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Redox enzyme-mimicking activities of CeO₂ nanostructures: Intrinsic influence of exposed facets

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CeO₂ nanoparticles (NPs) have been well demonstrated as an antioxidant in protecting against oxidative stress-induced cellular damages and a potential therapeutic agent for various diseases thanks to their redox enzyme-mimicking activities. The Ce³⁺/Ce⁴⁺ ratio and oxygen vacancies on the surface have been considered as the major originations responsible for the redox enzyme-mimicking activities of CeO₂ NPs. Herein, CeO₂ nanostructures (nanocubes and nanorods) exposed different facets were synthesized via a facile hydrothermal method. The characterizations by X-ray photoelectron spectroscopy, Raman spectroscopy, and UV-Vis spectroscopy show that the Ce³⁺/Ce⁴⁺ ratio and oxygen vacancy content on the surfaces of as-synthesized CeO₂ nanostructures are nearly at the same levels. Meanwhile, the enzymatic activity measurements indicate that the redox enzyme-mimicking activities of as-synthesized CeO₂ nanostructures are greatly dependent on their exposed facets. CeO₂ nanocubes with exposed {100} facets exhibit a higher peroxidase but lower superoxide dismutase activity than those of the CeO₂ nanorods with exposed {110} facets. Our results provide new insights into the redox enzyme-mimicking activities of CeO₂ nanostructures, as well as the design and synthesis of inorganic nanomaterials-based artificial enzymes.

Enzyme-mimicking activities of various nanomaterials have attracted considerable interest in industrial catalysis and biomedical fields. CeO₂ nanoparticles (NPs) have been well demonstrated as an antioxidant highly effective in protecting against oxidative stress-induced cellular damages and a potential therapeutic agent for various diseases, such as cancer, diabetes, chronic inflammation, brain autoimmune degenerative disease, and retinal pathologies^{1–4}. Numerous studies have shown that CeO₂ NPs display high activities mimicking a series of redox enzymes, including superoxide dismutase (SOD)^{5–7}, catalase^{8–10}, peroxidase^{11–15}, phosphotriesterase¹⁶, phosphatase^{17,18}, and oxidase¹⁹, which can scavenge reactive oxygen and nitrogen species. In CeO₂, the intrinsic defects derived from the existence of Ce³⁺ result in the formation of oxygen vacancies compensating for the positive charge deficiency. The enzyme-mimicking activities of CeO₂ NPs have been attributed to the auto-regenerative cycle of Ce³⁺/Ce⁴⁺ and oxygen vacancies on the surface of CeO₂. Hence, the redox state of Ce ions on the surface plays a crucial role in the redox enzyme-mimicking activities of CeO₂ NPs. Studies by Self *et al.*^{5–10} indicated that the high Ce³⁺/Ce⁴⁺ ratio on the surface of CeO₂ NPs corresponded to high SOD mimetic activity, which was inhibited and transferred to catalase/peroxidase mimetic activities when the Ce³⁺/Ce⁴⁺ ratio decreased. On the other hand, some results suggested that increasing the Ce³⁺ concentration on the surface of CeO₂ NPs was beneficial to peroxidase mimetic activity^{11,12}.

The exposed facets have been demonstrated to considerably contribute to the catalytic performances of various metals, alloys, and metal oxides^{20–23}. Recent studies by Shen *et al.* based on theoretical calculations and experimental data confirmed that redox enzyme-mimicking activities of noble metals (Au, Ag, Pd, and Pt) were critically dependent on exposed facets²⁴. Previous studies also showed that the catalytic activities of CeO₂ NPs in the oxidation of CO, NO, and propane were substantially related to the exposed facets^{25–31}. So far, the relationship between the exposed facets of CeO₂ NPs and their redox enzyme-mimicking activities has not yet been explored.

Herein, CeO₂ nanocubes with exposed {100} facets and nanorods with exposed {110} facets were synthesized through a facile hydrothermal method. Phase structures and morphologies were characterized by X-ray

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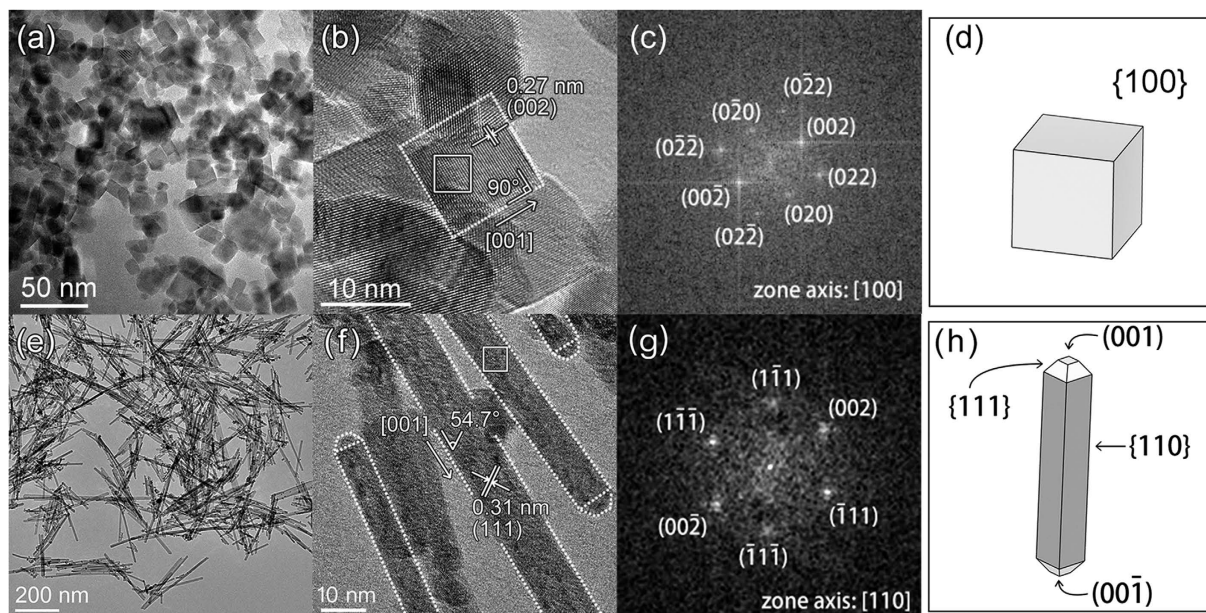


