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Integration of the Transcription Factor-Regulated and Epigenetic Mechanisms in the Control of Keratinocyte Differentiation

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Abstract

The epidermal differentiation program is regulated at several levels including signaling pathways, lineage-specific transcription factors, and epigenetic regulators that establish well-coordinated process of terminal differentiation resulting in formation of the epidermal barrier. The epigenetic regulatory machinery operates at several levels including modulation of covalent DNA/histone modifications, as well as through higher-order chromatin remodeling to establish long-range topological interactions between the genes and their enhancer elements. Epigenetic regulators exhibit both activating and repressive effects on chromatin in keratinocytes (KCs): whereas some of them promote terminal differentiation, the others stimulate proliferation of progenitor cells, as well as inhibit premature activation of terminal differentiation-associated genes. Transcription factor-regulated and epigenetic mechanisms are highly connected, and the p63 transcription factor has an important role in the higher-order chromatin remodeling of the KC-specific gene loci via direct control of the genome organizer Satb1 and ATP-dependent chromatin remodeler Brg1. However, additional efforts are required to fully understand the complexity of interactions between distinct transcription factors and epigenetic regulators in the control of KC differentiation. Further understanding of these interactions and their alterations in different pathological skin conditions will help to progress toward the development of novel approaches for the treatment of skin disorders by targeting epigenetic regulators and modulating chromatin organization in KCs.

INTRODUCTION

Establishment of the functional epidermal barrier is one of the major goals of the epidermal differentiation program, which includes a tightly regulated process of keratinocyte (KC) proliferation, terminal differentiation, apoptosis, and shedding. The program of epidermal development and KC differentiation is governed by coordinated involvement of several transcription factors (p63, AP-1, Klf4, Arnt, and so on), signaling pathways (Wnt, Bmp, Hedgehog, EGF, Notch, FGF, and so on) and epigenetic regulators (DNA/histone-modifying

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enzymes, Polycomb genes, higher-order and ATP-dependent chromatin remodelers, and non-coding RNAs) that control expression of lineage-specific genes (reviewed in Botchkarev *et al.*, 2012; Frye and Benitah, 2012; Perdigoto *et al.*, 2014).

MOLECULAR CONTROL OF CELL DIFFERENTIATION

The p63 transcription factor operates as master regulator of this program, and subsequently and stage-dependently induces in the epidermal progenitor cells expression of several groups of genes encoding the essential components of the cytoskeleton (keratins 5 and 14), cell adhesion molecules (P-cadherin, integrin- α 3, Perp, dystonin, and so on), cell matrix regulators (Fras-1), transcription factors (AP-2 γ , IKK α , IRF6), and epigenetic regulators (Satb1, Brg1) involved in the control of epidermal differentiation (Koster and Roop, 2007; Vanbokhoven *et al.*, 2011; Botchkarev and Flores, 2014).

Genetic p63 ablation in mice results in the failure of stratification of the epidermis and other squamous epithelia, lack of the formation of epidermal appendages (hair follicles, glands, teeth) and in severe abnormalities in the development of limbs and external genitalia (Mills *et al.*, 1999; Yang *et al.*, 1999; Ince *et al.*, 2002). Moreover, heterozygous mutations in the human p63 gene underlie several ectodermal dysplasia syndromes, also characterized by abnormalities in the development of digits, teeth, hairs, nails, and sweat glands (reviewed in Rinne *et al.*, 2007; Koster, 2010).

p63 has multiple isoforms, including TAp63 and Np63, which show distinct, although partially overlapping, roles in the control of epidermal differentiation and stratification (Koster and Roop, 2007; Vanbokhoven *et al.*, 2011; Botchkarev and Flores, 2014). The Np63 isoform is more abundant in the epidermis compared with TAp63, is strongly expressed in basal epidermal KCs, and is markedly downregulated in the spinous layer (Romano *et al.*, 2012). Np63 has a major role in mediating the effects of p63 on epidermal development, whereas TAp63 protects KCs from senescence and suppress tumorigenesis in postnatal epidermis (Guo *et al.*, 2009; Romano *et al.*, 2012).

EPIGENETIC MECHANISMS

Epigenetic mechanisms have an important role in the control of cellular functions in living organisms and are considered as a driving force of the phenotypic plasticity and evolutionary adaptation (Feinberg, 2007). Epigenetic regulatory machinery operates at several levels including modulation of covalent DNA/histone modifications, as well as through higher-order chromatin remodeling to establish long-range topological interactions between the genes and their enhancer elements in three-dimensional (3D) nuclear space (Ho and Crabtree, 2010; Bickmore and van Steensel, 2013).

