

Different Divalent Cations Alter the Kinetics and Fidelity of DNA Polymerases*

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Divalent metal ions are essential components of DNA polymerases both for catalysis of the nucleotidyl transfer reaction and for base excision. They occupy two sites, A and B, for DNA synthesis. Recently, a third metal ion was shown to be essential for phosphoryl transfer reaction. The metal ion in the A site is coordinated by the carboxylate of two highly conserved acidic residues, water molecules, and the 3'-hydroxyl group of the primer so that the A metal is in an octahedral complex. Its catalytic function is to lower the pK_a of the hydroxyl group, making it a highly effective nucleophile that can attack the α phosphorous atom of the incoming dNTP. The metal ion in the B site is coordinated by the same two carboxylates that are affixed to the A metal ion as well as the non-bridging oxygen atoms of the incoming dNTP. The carboxyl oxygen of an adjacent peptide bond serves as the sixth ligand that completes the octahedral coordination geometry of the B metal ion. Similarly, two metal ions are required for proofreading; one helps to lower the pK_a of the attacking water molecule, and the other helps to stabilize the transition state for nucleotide excision. The role of different divalent cations are discussed in relation to these two activities as well as their influence on base selectivity and misincorporation by DNA polymerases. Some, but not all, of the effects of these different metal ions can be rationalized based on their intrinsic properties, which are tabulated in this review.

All DNA polymerases (DNA pols)² require Mg^{2+} or Mn^{2+} for primer extension and for excision of incorrectly incorporated dNTPs via intrinsic 3'→5' exonuclease activity (1–5). A two-metal-ion mechanism is used by all DNA pols to catalyze

nucleotide addition to a growing primer strand (6). Although DNA polymerases employ the physiologically relevant Mg^{2+} , other divalent metal ions can substitute for Mg^{2+} , although they tend to reduce the fidelity of DNA replication (7–10). The effect of metal ion cofactors on the fidelity of DNA replication has been studied for various DNA pols including *E. coli* DNA pol I (11), AMV DNA pol (12), Klenow fragment of *E. coli* DNA pol I (13), T4 pol (7), T7 pol (7), human pol α (7), pol β (7), and Dpo4 (8). Some metal ions have been shown to be mutagens and carcinogens probably because they reduce the base selectivity of DNA pols (7, 8, 11–15). Different divalent cations influence fidelity check points in the minimal kinetic scheme for the nucleotidyl transfer reaction (Scheme 1). Cations that can substitute for Mg^{2+} affect DNA pols by: 1) altering the ground-state binding affinity of incoming dNTPs to pol·DNA binary complexes (16); 2) decreasing base selectivity by promoting misincorporation during primer extension (8); 3) decreasing the rate of base excision (17); 4) altering primer extension past a mismatch at the primer-template (P/T) terminus (17). This review will address the way various metal ions increase misincorporation based on their physical properties. Our emphasis will be on the effect of different cations on the behavior of RB69 pol, which we have studied extensively in our lab (18). There are other reviews that deal with pol β , another DNA pol that has been thoroughly studied with respect to the influence of different metal ions on its structure and function (19–21).

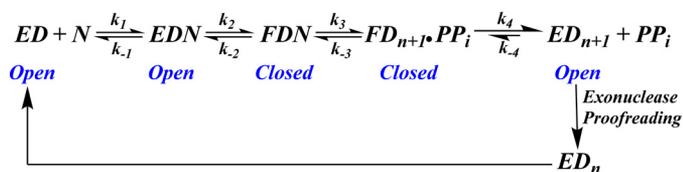
The Two-metal-ion Mechanism for Nucleotidyl Transfer Reaction

All DNA pols require two divalent metal ions for primer extension (6). One metal ion occupies the “A site” and helps to lower the pK_a of the terminal 3'-OH group on the primer and coordinates both the α -phosphate of the incoming dNTP and the 3'-OH of the primer strand, which facilitates its nucleophilic attack on the α -phosphorous atom of the incoming dNTP (6). The other metal ion, occupying the “B site,” coordinates the α -, β -, and γ - non-bridging phosphate oxygens of the incoming dNTP, helping to neutralize the developing negative charge as the ternary complex approaches the transition state in the nucleotidyl transfer reaction, and assists in the departure of the PP_i product. Yang *et al.* (22) have shown that for pol β , the dNTP·metal ion complex in the “B site” alone is unable to induce closing of the fingers, a necessary step for the phosphoryl transfer reaction to proceed. Both A and B metal ions are thus required to prepare the active site for nucleotidyl transfer (22). Tsai and co-workers (23) have used Rh^{+3} as an exchange-inert cation complexed with an incoming dNTP to selectively fill the “B site” in pol β so that the effect of binding a metal ion in the “A site” could be studied independently of the cation in the B site. Their results showed that the closing of the fingers could occur before occupancy of the A site but that Mg^{2+} could diffuse into the A site, although pol β was in the closed form (23). In contrast, studies with RB69 pol by Wang and co-workers (16) showed that although the fingers can close in the absence of an “A” metal ion, the fingers have to reopen for

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² The abbreviations used are: pol, polymerase; AMV, avian myeloblastosis virus; 2AP, 2-aminopurine; Dpo4, DNA polymerase IV from *Sulfolobus solfataricus*; exo, exonuclease; ptO3', terminal 3'-OH group on the primer; RB69 pol, bacteriophage RB69 DNA polymerase; tm, triple mutant of RB69 pol containing L561A, S565G, and Y567A mutations in the active site; P/T, primer-template; dUPNpp, 2'-deoxyuridine 5'-(α,β -imido)triphosphate.