Figure 1. TEM and HRTEM images of the as-synthesized CeO₂ nanostructures. (a–f) CeO₂ nanocubes; (e–h) CeO₂ nanorods. (c,g) show the FFT patterns of the areas marked with white lines in (b,f), respectively. (e,h) display the proposed models of the CeO₂ nanocubes and nanorods, respectively. The CeO₂ models were drawn using software VESTA³².

diffraction (XRD), transmission electron microscopy (TEM), and high resolution transmission electron microscopy (HRTEM). The surface information of the cerium element located on the surface of the CeO₂ nanostructures was analyzed by X-ray photoelectron spectroscopy (XPS), Raman spectroscopy, and UV-Vis spectroscopy. The specific surface areas of the CeO₂ nanostructures were measured by nitrogen adsorption in accordance with the Brunauer-Emmett-Teller (BET) method. Furthermore, the peroxidase and SOD mimetic activities of the CeO₂ nanostructures were evaluated.

Results and Discussion

Figure 1 shows the TEM and HRTEM images of the as-synthesized CeO₂ nanostructures. Homogeneous CeO₂ nanocubes were obtained at an average size of approximately 13 ± 3 nm (Fig. 1a). The HRTEM image displays the well-aligned crystal planes, indicating the single-crystalline nature of CeO₂ nanocubes (Fig. 1b). The interplanar spacing of approximately 0.27 nm could be indexed to the (200) plane of the face-centered cubic CeO₂. The spots that appeared on the fast Fourier transformation (FFT) pattern of the area marked with white lines in Fig. 1b were indexed, and the zone axis was found to be [100] (Fig. 1c). Accordingly, a three-dimensional model describing the nanocubes is shown in Fig. 1d, representing a cubic structure enclosed with {100} facets. CeO₂ nanorods with an average diameter of approximately 10 ± 3 nm were obtained by adjusting the synthesis parameters (Fig. 1e). The well-aligned crystal planes observed from the HRTEM image in Fig. 1f also indicate the single-crystalline structure of CeO₂ nanorods.

The interplanar spacing of approximately 0.31 nm could be indexed to the (111) plane. The spots that appeared on the FFT pattern of the area marked with white lines in Fig. 1f were indexed, and the zone axis was found to be [110] (Fig. 1g). Accordingly, a three-dimensional model describing the nanorods is shown in Fig. 1h, suggesting a rod-like structure enclosed with {110} and {111} facets and truncated by (100) and (00 $\bar{1}$) facets at the z-axis.

The phase structure of the as-synthesized CeO₂ nanostructures was identified by XRD characterization. All of the diffraction peaks in the XRD patterns could be indexed to the cubic fluorite structure of CeO₂ (JCPDS Card 34-0394) (Fig. 2a). Pawley method³⁴ was adopted to obtain the full width at half maximum (FWHM) of the individual peak in the XRD patterns. The result is illustrated as Williamson-Hall plots (Fig. 2b)³⁵. The coefficient of the fitted lines reflects the magnitude of the microstrain (a higher coefficient denotes a higher microstrain), and the intercept of the fitted lines indicates the crystallite size. No microstrain existed in the CeO₂ nanocubes, but a large microstrain was observed in the CeO₂ nanorods (0% vs. 0.98%). Meanwhile, the crystallite size calculated from the intercept of the fitted lines was approximately 13.5 nm for the CeO₂ nanocubes and 10.7 nm for the CeO₂ nanorods, consistent with the average size obtained from the TEM images. In addition, the BET specific surface area measured by N₂ adsorption-desorption isotherms was approximately 70.9 m²/g for the CeO₂ nanocubes and 95.4 m²/g for the CeO₂ nanorods, indicating the larger specific surface area of the CeO₂ nanorods compared with that of the CeO₂ nanocubes.