Epigenetic regulators exhibit both activating and repressive effects on chromatin in KCs: histone demethylase Jmjd3, ATP-dependent chromatin remodeler Brg1, and genome organizer Satb1 promote terminal KC differentiation, whereas DNA methyltransferase (DNMT1), histone deacetylases (HDAC1/2), and polycomp components (Bmi1 and Ezh1/2) stimulate proliferation of progenitor cells via repression of the genes encoding cell-cycle inhibitors, as well as inhibit premature activation of terminal differentiation-associated genes

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(reviewed in Eckert *et al.*, 2011; Botchkarev *et al.*, 2012; Frye and Benitah, 2012; Perdigoto *et al.*, 2014). In addition, epigenetic regulators might control expression of transcription factors in KCs via formation of the active or repressive local chromatin structure at their promoter regions: for instance, p63 expression in KCs is inhibited by histone methyltransferase Setd8 and HDAC2, which interfere with the Np63-regulated gene expression program in KCs (LeBoeuf *et al.*, 2010; Driskell *et al.*, 2012).

The KC nucleus is a complex and highly compartmentalized organelle, whose 3D organization undergoes major changes during terminal differentiation in the epidermis (Gdula *et al.*, 2013). The process of terminal differentiation in epidermal cells is accompanied by sequential changes of gene expression in the keratin type I/II loci, followed by the onset of expression of the epidermal differentiation complex (EDC) locus genes encoding the essential components of the epidermal barrier (Fuchs and Horsley, 2011).

Our recent studies revealed that transcription factor-dependent and epigenetic regulatory mechanisms in KCs are highly connected, and p63 has a hitherto unrecognized role in the higher-order chromatin remodeling of the EDC locus via direct control of the genome organizer *Satb1* and ATP-dependent chromatin remodeler *Brg1* (Fessing *et al.*, 2011; Mardaryev *et al.*, 2014).

Among different epigenetic regulators, Satb1 has a unique role in the execution of lineagespecific gene expression programs by integrating high-order chromatin organization with regulation of gene expression (Cai *et al.*, 2006). Satb1 binds specialized DNA regions with an ATC-sequence context and folds chromatin into loops involving tissue-specific gene loci (T_{H2} -cytokine and MHC class I loci, globin locus, and so on), as well as targets chromatin remodelers/transcription factors to gene loci (Kumar *et al.*, 2007). Satb1 also contributes to the progression of tumors and promotes reprogramming of the genome of breast cancer cells towards metastasis (Kohwi-Shigematsu *et al.*, 2013).

Satb1 is expressed in basal epidermal KCs and promotes cell differentiation via establishment of specific conformation of the EDC locus, whereas its ablation in mice results in the marked elongation of the EDC central domain associated with alterations in expression of the EDC genes and in epidermal morphology (Fessing *et al.*, 2011).

ATP-dependent chromatin remodeler *Brg1*, on the other hand, promotes developmentally regulated relocation of the EDC locus from the nuclear periphery towards nuclear interior into the compartment enriched by nuclear speckles, which is associated with marked increase in expression of the EDC genes (Mardaryev *et al.*, 2014). Importantly, conditional ablation of Brg1 in the epidermis results in failure to form a functional barrier, thus partially resembling the phenotype of p63 knock-out mice (Indra *et al.*, 2005). These data suggest that chromatin remodeling genes represent a novel cohort of p63 targets that mediate its effects on execution of lineagespecific gene expression program in KCs (Botchkarev *et al.*, 2012; Fessing, 2014).

CONCLUSIONS AND FUTURE DIRECTIONS

However, additional efforts are required to fully understand the complexity of interactions between distinct transcription factors and epigenetic regulatory machinery in the control of epidermal development, regeneration, and stem cell activity. Recently, a number of molecules that are capable of modulating distinct components of the epigenetic machinery have been developed, and some of them are already approved for treatment of the distinct neoplastic conditions or under clinical trials (Heightman, 2011). Thus, understanding of epigenetic mechanisms controlling epidermal differentiation and skin stem cell activity and their alterations in different pathological skin conditions will help to develop a novel cohort of epigenetic drugs for the treatment of skin disorders.

