Combination of Two Mutant Alpha Spectrin Alleles Underlies a Severe Spherocytic Hemolytic Anemia

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Abstract

We studied a patient with a severe spherocytic hemolytic anemia without family history of spherocytosis. Analysis of patient's erythrocyte membrane proteins revealed spectrin deficiency and a truncated α spectrin protein. We determined that the patient is a compound heterozygote with two mutations in α spectrin gene. Mutation in the paternal allele, designated α spectrin^{PRAGUE}, is a transition A to G in the penultimate position of intron 36 that leads to skipping of exon 37, frameshift, and production of the truncated α spectrin protein. The maternal allele, designated α spectrin-LEPRA, contains transition C \rightarrow T in position -99 of intron 30. This mutation enhances an alternative acceptor splice site 70 nucleotides upstream from the regular site. The alternative splicing causes a frameshift and premature termination of translation leading to a significant decrease in α spectrin production. The α^{LEPRA} mutation is linked to a spectrin αIIa marker that was found to be associated with recessive or nondominant spectrin-deficient hereditary spherocytosis in \sim 50% of studied families. We conclude that the α^{LEPRA} mutation combined in trans with the α^{PRAGUE} mutation underlie the severe hemolytic anemia in the proband. We suggest that allele α spectrin^{LEPRA} may be frequently involved in pathogenesis of recessive or nondominant spectrin-deficient hereditary spherocytosis. (J. Clin. Invest. 1996. 98:2300-2307.) Key words: congenital hemolytic anemia • DNA mutational analysis • messenger RNA • alternative splicing • protein synthesis

Introduction

Hereditary spherocytosis (HS)¹ is an inherited hemolytic anemia characterized by the presence of spherical erythrocytes in

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peripheral blood smears and increased osmotic fragility of red blood cells(RBC). The disease prevalence is highest in Northern Europe where it affects $\sim 0.02\,\%$ of population. The molecular basis of HS is heterogeneous, with primary defects in several proteins of the erythrocyte membrane skeleton, a two-dimensional flexible network of spectrin and other proteins laminating the cytoplasmic side of RBC plasma membrane. Several groups identified mutations in band 3 protein (1–7), ankyrin (7–10), and β spectrin (11–15) associated mostly with autosomal dominant HS, and in protein 4.2 (16–20) associated mostly with mild to moderate recessive (r) or nondominant (nd) HS.

Besides the typical autosomal dominant HS and the relatively rare, recessive, protein 4.2-deficient HS, Agre and colleagues identified a subset of patients affected by a more severe anemia inherited in an autosomal recessive or nondominant pattern (r/ndHS) (21–23). Clinical severity of this type of anemia was proportional to a degree of spectrin deficiency, ranging from 53% of normal spectrin content in severely anemic patients to 31% of normal in nearly lethal cases (22). Analysis of red cell membrane proteins isolated from r/ndHS patients demonstrated that in $\sim 50\%$ cases the disease was linked to an α spectrin variant α IIa, whose prevalence in the normal population ranges from 2 to 7% (24-27). This linkage disequilibrium indicates that the α spectrin allele carrying the αIIa mutation plays a role in the pathogenesis of r/ndHS, but there is no direct evidence that the mutation itself has any effect on the phenotype (28).

Spectrin, the predominant constituent of RBC membrane skeleton, is a filamentous protein composed of two subunits: α spectrin (281 kD) and β spectrin (246 kD). The formation of the spectrin heterodimer starts by primary interaction of the two subunits at nucleation sites localized in αV (COOH-terminal) and βIV (NH₂-terminal) domains and proceeds by zipping the two chains together in antiparallel manner (29). The α spectrin is synthesized in more than twice the amount of β spectrin during maturation of mammalian erythrocytes (30, 31), suggesting that in the majority of cases both α spectrin alleles have to be mutated to produce an abnormal RBC phenotype.

This report represents an initial attempt to define molecular basis underlying the spectrin deficiency in patients with r/ndHS. We studied one subject heterozygous for α IIa mutation and affected by the disease. We present two mutations identified in the α spectrin gene that very likely account for patient's phenotype. In addition we suggest that the mutation found in cis with the α IIa marker may represent a defect frequently involved in pathogenesis of spectrin deficient r/ndHS.

