BEDSIDE TO BENCH REPORT



Complete and sustained response of adult medulloblastoma to first-line sonic hedgehog inhibition with vismodegib

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ABSTRACT

Medulloblastoma is an aggressive primitive neuroectodermal tumor of the cerebellum that is rare in adults. Medulloblastomas fall into 4 prognostically significant molecular subgroups that are best defined by experimental gene expression profiles: the WNT pathway, sonic hedgehog (SHH) pathway, and subgroups 3 and 4 (non-SHH/WNT). Medulloblastoma of adults belong primarily to the SHH category. Vismodegib, an SHH-pathway inhibitor FDA-approved in 2012 for treatment of basal cell carcinoma, has been used successfully in the setting of chemorefractory medulloblastoma, but not as a first-line therapy. In this report, we describe a sustained response of an unresectable multifocal form of adult medulloblastoma to vismodegib. Molecular analysis in this case revealed mutations in *TP53* and a cytogenetic abnormality, i17q, that is prevalent and most often associated with subgroup 4 rather than the SHH-activated form of medulloblastoma. Our findings indicate that vismodegib may also block alternate, non-canonical forms of downstream SHH pathway activation. These findings provide strong impetus for further investigation of vismodegib in clinical trials in the first-line setting for pediatric and adult forms of medulloblastoma.

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Introduction

Medulloblastoma is an aggressive primitive neuroectodermal tumor of the cerebellum with a propensity for distant metastasis and a high recurrence rate.^{1,2} It is the most common primary intracranial malignancy in the pediatric population, but it is quite rare in adults, with an approximate incidence of 0.5 cases per 1,000,000 per year.^{2,3} Due to its rarity in adults, treatment modalities have been adapted from pediatric protocols, and there have been no randomized trials to establish standard-of-care treatment of adult medulloblastoma.

There is rising interest in the use of molecular targeted therapy, and very specifically SHH inhibitors, to treat medulloblastomas. One such inhibitor, vismodegib (GDC-0449), acts by inhibiting mutated smoothened (SMO) receptors, and thus overcoming regulation by PTCH1 (Fig. 1). To date, several phase I studies using vismodegib have reported temporary response in patients with chemorefractory medulloblastoma.^{4,5} In these reports, response has been limited to several months. In a recent phase II study of 31 adults with recurrent medulloblastoma receiving vismodegib, 3 patients with SHH-subgroup type displayed progression-defined response (defined as radiologic response lasting 8 weeks).⁶ Here, we present the case of an exceptional responder, a patient with an unresectable/multifocal form of adult medulloblastoma that has responded completely to SHH inhibition with vismodegib in the first-line setting, with sustained response lasting nearly 3 years to date.

Clinical case report

A 51-year-old male with history of basal cell carcinoma presented with a 3-month history of unremitting headaches, dizziness, expressive aphasia, agraphia, difficulty with balance, and tinnitus. Neurologic evaluation revealed dysmetria of the left upper extremity; no nystagmus was noted. Brain magnetic resonance imaging (MRI) revealed a heterogeneous abnormality in the cerebellum, presenting as a poorly defined mass involving both cerebellar hemispheres and the vermis (Fig. 2A & C). There was no mass effect on the fourth ventricle or evidence of hydrocephalus.

Biopsy of the mass showed a small round blue cell tumor infiltrating the surrounding cerebellar parenchyma (Fig. 3A–F). No desmoplastic features were present, but the small biopsy precluded definitive assessment. Immunohistochemical stains were strongly positive for synaptophysin, with weak glial fibrillary acidic protein (GFAP) and patchy chromogranin positivity. Nuclear staining for β -catenin was absent. CD3, CD20, LCA, cytokeratin, and TTF-1 were negative. Ki-67 showed a high proliferation index (30–50% of tumor cells). A diagnosis of medulloblastoma was made based on morphology, immunophenotype, and anatomic location.

The amount of tissue was insufficient to initiate cultures for G-banding, but fluorescence *in situ* hybridization (FISH) on limited touch preps demonstrated 3 copies of *RAI1* (17p11.2) and *RARA* (17q21) in 161 of 200 cells, which likely represented an isodicentric chromosome with a breakpoint in proximal 17p and deletion of distal 17p (Fig. 3G). Isochromosome

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B Supplemental data for this article can be accessed on the publisher's website.