SCHEME 1. Minimal kinetic scheme for DNA polymerases depicting various fidelity checkpoints along the reaction pathway. *EDN* represents the open conformation of the ternary collision complex, whereas *FDN* represents the closed conformation. Additional details are provided in the text.

a divalent cation to bind in the “A site.” Recent studies by Yang and co-workers (24) observed the transient presence of a third metal ion of pol η after the nucleotidyl transfer reaction was initiated, but before release of the products, using time-resolved x-ray crystallography (24). The third transient metal ion was coordinated to four water molecules in addition to an oxygen atom (which acts as a bridge between the α - and β -phosphorous atoms) and to the non-bridging oxygen of the α -phosphate. Yang and co-workers (24) proposed that in addition to the A and B metal ions, the third metal ion (Mg^{2+}) participates in neutralizing the negative charges built up in the transition state and is likely involved in facilitating the protonation of the pyrophosphate leaving group. Recent studies by Gao and Yang (25) have shown that the third metal ion is indeed required for catalysis by pol η .

The Nature of Metal Ion Coordination Complexes with Various DNA pols

Several groups have solved the crystal structures of DNA pols with metal ions and an incoming dNTP bound in the polymerase active site (6, 9, 26–29). RB69 pol has been one of the most extensively studied DNA pols in the B family and is the only DNA pol where all combinations of mispaired bases were captured in ternary complexes (26). For this purpose, four mutations were required adjacent to the nucleotide binding pocket. Among these the triple mutant (tm) was used to capture a ternary complex in the presence of Mn^{2+} and dUpNpp, a non-hydrolyzable dNTP analogue (Fig. 1) (6). In this structure, Mn^{2+} , bound in the “B site,” has nearly ideal octahedral geometry and is coordinated by the triphosphate tail of the incoming dUpNpp along with the carboxylate side chains of Asp⁴¹¹ and Asp⁶²³, and the backbone carbonyl oxygen of Leu⁴¹² (Fig. 1b). Mn^{2+} bound in the “A site,” however, has highly distorted octahedral geometry because in this structure, it is coordinated with the 3'-OH group of the primer, the two carboxylate side chains of Asp⁴¹¹ and Asp⁶²³, and the oxygen of the α -phosphate of the incoming dNTP. However, the distance between the 3'-OH group and P α of the incoming dUpNpp is too great to support phosphodiester bond formation (6). Xia *et al.* (6) proposed that as the reaction approaches the transition state, metal ion A helps to reduce the 3'-OH-P α distance, facilitating covalent bond formation (Fig. 1c). Similar results have been observed when Mg^{2+} occupies both the A sites and B sites, suggesting that Mg^{2+} and Mn^{2+} share similar coordination geometries in RB69 pol ternary complexes (6).

In addition to RB69 pol (6, 30), T7 pol (31, 32), Klenow fragment (33–35), and Dpo4 (8, 10, 36), pol β has also been extensively studied (9, 19–21, 23). Although the topology of the Palm

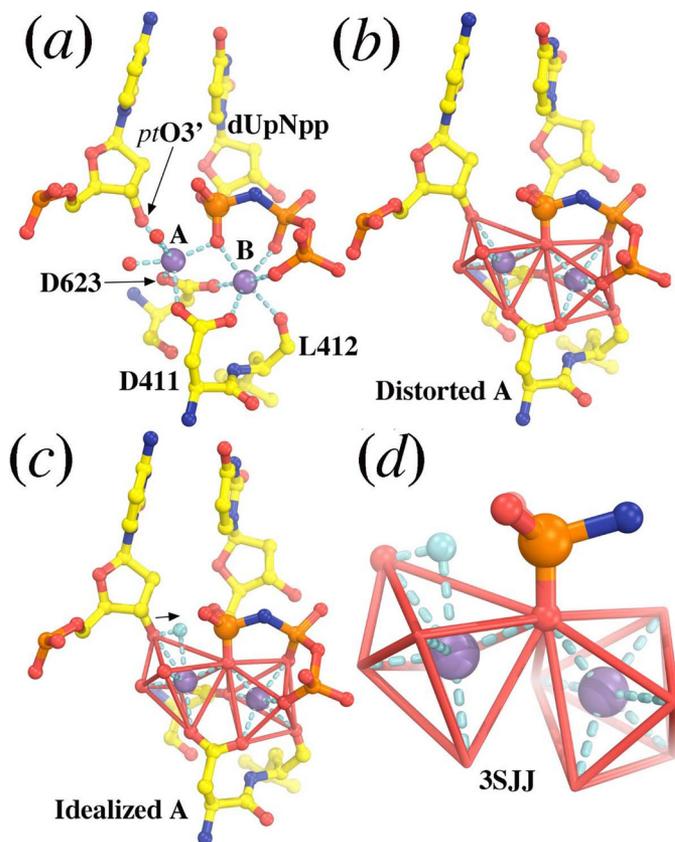


FIGURE 1. The structure of the tm RB69 pol ternary complex (Protein Data Bank (PDB) accession number 3SJJ). *a*, ternary complex showing dUpNpp bound in the active site with metal ions A and B. *b*, close-up of the tm RB69 pol active site showing the B metal ion in perfect octahedral geometry and the A metal ion in a distorted octahedral geometry. *c*, predicted position of 3'-OH group in an idealized octahedron during the transition state. *d*, a close-up of the coordination complex showing the two metal ions, the 3'-OH group, and the α phosphate of the incoming dNTP.

domain of pol β , which contains the two conserved catalytic carboxylates, differs between RB69 pol and pol β , the metal ion coordination geometries are nearly identical. In fact, crystal structures of all DNA pols have the same coordination geometry of the A and B metal ions (for examples, see Fig. 2).