Methods

Case report. The proband was splenectomized at the age of 6 because of a severe spherocytic hemolytic anemia. Basic hematological parameters are shown in Table I. In addition, before splenectomy, the

^{1.} Abbreviations used in this paper: HS, hereditary spherocytosis; LEPRA, low expression allele Prague; nt, nucleotide; r/ndHS, recessive nondominant HS; SSCP, single-strand conformational polymorphism.

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Table I. Basic Hematological Parameters of the Studied Subjects

	Date of birth	Splenectomy	Hemoglobin	Hematocrit	MCV	RDW	Retics	Osmotic fragility [‡]
			g/dl	%	fl	%*	%	
Proband	1985		8.5	23.0	80.0		13.4	0.76-0.36
		1991	15.28	44.0§	84.8§	13.98	3.18	0.70-0.38§
Father	1960	No	15.2	46.0	84.9	12.2	1.0	0.46-0.32
Mother	1965	No	14.7	44.0	90.4	12.6	1.2	0.47-0.34

^{*}Normal range 11.5–14.5. *Osmotic fragility is given as an interval between minimum osmotic resistance (normal range 0.48–0.44) and complete hemolysis (normal 0.34–0.30). *Data after splenectomy.

proband was subicteric with total bilirubin 2.9 mg/dl. Autohemolysis without glucose was 13.1%, with glucose 7.9%, spleen was palpable 3 cm over the costal margin, and bone marrow aspirate revealed a marked hyperplasia of the erythroid lineage that comprised 50.4% of total bone marrow cellularity. After splenectomy, the patient had normal red cell count; however, his reticulocyte count remained elevated and osmotic fragility remained increased. Patient's peripheral blood smear contained numerous spherocytes and microspherocytes. There is no family history of spherocytosis and the hematological parameters as well as red cell morphology of both parents are normal.

Membrane protein analysis. Freshly drawn blood anticoagulated in acid citrate/dextrose was shipped on ice from Prague to Boston. The blood samples were processed within 48 h of phlebotomy. Erythrocyte ghosts were prepared by the method of Dodge et al. (32). Proteins were analyzed by SDS-PAGE in 3.5-17% gradient gels according to Agre's modification (22) of the original Fairbanks method (33) and in 12% Laemmli gels (34). The relative abundance of the major red cell membrane proteins was analyzed, using the Eagle Eye II system (Stratagene, La Jolla, CA) and the ONE-Dscan 1.0 program (Scanalytics, Billerica, MA). For immunoblotting, the electrophoresed proteins were transferred to a nitrocellulose membrane, incubated with rabbit anti-human α spectrin antiserum, peroxidase-conjugated goat anti-rabbit antibody (Sigma Chemical Co., St. Louis, MO) and visualized by chloronaphtol staining. For quantitation of band 3 by flow cytometry, washed RBC were incubated with eosin-5-maleimide (Molecular Probes Inc., Eugene, OR) and analyzed using a flow cytometer (PROFILE II; Coulter Electronics, Hialeah, FL) as described (2, 35). For study of the spectrin oligomerization state, spectrin was extracted and the percentage of spectrin dimers was calculated from densitometric tracing of nondenaturing tube gels as described (36).

cDNA analysis. Total reticulocyte RNA was isolated by ammonium chloride lysis (37) and reverse-transcribed using random primers. The structure of α spectrin cDNA was analyzed by PCR amplification of overlapping segments with primers derived from α spectrin sequence (38). To study the aberrant exon 37 region, cDNA from the proband and a control subject were PCR amplified using primers SA39 (5'-CCTATGCCTTGTTCCAGTTC-3'; within exon 36) and 405 (5'-GGATGTCTTCTCCTTGTGCA-3'; within exon 40) (30 cycles; 10 s at 96°C, 1 min at 64°C, and 30 s at 72°C). An aliquot of the PCR product was cloned into plasmid pCR II using the TA Cloning Kit (Invitrogen, San Diego, CA). One clone of the expected size and one clone containing the truncated cDNA were sequenced using the Sequenase Version 2.0 DNA Sequencing Kit (U.S. Biochemical Corp., Cleveland, OH) and a sequencing primer SA39. To analyze the aberrant exon 30 region, cDNA was PCR amplified using primers SA9 (5'-AGAGGACTTCAGTGCAGAAC-3'; within exon 29) and SA10 (5'-CCTGTCTAGTACACGTTGGA-3'; within exon 31) in the DNA Thermal Cycler (Perkin-Elmer Cetus, Norwalk, CT) programmed for 35 cycles (10 s at 96°C, 30 s at 64°C, and 10 s at 72°C). An aliquot of the product was cloned into plasmid; one clone of the expected size and one clone containing the larger cDNA were sequenced using a sequencing primer SA10 as described above. To analyze the alternative splicing, the two isoforms were separated on 1.5% agarose minigel (30 min, 110 V). The gel was stained with ethidium bromide and the relative abundance of the isoforms was analyzed by densitometry as described above. To study the αIIa mutation, cDNA was PCR amplified using primers SA29 (5'-ATGAGGCCTTTC-TATTAGATC-3'; within exon 20) and K2 (5'-AGCTGGGACAAT-GCCCTGAT-3'; within exon 22) (40 cycles; 10 s at 96°C, 30 s at 64°C, and 10 s at 72°C). The PCR product was ethanol precipitated, resuspended in water, digested with restriction endonuclease FokI and analyzed on 2% agarose gel.

