

## Possible Anticancer Mechanisms of Some *Costus speciosus* Active Ingredients Concerning Drug Discovery



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**Abstract:** *Costus speciosus* is native to South East Asia, especially found in India, Srilanka, Indonesia and Malaysia. *C. speciosus* have numerous therapeutic potentials against a wide variety of complains. The therapeutic properties of *C. speciosus* are attributed to the presence of various ingredients such as alkaloids, flavonoids, glycosides, phenols, saponins, sterols and sesquiterpenes. This review presented the past, present, and the future status of *C. speciosus* active ingredients to propose a future use as a potential anticancer agent. All possible up-regulation of cellular apoptotic molecules as p53, p21, p27, caspases, reactive oxygen species (ROS) generation and others attribute to the anticancer activity of *C. speciosus* along the down-regulation of anti-apoptotic agents such as Akt, Bcl2, NFκB, STAT3, JAK, MMPs, actin, surviving and vimentin. Eventually, we recommend further investigation of different *C. speciosus* extracts, using some active ingredients and evaluate the anticancer effect of these chemicals against different cancers.



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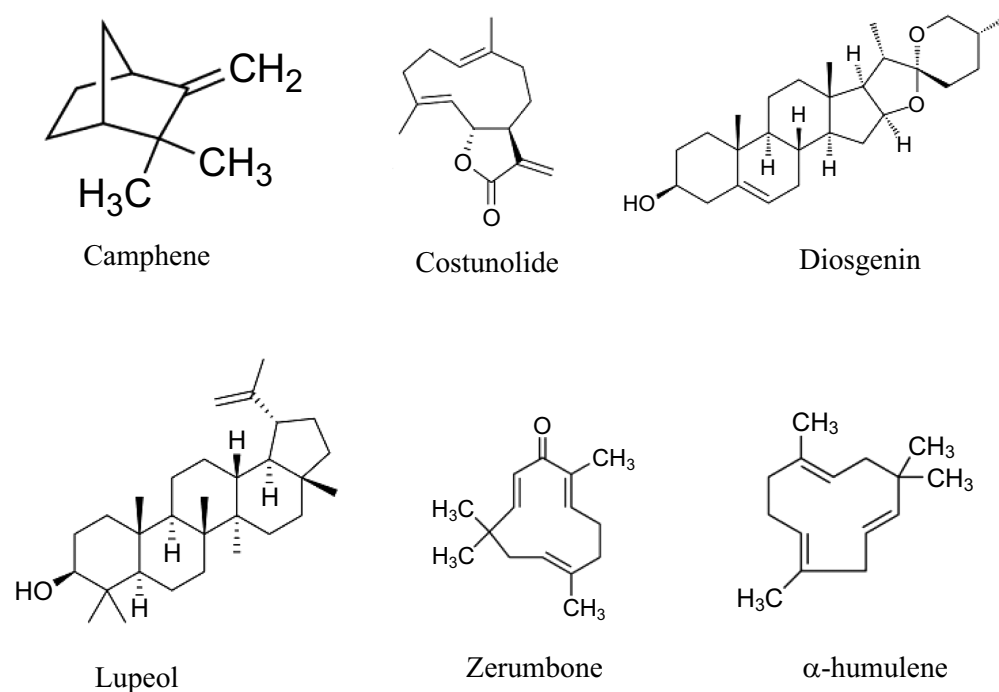
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### INTRODUCTION

Plant active principles are important task for developing therapeutic agents. Herbal products are greatly safe in comparison to the synthetics. Herbal natural products are a components of different parts of medicinal herb [1]. From the important medicinal plant families is the *Zingiberaceae* that distributed throughout tropical Africa, Asia, Americas and Indo-Malayan region, Sri Lanka and in India. It is commonly grown along road sides and streams [2]. The rhizomes and roots are ascribed to have an anthelmintic, expectorant, tonic, aphrodisiac, flatulence, anti-inflammatory, antidiabetic,

hepatoprotective, antihyperlipidemic, antispasmodic and antimicrobial activities [3]. Indeed, leaf extract of *C. speciosus* shows potential *in vitro* anticancer activity toward liver cancer [4]. *C. speciosus* serves as an important source of numerous compounds owning many pharmacological benefits as diosgenin, tigogenin, saponins and β-sitosterol; diosgenin, 5α-stigmast-9 (11)-en-3β-ol, β-sitosterol-β-D-glucoside, dioscin, prosapogenins A and B of dioscin, gracillin, α-tocopherol; diosgenone, cycloartanol, 25-en-cycloartenol and octacosanoic acid [5]. The major compounds *C. speciosus* oils such as α-humulene, zerumbone, camphene, α-amyrin stearate, β-amyrin, costunolide and lupeol have been isolated from its rhizomes and their structural formula are illustrated in Fig. 1 [6,7].