The surface chemical information of the CeO₂ nanostructures was analyzed by XPS, Raman spectroscopy, and UV-Vis spectroscopy. The peaks in the XPS spectra were fitted using the Gaussian function (Fig. 3a,b). The areas of the fitting peaks ascribed to Ce³⁺ and Ce⁴⁺ were used to calculate the contents of Ce³⁺ and Ce⁴⁺ on the surface of the CeO₂ nanostructures^{15,36}. The calculations yielded approximately 31.8% of Ce³⁺ for the CeO₂

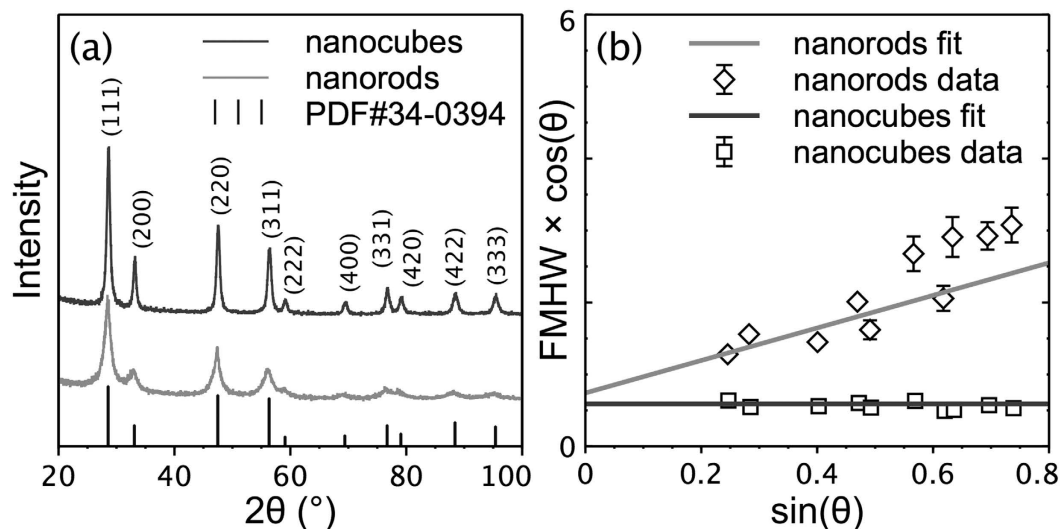


Figure 2. (a) XRD patterns of the as-synthesized CeO₂ nanostructures. (b) Williamson-Hall plots of the CeO₂ nanostructures derived from the XRD patterns. The coefficient of the fitted lines indicates the microstrain (a higher coefficient denotes a higher microstrain), and the intercept of the fitted lines indicates the crystallite size (a higher intercept means a smaller size). The linear and nonlinear fittings were performed using software Fityk³³.

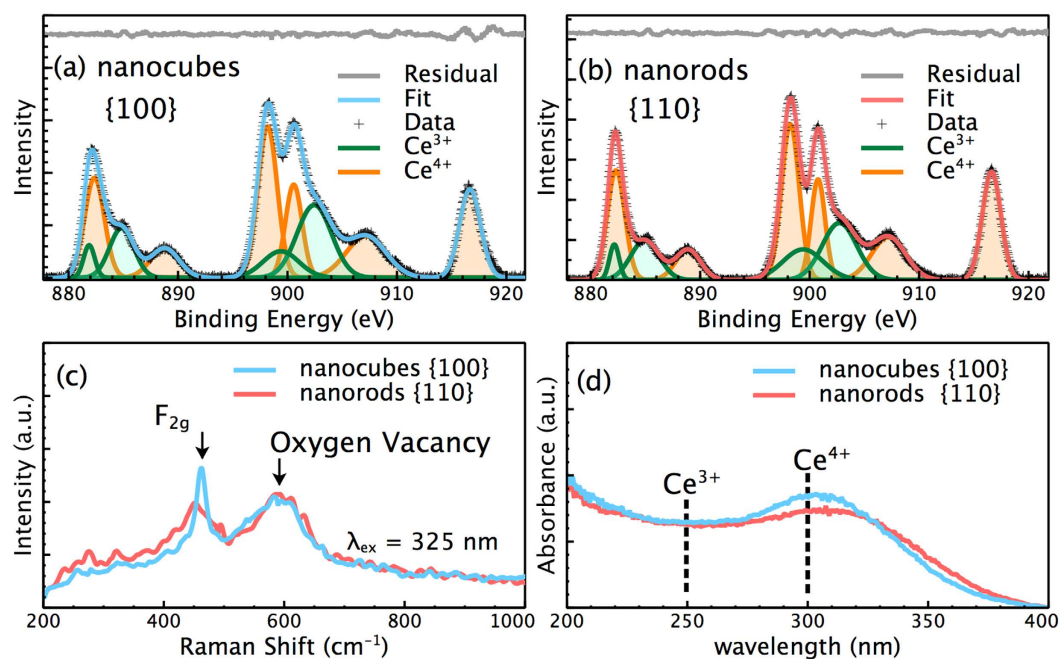


Figure 3. (a,b) XPS spectra of the as-synthesized CeO₂ nanostructures. The peaks in XPS spectra were fitted using Gaussian function and ascribed to Ce³⁺ and Ce⁴⁺, respectively. (c) Raman spectra of the as-synthesized CeO₂ nanostructures at the excitation wavelength of 325 nm. (d) UV-Vis spectra of the as-synthesized CeO₂ nanostructures.

nanocubes and 31.2% of Ce³⁺ for the CeO₂ nanorods, suggesting the similar Ce³⁺ contents on the surfaces of the as-synthesized CeO₂ nanostructures. Figure 3c shows the Raman spectra of CeO₂ nanostructures at the excitation wavelength of 325 nm. The strong peak at 462 cm⁻¹ could be ascribed to the F_{2g} mode^{37,38}. The asymmetrical and broader peak in the Raman spectrum of the CeO₂ nanorods could be attributed to the smaller crystallite size and larger microstrain compared with those of the CeO₂ nanocubes³⁹. Notably, the two CeO₂ nanostructures exhibited similar Raman absorption at 595 cm⁻¹, which corresponds to the characteristic peak of oxygen vacancy^{37,38}.