Genomic DNA analysis. Genomic DNA was isolated from the proband as described (39). To investigate the exon 37 skipping event, the genomic DNA was PCR amplified using primers SA39 and SA40 (5'-ATCTCCTCTTGCCCCACAGC-3'; within exon 38) (35 cycles; 10 s at 98°C; 1 min at 60°C; and 4 min at 68°C). The PCR product was cloned as described above and six clones together with genomic DNA from the proband and his parents were PCR amplified using primers SA41 (5'-AAGCTATTGCTGTAGGATGAC-3'; within intron 36) and SA18 (5'-GGATGGCAGGCTCATGGG-3'; within exon 37) (25 cycles; 10 s at 96°C, 30 s at 64°C, and 1 s at 72°C) and screened for single-strand conformational polymorphism (SSCP) on 10% glycerol MDE gel (AT Biochem, Malvern, PA) as described (2, 40). The mutant and the normal clone were sequenced as described above, using nested sequencing primer SA18. To study the alternative splicing of intron 30, proband's genomic DNA was PCR amplified using primers SA45 (5'-TGATGAAGAAACGGGACGAT-3'; within exon 30) and SA10 (35 cycles; 10 s at 95°C, 30 s at 60°C, and 2 min at 68°C). The PCR product was cloned and 4 individual and 20 pooled clones were sequenced as described above, using sequencing primers SA42 (5'-AAGAGCCCAGCTGCTTGGAG-3', within alternatively spliced part of intron 30). To screen subjects for the mutation introducing EcoRI site, genomic DNA was PCR amplified using primers SA55 (5'-GACCTCAAGTGATCTACCT-3'; within intron 30) and SA10 (35 cycles; 10 s at 96°C, 30 s at 60°C, and 30 s at 72°C) and the PCR product was digested with restriction endonuclease EcoRI and analyzed on 1.5% agarose gel.

Study of spectrin synthesis in erythroblasts. Erythroblasts were prepared from peripheral blood in a two-phase liquid culture system as described (31, 41). An aliquot of erythroblasts was metabolically labeled for 60 min with [35 S]methionine. The soluble (cytoplasmic) and insoluble (membrane) fractions were separated, spectrin was immunoprecipitated using polyclonal antispectrin antibodies, electropho-resed in 8% Laemmli gel, and the gel was dried and exposed to Hyperfilm (Amersham Corp., Arlington Heights, IL). The intensity of the α and β spectrin bands was analyzed by densitometry as described above.

Results

Biochemical characterization of membrane proteins. To determine defects underlying the severe spherocytic anemia diagnosed in the patient, we analyzed RBC membrane proteins isolated from the patient, his parents, and a control subject on

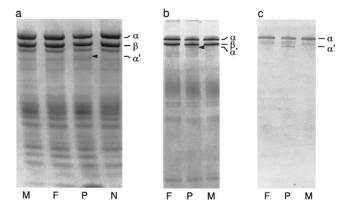


Figure 1. Electrophoretic analysis of RBC membrane proteins. Coomassie blue–stained erythrocyte membrane proteins from the proband (P), his parents (M, F) and a normal subject (N) were electrophoresed (a) in the 3.5–17% exponential gradient polyacrylamide gel and (b) in 12% Laemmli gel. Both gel systems reveal presence of an abnormal band designated as α' (arrowhead). (c) Western blot of the gel in (b) shows immunoreactivity of the truncated band with polyclonal anti– α spectrin antibodies.