DNA Polymerases Can Use Different Metal Ions for Catalyzing Phosphoryl Transfer

Early studies by Sirover and Loeb (15, 37) measured perturbations in the fidelity of DNA synthesis using AMV DNA pol. They identified several metal ions as being mutagenic or carcinogenic including Ag^+ , Be^{2+} , Cd^{2+} , Co^{2+} , Cr^{3+} , Mn^{2+} , Ni^{2+} , and Pb^{2+} (15). Subsequent studies with *E. coli* DNA pol I (11, 38) showed that Co^{2+} and Mn^{2+} could effectively replace Mg^{2+} but caused an increase in misincorporation. Similar results were reported with Mn^{2+} and Co^{2+} for human pol α and pol β (39). Later studies by Snow *et al.* (7) showed that Ni^{2+} , albeit active, was an inefficient activator for a variety of DNA pols including AMV pol, human pol α , T4 pol, Klenow fragment, and T7 pol.

Pelletier *et al.* (9) have reported structures of pol β ternary complexes in the presence of several different metal ions and showed that apart from Mg^{2+} and Mn^{2+} , only Cd^{2+} and Zn^{2+} catalyzed primer extension with a blunt-end DNA substrate.

Egli and co-workers (36) carried out kinetic studies using Dpo4 pol with a variety of divalent cations and found that only Mg^{2+} , Mn^{2+} , and Ca^{2+} could support Dpo4-catalyzed polymerization but that Sr^{2+} , Ba^{2+} , Zn^{2+} , Cu^{2+} , Ni^{2+} , and Co^{2+} were inactive. Recent work by Vashishtha and Konigsberg (30) on RB69 pol showed that, apart from Mg^{2+} and Mn^{2+} , Co^{2+} , and to a lesser extent Ni^{2+} , were the only divalent cations that could support both pol and exo activities. The metal ion preferences for different DNA pols are summarized in Table 1. Thus, DNA pols from different families can utilize different metal ions as cofactors, but they do not follow a pattern that can be predicted from their physical properties.

Metal Ions Affect Various Fidelity Checkpoints during DNA Replication

Divalent metal ions can alter the fidelity of DNA replication at various points along the reaction pathway by: 1) affecting the ground-state binding affinity of correct and incorrect dNTPs to DNA pol:P/T binary complexes (16); 2) promoting misincorporation during primer extension (8); and 3) influencing exonuclease activity (17). Scheme 1 shows the various fidelity checkpoints employed by DNA pols that minimize dNMP misincorporation. The effect of divalent cations on each of these checkpoints will be addressed in the following sections.

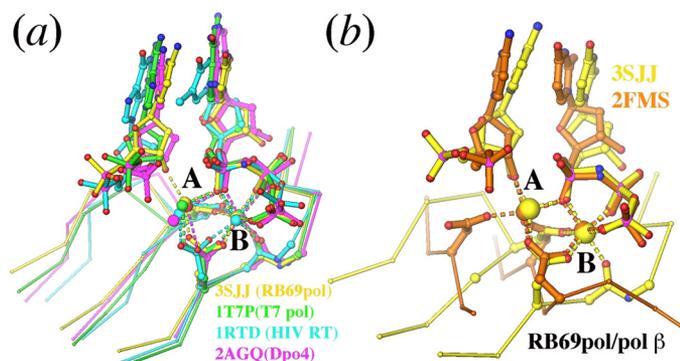


FIGURE 2. Comparison of metal ion bound structures of the tm RB69 pol with other DNA polymerases. *a*, superposition of the tm RB69 pol with T7 DNA pol (PDB accession number: 1T7P, green), HIV reverse transcriptase (HIV RT) (PDB accession number: 1RTD, cyan), and Dpo4 (PDB accession number: 2AGQ, magenta). *b*, superposition of the tm RB69 pol with pol β (PDB accession number: 2FMS, golden).

TABLE 1

Summary of metal ion preferences of different DNA polymerases

Metal ions that activate the respective DNA polymerase are shown with +, and those that are not able to support the polymerase activity are shown with -. Bst, *B. stearothermophilus*.

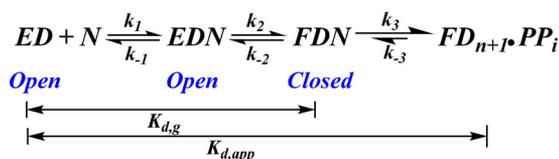
DNA polymerase	Metal ion										
	Mn^{2+}	Co^{2+}	Fe^{2+}	Ni^{2+}	Zn^{2+}	Cd^{2+}	Sr^{2+}	Ba^{2+}	Cu^{2+}	Cr^{3+}	Ca^{2+}
DNA pol I	+	+	-	+	-	-	-	-	-	-	-
Human pol α	+	+	-	-	-	-	-	-	-	-	-
pol β	+	+ ^a	-	-	+	+	-	-	-	-	-
AMV DNA pol	+	+	-	+	-	-	-	-	-	-	-
T4 pol	+	+	-	+	-	-	-	-	-	-	-
T7 pol	+	+	-	+	-	-	-	-	-	-	-
RB69pol	+	+	-	+	-	-	-	-	-	-	-
Bst pol	+	+ ^b	-	+ ^b	+ ^b	+ ^b	-	-	+ ^b	-	-
Dpo4 pol	+	+ ^a	-	-	-	-	-	-	-	-	+

^a Studies by Pelletier *et al.* (9) and Egli and co-workers (36) previously claimed that human pol β and Dpo4 were not able to utilize Co^{2+} as cofactor. However, recent studies by Vashishtha and Konigsberg (30) showed that both pol β and Dpo4 can catalyze primer extension in the presence of Co^{2+} as explained in the text.

^b See Footnote 3.