This result implies that oxygen vacancies on the surfaces of both CeO₂ nanostructures were at the same level. The absorption peak at 250 nm in UV-Vis spectra of CeO₂ has been ascribed to Ce³⁺, and the absorption peak at

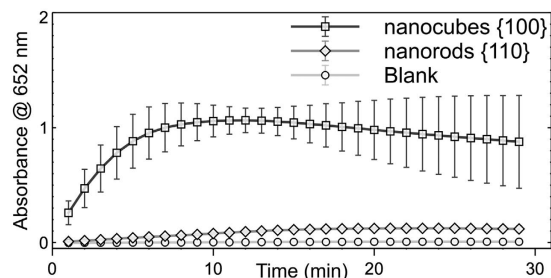


Figure 4. Peroxidase mimetic activity of the CeO₂ nanostructures. The change in absorbance at 652 nm represents the conversion of TMB to oxidized TMB (TMB_{ox}).

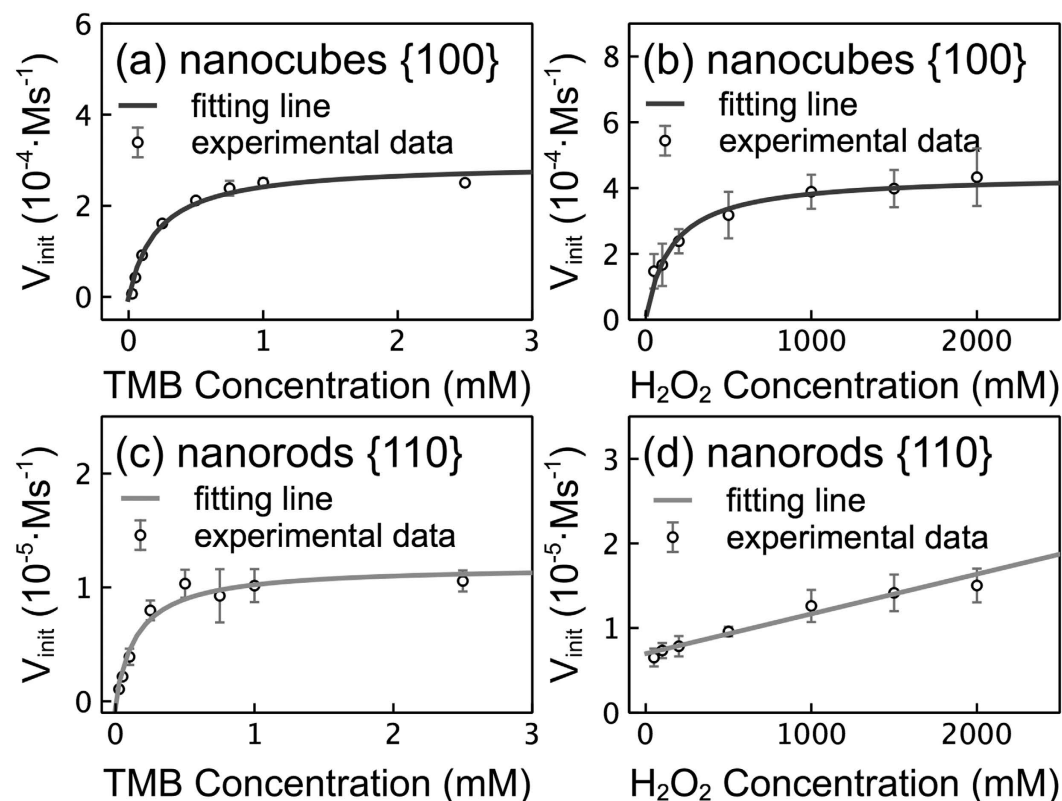
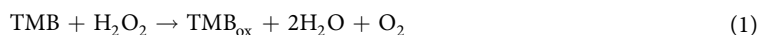


Figure 5. Steady-state kinetic assay of peroxidase mimetic activity.

295 nm has been ascribed to Ce⁴⁺^{6,10}. No obvious absorption peak belonging to Ce³⁺ was observed for both of the CeO₂ nanostructures (Fig. 3d), implying the low Ce³⁺ content in the crystal structure of the CeO₂ nanostructures. Therefore, these results strongly demonstrated that both the Ce³⁺/Ce⁴⁺ ratio and oxygen vacancies on the surface of the as-synthesized CeO₂ nanostructures were nearly at the same levels.