the Fairbanks and Laemmli SDS-PAGE. The analysis revealed the presence of an additional band of apparent molecular mass of ~ 195 kD in proband's membranes (Fig. 1, a and b). On Western blots the protein reacted with antibodies against α spectrin (Fig. 1 c) but did not react with antibodies against β spectrin or ankyrin. The band, designated as α' spectrin, constituted $\sim 4\%$ of total α spectrin. In addition to the truncated protein detected in proband, densitometric tracing of Fairbanks gels revealed an \sim 34% decrease in the spectrin to band 3 ratio and an \sim 27% decrease in the ankyrin to band 3 ratio in patient's membrane skeleton. Measurement of the relative number of band 3 copies per cell by flow cytometry revealed a decrease in the band 3 content to 70% of normal. This result allowed us to correct the relative spectrin to band 3 ratio obtained by gel densitometry and to estimate the actual spectrin content in proband's erythrocytes as $\sim 46\%$ of normal (Table II). Functional study of the spectrin dimer-dimer association site by separation of spectrin dimers, tetramers, and oligomers on nondenaturing gel revealed normal ratio of dimers and tetramers (data not shown).

Splice site mutation in paternal allele. To identify the molecular defect responsible for the truncation of α spectrin protein, we analyzed structure of proband's α spectrin mRNA. Amplification and agarose gel analysis of a region spanning exon 37 revealed a doublet in the proband and in his father and a single band in his mother and in five control subjects. The single band of the expected size corresponded to the longer band in the doublet. Cloning and sequencing of the shorter fragment showed that the entire exon 37 was missing from the mRNA (data not shown).

To determine the basis of the aberrant splicing event, proband's genomic DNA spanning intron 36, exon 37, and intron 37 was PCR amplified, cloned, and the clones together with proband's and his parents' genomic DNA were screened for an SSCP in exon–intron boundaries. Sequencing of the polymorphic intron 36–exon 37 boundary revealed an A to G transition at the penultimate position of the acceptor splice site upstream of exon 37: ttgattttag/GGA to ttgattttgg/GGA (Fig.

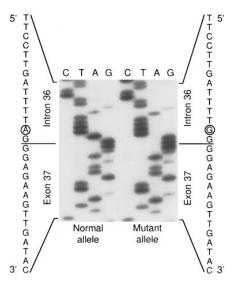


Figure 2. Nucleotide sequences of intron 36–exon 37 junction corresponding to the normal α spectrin allele and to the mutated paternal allele. Sequences of two different subclones derived from proband's genomic DNA are shown. The nucleotide in -2 position of the acceptor splice site of intron 36, mutated in the paternal allele, is circled.

2). The same mutation was identified in father's genomic DNA. Schematic representation of the defective allele designated as α spectrin^{PRAGUE} and the ensuing aberrant splicing leading to a frameshift and premature termination of translation is shown in Fig. 3. 700 amino acid residues from α spectrin COOH terminus (corresponding to a half of the α IV and the entire α V domains) are replaced by eight new amino acids (MCWIWQRS) in the putative truncated protein. The calculated molecular mass of the protein (202 kD) corresponds well to the apparent molecular mass deduced from electrophoretic mobility of the α' spectrin detected in subject's RBC membranes (\sim 195 kD).

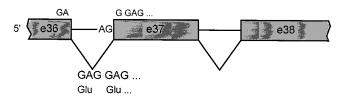
Aberrant alternative splicing in the maternal allele. Detection of one α spectrin PRAGUE allele in the asymptomatic father suggested that the mutation is recessive. Since the propositus is also heterozygous for this recessive mutation, we searched for an additional α spectrin gene defect, which would explain the subject's spherocytic phenotype. PCR amplification and agarose gel analysis of a region spanning exon 30 revealed a doublet with a shorter band of the expected size (Fig. 4). Cloning and sequencing of the longer fragment demonstrated that 70 nucleotides (nt) from the 3' end of intron 30 were inserted into

Table II. Quantitation of Red Cell Membrane Proteins Expressed in Percentages of Control Values

	$\alpha~Sp/\beta~Sp*$	Sp/B3	2.1/B3	B3/cell‡	Sp/cell§
	%	%	%	%	
Proband	93	66	73	70	46
Father	101	97	93	95	92
Mother	99	100	98	92	92

Sp, spectrin; B3, band 3; 2.1, ankyrin isoform 2.1. * α spectrin in the proband includes the truncated α' spectrin (α' contains 4% of α spectrin). *Band 3 content per cell measured by flow cytometry. *Sp/B3 ratio corrected for the band 3 content per cell.