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**Fig. (1).** Shows the structural formulae of some active ingredients present in *C. speciosus*.

## Anticancer Activity Evaluation and Mechanism of *Costus speciosus* Active Ingredients

### *In Vitro* Anticancer Activity

#### Up-Regulation of p53, p21 and p27

The tumor suppressor p53 is a transcription factor that responds to diverse cases of cellular stress. It is recognized as the guardian of the genome [8]. P53 promotes growth arrest genes as p21. The p21 is a tumor suppressor able to suppress cancer cell proliferation [9]. The pro-apoptotic gene products such as the PUMA, Noxa, BAX and p53AIP1 localize to the mitochondria and promote the loss of mitochondrial membrane potential and cytochrome c release. Moreover, Fas or DR5/KILLER, the components of the extrinsic pathway of apoptosis were regulated by p53. Finally, p53 induces in reactive oxygen species (ROS) production that damage the mitochondria, leading to apoptosis [10].

Cyclin-dependent kinase inhibitor 1B (p27Kip1) is an enzyme inhibitor that binds to and inhibits the activation of cyclin E-CDK2 and cyclin D-CDK4 complexes, and thus induces G1 phase arrest that may be stop or slow down the cancer cell growth [11]. The up-regulation of p53, p21 and p27 by *C. speciosus* active ingredients in different cancer cells is illustrated in Table 1.

### Up-Regulation of Caspases

Caspases are endo-proteases that accomplish their activity by hydrolysing cell protein peptide bonds. The apoptotic caspases have been sub-classified by their mechanism of action into initiator caspases (caspase-8 and -9) or executioner caspases (caspase-3, -6, and -7) [12]. They are activated in both main apoptotic pathways: extrinsic, mediated by death receptors, and intrinsic, where mitochondria play a central role. The mitochondrial pathway activates caspase-9, which, when activated, forms an apoptosome in the cytosol, together with cytochrome c, Apaf-1 and deoxyadenosine triphosphate (dATP). The apoptosome activates caspase-3 [13]. Whereas, the extrinsic death receptor Fas pathway is activated by Fas ligand interaction with Fas complexes those activate caspase 3 and induce apoptosis [14]. The up-regulation of apoptotic initiators and executioner caspases by *C. speciosus* active ingredients in numerous cancer cells are illustrated in Table 2.

### Calcium Overload Induce Apoptosis

Variation in cytosolic calcium concentration promotes numerous cellular functions as contraction of myofilaments, secretion of hormonal secretion and metabolic regulation [15]. However, it has become clear that cellular  $\text{Ca}^{2+}$  overload can cause cytotoxicity and trigger apoptosis [16]. The up-regulation of intracellular  $\text{Ca}^{2+}$  by *C. speciosus* active ingredients presented in Table 3.

Table 1. Up-regulation of p53, p21 and p27 by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Up-regulation of p53	Diosgenin	Human osteosarcoma (1547)	[21]
		Cervix carcinoma (HEp-2)	
		Human melanoma (M4Beu)	
		Human osteosarcoma (1547)	[22]
	Zerumbone	Human lung cancer (NSCLC)	[24]
		Human pancreatic carcinoma (PANC-1)	[25]
Up-regulation of p21	Costunolide	Human prostate cancer (PC-3)	[26]
		Breast cancer (MDA-MB-231)	[27]
	Diosgenin	Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
		Human hepatoma cells (SMMC-7721)	
		Human osteosarcoma (1547)	[23]
		Human erythromyeloblastoid leukemia (K562)	[29]
	Lupeol	Human colon carcinoma (HCT-116)	[30]
		Human osteosarcoma cells (MNNG/HOS)	[31]
		Human osteosarcoma cells (MG-63)	
		Human pancreatic cancer (PCNA-1)	[32]
Zerumbone	Melanoma (451Lu)	[33]	
Zerumbone	Human pancreatic carcinoma (PANC-1)	[25]	
Up-regulation of p27	Diosgenin	Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
		Human hepatoma cells (SMMC-7721)	
	Lupeol	Human osteosarcoma cells (MNNG/HOS)	[31]
		Human osteosarcoma cells (MG-63)	
		Human pancreatic cancer (PCNA-1)	[32]

Table 2. Up-regulation of caspases by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Up-regulation of caspase-3	Camphene	Murine melanoma cell (B16F10-Nex2)	[34]
		Human pancreatic carcinoma (MIA PaCa-2)	[35]
		Human hepatocellular carcinoma (HepG2)	
		Human colon adenocarcinoma (SW-480)	
	Costunolide	Human promyelocytic leukemia (HL-60)	[36]
		Breast cancer (MDA-MB-231)	[27]

Table 2. contd....