The Effect of Metal Ions on Ground-state Binding Affinity of Incoming dNTPs for pol:P/T Binary Complexes

Different divalent cations can affect the ground-state equilibrium dissociation constant ($K_{d,g}$) for incoming dNTPs (Scheme 2). Zhang *et al.* (40) used a dideoxy P/T containing 2AP opposite an incoming dNTP (the templating position) and measured the $K_{d,g}$ for dTTP binding, which was $9 \mu M$ in the presence of Mg^{2+} . This equilibrium binding assay was based on the observation that, upon formation of the pol:P/T binary complex, 2AP becomes unstacked and exists in a high fluorescence state (41, 42). The addition of dTTP results in 2AP stacking (2AP is translocated from n position to $n - 1$ position) and quenching of the 2AP fluorescence (41). In the same study (40), there was no change in 2AP fluorescence when it was present at a location one residue downstream from the templating position when dCTP was added opposite dG as the templating base; therefore the $K_{d,g}$ for binding of the incoming dNTPs could not be determined. In a similar study carried out by Wang and co-workers (16) in the presence of Ca^{2+} , the $K_{d,g}$ values were determined to be $53 nM$ and $53 \mu M$ for dTTP (correct) and dCTP (incorrect) binding, respectively, opposite 2AP (when 2AP was present in the templating position), suggesting that the identity of the divalent cation cofactor has a profound effect on the ground-state equilibrium dissociation constant. Hariharan *et al.* (41) also reported a $K_{d,g}$ value of $31 \mu M$ for dTTP binding opposite 2AP with T4 pol in the presence of Mg^{2+} , similar to the value reported for RB69 pol (16). A more comprehensive study on the effect of different divalent cations on ground-state binding affinity for incoming dNTPs was carried out by Vashishtha and Konigsberg (30), where the $K_{d,g}$ values were measured in the presence of Mg^{2+} , Mn^{2+} , Co^{2+} , and Ca^{2+} with RB69 pol. Their results showed that the dissociation constants for dTTP binding opposite 2AP in RB69 pol ternary



SCHEME 2. Minimal Kinetic scheme for DNA polymerases depicting the ground-state binding affinity ($K_{d,g}$) and apparent binding affinity ($K_{d,app}$) for an incoming dNTP. *EDN* represents the open conformation of the ternary collision complex, whereas *FDN* represents the closed conformation.

complexes decreased from Mg^{2+} , Co^{2+} , Mn^{2+} , and Ca^{2+} in that order. A similar pattern of dissociation constants was observed for dCTP binding opposite 2AP. In general, the $K_{d,app}$ values were substantially lower with Mn^{2+} as compared with Mg^{2+} (30). This may be due to the ability of Mn^{2+} , in contrast to Mg^{2+} , to accommodate base pairs other than the regular Watson-Crick base pairs in the nucleotide binding pocket. The possible reasons for this are discussed in the following sections.

Effect of Metal Ions on Base Selectivity

Numerous studies have shown that various divalent cations affect the base selectivity of pols to different extents (13, 14, 43–46). Substitution of Mg^{2+} by Mn^{2+} generally results in a decrease in the fidelity of DNA pols including T4 pol (14, 47), T7 pol (31), *E. coli* DNA pol I (13, 48), AMV DNA pol (45), and pol β (9). Beckman *et al.* (48) showed that at very low [Mn^{2+}] ($<1 \mu M$), the fidelity of DNA replication is similar to that observed with Mg^{2+} and that the decrease in fidelity is only observed at elevated [Mn^{2+}] ($<100 \mu M$). The possible reasons for this behavior include the binding of Mn^{2+} to the DNA template at elevated [Mn^{2+}]. Other divalent cations including Co^{2+} and Ni^{2+} have been reported to have similar effects on base selectivity as a function of the metal ion concentration (12). Pre-steady-state kinetic studies by Vashishtha and Konigsberg (30) showed that Mn^{2+} and Co^{2+} are cofactors for RB69 pol-catalyzed reactions. Surprisingly, the incorporation efficiency for correct incoming dNTPs was higher with Co^{2+} than with Mg^{2+} or Mn^{2+} . In contrast, base selectivity was decreased with Co^{2+} versus Mg^{2+} but not nearly as much as when Mn^{2+} replaced Mg^{2+} (30). Of all the divalent cations tested, Mn^{2+} is the most highly mutagenic as Mn^{2+} promotes misincorporation by increasing the rate of incorporation (k_{pol} or V_{max}) as well as by decreasing the $K_{d,app}$ or K_m values for incorrect incoming dNTPs (14, 30) (where k_{pol} is the maximum rate of dNMP incorporation, and $K_{d,app}$ is the apparent equilibrium dissociation constant for [dNTP] that supports the half-maximal rate of dNMP incorporation).

Rare Tautomer Hypothesis for Mutagenesis When Mn^{2+} Is Present

Occasionally, replicative DNA pols incorporate mismatched nucleotides (49) via the formation of high-energy tautomers (50–52). In fact, Beese and co-workers (53) provided structural evidence for the presence of these rare tautomers using a D598A/F710Y double mutant of a catalytically competent fragment of *Bacillus stearothermophilus* pol. Beese and co-workers (53) showed that, in the presence of Mn^{2+} , the C/A mismatch adopts a tautomeric cognate base pair shape that is virtually indistinguishable from the canonical, Watson-Crick base pair in double-stranded DNA at the insertion site. With Mn^{2+} , the triphosphate tail was properly aligned for catalysis, and the polymerase was in a closed conformation, facilitating the misincorporation. In contrast, in the presence of Mg^{2+} , the C/A mismatch forms a non-cognate wobble base pair, and the polymerase adopts an “ajar” or partially closed conformation, which prevented mismatch incorporation. In addition, the triphos-

phate tail of the incoming incorrect dNTP was not properly aligned for catalysis, which helped to prevent misincorporation. These results provide a structural rationale for the mutagenic behavior of Mn^{2+} in this situation.

Effect of Metal Ions on the Exonuclease Activity

Similar to the polymerase active site, the exonuclease site also requires divalent cations to catalyze the 3'-5'-exonuclease activity associated with several DNA pols (5, 30, 32, 35, 54, 55). Divalent cations are required but have a varying effect on the exonuclease activity. Results with RB69 pol (30) showed that, as compared with Mn^{2+} and Co^{2+} , Mg^{2+} was most effective in promoting base excision but the exo rates varied only slightly among these three metal ions. Ni^{2+} on the other hand caused a dramatic decrease in exo activity (33-fold with Ni^{2+} versus Mg^{2+}). Similarly, the rates of base excision were reported to be nearly identical for *E. coli* DNA pol I with Mg^{2+} , Mn^{2+} , and Co^{2+} (11).