Normal allele



Mutant allele

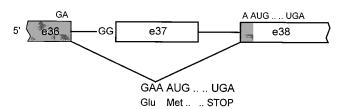


Figure 3. Scheme of exon 37 splicing in normal and α spectrin PRAGUE allele. Mutation of the acceptor splice site of intron 36 leads to skipping of exon 37. Translation of the mutated mRNA is prematurely terminated at the beginning of exon 38. Rectangles correspond to α spectrin exons, coding region is shaded. Not drawn to scale.

the exon 30–exon 31 boundary. Although the doublet was present in all studied subjects, the proband and his mother had disproportionately increased intensity of the longer alternatively spliced isoform (Fig. 4). To exclude the possibility that the varying intensities were artifacts, we blotted the gel and hybridized it with a probe corresponding to the region of α spectrin gene containing exon 30. Ratios of the two isoforms obtained by densitometric scanning of autoradiographs of Southern blots were consistent with ratios calculated from densitometric tracing of ethidium bromide stained agarose gels.

To identify molecular basis of increased aberrant alternative splicing, we cloned and sequenced the entire intron 30 with adjacent exons from proband's and a control genomic DNA. The sequencing revealed an alternative acceptor splice site located 70 nt upstream from the intron 30–exon 31 boundary and a single point mutation (C→T) introducing an EcoRI restriction site 29 nt upstream from the alternative splice site in one patient's allele (Fig. 5). The insertion of 70 nt in the alternatively spliced mRNA leads to a frameshift and premature termination of translation. In the putative truncated protein 983 amino acid residues from α spectrin COOH terminus are replaced by 28 new amino acids (EGVWDCTKSLISKHLGSFLFLY-

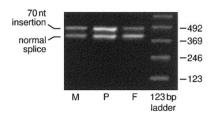


Figure 4. Alternative splicing of intron 30. PCR amplification of cDNA spanning the exon 30–exon 31 boundary reveals alternative splicing of intron

30. The shorter band corresponds to the correct isoform and the longer band corresponds to the alternatively spliced isoform with 70 nt insertion. The alternative isoform is significantly stronger in the proband (P) and in his mother (M) than in his father (F).

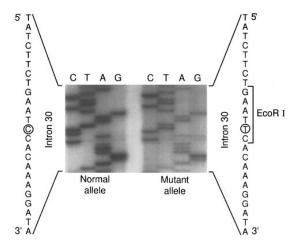
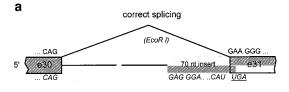
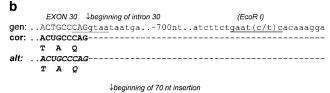


Figure 5. Nucleotide sequence of intron 30 upstream from the alternative acceptor splice site. Sequences of two subclones derived from proband's genomic DNA correspond to the normal α spectrin allele (*left*) and to the mutated α spectrin allele (*right*). The mutated nucleotide in position -99 of intron 30 is circled. The mutation introduces a new EcoRI restriction site into genomic DNA.



alternative splicing



tacc<u>ttcattctttctagg</u>agggagtctgggattgcaccaagagcctaatctccaagca



| beginning of EXON 31
| totgggctotttcottttotttattagGAAGGGAAGATCACTGACCTAGAACATTT..-GAAGGGAAGAUCACUGACCUAGAACAUUU..