		Human bladder carcinoma (T24)	[6]
		Ovarian cancer (MPSC1)	[37]
		Human ovarian carcinoma (A2780)	
		Human ovarian carcinoma (SKOV3)	
		Human breast adenocarcinoma (MCF-7)	[38]
		Breast cancer (MDA-MB-231)	
	Diosgenin	Human erythromyeloblastoid leukemia (K562)	[39]
		Human promyelocytic leukemia (HL-60)	
		Human osteosarcoma (1547)	[21]
		Cervix carcinoma (HEp-2)	
		Human melanoma (M4Beu)	
		Human lung carcinoma (A549)	[40]
		Human erythroleukemia (HEL)	[41]
		Human hepatocellular carcinoma (HepG2)	[42]
		Human breast adenocarcinoma (MCF-7)	
		Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
		Human hepatoma cells (SMMC-7721)	
		Human epidermoid carcinoma (A431)	
		Human hepatocellular carcinoma (Hep2)	[43]
Human erythroleukemia (HEL)	[44]		
Human erythromyeloblastoid leukemia (K562)	[29]		
Human colon adenocarcinoma (HT-29)	[45]		
Up-regulation of caspase-3	Lupeol	Melanoma (451Lu)	[33]
		Head and neck squamous cell carcinoma (HNSCC)	[46]
		Human hepatoma cells (SMMC-7721)	[47]
		Human hepatocellular carcinoma (HepG2)	
	Zerumbone	Acute promyelocytic leukemia (NB4)	[48]
		Chronic myeloid leukemia (CML)	[49]
		Human erythromyeloblastoid leukemia (K562)	
		Human T-cell (Jurkat)	[50]
		Human lung cancer (NSCLC)	[24]
		Human renal carcinoma (786-0)	[51]
		Human renal carcinoma (769-P)	
		Human brain malignant glioma (GBM8401)	[52]
		Human pancreatic carcinoma (PANC-1)	[25]
	Human epithelioid cervical carcinoma (HeLa)	[53]	
leaves methanol extract	Human hepatocellular carcinoma (HepG2)	[4]	

Table 2. contd....

Up-regulation of caspase-7	Camphene	Human pancreatic carcinoma (MIA PaCa-2)	[35]
		Human hepatocellular carcinoma (HepG2)	
		Human colon adenocarcinoma (SW-480)	
	Costunolide	Human promyelocytic leukemia (HL-60)	[36]
		Human neuroblastoma (IMR-32)	[54]
		Human neuroblastoma (NB-39)	
		Human neuroblastoma (SK-N-SH)	
Human neuroblastoma (LA-N-1)			
Up-regulation of caspase-6	Costunolide	Human promyelocytic leukemia (HL-60)	[36]
Up-regulation of caspase-8	Costunolide	Breast cancer (MDA-MB-231)	[27]
		Ovarian cancer (MPSC1)	[37]
		Human ovarian carcinoma (A2780)	
		Human ovarian carcinoma (SKOV3)	
	Diosgenin	Human lung carcinoma (A549)	[40]
		Human erythroleukemia (HEL)	[41]
		Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
	Human hepatoma cells (SMMC-7721)		
	Lupeol	Pancreatic cancer (PaCa)	[55]
Zerumbone	Acute promyelocytic leukemia (NB4)	[48]	
Up-regulation of caspase-9	Costunolide	Ovarian cancer (MPSC1)	[37]
		Human ovarian carcinoma (A2780)	
		Human ovarian carcinoma (SKOV3)	
		Human breast adenocarcinoma (MCF-7)	[38]
		Breast cancer (MDA-MB-231)	
	Diosgenin	Human lung carcinoma (A549)	[40]
		Human erythroleukemia (HEL)	[41]
		Human erythromyeloblastoid leukemia (K562)	[39]
		Human promyelocytic leukemia (HL-60)	
		Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
	Human hepatoma cells (SMMC-7721)		
	Lupeol	Human hepatoma cells (SMMC-7721)	[56]
	Zerumbone	Chronic myeloid leukemia (CML)	[49]
		Human erythromyeloblastoid leukemia (K562)	
		Human T-cell (Jurkat)	[50]
Human lung cancer (NSCLC)		[24]	
Human renal carcinoma (786-0)		[51]	
Human renal carcinoma (769-P)			
Acute promyelocytic leukemia (NB4)		[48]	