All DNA pols Can Utilize Co^{2+} as Cofactor

In addition to Mg^{2+} , most other DNA pols can also use Mn^{2+} and Co^{2+} , albeit with reduced fidelity (11–13, 15, 30). Pelletier *et al.* (9) and Egli and co-workers (36) have shown that pol β and Dpo4 are the two known exceptions that cannot be activated by Co^{2+} . In contrast, Vashishtha and Konigsberg (30) showed that these DNA pols can actually catalyze primer extension in the presence of Co^{2+} . The apparent conflicting results can be rationalized based on the fact that different assay conditions were used by each of these groups. For example, Pelletier *et al.* (9) used blunt-ended DNA with pol β , whereas Vashishtha and Konigsberg (30) used a P/T with a four-base overhang 5' to the templating base. In the Egli and co-workers (36) experiments, 2 mM DTT was included in their assay with Dpo4, which reduced Co^{2+} to Co^{1+} ($E^0 = -0.33$ V for DTT versus -0.28 V for Co^{2+}). DTT was omitted by Vashishtha and Konigsberg (30) in their assays of Dpo4, so cobalt remained as Co^{2+} . Based on these results, it appears that Co^{2+} can support catalysis for all DNA pols that have been studied to date.

Properties of Divalent Cations That Can Activate DNA Polymerases

Magnesium is in the second row of the periodic table and has 2s electrons that are typically lost when it becomes Mg^{2+} (56). Together with two 3d orbitals, Mg^{2+} forms unoccupied stable sp^3d^2 hybrid orbitals for six coordination ligands. Because of the involvement of 3d orbitals, the coordination bonds are very strong. The average covalent length is 2.09 Å when all high-resolution Mg^{2+} -containing protein structures are compared (6, 29, 57). When carboxylate groups are ligands, the coordination bond lengths are typically reduced slightly relative to non-carboxylate ligands. Manganese is in the third row of the periodic table and has two electrons in 3d orbitals, which are more stable than its 3s electrons. When it loses two 3s electrons, it becomes Mn^{2+} , and leaves two 3d electrons in parallel configurations with two separate 3d orbitals in a high-spin state (56). Upon hybridization in sp^3d^2 orbitals, the coordination geometry is also octahedral. Due to the involvement of 3s orbitals, the coordination bond lengths increase to 2.22 Å when all the

TABLE 2

Ionic radii, coordination geometry, and pK_a of water molecules coordinated to Mg^{2+} , Mn^{2+} , Co^{2+} , Ni^{2+} , Zn^{2+} , Cd^{2+} , and Ca^{2+}

Td, tetrahedral; Sq, square planar; TBP, trigonal bipyramidal; Oct, octahedral; PBP, pentagonal bipyramid; HBP, hexagonal bipyramidal.

	Metal ion						
	Mg^{2+}	Mn^{2+}	Co^{2+}	Ni^{2+}	Zn^{2+}	Cd^{2+}	Ca^{2+}
Ionic radius (Å)	0.86	0.81	0.89	0.83	0.88	0.95	1.1
Coordination	Oct	Oct	Oct	Oct	Oct	Oct	Oct
Geometry	Td	Td	Td	Td	Td ^a	Td	PBP
	Sq	Sq	Sq	TBP	TBP	HBP	
	TBP	TBP					
pK_a of the water molecule	11.4	11.5	10.0	10.6	7.0	9.0	12.8

^a Although Zn^{2+} can form octahedral complexes, the majority of Zn^{2+} complexes are tetrahedral.

Mn^{2+} -containing high-resolution protein structures are compared (6, 9, 57). The increased distance between adjacent oxygen ligands of Mn^{2+} (2.22 Å relative to 2.09 Å for Mg^{2+}) permits the Mn^{2+} coordination octahedron to access ligands that have larger deviations of coordination bond lengths than Mg^{2+} . This is also true for ligands from the triphosphate moieties of mismatched dNTPs.

Proceeding from Mg^{2+} , to Mn^{2+} , to Ca^{2+} , the coordination bond lengths continue to increase from 2.09, 2.22, and 2.40 Å for corresponding metal ion-containing high-resolution protein structures (6). However, increased coordination bond lengths also pose problems for reducing the apical $ptO3'$ - $P\alpha$ distance in the transition state. In fact, RB69 pol is completely inactive with Ca^{2+} , because the shortest estimated $ptO3'$ - $P\alpha$ distance estimated would be about 3.3 Å, which is too great for nucleophilic attack of $ptO3'$ on the $P\alpha$ center (6). The reason why Mn^{2+} is catalytically active is that certain Mn^{2+} coordination bond lengths can be reduced to those of Mg^{2+} at the expense of increasing the bond lengths of its adjacent ligands, *i.e.* distortion of the octahedrons (6). Although Mn^{2+} remains in a high-spin state in most protein structures, it can be converted to low spin in the transition state where two of its 3d electrons occupy the same orbital and the coordination bond lengths are reduced to those of Mg^{2+} (57). This conversion can accelerate incorporation of any dNMP, correct or incorrect, once they are stabilized in a closed ternary complex. When going from Mn^{2+} to Co^{2+} , two more electrons are added to the 3d orbital, which results in a low-spin state. As a consequence, the Co^{2+} coordination bond lengths are reduced to 2.09 Å, almost identical to those of Mg^{2+} , which could account for activation of various DNA pols by Co^{2+} . From Co^{2+} to Zn^{2+} , the preferred coordination geometry becomes tetrahedral instead of octahedral, notably in Zn^{2+} -binding motifs that involve Cys and His residues. Thus, neither Zn^{2+} nor Cd^{2+} can be used by most DNA pols as catalytic metal ions. pol β (9) and *B. stearothermophilus* DNA pol I large fragment³ are the only two DNA pols that can be activated by Zn^{2+} and Cd^{2+} , but the reason for this is not known.