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Abbreviations

КС	keratinocyte
3D	three-dimensional
EDC	epidermal differentiation complex

REFERENCES

- Bickmore WA, van Steensel B. Genome architecture: domain organization of interphase chromosomes. Cell. 2013; 152:1270–84. [PubMed: 23498936]
- Botchkarev VA, Flores ER. p53/p63/p73 in the epidermis in health and disease. Cold Spring Harb Perspect Med. 2014; 4:ii.
- Botchkarev VA, Gdula MR, Mardaryev AN, et al. Epigenetic regulation of gene expression in keratinocytes. J Investig Dermatol Symp Proc. 2012; 132:2505–21.
- Cai S, Lee CC, Kohwi-Shigematsu T. SATB1 packages densely looped, transcriptionally active chromatin for coordinated expression of cytokine genes. Nat Genet. 2006; 38:1278–88. [PubMed: 17057718]
- Driskell I, Oda H, Blanco S, et al. The histone methyltransferase Setd8 acts in concert with c-Myc and is required to maintain skin. EMBO J. 2012; 31:616–29. [PubMed: 22117221]
- Eckert RL, Adhikary G, Rorke EA, et al. Polycomb group proteins are key regulators of keratinocyte function. J Investig Dermatol Symp Proc. 2011; 131:295–301.
- Feinberg AP. Phenotypic plasticity and the epigenetics of human disease. Nature. 2007; 447:433–40. [PubMed: 17522677]
- Fessing MY. Gene regulation at a distance: higher-order chromatin folding and the coordinated control of gene transcription at the epidermal differentiation complex locus. J Investig Dermatol Symp Proc. 2014; 134:2307–2310.
- Fessing MY, Mardaryev AN, Gdula MR, et al. p63 regulates Satb1 to control tissue-specific chromatin remodeling during development of the epidermis. J Cell Biol. 2011; 194:825–39. [PubMed: 21930775]
- Frye M, Benitah SA. Chromatin regulators in mammalian epidermis. Semin Cell Dev Biol. 2012; 23:897–905. [PubMed: 22944592]

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- Fuchs E, Horsley V. Ferreting out stem cells from their niches. Nat Cell Biol. 2011; 13:513–8. [PubMed: 21540847]
- Gdula MR, Poterlowicz K, Mardaryev AN, et al. Remodelling of three-dimensional organization of the nucleus during terminal keratinocyte differentiation in the epidermis. J Investig Dermatol Symp Proc. 2013; 133:2191–201.
- Guo X, Keyes WM, Papazoglu C, et al. TAp63 induces senescence and suppresses tumorigenesis *in vivo*. Nat Cell Biol. 2009; 11:1451–7. [PubMed: 19898465]
- Heightman TD. Therapeutic prospects for epigenetic modulation. Expert Opin Ther Targets. 2011; 15:729–40. [PubMed: 21366500]
- Ho L, Crabtree GR. Chromatin remodelling during development. Nature. 2010; 463:474–84. [PubMed: 20110991]
- Ince TA, Cviko AP, Quade BJ, et al. p63 Coordinates anogenital modeling and epithelial cell differentiation in the developing female urogenital tract. Am J Pathol. 2002; 161:1111–7. [PubMed: 12368184]
- Indra A, Dupe V, Bornert JM, et al. Temporally controlled targeted somatic mutagenesis in embryonic surface ectoderm and fetal epidermal keratinocytes unveils two distinct developmental functions of BRG1 in limb morphogenesis and skin barrier formation. Development. 2005; 132:4533–44. [PubMed: 16192310]
- Kohwi-Shigematsu T, Poterlowicz K, Ordinario E, et al. Genome organizing function of SATB1 in tumor progression. Semin Cancer Biol. 2013; 23:72–9. [PubMed: 22771615]
- Koster MI. p63 in skin development and ectodermal dysplasias. J Investig Dermatol Symp Proc. 2010; 130:2352–8.
- Koster MI, Roop DR. Mechanisms regulating epithelial stratification. Annu Rev Cell Dev Biol. 2007; 23:93–113. [PubMed: 17489688]
- Kumar PP, Bischof O, Purbey PK, et al. Functional interaction between PML and SATB1 regulates chromatin-loop architecture and transcription of the MHC class I locus. Nat Cell Biol. 2007; 9:45– 56. [PubMed: 17173041]
- LeBoeuf M, Terrell A, Trivedi S, et al. Hdac1 and Hdac2 act redundantly to control p63 and p53 functions in epidermal progenitor cells. Dev Cell. 2010; 19:807–18. [PubMed: 21093383]
- Mardaryev AN, Gdula MR, Yarker JL, et al. p63 and Brg1 control developmentally regulated higherorder chromatin remodelling at the epidermal differentiation complex locus in epidermal progenitor cells. Development. 2014; 141:101–11. [PubMed: 24346698]
- Mills AA, Zheng B, Wang XJ, et al. p63 is a p53 homologue required for limb and epidermal morphogenesis. Nature. 1999; 398:708–13. [PubMed: 10227293]
- Perdigoto CN, Valdes VJ, Bardot ES, et al. Epigenetic regulation of epidermal differentiation. Cold Spring Harb Perspect Med. 2014; 4:ii.
- Rinne T, Brunner HG, van Bokhoven H. p63-associated disorders. Cell Cycle. 2007; 6:262–8. [PubMed: 17224651]
- Romano RA, Smalley K, Magraw C, et al. DeltaNp63 knockout mice reveal its indispensable role as a master regulator of epithelial development and differentiation. Development. 2012; 139:772–82. [PubMed: 22274697]
- Vanbokhoven H, Melino G, Candi E, et al. p63, a story of mice and men. J Investig Dermatol Symp Proc. 2011; 131:1196–207.
- Yang A, Schweitzer R, Sun D, et al. p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. Nature. 1999; 398:714–8. [PubMed: 10227294]