**Table 3.** Up-regulation of Bax by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Up-regulation of Bax	Costunolide	Human bladder carcinoma (T24)	[6]
	Diosgenin	Human erythromyeloblastoid leukemia (K562)	[29]
		Human erythroleukemia (HEL)	[57]
		Human lung carcinoma (A549)	[40]
		Human epidermoid carcinoma (A431)	[43]
		Human hepatocellular carcinoma (Hep2)	
		Human erythroleukemia (HEL)	[41]
	Lupeol	Human epidermoid carcinoma (A431)	[58]
		Melanoma (451Lu)	[33]
		Head and neck squamous cell carcinoma (HNSCC)	[46]
	Zerumbone	Human lung cancer (NSCLC)	[24]
Human hepatocellular carcinoma (HepG2)		[59]	
Intracellular Ca <sup>2+</sup> increase	Camphene	Murine melanoma cell (B16F10-Nex2)	[34]
	Zerumbone	Human prostate cancer (PC-3)	[60]
		Human prostate cancer (DU-145)	
		Chronic myeloid leukemia (CML)	[49]
Human erythromyeloblastoid leukemia (K562)			
Overload of nuclear Ca <sup>2+</sup>	Costunolide	Human prostate cancer (PC-3)	[26]
		Human prostate cancer (DU-145)	
		Human prostate adenocarcinoma (LNCaP)	

### **Up-regulation of ROS Generation**

Nitric oxide synthase (nNOS) is a Ca<sup>2+</sup>-dependent cytosolic enzyme that forms nitric oxide (NO) from l-arginine, and NO reacts with the free superoxide radical (O<sub>2</sub><sup>-</sup>) to form the toxic free peroxynitrite radical (ONOO<sup>-</sup>). These free radicals predispose the damage of cellular membranes and intracellular proteins, enzymes and DNA. COX-2-dependent reactions generate ROS during the conversion of arachidonic acid to prostaglandin G<sub>2</sub>, causing direct oxidative damage to DNA and favour apoptosis [17]. The ROS generation in cancer cells and the antioxidant status augmentation of cancer bearing animal by *C. speciosus* active ingredients are tabulated in Table 4.

### **Induction of Apoptosis and Oppose Metastasis**

The up-regulation of the following mentioned apoptotic molecules by *C. speciosus* active ingredients is illustrated in Table 5. In which, the

apoptosis inducing factor (AIF) is a mitochondrial intermembrane flavoprotein that induce chromatin condensation and DNA cleavage. AIF can also participate in the regulation of apoptosis by means of mitochondrial membrane permeabilization [18].

E-cadherin plays important roles in cell-cell adhesion. Cancer cell metastasis include loss of cell-cell adhesion that leads to increased invasiveness, entry into the circulation, dispersion to distant anatomic sites, extravasation and colonization. Therefore, down-regulation of E-cadherin facilitates metastasis. The combination of diosgenin and HIF-1 $\alpha$  silencing RNAs can enhance the expression of E-cadherin [19]. Phosphatase and tensin homolog (PTEN) inhibits p-Akt and mouse double minute 2 homolog (MDM2), and then increases the level of p53, thereby inducing G1 phase arrest and apoptosis. PTEN functions by dephosphorylation of

Table 4. Up-regulation of antioxidant status by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Intracellular thiols depletion	Costunolide	Human prostate cancer (PC-3)	[26]
		Human prostate cancer (DU-145)	
		Human prostate adenocarcinoma (LNCaP)	
Up-regulation of 5-LOX	Diosgenin	Human colon carcinoma (HCT-116)	[61]
		Human colon adenocarcinoma (HT-29)	
Up-regulation of COX-2	Diosgenin	Human colon adenocarcinoma (HT-29)	[61]
		Human colon carcinoma (HCT-116)	
		Human colon carcinoma (HCT-116)	[62]
		Human colon adenocarcinoma (HT-29)	
ROS generation	Costunolide	Breast cancer (MDA-MB-231)	[27]
		Human promyelocytic leukemia (HL-60)	[36]
		Human bladder carcinoma (T24)	[6]
		Ovarian cancer (MPSC1)	[37]
		Human ovarian carcinoma (A2780)	
		Human ovarian carcinoma (SKOV3)	
	Diosgenin	Human erythromyeloblastoid leukemia (K562)	[29]
	Lupeol	Human prostate adenocarcinoma (LNCaP)	[63]
		Human epidermoid carcinoma (A431)	[58]
	Zerumbone	Human lung cancer (NSCLC)	[24]
		Chronic myeloid leukemia (CML)	[49]
		Human erythromyeloblastoid leukemia (K562)	
		Human colon carcinoma (HCT116)	[64]
		Human pancreatic carcinoma (PANC-1)	[25]
$\alpha$ -Humulene	Human colon carcinoma (CaCo-2)	[65]	
$\beta$ -amyrin	Human bladder carcinoma (NTUB1)	[66]	

Table 5. Up-regulation of some apoptotic molecules by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Up-regulation of Apaf1	Lupeol	Human epidermoid carcinoma (A431)	[58]
Up-regulation of AIF	Diosgenin	Human osteosarcoma (1547)	[21]
		Cervix carcinoma (HEp-2)	
		Human melanoma (M4Beu)	
Up-regulation of ATF3	Zerumbone	Human colon carcinoma (HCT116)	[67]
		Human colon adenocarcinoma (SW-480)	
Up-regulation of E-cadherin	Diosgenin	Human gastric cancer (BGC-823)	[68]
Up-regulation of FADD	Lupeol	Human hepatoma cells (SMMC-7721)	[69]

Table 5. contd....