Figure 1. Schematic of molecular subgroups of medulloblastoma, including the SHH and alternate molecular pathways. Signaling in the SHH subgroup of medulloblastoma and recurring genomic abnormalities observed in other subtypes. Sonic hedgehog (SHH) ligand binds *PTCH1* receptor, relieving this receptor's baseline inhibition of *SMO*. Active *SMO* prevents *SUFU* inhibition of *GLI* and leads to *GLI* nuclear translocation and transcriptional activation as well as transcription-independent upregulation of *MYCN* with subsequent downstream transcriptional regulation. The FDA-approved molecular inhibitors vismodegib and sonidegib block SHH-mediated downstream activation by direct inhibition of SMO receptor. Common genomic aberrations in the SHH subtype include: 1) gernline or somatic mutations of *PTCH1*, *SMO*, or *SUFU*; 2) amplification of 2q (*GLI2*) or 2p (*MYCN*); or deletion of 9q (*PTCH1*), 10q (*SUFU*), or 17p (*TP53*). Note that i17q functionally leads to deletion of 17p. Other genomic alterations profiling, and many of the genomic alterations are not specific for an individual molecular subtype.

17q (i17q) is one of the most frequent recurring abnormalities in medulloblastoma, occurring in approximately 40–50% of cases, predominantly within the Group 4/D subtype in adults.⁷ It has been reported to be a negative prognostic factor;^{8,9} in childhood medulloblastoma, it has been commonly associated with mutations in *TP53* and desmoplastic histology¹⁰, which was consistent with what we found in our case despite its occurrence in an adult. A predominant difference is that studies in the childhood form associated 17p loss with the SHH subgroup.¹⁰

The patient's clinical workup found no evidence of systemic disease at the time of diagnosis, including cytologic evaluation of cerebrospinal fluid. Initial therapy included craniospinal irradiation (1980 cGy to the brain in 11 fractions; 3600 cGy to the spine in 20 fractions). One week into this course, the patient developed headaches and lethargy. Cranial computed tomography scanning demonstrated ventriculomegaly requiring endoscopic third ventriculostomy and placement of an external ventriculostomy drain. Further complications included intractable nausea and vomiting, malnutrition, anorexia, radiation-induced pancytopenia, and lower extremity deep venous thrombosis requiring anticoagulation.

Brain MRI following completion of radiation therapy showed a marked decrease in the size and enhancement of the tumor, with no evidence of spinal metastasis. However, as the patient had continued neurologic compromise and poor performance status, he was not considered to be a candidate for platinum-based chemotherapy. In light of the preponderance of Hedgehog pathway activation in adult medulloblastoma, the Hedgehog inhibitor vismodegib was initiated on the basis of compassionate use. Within four months, there was no evidence of abnormal enhancing lesions at previous sites of known medulloblastoma (Fig. 2B & D). The patient remained on vismodegib for 22 months without evidence of disease recurrence before he elected to stop treatment. He remains disease-free as of 7 months after cessation of therapy. Given his remarkable response, clinical testing of his diagnostic biopsy for somatic mutations in the SHH pathway was considered, but insufficient tissue remained in the archived block.

Retrospective analysis of residual diagnostic cytologic preparations

Although clinical analyses were hampered by the limited quantity of specimen, a residual cytologic touch preparation from the patient's initial diagnosis was available for additional research testing after appropriate patient consent. DNA extracted from this specimen was sequenced on a targeted Next Generation Sequencing (NGS) assay for 21 genes commonly mutated in solid and hematologic malignancies, but no sequence mutations were identified by this panel including within *TP53*.

A qPCR assay was used to assess copy number alterations (CNAs) commonly observed in medulloblastoma. This testing confirmed the deletion of one copy of *TP53* inferred by the clinical FISH results, but showed no CNAs in *SUFU* (10q), *PTCH1* (9q), or *GLI2* (2q), which are known drivers of the SHH molecular subtype (Fig. 4A). Furthermore, no clear CNAs were observed in *MYC* (8q), *MYCN* (2p), or *OTX2* (14q), which also show enrichment in particular molecular subtypes (Fig. 4B).

Finally, somatic activating mutations within *SMO* have been reported to occur in several hotspots within exons 4, 6, and 9¹⁰. Targeted Sanger sequencing was performed on the residual specimen, but no mutations were identified, including the most commonly identified alteration at p.L412.

Clinical medical genetics evaluation

Due to his personal history of basal cell carcinoma, the patient was evaluated for family history, clinical evidence, and genetic testing for Gorlin syndrome/ Nevoid Basal Cell Carcinoma Syndrome



Figure 2. MRI images of unresectable multifocal medulloblastoma at time of diagnosis (A and C) and after 4 months of vismodegib therapy (B and D). Images are shown in axial sections with contrast (top) and coronal sections (bottom).

(NBCCS). This is an autosomal dominant syndrome characterized by basal cell carcinomas and jaw keratocysts associated with a spectrum of facial and skeletal abnormalities. The incidence may be as high as 1 in 20,000 births,¹¹ and sequence mutations or deletions of *PTCH1* are identified in up to 85–90% of cases.¹² The clinical assessment is summarized in Table 1, and germline sequencing with deletion/duplication analysis of *PTCH1* performed at a commercial reference laboratory was reported negative. In light of these findings, a clinical diagnosis of Gorlin syndrome was deemed unlikely but not definitively excluded.