E G K I T D L E H F ...
| UCUGGGCUCUUUUCCUUUAUUUAGGAAGGGAAGAUCACUGA

Figure 6. Scheme of the alternative splicing in intron 30. (a) The alternative splicing inserts 70 nt from the 3' end of intron 30 into the mRNA. Translation of the alternatively spliced mRNA is prematurely terminated. Rectangles correspond to α spectrin exons, shaded boxes are the coding sequences. Position of the mutation enhancing the alternative splicing and introducing EcoRI site in α spectrin LEPRA is shown by arrow. Alternatively spliced sequence is typed in italic. (b) Nucleotide sequence of parts of intron 30 with neighboring exons corresponding to the genomic DNA (gen), correctly (cor), and alternatively (alt) spliced RNA. Single underlined are nucleotides matching the consensus sequences for splicing, double underlined is the position of the EcoRI restriction site in the α spectrin lepra allele, and dotted underlined is the termination codon in the alternatively spliced RNA. Amino acid sequences were derived from the two mRNA isoforms (one letter coding system).

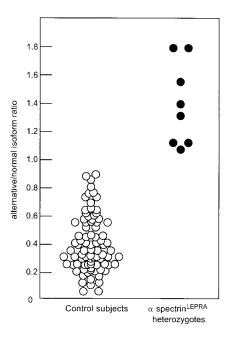


Figure 7. Correlation between the presence of α^{LEPRA} mutation and increased aberrant alternative splicing in intron 30. cDNA from control subjects was PCR amplified and the ratios of alternatively to correctly spliced mRNA were calculated from densitometric tracing of ethidium bromide–stained agarose gels. Based on screening for the EcoRI site created by the mutation the subjects were divided into two groups: α spectrin homozygotes (n=86) represented by open circles and the α spectrin lepra heterozygotes (n=8) represented by filled circles.

LGREDH). The calculated molecular mass of the aberrant protein is 171 kD. The allele carrying the EcoRI mutation was designated as α spectrin^{LEPRA} (low expression allele Prague). Schematic representation of the correct and alternative splicing together with underlying sequences is shown in Fig. 6.

Screening for aberrant alternative splicing and α^{LEPRA} mutation. To pursue our hypothesis that the α^{LEPRA} mutation enhances the alternative acceptor splice site and increases production of the alternatively spliced mRNA, we screened, in addition to the studied family, 91 Czech subjects from 55 families for the ratios of the alternatively to correctly spliced isoform of α spectrin and for the presence of the EcoRI restriction site in intron 30. The studied individuals included both control subjects and patients with autosomal dominant hereditary spherocytosis. We found that six other subjects from four different kindred are heterozygous for the α^{LEPRA} mutation and have an increased alternative/normal isoform ratio (Fig. 7 and Table III). Screening for the α IIa mutation by FokI restriction analysis of 3' region of αII domain revealed that all eight subjects heterozygous for α^{LEPRA} mutation (including the proband and his mother) were also heterozygous for the αIIa marker. The linkage of the two mutations in the studied family, shown in Fig. 8, was confirmed in two other families in whom we could study a child and both parents.

Study of spectrin synthesis in erythroblasts. PCR amplification of cDNA from the propositus and his parents spanning various polymorphic sites as well as the exon 37 deletion for decreasing number of cycles, allowed us to conclude that there is not a major difference in levels of mRNA produced from individual α spectrin alleles in reticulocytes (data not shown). To

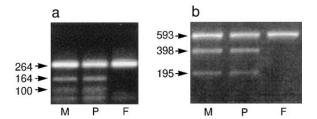


Figure 8. Coinheritance of the α IIa mutation and α spectrin^{LEPRA} mutation in the proband's family. (a) cDNA prepared from the proband (P), his mother (M), and father (F) was PCR amplified with primers specific for 3' region of the α II domain and digested with FokI restriction endonuclease. The 264-bp undigested PCR product corresponds to the common α spectrin allele. The PCR product of α IIa allele is cut with FokI restriction endonuclease into two pieces (164 bp and 100 bp). (b) EcoRI digest of amplified genomic DNA isolated from the proband and his parents. Normal α spectrin allele produces undigested 593-bp-long fragment. The PCR product of the α spectrin^{LEPRA} allele is cut with EcoRI restriction endonuclease into two fragments (398 bp and 195 bp).