Up-regulation of Fas	Costunolide	Breast cancer (MDA-MB-231)	[27]
	Zerumbone	Acute promyelocytic leukemia (NB4)	[48]
Up-regulation of integrin $\alpha 5$	Diosgenin	Human gastric cancer (BGC-823)	[68]
Up-regulation of integrin $\beta 6$ .	Diosgenin	Human gastric cancer (BGC-823)	[68]
Up-regulation of Notch2	Zerumbone	Human breast adenocarcinoma (MCF-7)	[70]
		Breast cancer (MDA-MB-231)	[70]
Up-regulation of PTEN	Lupeol	Hepatocellular carcinoma (MHCC-LM3 HCC)	[71]
Up-regulation of Rab27a	Lupeol	Mouse melanoma (B16 2F2)	[72]
Up-regulation of thromboxane synthase	Diosgenin	Human erythroleukemia (HEL)	[41]
Up-regulation of DR4	Zerumbone	Human colon carcinoma (HCT116)	[64]

phosphatidyl inositol 3-phosphate (PIP3) and negatively regulating survival signalling mediated by protein kinase B/Akt (PKB/Akt) [20].

#### **Down-Regulation of Akt**

Akt is a serine-threonine kinase which regulates cell growth, survival and proliferation. The phosphatidylinositol 3-kinase/Akt pathway plays a key role in cancer cell survival [73]. Foxo inhibits tumor growth in breast cancer, and cytoplasmic localization of Foxo interrelated with poorer cancer cell survival. Phosphorylation of Foxos by Akt inhibits transcriptional functions of Foxos and contributes to cell survival, growth and proliferation [73]. The cell survival encouraged by Akt was diminished by *C. speciosus* active ingredients (Table 6).

#### **Cell Cycle Arrest**

The cell cycle starts by G1 phase, during which cytoplasmic organelles are replicated. Afterward, the cell enters into the S phase where the DNA is replicated. After which cell reaches the second phase, G2 where proteins and other cellular elements are synthesized. Eventually, the cell enters M phase where it splits into two daughter cells [74]. Cell cycle progression is forcefully regulated by interaction between cyclin-dependent kinases (Cdk1, 2, 4, or 6) and regulatory cyclin subunits (cyclin A, B, Ds, or E). The cell arrest is accompanied by micro-nucleation resulting from chromosome fragments [75]. This cycle arrest was

accomplished by *C. speciosus* active ingredients in different cell cycle phases as presented in Table 7.

#### **Down-Regulation of BCL2**

B cell lymphoma-2 (BCL2) family proteins are key regulators of the apoptotic process and classified into three subgroups anti-apoptotic (BCL2, BCL-XL, and BCL2L10), pro-apoptotic (e.g. BAX, BAK, and BOK) and BH3-only pro-apoptotic members (e.g. BID, BAD, and BIM) [76]. BCL2 and the BCL2-associated X protein gene (BAX) are an oncogene and a cancer suppressor gene, respectively. Overexpression of BCL2 promotes cell survival *in vitro* and *in vivo*. When Bax is overexpressed, cell apoptosis will be hastened. Hence, the ratio BCL2/Bax governs the cell survival or death [77]. Moreover, NF- $\kappa$ B p65/p52 signalling mediated the effects of Glial-cell-line-derived neurotrophic factor (GDNF) on BCL2 and BCL2-w expressions [78]. The up-regulation of Bax by *C. speciosus* active ingredients in different cancer cells is illustrated in Table 3. Whereas BCL2 down-regulations is presented in Table 8.

#### **Down-Regulation of NF $\kappa$ B**

Nuclear factor  $\kappa$ B (NF $\kappa$ B) is a transcription factor that activates its own inhibitor (I $\kappa$ B) as well as groups of pro-apoptotic and anti-apoptotic genes [79]. NF $\kappa$ B activates the inhibitor of apoptosis protein (IAP) gene transcription and down-regulate the activity of the caspase cascade.