The peroxidase-mimicking activity of the CeO₂ nanostructures was evaluated by monitoring the catalytic reaction between H₂O₂ and tetramethylbenzidine (TMB) in acetic buffer solution (equation 1).



The change in absorbance at 652 nm indicates the formation of TMB_{ox}. Almost no change in absorbance was observed within 30 min (Fig. 4), implying that H₂O₂ failed to oxidize TMB in the absence of catalysts. Subsequently, in the presence of CeO₂ nanorods with exposed {110} facets, a slight increase in absorbance was observed with reaction time extension, suggesting that the TMB was slowly oxidized by H₂O₂. The absorbance was dramatically increased with the reaction prolongation in the presence of CeO₂ nanocubes, demonstrating that the oxidation of TMB by H₂O₂ was greatly accelerated by the CeO₂ nanocubes with exposed {100} facets.

Figure 5 shows the results of a steady-state kinetic assay conducted by varying the concentrations of TMB and H₂O₂. Kinetic parameters were calculated by fitting experimental data to the Michaelis-Menten equation. V_{max} is the maximal reaction velocity, and K_m is the Michaelis constant. A higher V_{max} value represents a higher conversion rate from substrate to product, and a higher K_m denotes a smaller catalyst affinity to the substrate. Under

varied TMB concentrations (Fig. 5a,c), the K_m value was approximately 0.217 mM for the CeO₂ nanocubes and approximately 0.240 mM for the CeO₂ nanorods, consistent with the values reported previously^{14,40}. These results suggest the similar affinities of TMB to both of the CeO₂ nanostructures. However, V_{max} value was approximately 8.2×10^{-8} M/s for the CeO₂ nanocubes and 0.4×10^{-8} M/s for the CeO₂ nanorods, indicating that the rate of the reaction catalyzed by the CeO₂ nanocubes with exposed {100} facets was 23 times higher than that catalyzed by the CeO₂ nanorods with exposed {110} facets. Meanwhile, under varying H₂O₂ concentrations, the K_m and V_{max} for the CeO₂ nanocubes were approximately 153.6 mM and 12.2×10^{-8} M/s, respectively. In such cases, the K_m obtained is far greater than that obtained from the varied TMB levels, suggesting that the affinity of H₂O₂ on the surface of the CeO₂ nanocubes was far smaller than that of TMB. While the larger V_{max} in the case of varying H₂O₂ concentrations than that in the case of varying TMB concentrations suggests that the catalytic reaction was more sensitive to the H₂O₂ concentration. Notably, the Michaelis-Menten equation failed to describe the relation between the concentration of H₂O₂ and the initial velocity of the reaction in the presence of the CeO₂ nanorods (Fig. 5d). Instead, a linear relation was observed, implying a simple first-order reaction dependent on the concentration of the H₂O₂ catalyzed by the CeO₂ nanorods.

The peroxidase-mimicking activity of the CeO₂ nanorods was also measured after a heating treatment at 650 °C for 48 h because the fitting of the XRD patterns reveals the existence of microstrain in the CeO₂ nanorods. However, no significant increase in enzyme activity was observed in spite of the disappearance of the microstrain in the CeO₂ nanorods after annealing (see Supplementary Figs S1 and S2). Hence, the peroxidase mimetic activities of the CeO₂ nanostructures were influenced by the exposed facets instead of the microstrain.

The SOD mimetic activity of the CeO₂ nanostructures was also evaluated with colorimetric assay kits. The results indicate that the SOD mimetic activity of the CeO₂ nanorods with exposed {110} facets was four times higher than that of the CeO₂ nanocubes with exposed {100} facets (see Supplementary Fig. S3). After annealing to remove the microstrain, the CeO₂ nanorods still exhibited a higher SOD mimetic activity than that of the CeO₂ nanocubes. Thus, the SOD mimetic activity of the CeO₂ nanostructures may have also depended on the exposed facets instead of the microstrain.

Theoretical calculations and experimental data have indicated that redox enzymes-mimicking activities of CeO₂ could be ascribed to the transformation between Ce⁴⁺ and Ce³⁺ ions and the generation of oxygen vacancies compensating for the positive charge deficiency on the surfaces^{41–43}. However, in the present study, both the Ce³⁺/Ce⁴⁺ ratio and oxygen vacancies were of the same level on the surfaces of the two CeO₂ nanostructures, which displayed different redox enzyme-mimicking activities. Therefore, the different exposed facets should be responsible for such different catalytic performances. It is generally known that CeO₂ possesses three low-index facets: {111}, {110}, and {100} facet. Among which, CeO₂ {110} surface is a type I ionic crystal surface with neutral atomic planes due to a stoichiometric balance between anions and cations, while CeO₂ {111} surface is a type II ionic crystal surface with charged planes but without net dipole moment perpendicular to the surface. Both CeO₂ {110} and {111} surfaces have relatively low surface energies and display modest relaxations on the surfaces compared to the bulk. By comparison, CeO₂ {100} surface has a nearly infinite free energy and therefore require a major reconstruction compared to the bulk because such a surface consists of alternatively charged planes and thus produces a dipole moment perpendicular to the surface. Accordingly, the stability of three facets decreased in an order of {111} > {110} > {100}, which means the higher catalytic activity of {100} facets than that of {110} and {111} facets^{41–43}. According to an energy-based model describing the facet-dependent redox enzyme-mimicking activities of noble metal NPs, the dissociative adsorption of O₂ on the metal surfaces was proposed to be the key step that provided the surfaces with oxidase-like activities²⁴. Meanwhile, the protonation of O₂[−] and adsorption and rearrangement of HO₂ on metal surfaces were mainly responsible for SOD-like activity of these metals²⁴. Therefore, it is reasonable to speculate that the facet-dependent adsorption, activation, and rearrangement processes between the reacting species and the exposed facets may play crucial roles on the redox enzyme-mimicking activities of CeO₂ nanostructures in addition to the transformation between Ce⁴⁺ and Ce³⁺ ions ratio and the generation of oxygen vacancies on the surfaces. Unfortunately, although numerous theoretical and experimental investigations have been performed in order to interpret the relationship between the catalytic activity and the surface compositions and crystal structures, the accurate atomic structures of CeO₂ nanostructures, particularly for the CeO₂ with exposed {110} and {100} facets, still remain indistinct^{41–44}. Therefore, more theoretical and experimental investigations are necessary to elucidate the precise mechanisms on the influences of redox enzyme-mimicking activities of CeO₂ nanostructures by their exposed facets.