determine the ratio of spectrin synthesis, erythroblasts were cultured from peripheral blood of the proband, his parents, and one control subject. Metabolically labeled spectrin was immunoprecipitated from soluble (cytosolic) and insoluble (membrane) fraction and analyzed on SDS-PAGE (Fig. 9). The ratio of α spectrin to β spectrin was decreased in cytosolic fractions of both parents (father: 1.06; mother: 1.22) to $\sim 50\%$ of the control ratio (2.16) and strikingly reduced to 10% (0.21)of the control ratio in the cytosolic fraction from proband's erythroblasts. Careful examination of an overexposed autoradiogram revealed that while no extra band corresponding to the putative alternatively spliced α spectrin^{LEPRA} was detected in cytosolic fractions of the propositus and his mother, there was a suggestion of a weak band migrating at the position of spectrin α' in cytosolic fractions of the propositus and his father. Analysis of membrane fractions from control and proband demonstrated that spectrin is assembled into the membrane skeleton in approximately equimolar amounts (Fig. 9). No truncated α spectrin is detectable in proband's membranes (expected $\sim 4\%$ of total α spectrin is below the detection limit in the membrane fraction). The rates of α spectrin synthesis are consistent with our hypothesis that both parents have a defect in α spectrin synthesis and that the proband inherited both defective alleles from his parents (Fig. 10).

Table III. Frequency of the α^{LEPRA} Mutation

	Subjects	Families
α/α	85	51
α/α^{LEPRA}	6*	4^{\ddagger}
$\alpha^{LEPRA}/\alpha^{LEPRA}$	0	0
$\alpha^{\text{LEPRA}}\text{frequency}$	3.3%	3.6%

Subjects included in the screening were either healthy controls or patients with dominant HS. *All six subjects heterozygous for the α^{LEPRA} mutation were also heterozygous for the α IIa mutation. ‡ In addition to the family of the propositus, the linkage of the α^{LEPRA} and α IIa mutations was demonstrated in two families where both parents and a child were available for study.

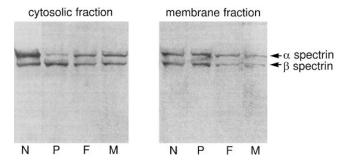


Figure 9. Spectrin synthesis in erythroblasts. Erythroblasts were cultured from peripheral blood from a normal subject (N), the patient (P), and his parents (F, M). Spectrin was immunoprecipitated from soluble (cytosolic) and insoluble (membrane) fractions of metabolically labeled erythroblasts and analyzed on 8% Laemmli gel. The upper band of the autoradiograph corresponds to α spectrin and the lower band to β spectrin.

Discussion

We studied a patient with severe spherocytic hemolytic anemia without family history of spherocytosis. Spectrin deficiency and the spectrin αHa marker detected in one α spectrin allele indicated that this was a case of typical r/ndHS. Identification of an additional truncated α spectrin band in proband's RBC membrane skeleton suggested that the defect underlying the disease resides in the α spectrin gene.

Analysis of the 3' region of α spectrin gene revealed a mutation in the penultimate position of intron 36. Mutations of the conserved dinucleotide in the acceptor splice site lead either to use of a cryptic splice site or to skipping of the following exon (42–46). The latter occurs in the mutated allele designated as α spectrin PRAGUE. Skipping of exon 37 from α spectrin mRNA leads to a frameshift and premature termination of translation at the 5' end of exon 38. The putative truncated protein corresponds in size to the α' spectrin detected in proband's RBC membranes and we propose that the α' spectrin is expressed from the α spectrin α' spectrin is expressed from the α' spectrin α' spectrin detected in probability α' spectrin α' s

Presence of apparently normal α spectrin in patient's membranes, which cannot be synthesized from the paternal α spectrin^{PRAGUE} allele, suggests that the second mutation will reduce expression of the maternal α spectrin allele. One such mutation designated as α^{LELY} was recently discovered by Wilmotte and colleagues in the α spectrin nucleation site (47). Analysis of the nucleation site, however, revealed that none of the family members carries the α^{LELY} mutation. Search for the potential maternal defect brought to our attention alternative splicing in intron 30 of the α spectrin gene. Splicing of intron 30 can proceed either normally by using the regular acceptor splice site and producing a normal mRNA transcript or alternatively by using a conserved alternative acceptor splice site detected 70 nt upstream from the regular site. The alternatively processed α spectrin mRNA retains the last 70 nt from the 3'-end of intron 30, which causes a frameshift and leads to premature termination of translation in exon 31.

While most of the studied control subjects used predominantly the regular splice site, the patient, his mother, and several other subjects produced more of the alternative isoform. We identified a mutation 29 nt upstream from the alternative splice site which is linked to the observed increase in alternative splicing. Since this mutation does not involve any of the known conserved splicing sequences, we suggest that it causes a structural change in pre-mRNA, making the alternative acceptor splice site more accessible for a spliceosome. We designate the maternal allele carrying the mutation α spectrin LEPRA.