Table 6. Down-regulation of PI3-kinase/Akt by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Down-regulation of PI3-kinase/Akt	Lupeol	Human hepatocellular carcinoma (HepG2)	[86]
		Human hepatoma cells (SMMC-7721)	
	Diosgenin	Human prostate cancer (PC-3)	[87]
Down-regulation of Akt	Diosgenin	Human erythroleukemia (HEL)	[88]
		Mouse melanoma (B16)	[89]
		Human prostate cancer (DU145)	[90]
		Human epidermoid carcinoma (A431)	[43]
		Human hepatocellular carcinoma (Hep2)	
		Breast cancer (HER2)	[91]
		Human breast carcinoma (BCa)	[92]
	Human breast carcinoma (BCa)		
	Lupeol	Human epidermoid carcinoma (A431)	[58]
	Zerumbone	Human colon carcinoma (HCT116)	[93]
Human brain malignant glioma (GBM8401)		[52]	
Non-Small Cell Lung Cancer (A549)		[94]	
Down-regulation of (p-PI3K)	Lupeol	Human osteosarcoma cells (MNNG/HOS)	[31]
		Human osteosarcoma cells (MG-63)	
		Human pancreatic cancer (PCNA-1)	[32]
Down-regulation of p-AKT	Lupeol	Gallbladder carcinoma (GBC-SD)	[95]
		Human osteosarcoma cells (MNNG/HOS)	[31]
		Human osteosarcoma cells (MG-63)	
		Human pancreatic cancer (PCNA-1)	[32]

Table 7. Down-regulation of cell cycle components by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Down-regulation of cdc25B	Lupeol	Human prostate cancer (PC-3)	[96]
		Human prostate cancer (PC-3)	[60]
	Human prostate cancer (DU-145)		
	Zerumbone	Human breast adenocarcinoma (MCF-7)	
		Breast cancer (MDA-MB-231)	
		Human breast adenocarcinoma (MCF-7)	
		Breast cancer (MDA-MB-231)	
Down-regulation of cdc42	Diosgenin	Breast cancer (MDA-MB-231)	[98]
Down-regulation of cdk 1	Zerumbone	Human breast adenocarcinoma (MCF-7)	[97]
		Breast cancer (MDA-MB-231)	

Table 7. contd....

Down-regulation of cdk 2	Diosgenin	Human breast carcinoma (BCa)	[92]
	Lupeol	Melanoma (451Lu)	[33]
		Human prostate adenocarcinoma (LNCaP)	[99]
Down-regulation of cdk 4	Diosgenin	Human breast carcinoma (BCa)	[92]
Down-regulation of cyclin A	Lupeol	Human prostate adenocarcinoma (LNCaP)	[99]
		Human prostate cancer (DU145)	
Down-regulation of cyclin B	Lupeol	Swiss albino mice	[100]
		Human prostate cancer (PC-3)	[96]
Down-regulation of cyclin B1	Diosgenin	Human erythromyeloblastoid leukemia (K562)	[29]
	Lupeol	Human prostate adenocarcinoma (LNCaP)	[99]
		Human prostate cancer (DU145)	
	Zerumbone	Human breast adenocarcinoma (MCF-7)	[97]
		Breast cancer (MDA-MB-231)	
Acute promyelocytic leukemia (NB4)	[48]		
Down-regulation of cyclin D1	Diosgenin	Human breast carcinoma (BCa)	[92]
	Lupeol	Melanoma (451Lu)	[33]
		Human prostate adenocarcinoma (LNCaP)	[99]
		Human prostate cancer (DU145)	
		Human osteosarcoma cells (MNNG/HOS)	[31]
		Human osteosarcoma cells (MG-63)	
		Human pancreatic cancer (PCNA-1)	[32]
Down-regulation of cyclin D2	Lupeol	Melanoma (451Lu)	[33]
		Human prostate adenocarcinoma (LNCaP)	
		Human prostate cancer (DU145)	[99]
		Human prostate adenocarcinoma (LNCaP)	
		Human prostate cancer (DU145)	
G0/G1 phase arrest	Lupeol	Human osteosarcoma cells (MG-63)	[31]
		Human osteosarcoma cells (MNNG/HOS)	
		Human pancreatic cancer (PCNA-1)	[32]
	Zerumbone	Human prostate cancer (DU145)	[101]
		Human prostate cancer (PC-3)	
Human colon adenocarcinoma (HT-29)	[102]		
G1 phase arrest	Costunolide	Human prostate cancer (PC-3)	
		Human prostate cancer (DU-145)	[26]
		Human prostate adenocarcinoma (LNCaP)	
	Diosgenin	Human erythroleukemia (HEL)	[103]
		Human osteosarcoma (1547)	[23]
Human breast carcinoma (BCa)	[92]		

Table 7. contd....