Conclusion

In summary, we have shown that the redox enzyme mimetic activities of the CeO₂ nanostructures were determined by the exposed facets. At the same levels of Ce³⁺/Ce⁴⁺ ions and oxygen vacancies on the surfaces, CeO₂ nanocubes with exposed {100} facets displayed a higher peroxidase but lower SOD mimetic activity than those of the CeO₂ nanorods with exposed {110} facets. Therefore, integrated factors, including exposed facets, Ce³⁺/Ce⁴⁺ ratio, and oxygen vacancy surface content, should be carefully considered when CeO₂ nanostructures are employed as antioxidant and therapeutic agent for various diseases on the basis of their redox enzyme-mimicking activities. Our results provide new insight into the redox enzyme-mimicking activities of CeO₂ nanostructures, as well as the design and synthesis of other inorganic nanomaterials-based artificial enzymes.

Methods

Materials. Sodium hydroxide (NaOH, 96%), hydrogen peroxide (H₂O₂, 30%), acetic acid (99.5%) and cerium nitrate hexahydrate (Ce(NO₃)₃·6H₂O, 99%) were of analytical grade and purchased from Sinopharm Chemical Reagent Co. (Shanghai, China). 3,3',5,5'-tetramethyl-benzidine (TMB, 98%) was obtained from Aladdin

(Shanghai, China). Silicon (SRM™ 640e) was obtained from National Institute of Standards and Technology (NIST). All reagents were used as received without further purification.

Synthesis of CeO₂ nanostructures. In a typical synthesis, 20 mL of Ce(NO₃)₃·6H₂O solution (0.1 mol/L) was added dropwise into NaOH solution (0.1 mol/L for nanocubes and 12 mol/L for nanorods). The mixed solution was then transferred and sealed in a 50 mL Teflon-lined stainless steel autoclave, with subsequent heating for 24 h. The heating temperature was kept at 140 °C for nanocubes and 100 °C for nanorods. After the reaction, the precipitate was collected by centrifugation (9000 rpm, 5 min) and washed alternately with ethanol and deionized water several times, and then dried at 60 °C in the air.

Characterization of CeO₂ nanostructures. The phase structure of the samples was identified by powder X-ray diffraction (XRD) on a D8 Advance diffractometer using Cu K α radiation ($\lambda = 1.5418 \text{ \AA}$). NIST SRM™ 640e was used to obtain the instrumental broadening profile. X-ray photoelectron spectroscopy (XPS) measurements were performed on a spectrometer (Axis Ultra DLD-600W, Kratos Analytical Ltd.) using Al K α radiation as the excitation source. The morphologies of the samples were observed using high resolution transmission electron microscopy (HRTEM, JEM-2100F STEM/EDS, JEOL Corp, Japan). The Raman spectra of the samples were recorded by a Raman spectrometer with a 325 nm laser excitation (VERTEX 70, Bruker, Germany). The UV-Vis spectra of the samples were recorded with an ultraviolet–visible spectrometer (Shimadzu Corp., UV-2550 PC).

Enzyme mimetic activity of CeO₂ nanostructures. The peroxidase mimetic activity of CeO₂ was evaluated by monitoring the redox reaction between TMB and H₂O₂ in the presence of the CeO₂ nanostructures. Typically, 1 mL acetate buffer solution (50 mM, pH = 4.0) containing 25 μg CeO₂, 0.5 μmol TMB, and 1 mol H₂O₂ was added in a cuvette. The reaction was monitored using an ultraviolet–visible spectroscopy (Shimadzu Corp., UV-2550 PC) at wavelength of 652 nm. The measurement was recorded at an interval of one minute and the temperature was kept at 25 °C. In order to obtain the apparent kinetic parameters, experiments varying concentrations of the substrates were carried out. The measurement was recorded at an interval of 10 seconds in order to obtain a higher accuracy. All the rest reaction conditions were unchanged. More details on the calculation of apparent kinetic parameters can be found in the Supplementary Information. The SOD mimetic activity of CeO₂ nanostructures was determined with colorimetric assay kits (Nanjing Jiancheng Bioengineering Institute, China).