The ability to reduce the pK_a of bound water is very similar for Mg^{2+} and Mn^{2+} but is considerably higher for Co^{2+} , Ni^{2+} , Zn^{2+} , and Cd^{2+} (Table 2). Based on this property alone, Co^{2+} ,

Ni^{2+} , Zn^{2+} , and Cd^{2+} should be more effective as cofactors than Mg^{2+} and Mn^{2+} , but this does not agree with the experimental data obtained with nearly all DNA pols (11–13, 15, 30, 36–38, 58), so other issues must be involved. The ionic radii of metal ion A play a crucial role in determining the proximal distance between the 3'-hydroxyl group and α -phosphorous atom of the incoming dNTP as the transition state is approached. The ionic radii of Mn^{2+} , Co^{2+} , Ni^{2+} , and Zn^{2+} are very close to that of Mg^{2+} (Table 2), enabling all these metal ions to potentially bring the 3'-hydroxyl group and α -phosphate atom of the incoming dNTP close enough for reaction as opposed to Cd^{2+} and Ca^{2+} , whose ionic radii are larger than that Mg^{2+} (56). Despite its ability to lower the pK_a of bound water and despite having similar ionic radii to Mg^{2+} , Zn^{2+} is not able to activate most DNA pols. Based on its properties (Table 2), Ni^{2+} would be expected to substitute for Mg^{2+} , but it is not clear why Ni^{2+} is such a poor cofactor for DNA pols despite having similar physical properties as Mg^{2+} .

Ca^{2+} does not support primer extension with DNA pols with the exception of Dpo4 (6). One possible reason for this is the inability of Ca^{2+} to lower the pK_a of the 3'-hydroxyl group of the primer as compared with Mg^{2+} (12.8 versus 11.4). The ionic radius of Ca^{2+} is also significantly larger than that of Mg^{2+} (1.1 Å versus 0.86 Å), which renders Ca^{2+} ineffective in polarizing the hydroxyl group for nucleophilic attack. Dpo4 is the only DNA polymerase that can be activated by Ca^{2+} , although its ability to act as a cofactor is much reduced as compared with Mg^{2+} (36).

Mn^{2+} has been reported to be highly mutagenic, and this behavior can be rationalized based on the fact that Mn^{2+} is a softer metal ion than Mg^{2+} , suggesting that Mn^{2+} is more polarizable than Mg^{2+} (44). In terms of a hexahydrated complex of $Mn[H_2O]_6^{2+}$ and $Mg[H_2O]_6^{2+}$, there is a greater energy penalty with Mg^{2+} as compared with Mn^{2+} when the inner sphere coordination number is changed from 6→5→4, indicating more rigid coordination requirements for Mg^{2+} complexes, which allows less freedom for mismatched dNTPs to be accessible to the nucleotide binding pocket (44). Transition metal ions such as Mn^{2+} bind more tightly to carboxylate groups and the triphosphate moiety of dNTPs as compared with Mg^{2+} (9), which could explain its ability to reduce base selectivity.

To summarize, DNA pols from different families are able to utilize different divalent cations as cofactors to catalyze primer extension. Crystal soaking experiments with pol β have shown that Zn^{2+} and Cd^{2+} were active (9), whereas these metal ions were not able to activate RB69 pol for catalysis (30). Ni^{2+} can support primer extension with all DNA polymerases, albeit with greatly reduced activity except for Dpo4 (36), human pol α (43), and pol β (9). Moreover, Dpo4 is the only polymerase that can utilize Ca^{2+} , although Ca^{2+} is much less effective than Mg^{2+} (36). Thus, it seems that the abilities of different metal ions to lower the pK_a of the primer's 3'-OH group, the respective coordination geometries, and size are the main but not the only determinants of metal ion activation of DNA pols for catalysis of nucleotidyl transfer and base excision.

³ A. K. Vashishtha, J. Wang, and W. H. Konigsberg, unpublished data.