G1/S phase arrest	Lupeol	Melanoma (451Lu)	[33]
G2/M phase arrest	Costunolide	Breast cancer (MDA-MB-231)	[27]
		Human bladder carcinoma (T24)	[6]
		Human breast adenocarcinoma (MCF-7)	[38]
		Breast cancer (MDA-MB-231)	
	Diosgenin	Human erythroleukemia (HEL)	[57]
		Human hepatoma (Bel-7402)	[28]
		Human hepatocellular carcinoma (HepG2)	
		Human hepatoma cells (SMMC-7721)	
		Human erythromyeloblastoid leukemia (K562)	[39]
		Human promyelocytic leukemia (HL-60)	
		Human erythromyeloblastoid leukemia (K562)	[29]
	Lupeol	Human prostate cancer (PC-3)	[96]
	Zerumbone	Human ovarian cancer (Caov-3)	[104]
		Human epithelioid cervical carcinoma (HeLa)	[104]
		Human colorectal cancer (CRC)	[105]
Human colon adenocarcinoma (HT-29)		[102]	
Human breast adenocarcinoma (MCF-7)		[97]	
Breast cancer (MDA-MB-231)			
Acute promyelocytic leukemia (NB4)		[48]	

Table 8. Down-regulation of Bcl-2 and Bcl-xL by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Down-regulation of Bcl-2	Costunolide	Human bladder carcinoma (T24)	[6]
		Human ovarian carcinoma (SKOV3)	[37]
		Human ovarian carcinoma (A2780)	
		Human ovarian carcinoma (SKOV3)	
	Diosgenin	Human lung carcinoma (A549)	[40]
		Human epidermoid carcinoma (A431)	[43]
		Human hepatocellular carcinoma (Hep2)	
		Human erythroleukemia (HEL)	[41]
		Human breast carcinoma (BCa)	[92]
		Human erythromyeloblastoid leukemia (K562)	[29]
		Human colon adenocarcinoma (HT-29)	[45]
		Human erythroleukemia (HEL)	[57]
		Human erythromyeloblastoid leukemia (K562)	[39]
	Human promyelocytic leukemia (HL-60)		
Lupeol	Human epidermoid carcinoma (A431)	[58]	

Table 8. contd....

		Human breast adenocarcinoma (MCF-7)	[106]
		Melanoma (451Lu)	[33]
		Head and neck squamous cell carcinoma (HNSCC)	[46]
	Zerumbone	Human renal carcinoma (786-0)	[51]
		Human renal carcinoma (769-P)	
		Human hepatocellular carcinoma (HepG2)	[59]
Down-regulation of Bcl-xL	Diosgenin	Human erythroleukemia (HEL)	[44]
		Human erythromyeloblastoid leukemia (K562)	[29]
	Lupeol	Human breast adenocarcinoma (MCF-7)	[106]
	Zerumbone	Human prostate cancer (PC-3)	[60]
		Human prostate cancer (DU-145)	
		Human colon carcinoma (HCT116)	[93]

Following stimulation of the cell by a variety of agents, I $\kappa$ B is degraded, allowing NF- $\kappa$ B to translocate to the nucleus and bind to the promoter regions of its multiple target genes to promote cell survival [80,81]. The cell survival induced by NF- $\kappa$ B was down-regulated by *C. speciosus* active ingredients (Table 9).

#### **PARP Cleavage**

DNA damage activates nuclear poly (ADP-ribose) polymerase-1 (PARP-1) to repair DNA. The activated PARP-1 uses NAD<sup>+</sup> to form polymers of ADP-ribose that amend PARP-1 and DNA repair proteins [82]. PARP-1 was cleaved by *C. speciosus* active ingredients that inhibit DNA repair of cancer cells apoptosis (Table 10).

#### **Down-Regulation of STAT3, JAK and MMPs**

Signal transducer and activator of transcription 3 (STAT3) is a transcription factor which in humans is encoded by the STAT3 gene. The Janus kinase/signal transducers and activators of transcription (JAK/STAT) pathway regulate signals for development and homeostasis. JAK activation stimulates cell proliferation, differentiation, cell migration and apoptosis [83]. The down-regulation of STAT3 and JAKs by *C. speciosus* active ingredients was presented in Table 10.

The expression of matrix metalloproteinases (MMPs) correlates with the extracellular matrix degradation and tumor metastasis. The expression

of MMP-2 and MMP-9 is associated with metastasis of numerous human cancers because they play an important role in the degradation of type IV collagen, which is a major component of the basement membrane [84]. Therefore, MMP-2, -3 and -9 may be involved in the process of metastasis of breast cancer to the brain. MMPs were down-regulated by *C. speciosus* active ingredients that inhibit DNA repair of cancer cells apoptosis (Table 10).

#### **Down-Regulation of p38 MAPK**

The p38 is a member of Ser/Thr kinases family called the mitogen-activated protein kinase (MAPKs) (Table 12). The p38 MAPK signalling coordinates cellular responses during erythropoiesis in which the proliferation and differentiation of erythroid progenitors are controlled by erythropoietin through the p38 (Table 11) and Jun N-terminal kinase (JNK) (Table 9) signalling cascades [85].

#### **Down-Regulation of Cell Survival and Angiogenesis Molecules**

The down-regulation of numerous anti-apoptotic molecules such as actin, survivin, vimentin and others by *C. speciosus* active ingredients is illustrated in Tables (12).