Data process. All the non-linear curve fittings (XRD, XPS and Michaelis-Menten Kinetic) were processed using software Fityk (version 1.3.0)³³. All the experiments were repeated at least triplicated to obtain the standard deviation.

References

- Celardo, I., Pedersen, J. Z., Traversa, E. & Ghibelli, L. Pharmacological potential of cerium oxide nanoparticles. *Nanoscale* **3**, 1411–1420 (2011).
- Wason, M. S. & Zhao, J. Cerium oxide nanoparticles: Potential applications for cancer and other diseases. *Am. J. Transl. Res.* **5**, 126–131 (2013).
- Xu, C. & Qu, X. Cerium oxide nanoparticle: a remarkably versatile rare earth nanomaterial for biological applications. *NPG Asia Mater.* **6**, e90 (2014).
- Heckman, K. L. *et al.* Custom Cerium Oxide Nanoparticles Protect against a Free Radical Mediated Autoimmune Degenerative Disease in the Brain. *ACS Nano* **7**, 10582–10596 (2013).
- Korsvik, C., Patil, S., Seal, S. & Self, W. T. Superoxide dismutase mimetic properties exhibited by vacancy engineered ceria nanoparticles. *Chem. Commun.* 1056–1058 (2007).
- Heckert, E. G., Karakoti, A. S., Seal, S. & Self, W. T. The role of cerium redox state in the SOD mimetic activity of nanoceria. *Biomaterials* **29**, 2705–2709 (2008).
- Liu, X. *et al.* Apoferritin–CeO₂ nanotruffle that has excellent artificial redox enzyme activity. *Chem. Commun.* **48**, 3155–3157 (2012).
- Pirmohamed, T. *et al.* Nanoceria exhibit redox state-dependent catalase mimetic activity. *Chem. Commun.* **46**, 2736–2738 (2010).
- Singh, S. *et al.* A phosphate-dependent shift in redox state of cerium oxide nanoparticles and its effects on catalytic properties. *Biomaterials* **32**, 6745–6753 (2011).
- Singh, R. & Singh, S. Role of Phosphate on Stability and Catalase Mimetic Activity of Cerium Oxide Nanoparticles. *Colloids Surfaces B Biointerfaces* **132**, 78–84 (2015).
- Xiao, X., Luan, Q., Yao, X. & Zhou, K. Single-crystal CeO₂ nanocubes used for the direct electron transfer and electrocatalysis of horseradish peroxidase. *Biosens. Bioelectron.* **24**, 2447–2451 (2009).
- Li, X., Zhang, Z., Tao, L., Li, Y. & Li, Y. Y. A chemiluminescence microarray based on catalysis by CeO₂ nanoparticles and its application to determine the rate of removal of hydrogen peroxide by human erythrocytes. *Appl. Biochem. Biotechnol.* **171**, 63–71 (2013).
- Wang, N., Sun, J., Chen, L., Fan, H. & Ai, S. A Cu₂(OH)₃Cl–CeO₂ nanocomposite with peroxidase-like activity, and its application to the determination of hydrogen peroxide, glucose and cholesterol. *Microchim. Acta* **182**, 1733–1738 (2015).
- Tian, Z. *et al.* Highly sensitive and robust peroxidase-like activity of porous nanorods of ceria and their application for breast cancer detection. *Biomaterials* **59**, 116–124 (2015).
- Zhao, H., Dong, Y., Jiang, P., Wang, G. & Zhang, J. Highly Dispersed CeO₂ on TiO₂ Nanotube: A Synergistic Nanocomposite with Superior Peroxidase-Like Activity. *ACS Appl. Mater. Interfaces* **7**, 6451–6461 (2015).
- Vernekar, A. A., Das, T. & Mughes, G. Vacancy-Engineered Nanoceria: Enzyme Mimetic Hotspots for the Degradation of Nerve Agents. *Angew. Chemie Int. Ed.* **54**, 1–6 (2015).
- Kuchma, M. H. *et al.* Phosphate ester hydrolysis of biologically relevant molecules by cerium oxide nanoparticles. *Nanomedicine Nanotechnology, Biol. Med.* **6**, 738–744 (2010).
- Tan, F. *et al.* An efficient method for dephosphorylation of phosphopeptides by cerium oxide. *J. Mass Spectrom.* **43**, 628–632 (2008).
- Asati, M. A., Santra, D. S., Kaittanis, M. C., Nath, D. S. & Perez, P. J. M. Oxidase Activity of Polymer-Coated Cerium Oxide Nanoparticles. *Angew. Chem. Int. Ed.* **48**, 2308–2312 (2010).
- Tian, N., Zhou, Z.-Y., Sun, S.-G., Ding, Y. & Wang, Z. L. Synthesis of Tetrahedral Platinum Nanocrystals with High-Index Facets and High Electro-Oxidation Activity. *Science* **316**, 732–735 (2007).
- Mu, J., Zhang, L., Zhao, G. & Wang, Y. The crystal plane effect on the peroxidase-like catalytic properties of Co₃O₄ nanomaterials. *Phys. Chem. Chem. Phys.* **16**, 15709 (2014).