Methods

Consent and clinical testing

Informed consent was obtained directly from the patient for chart review and all research testing performed, with concurrent waiver from the Institutional Review Board. Germline testing for Gorlin syndrome (*PTCH1*) was sent to Ambry Genetics (Aliso Viejo, CA).

Fluorescence in situ hybridization (FISH)

Due to the very small size of the submitted tissue, insufficient tissue remained after the preparation of touch imprints to initiate cultures for a G-banded chromosome analysis. FISH was prepared on a touch imprint slide with probes to 17p11.2 (SpectrumOrange) and 17q21 (Spectrum-Green) (Abbott Molecular, Des Plaines, IL, USA).

Targeted next generation sequencing

A sequencing library was prepared using a custom designed microfluidics-based (Fluidigm Corporation, South San Francisco, CA) PCR amplicon target enrichment for the following genes: TP53, BRAF, KRAS, NRAS, HRAS, EGFR, PIK3CA, MET, IDH1, IDH2, KIT, PDGFRA, ERBB2, WT1, FLT3, CALR,



Figure 3. Histopathology staining of biopsied tumor from this patient. Scale bars = 20 μ m (lower right-hand corner of each image). (A) 40x original magnification of β -catenin immunoperoxidase stain: Diffuse moderately strong cytoplasmic immunostaining of tumor cells for β -catenin. No nuclear staining is present. (B) 40x original magnification of GFAP immunoperoxidase stain: Patchy woderate granular cytoplasmic staining of neoplastic cells for chromogranin. (C) 40x original magnification of GFAP immunoperoxidase stain: Patchy weak to moderately strong cytoplasmic staining of neoplastic cells for GFAP. (D) 40x original magnification of hematoxylin and eosin; Primitive appearing cells with high nuclear to cytoplasmic ratios, focally arranged in a vaguely nested pattern, infiltrate the cerebellar molecular layer. (E) 40x original magnification of KI-67 (30 – 50% of tumor cells) indicating a high proliferative fraction. (F) 40x original magnification of Synaptophysin immunoperoxidase stain: Strong nuclear staining of neoplastic strong cytoplasmic staining of the background infiltrated neuropil. (G) FISH performed on a touch imprint slide shows 3 signals each for 17p11.2 (orange) and 17q21 (green). Of the interphase cells examined, 80.5% had 3 signals each for the 17p11.2 and 17q21 probes. This signal pattern is consistent with either trisomy 17 or with an isodicentric chromosome composed of 2 copies of a chromosome 17 joined in mirror image in their short arms, distal to the 17p11.2 probe.

JAK2, MPL, NPM1,GATA1, and *MYD88*. Sequencing was performed on an Illumina MiSeq (San Diego, CA) with 300 base pair paired end reads and analyzed using a custom somatic variant calling pipeline.¹³

Copy number analysis

A qPCR method to confirm CNV calls has been described recently.¹⁴ Briefly, primers are designed for targets of interest and *GAPDH* is used for normalization. A pooled, sex-matched control (of peripheral blood from 10 healthy individuals) is compared to the analytic sample using a delta-delta method

normalizing target Ct to *GAPDH* Ct. To verify that individual results are not spurious, a negative control consisting of a single sex-match healthy individual is also compared to the pooled control. The pooled control is normalized to a CNA ratio = 1; experimental ratios that drop below 0.7 or rise above 1.3 are considered to be deleted or amplified, respectively.

Sanger sequencing

Primers flanking exons 4, 6, and 9 of *SMO* were designed and appended with M13 sequencing tags. Cycling parameters followed a touchdown PCR approach.¹⁵ Amplification products



Figure 4. Copy Number Alteration Analysis. Normalized copy number (CN) ratios for genes commonly deleted or amplified in medulloblastoma. A negative control and the tumor sample were compared by qPCR to a pooled (10 individual) control as a copy number benchmark. The pooled control is normalized to a CN ratio = 1; experimental ratios that drop below 0.7 or rise above 1.3 are considered to be deleted or amplified, respectively. The 0.53 ratio for *TP53* in the medulloblastoma correlates with FISH results indicating deletion of distal 17p. No other genes passed cut-off criteria for copy number alteration.

were separated on an ethidium bromide stained 1% agarose gel and purified with the QIAquick Gel Extractin Kit (Qiagen). Cycle sequencing products were generated using BigDye Ter-

 Table 1. Evaluation of major and minor criteria for Gorlin syndrome/Nevoid Basal

 Cell Carcinoma Syndrome (NBCCS).