References

- Drake, J. W. (1969) Comparative rates of spontaneous mutation. *Nature* **221**, 1132
- Goodman, M. F., and Tippin, B. (2000) The expanding polymerase universe. *Nat. Rev. Mol. Cell Biol.* **1**, 101–109
- Fil e, J., Forterre, P., Sen-Lin, T., and Laurent, J. (2002) Evolution of DNA polymerase families: evidences for multiple gene exchange between cellular and viral proteins. *J. Mol. Evol.* **54**, 763–773
- Vashishtha, A. K., and Kuchta, R. D. (2016) Effects of acyclovir, Foscarnet, and ribonucleotides on herpes simplex virus-1 DNA polymerase: mechanistic insights and a novel mechanism for preventing stable incorporation of ribonucleotides into DNA. *Biochemistry* **55**, 1168–1177
- Vashishtha, A. K., and Kuchta, R. D. (2015) Polymerase and exonuclease activities in herpes simplex virus type 1 DNA polymerase are not highly coordinated. *Biochemistry* **54**, 240–249
- Xia, S., Wang, M., Blaha, G., Konigsberg, W. H., and Wang, J. (2011) Structural insights into complete metal ion coordination from ternary complexes of B family RB69 DNA polymerase. *Biochemistry* **50**, 9114–9124
- Snow, E. T., Xu, L. S., and Kinney, P. L. (1993) Effects of nickel ions on polymerase activity and fidelity during DNA replication *in vitro*. *Chem. Biol. Interact* **88**, 155–173
- Vaisman, A., Ling, H., Woodgate, R., and Yang, W. (2005) Fidelity of Dpo4: effect of metal ions, nucleotide selection and pyrophosphorolysis. *EMBO J.* **24**, 2957–2967
- Pelletier, H., Sawaya, M. R., Wolffe, W., Wilson, S. H., and Kraut, J. (1996) A structural basis for metal ion mutagenicity and nucleotide selectivity in human DNA polymerase β . *Biochemistry* **35**, 12762–12777
- Irimia, A., Loukachevitch, L. V., Eoff, R. L., Guengerich, F. P., and Egli, M. (2010) Metal-ion dependence of the active-site conformation of the translesion DNA polymerase Dpo4 from *Sulfolobus solfataricus*. *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.* **66**, 1013–1018
- Sirover, M. A., Dube, D. K., and Loeb, L. A. (1979) On the fidelity of DNA replication: metal activation of *Escherichia coli* DNA polymerase I. *J. Biol. Chem.* **254**, 107–111
- Sirover, M. A., and Loeb, L. A. (1977) On the fidelity of DNA replication. Effect of metal activators during synthesis with avian myeloblastosis virus DNA polymerase. *J. Biol. Chem.* **252**, 3605–3610
- Miyaki, M., Murata, I., Osabe, M., and Ono, T. (1977) Effect of metal cations on misincorporation by *E. coli* DNA polymerases. *Biochem. Biophys. Res. Commun.* **77**, 854–860
- Goodman, M. F., Keener, S., Guidotti, S., and Branscomb, E. W. (1983) On the enzymatic basis for mutagenesis by manganese. *J. Biol. Chem.* **258**, 3469–3475
- Sirover, M. A., and Loeb, L. A. (1976) Infidelity of DNA synthesis *in vitro*: screening for potential metal mutagens or carcinogens. *Science* **194**, 1434–1436
- Lee, H. R., Wang, M., and Konigsberg, W. (2009) The reopening rate of the fingers domain is a determinant of base selectivity for RB69 DNA polymerase. *Biochemistry* **48**, 2087–2098
- Villani, G., Tanguy Le Gac, N., Wasungu, L., Burnouf, D., Fuchs, R. P., and Boehmer, P. E. (2002) Effect of manganese on *in vitro* replication of damaged DNA catalyzed by the herpes simplex virus type-1 DNA polymerase. *Nucleic Acids Res.* **30**, 3323–3332
- Xia, S., and Konigsberg, W. H. (2014) RB69 DNA polymerase structure, kinetics, and fidelity. *Biochemistry* **53**, 2752–2767
- Beard, W. A., and Wilson, S. H. (2014) Structure and mechanism of DNA polymerase β . *Biochemistry* **53**, 2768–2780
- Sobol, R. W., and Wilson, S. H. (2001) Mammalian DNA β -polymerase in base excision repair of alkylation damage. *Prog. Nucleic Acid Res. Mol. Biol.* **68**, 57–74
- Beard, W. A., and Wilson, S. H. (2000) Structural design of a eukaryotic DNA repair polymerase: DNA polymerase β . *Mutat Res.* **460**, 231–244
- Yang, L., Arora, K., Beard, W. A., Wilson, S. H., and Schlick, T. (2004) Critical role of magnesium ions in DNA polymerase β 's closing and active site assembly. *J. Am. Chem. Soc.* **126**, 8441–8453
- Bakhtina, M., Lee, S., Wang, Y., Dunlap, C., Lamarche, B., and Tsai, M. D. (2005) Use of viscogens, dNTP α S, and rhodium(III) as probes in stopped-flow experiments to obtain new evidence for the mechanism of catalysis by DNA polymerase β . *Biochemistry* **44**, 5177–5187
- Nakamura, T., Zhao, Y., Yamagata, Y., Hua, Y. J., and Yang, W. (2012) Watching DNA polymerase η make a phosphodiester bond. *Nature* **487**, 196–201
- Gao, Y., and Yang, W. (2016) Capture of a third Mg²⁺ is essential for catalyzing DNA synthesis. *Science* **352**, 1334–1337
- Xia, S., Wang, J., and Konigsberg, W. H. (2013) DNA mismatch synthesis complexes provide insights into base selectivity of a B family DNA polymerase. *J. Am. Chem. Soc.* **135**, 193–202
- Doubl e, S., Tabor, S., Long, A. M., Richardson, C. C., and Ellenberger, T. (1998) Crystal structure of a bacteriophage T7 DNA replication complex at 2.2   resolution. *Nature* **391**, 251–258
- Johnson, S. J., Taylor, J. S., and Beese, L. S. (2003) Processive DNA synthesis observed in a polymerase crystal suggests a mechanism for the prevention of frameshift mutations. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 3895–3900
- Jain, R., Rajashankar, K. R., Buku, A., Johnson, R. E., Prakash, L., Prakash, S., and Aggarwal, A. K. (2014) Crystal structure of yeast DNA polymerase ϵ catalytic domain. *PLoS ONE* **9**, e94835
- Vashishtha, A. K., and Konigsberg, W. H. (2016) Effect of different divalent cations on the kinetics and fidelity of RB69 DNA polymerase. *Biochemistry* **55**, 2661–2670
- Tabor, S., and Richardson, C. C. (1989) Effect of manganese ions on the incorporation of dideoxynucleotides by bacteriophage T7 DNA polymerase and *Escherichia coli* DNA polymerase I. *Proc. Natl. Acad. Sci. U.S.A.* **86**, 4076–4080
- Hori, K., Mark, D. F., and Richardson, C. C. (1979) Deoxyribonucleic acid polymerase of bacteriophage T7: characterization of the exonuclease activities of the gene 5 protein and the reconstituted polymerase. *J. Biol. Chem.* **254**, 11598–11604
- Kuchta, R. D., Mizrahi, V., Benkovic, P. A., Johnson, K. A., and Benkovic, S. J. (1987) Kinetic mechanism of DNA polymerase I (Klenow). *Biochemistry* **26**, 8410–8417
- Venkitaraman, A. R. (1989) Use of modified T7 DNA polymerase (sequenase version 2.0) for oligonucleotide site-directed mutagenesis. *Nucleic Acids Res.* **17**, 3314
- Joyce, C. M. (1989) How DNA travels between the separate polymerase and 3'-5'-exonuclease sites of DNA polymerase I (Klenow fragment). *J. Biol. Chem.* **264**, 10858–10866
- Irimia, A., Zang, H., Loukachevitch, L. V., Eoff, R. L., Guengerich, F. P., and Egli, M. (2006) Calcium is a cofactor of polymerization but inhibits pyrophosphorolysis by the *Sulfolobus solfataricus* DNA polymerase Dpo4. *Biochemistry* **45**, 5949–5956
- Sirover, M. A., and Loeb, L. A. (1976) Metal-induced infidelity during DNA synthesis. *Proc. Natl. Acad. Sci. U.S.A.* **73**, 2331–2335
- Sirover, M. A., and Loeb, L. A. (1976) Metal activation of DNA synthesis. *Biochem. Biophys. Res. Commun.* **70**, 812–817
- Seal, G., Shearman, C. W., and Loeb, L. A. (1979) On the fidelity of DNA replication: studies with human placenta DNA polymerases. *J. Biol. Chem.* **254**, 5229–5237
- Zhang, H., Cao, W., Zakharova, E., Konigsberg, W., and De La Cruz, E. M. (2007) Fluorescence of 2-aminopurine reveals rapid conformational changes in the RB69 DNA polymerase-primer/template complexes upon binding and incorporation of matched deoxynucleoside triphosphates. *Nucleic Acids Res.* **35**, 6052–6062
- Hariharan, C., Bloom, L. B., Helquist, S. A., Kool, E. T., and Reha-Krantz, L. J. (2006) Dynamics of nucleotide incorporation: snapshots revealed by 2-aminopurine fluorescence studies. *Biochemistry* **45**, 2836–2844
- Frey, M. W., Sowers, L. C., Millar, D. P., and Benkovic, S. J. (1995) The nucleotide analog 2-aminopurine as a spectroscopic probe of nucleotide incorporation by the Klenow fragment of *Escherichia coli* polymerase I and bacteriophage T4 DNA polymerase. *Biochemistry* **34**, 9185–9192
- Chin, Y. E., Snow, E. T., Cohen, M. D., and Christie, N. T. (1994) The effect of divalent nickel (Ni²⁺) on *in vitro* DNA replication by DNA polymerase α . *Cancer Res.* **54**, 2337–2341

