factor activates MEK/ERK cell signaling pathway and stimulates the proliferation of chicken primordial germ cells. *PLoS One* **5**, e12968 (2010).
25. Macdonald, J., Glover, J. D., Taylor, L., Sang, H. M. & McGrew, M. J. Characterisation and germline transmission of cultured avian primordial germ cells. *PLoS One* **5**, e15518 (2010).
26. Whyte, J. *et al.* FGF, Insulin, and SMAD Signaling Cooperate for Avian Primordial Germ Cell Self-Renewal. *Stem Cell Reports* **5**, 1171–1182 (2015).
27. Ding, V. M. *et al.* FGF-2 modulates Wnt signaling in undifferentiated hESC and iPSC cells through activated PI3-K/GSK3beta signaling. *J Cell Physiol* **225**, 417–428 (2010).
28. Israsena, N., Hu, M., Fu, W., Kan, L. & Kessler, J. A. The presence of FGF2 signaling determines whether beta-catenin exerts effects on proliferation or neuronal differentiation of neural stem cells. *Dev Biol* **268**, 220–231 (2004).
29. Katoh, M. Cross-talk of WNT and FGF signaling pathways at GSK3beta to regulate beta-catenin and SNAIL signaling cascades. *Cancer Biol Ther* **5**, 1059–1064 (2006).
30. Katoh, M. Networking of WNT, FGF, Notch, BMP, and Hedgehog signaling pathways during carcinogenesis. *Stem Cell Rev* **3**, 30–38 (2007).
31. Buchtova, M. *et al.* Fibroblast growth factor and canonical WNT/beta-catenin signaling cooperate in suppression of chondrocyte differentiation in experimental models of FGFR signaling in cartilage. *Biochim Biophys Acta* **1852**, 839–850 (2015).
32. Krejci, P. *et al.* Receptor tyrosine kinases activate canonical WNT/beta-catenin signaling via MAP kinase/LRP6 pathway and direct beta-catenin phosphorylation. *PLoS One* **7**, e35826 (2012).
33. Stulberg, M. J., Lin, A. P., Zhao, H. Y. & Holley, S. A. Crosstalk between Fgf and Wnt signaling in the zebrafish tailbud. *Developmental Biology* **369**, 298–307 (2012).
34. Chacon, M. A., Varela-Nallar, L. & Inestrosa, N. C. Frizzled-1 is involved in the neuroprotective effect of Wnt3a against Abeta oligomers. *J Cell Physiol* **217**, 215–227 (2008).
35. Endo, Y. *et al.* Wnt-3a and Dickkopf-1 stimulate neurite outgrowth in Ewing tumor cells via a Frizzled3- and c-Jun N-terminal kinase-dependent mechanism. *Mol Cell Biol* **28**, 2368–2379 (2008).
36. Badders, N. M. *et al.* The Wnt Receptor, Lrp5, Is Expressed by Mouse Mammary Stem Cells and Is Required to Maintain the Basal Lineage. *PLoS One* **4**, e6594 (2009).
37. Bikkavilli, R. K. & Malbon, C. C. Wnt3a-stimulated LRP6 phosphorylation is dependent upon arginine methylation of G3BP2. *J Cell Sci* **125**, 2446–2456 (2012).
38. MacDonald, B. T. & He, X. Frizzled and LRP5/6 receptors for Wnt/beta-catenin signaling. *Cold Spring Harb Perspect Biol* **4**, a007880 (2012).
39. Cao, H. *et al.* GSK3 inhibitor-BIO regulates proliferation of immortalized pancreatic mesenchymal stem cells (iPMSCs). *PLoS One* **7**, e31502 (2012).
40. Tanaka, S. S. *et al.* Dullard/Ctdnep1 modulates WNT signalling activity for the formation of primordial germ cells in the mouse embryo. *PLoS One* **8**, e57428 (2013).
41. Tevosian, S. G. Gone without the WNT: a requirement for WNT5A in germ cell migration and testis development. *Biol Reprod* **86**, 1–2 (2012).
42. Brennan, K. R. & Brown, A. M. Wnt proteins in mammary development and cancer. *J Mammary Gland Biol Neoplasia* **9**, 119–131 (2004).
43. Moravec, M. Colorectal cancer and canonical Wnt signalling pathway. *Cas Lek Cesk* **151**, 335–342 (2012).
44. Nusse, R. Wnt signaling. *Cold Spring Harb Perspect Biol* **4**, a011163 (2012).
45. Logan, C. Y. & Nusse, R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol* **20**, 781–810 (2004).
46. Sun, F. *et al.* Downregulation of CCND1 and CDK6 by miR-34a induces cell cycle arrest. *FEBS Lett* **582**, 1564–1568 (2008).
47. Otero, J. J., Fu, W. M., Kan, L. X., Cuadra, A. E. & Kessler, J. A. beta-catenin signaling is required for neural differentiation of embryonic stem cells. *Development* **131**, 3545–3557 (2004).
48. Ge, C., Yu, M. & Zhang, C. G protein-coupled receptor 30 mediates estrogen-induced proliferation of primordial germ cells via EGFR/Akt/beta-catenin signaling pathway. *Endocrinology* **153**, 3504–3516 (2012).
49. Kim, J. N. *et al.* Migration and proliferation of intact and genetically modified primordial germ cells and the generation of a transgenic chicken. *Biol Reprod* **82**, 257–262 (2010).
50. Park, T. S. & Han, J. Y. piggyBac transposition into primordial germ cells is an efficient tool for transgenesis in chickens. *Proc Natl Acad Sci USA* **109**, 9337–9341 (2012).

Acknowledgements

This work was supported by a National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIP) (No. 2015R1A3A2033826).

Author Contributions

H.C.L., S.L. and J.Y.H. designed the experiments; H.C.L. and S.L. performed the experiments; all authors analyzed the data and wrote the manuscript.

Additional Information

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Lee, H. C. *et al.* Wnt/ β -catenin signaling pathway activation is required for proliferation of chicken primordial germ cells *in vitro*. *Sci. Rep.* **6**, 34510; doi: 10.1038/srep34510 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016