Major Criteria	Patient Findings
Lamellar calcification of the falx cerebri	Negative
Jaw keratocyst	Unlikely based on dental x-rays, but not definitively excluded by a current orthopantogram
2 or more palmar/plantar pits	Negative
Multiple basal cell carcinomas (> 5) or diagnosis before age 30	Negative; patient diagnosed with one BCC at age of 48
First degree relative with Gorlin	Negative; but known family medical history was limited
Minor Criteria	Patient Findings
Childhood medulloblastoma	Negative; patient diagnosed at age of 51
Lympho-mesenteric or pleural cysts	Negative
Macrocephaly (OFC $>$ 97th centile)	Positive
Cleft lip/palate	Negative
Rib or vertebrae anomalies	Negative
Polydactyly	Negative
Cardiac fibroma	Negative
Cataracts or ocular developmental abnormalities	Negative

minator chemistry (Applied Biosystems) and analyzed on a 3130xl Genetic Analyzer (Applied Biosystems).

Discussion

Here, we present a case of unresected adult medulloblastoma that responded completely to SHH inhibition with vismodegib in the first-line setting after craniospinal irradiation. We identified a cytogenetic abnormality (i17q) that is not specific, but is more frequently associated with group 4/D medulloblastoma than with the SHH subtype. Furthermore, this abnormality has been reported as an independent poor prognostic factor in previous studies. Nonetheless, based on the preponderance of SHH subtype in adults and the relative lack of other options, vismodegib given on a compassionate use basis resulted in a phenomenal response in this patient. In context of our patient's extraordinary response to vismodegib, related pathways markers SMO, SUFU, and downstream transcription factor GLI2 were all unaffected in this patient's tumor. We did identify genomic deletion of one copy of TP53 via FISH and copy number analysis, suggesting that this tumor was activated through mechanisms mostly associated with group 4 medulloblastomas that carry intermediate prognosis and have a higher rate of metastasis than the SHH subgroup.¹⁶ Our findings thus suggest that vismodegib can have significant effects through an alternate non-canonical SHH-activation

pathway. TP53 mutations are associated with pediatric forms of medulloblastoma and are considered rare in adults.^{10,17} Loss of 17p (as seen here) is enriched in TP53 mutant cases, but has been most reported with childhood cases.¹⁰ Our evaluation was not exhaustive in comparison to gene expression profiling tools used to define categories of medulloblastoma in clinical research settings, and the limited amount of tissue available for testing highlights the difficulty in identifying sensitive and specific genomic biomarkers that accurately identify cases that may benefit from SHH inhibition.

Historically, the prognosis of medulloblastoma has been clinically assessed based on factors such as the histopathologic subtype (e.g., desmoplastic vs large cell anaplastic) and the presence of residual disease after resection. The recent identification of molecular drivers involved in medulloblastoma tumorigenesis, including Wnt and SHH, has enabled the identification of additional prognostic markers.¹⁸ SHH tumors are associated with desmoplastic histology in the vast majority of cases, in both infant (89%) and adult tumors (nearly all), but only 25% of cases in children.¹⁹ As many as 84% of adult medulloblastoma tumors may show evidence of activation of the SHH pathway.²⁰ When considering both pediatric and adult cases, most studies report a prevalence of approximately 28%.¹⁹ Notably, more than half of medulloblastoma cases are driven by the SHH pathway in the infant population as well as adults.¹⁹ Interestingly, WNT pathway activation is less frequent in adult medulloblastomas.²⁰ Further studies have substantiated these differences in pathway activation as compared to pediatric cases. Evidence to date suggests that SHH pathway activation in adults does not correlate with a favorable prognosis.²⁰ In a meta-analysis of 550 cases, Kool et al. reported an approximately 70% overall survival rate of adults with SHH medulloblastomas by 5 years, but with significant subsequent rise in mortality, evidenced by a 30% overall survival rate by 150 months (12.5 years).¹⁹

Agents targeting the sonic hedgehog (SHH) pathway have been investigated in recurrent adult and pediatric medulloblastomas because a significant subgroup of this disease is driven by this molecular pathway.^{6,16,19} In the case we describe in this report, our patient had a complete and sustained radiologic response over 2 y on vismodegib in the first-line setting after standard craniospinal irradiation. However, this exceptional response occurred in a patient whose tumor did not harbor alterations in SMO or in other canonical SHH pathway-associated drivers; this indicates that vismodegib may inhibit non-canonical alterations that activate the SHH pathway downstream (Fig. 1). In light of these findings, further exploration of vismodegib efficacy in other molecular subgroups of medulloblastoma is warranted. In addition, in the burgeoning era of molecularly targeted therapy, vismodegib merits strong consideration and investigation as an appropriate form of first-line treatment, replacing cytotoxic chemotherapies that have traditionally been used to treat adult forms of the disease based on data from trials performed in the pediatric population.

Disclosure of potential conflicts of interest

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