44. Bock, C. W., Katz, A. K., Markham, G. D., and Glusker, J. P. (1999) Manganese as a replacement for magnesium and zinc: functional comparison of the divalent ions. *J. Am. Chem. Soc.* **121**, 7360–7372
45. Dube, D. K., and Loeb, L. A. (1975) Manganese as a mutagenic agent during *in vitro* DNA synthesis. *Biochem. Biophys. Res. Commun.* **67**, 1041–1046
46. Jin, Y. H., Clark, A. B., Slebos, R. J., Al-Refai, H., Taylor, J. A., Kunkel, T. A., Resnick, M. A., and Gordenin, D. A. (2003) Cadmium is a mutagen that acts by inhibiting mismatch repair. *Nat. Genet.* **34**, 326–329
47. Hays, H., and Berdis, A. J. (2002) Manganese substantially alters the dynamics of translesion DNA synthesis. *Biochemistry* **41**, 4771–4778
48. Beckman, R. A., Mildvan, A. S., and Loeb, L. A. (1985) On the fidelity of DNA replication: manganese mutagenesis *in vitro*. *Biochemistry* **24**, 5810–5817
49. Kunkel, T. A., and Bebenek, K. (2000) DNA replication fidelity. *Annu. Rev. Biochem.* **69**, 497–529
50. Harris, V. H., Smith, C. L., Cummins, W. J., Hamilton, A. L., Adams, H., Dickman, M., Hornby, D. P., and Williams, D. M. (2003) The effect of tautomeric constant on the specificity of nucleotide incorporation during DNA replication: support for the rare tautomer hypothesis of substitution mutagenesis. *J. Mol. Biol.* **326**, 1389–1401
51. Topal, M. D., and Fresco, J. R. (1976) Complementary base pairing and the origin of substitution mutations. *Nature* **263**, 285–289
52. Bebenek, K., Pedersen, L. C., and Kunkel, T. A. (2011) Replication infidelity via a mismatch with Watson-Crick geometry. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 1862–1867
53. Wang, W., Hellinga, H. W., and Beese, L. S. (2011) Structural evidence for the rare tautomer hypothesis of spontaneous mutagenesis. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 17644–17648
54. Doetsch, P. W., Chan, G. L., and Haseltine, W. A. (1985) T4 DNA polymerase (3′-5′) exonuclease, an enzyme for the detection and quantitation of stable DNA lesions: the ultraviolet light example. *Nucleic Acids Res.* **13**, 3285–3304
55. Kunkel, T. A., and Soni, A. (1988) Exonucleolytic proofreading enhances the fidelity of DNA synthesis by chick embryo DNA polymerase- γ . *J. Biol. Chem.* **263**, 4450–4459
56. Cotton, F. A., Wilkinson, G., Murillo, C. A., and Bochmann, M. (1999) *Advanced Inorganic Chemistry*, 6th Ed., pp. 692–854, John Wiley & Sons, New York
57. Harding, M. M. (2006) Small revisions to predicted distances around metal sites in proteins. *Acta Crystallogr. D Biol. Crystallogr.* **62**, 678–682
58. Mildvan, A. S., and Loeb, L. A. (1979) The role of metal ions in the mechanisms of DNA and RNA polymerases. *CRC Crit. Rev. Biochem.* **6**, 219–244