22. Yu, J., Low, J., Xiao, W., Zhou, P. & Jaroniec, M. Enhanced photocatalytic CO₂-Reduction activity of anatase TiO₂ by Coexposed {001} and {101} facets. *J. Am. Chem. Soc.* **136**, 8839–8842 (2014).
23. Lin, Y., Ren, J. & Qu, X. Catalytically active nanomaterials: a promising candidate for artificial enzymes. *Acc. Chem. Res.* **47**, 1097–1105 (2014).
24. Shen, X. *et al.* Mechanisms of Oxidase and Superoxide Dismutation-like Activities of Gold, Silver, Platinum, and Palladium, and Their Alloys: A General Way to the Activation of Molecular Oxygen. *J. Am. Chem. Soc.* **137**, 15882–15891 (2015).
25. Aneggi, E., Llorca, J., Boaro, M. & Trovarelli, A. Surface-structure sensitivity of CO oxidation over polycrystalline ceria powders. *J. Catal.* **234**, 88–95 (2005).
26. Agarwal, S., Zhu, X., Hensen, E. J. M., Mojet, B. L. & Lefferts, L. Surface-Dependence of Defect Chemistry of Nanostructured Ceria. *J. Phys. Chem. C* **119**, 12423–12433 (2015).
27. Hu, Z. *et al.* Effect of Ceria Crystal Plane on the Physicochemical and Catalytic Properties of Pd/Ceria for CO and Propane Oxidation. *ACS Catal.* **6**, 2265–2279 (2016).
28. Tang, K. *et al.* The Effect of Exposed Facets of Ceria to the Nickel Species in Nickel-Ceria Catalysts and Their Performance in a NO+CO Reaction. *ACS Appl. Mater. Interfaces* **7**, 26839–26849 (2015).
29. Sun, C., Li, H. & Chen, L. Nanostructured ceria-based materials: synthesis, properties, and applications. *Energy Environ. Sci.* **5**, 8475–8505 (2012).
30. Zhang, D., Du, X., Shi, L. & Gao, R. Shape-controlled synthesis and catalytic application of ceria nanomaterials. *Dalton Trans.* **41**, 14455–14475 (2012).
31. Younis, A., Chu, D., Kaneti, Y. V. & Li, S. Tuning the surface oxygen concentration of {111} surrounded ceria nanocrystals for enhanced photocatalytic activities. *Nanoscale* **8**, 378–387 (2016).
32. Momma, K. & Izumi, F. VESTA 3 for three-dimensional visualization of crystal, volumetric and morphology data. *J. Appl. Crystallogr.* **44**, 1272–1276 (2011).
33. Wojdyr, M. *Fityk*. A general-purpose peak fitting program. *J. Appl. Crystallogr.* **43**, 1126–1128 (2010).
34. Pawley, G. S. Unit-cell refinement from powder diffraction scans. *J. Appl. Crystallogr.* **14**, 357–361 (1981).
35. Scardi, P., Leoni, M. & Delhez, R. Line broadening analysis using integral breadth methods: A critical review. *J. Appl. Crystallogr.* **37**, 381–390 (2004).
36. Natile, M. M. & Glisenti, A. CoO_x/CeO₂ Nanocomposite Powders: Synthesis, Characterization, and Reactivity. *Chem. Mater.* **17**, 3403–3414 (2005).
37. Reddy, B. M. *et al.* Surface Characterization of CeO₂/SiO₂ and V₂O₅/CeO₂/SiO₂ Catalysts by Raman, XPS, and Other Techniques. *J. Phys. Chem. B* **106**, 10964–10972 (2002).
38. Wu, Z., Li, M., Howe, J., Meyer, H. M. & Overbury, S. H. Probing defect sites on CeO₂ nanocrystals with well-defined surface planes by raman spectroscopy and O₂ adsorption. *Langmuir* **26**, 16595–16606 (2010).
39. Spanier, J. E., Robinson, R. D., Zhang, F., Chan, S.-W. & Herman, I. P. Size-dependent properties of CeO_{2-y} nanoparticles as studied by Raman scattering. *Phys. Rev. B* **64**, 245407 (2001).
40. Gao, L. *et al.* Intrinsic peroxidase-like activity of ferromagnetic nanoparticles. *Nat. Nanotechnol.* **2**, 577–583 (2007).
41. Huang, W. Oxide Nanocrystal Model Catalysts. *Acc. Chem. Res.* **49**, 520–527 (2016).
42. Mullins, D. R. The surface chemistry of cerium oxide. *Surf. Sci. Rep.* **70**, 42–85 (2015).
43. Paier, J., Penschke, C. & Sauer, J. Oxygen defects and surface chemistry of ceria: Quantum chemical studies compared to experiment. *Chem. Rev.* **113**, 3949–3985 (2013).
44. Conesa, J. Computer modeling of surfaces and defects on cerium dioxide. *Surf. Sci.* **339**, 337–352 (1995).

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Author Contributions

Y.Y., Z.M., W.H. and L.L. synthesized the CeO₂ nanostructures and evaluated the enzyme mimicking activities. Y.Y. analyzed the data. Y.Y., J.L. and Q.W. designed the whole work and drafted the manuscript. All the authors revised the manuscript.

Additional Information

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