Based on results obtained from metabolic labeling of erythroblasts, we calculated that the α spectrin^{LEPRA} allele produces only \sim 1/5 of full-length spectrin as compared with the normal α spectrin allele. Similarly, assuming that the transcription of α spectrin gene and the stability of the two mRNA isoforms are comparable, we can calculate, based on the average ratio of alternative to correct isoform in the normal α spectrin homozygotes (0.4) and the average ratio in the α spectrin^{LEPRA} heterozygotes (1.4), that the α spectrin^{LEPRA} allele produces only \sim 1/6 of the correctly spliced isoform as compared with the normal α spectrin allele. Our hypothesis that the α^{LEPRA} mutation is the actual defect involved in the disease is further supported by our discovery of a linkage between the α^{LEPRA} mutation and the α IIa polymorphism that was shown to be a marker in \sim 50% cases of r/ndHS (25–27). We conclude that

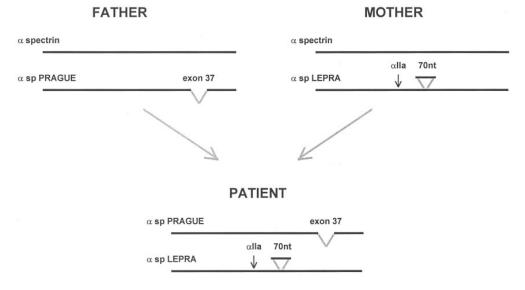


Figure 10. Scheme of inheritance of the two defects in the studied family. The subject is a compound heterozygote with two recessive mutations inherited from his parents. α spectrin-LEPRA allele, which inserts with higher frequency 70 nt from intron 30 into mRNA, was inherited from his mother, and α spectrin-PRAGUE allele, which skips exon 37 from mRNA, was inherited from his father.

the α^{LEPRA} mutation combined in *trans* with the α^{PRAGUE} mutation, are the two defects underlying patient's phenotype.

Spectrin synthesized in avian erythrocytes exists in four different forms: α homodimers, β homotetramers, α/β heterodimers, and α/β heterotetramers (48). The α homodimer form is turned over with a half-life of 2 h (49). Our results from metabolic labeling of human erythroblasts suggest that truncated α spectrin proteins lacking the nucleation site are rapidly turned over with half-lives undetectable by standard pulsechase experiment. Thus, it is likely that α spectrin missing the COOH terminus will be produced as a highly unstable monomeric α spectrin, which will be degraded by a fast pathway distinct from the degradation of dimeric species. The fact that we were able to detect some truncated α spectrin produced from the paternal α spectrin^{PRAGUE} allele in patient's RBC membrane skeleton indicates that even though the protein lacks the entire nucleation site, it still retains a limited ability to bind β spectrin. The rate of this binding is probably much slower than heterodimerization of normal α spectrin; therefore, the truncated protein can only be detected in the α spectrin-deficient environment of patient's erythrocytes. On the other hand, the notably shorter α spectrin produced from the maternal allele is not detectable even in patient's membrane skeleton indicating that the protein completely lost its ability to heterodimerize with β spectrin.

The amount of mammalian α spectrin synthesized during in vitro erythroid differentiation considerably exceeds that of β spectrin (30, 31). Consequently, a complete knockout of one α spectrin allele should not lead to a detectable spectrin deficiency, since one α spectrin allele would produce enough protein to associate with the available β spectrin. Our results prove this hypothesis by demonstrating the absence of the protein product of one α allele in the asymptomatic father of the propositus and by showing that an additional mutation in the other α spectrin allele is needed to produce a clinically apparent phenotype.

We propose that α spectrin^{LEPRA} might represent a wide-spread allele involved in pathogenesis of spectrin deficient, recessive, or nondominant HS. Based on the reported frequency of αIIa polymorphism in families with r/nd HS (26), we expect that $\sim 50\%$ of subjects with this phenotype will be either α spectrin LEPRA homozygotes or compound heterozygotes with another α spectrin defect found in *trans* with α spectrin LEPRA . To address this hypothesis we are currently analyzing patients carrying the αIIa mutation for the presence of the α^{LEPRA} mutation and for aberrant alternative splicing.

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