#### **In Vivo Anticancer Activity**

The *in vivo* anticancer activity of diosgenin, lupeol and zerumbone was conducted by up-

**Table 9. Down-regulation of NF- $\kappa$ B, JAKs and JNK by *C. speciosus* active ingredients.**

Mechanism	Ingredients	Cell	References
Down-regulation of NF- $\kappa$ B	Costunolide	Breast cancer (MDA-MB-231)	[107]
	Diosgenin	Human prostate cancer (PC-3)	[87]
		Human erythroleukemia (HEL)	[44]
		Human breast carcinoma (BCa)	[92]
	Lupeol	Head and neck squamous cell carcinoma (HNSCC)	[46]
		human pancreatic adenocarcinoma cells (AsPC-1)	[108]
	Zerumbone	Pancreatic cancer (PaCa)	[109]
		Breast cancer (HER2)	[110]
		Breast cancer (MDA-MB-231)	[111]
		Human gastric carcinoma (AGS)	[112]
Down-regulation of JAK1	Diosgenin	hepatocellular carcinoma (HCC)	[113]
Down-regulation of JAK2	Diosgenin	hepatocellular carcinoma (HCC)	[113]
	Zerumbone	Human prostate cancer (DU145)	[101]
Human prostate cancer (PC-3)			
Down-regulation of JAKs	Lupeol	Human hepatocellular carcinoma (HepG2)	[56]
		Human liver hepatoma (PLC/PRF5)	
		Human hepatoma-derived (C3A)	
		Hepatocarcinoma (HUH-7)	
		Human hepatoma (Hep3B)	
Down-regulation of JNK	Diosgenin	Breast cancer (HER2)	[91]
		Human prostate cancer (PC-3)	[87]
		Human epidermoid carcinoma (A431)	[43]
		Human hepatocellular carcinoma (Hep2)	

**Table 10. Down-regulation of PARP, STAT3 and MMPs by *C. speciosus* active ingredients.**

Mechanism	Ingredients	Cell	References
PARP cleavage	Costunolide	Breast cancer (MDA-MB-231)	[27]
		Human bladder carcinoma (T24)	[6]
		Human neuroblastoma (IMR-32)	[54]
	Diosgenin	Human erythroleukemia (HEL)	[44]
		Human lung carcinoma (A549)	[40]
	Lupeol	Human hepatoma cells (SMMC-7721)	[47]
		Human hepatocellular carcinoma (HepG2)	

Table 10. contd....

		Pancreatic cancer (PaCa)	[55]	
		Human epidermoid carcinoma (A431)	[58]	
		Human prostate cancer (CWR22Rnu1)	[108]	
		Melanoma (451Lu)	[33]	
	Zerumbone	Chronic myeloid leukemia (CML)	Human erythromyeloblastoid leukemia (K562)	[49]
		Human renal carcinoma (769-P)	[48]	
		Acute promyelocytic leukemia (NB4)		
Down-regulation of STAT3	Diosgenin	hepatocellular carcinoma (HCC)	[113]	
	Lupeol	Human hepatocellular carcinoma (HepG2)	[56]	
		Human liver hepatoma (PLC/PRF5)		
		Human hepatoma-derived (C3A)		
		Hepatocarcinoma (HUH-7)		
		Human hepatoma (Hep3B)		
	Zerumbone	Human prostate cancer (DU145)	[101]	
Breast cancer cells		[114]		
Down-regulation of MMP-2	Diosgenin	Human prostate cancer (PC-3)	[87]	
	Lupeol	Prostate cancer (CaP)	[115]	
Down-regulation of MMP-3	Zerumbone	Breast cancer (Hs578T)	[116]	
		Breast cancer (MDA-MB-231)		
Down-regulation of MMP-9	Costunolide	Breast cancer (MDA-MB-231)	[27]	
	Diosgenin	Human prostate cancer (PC-3)	[87]	
	Lupeol	Gallbladder carcinoma (GBC-SD)	[95]	

Table 11. Down-regulation of CXCR4, CXCL12, p52, p65, p70S6K and p100 by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Down-regulation of C-X-C chemokine receptor type 4 (CXCR-4)	Zerumbone	Breast cancer (HER2)	[110]
		Chronic Myelogenous Leukemia (KBM-5)	
		Human myeloma (U266)	
		Human squamous carcinoma (SCC4)	
		Human embryonic kidney (A293)	
		Human non-small cell lung carcinoma (H1299)	
		Human pancreatic carcinoma (PANC-1)	
		Pancreatic carcinoma (PANC-28)	
		Human pancreatic carcinoma (MIA PaCa-2)	
Down-regulation of C-X-C motif chemokine 12 (CXCL12)	Zerumbone	Breast cancer (HER2)	[110]
		Human pancreatic carcinoma (PANC-1)	
		Pancreatic carcinoma (PANC-28)	

Table 11. contd....

		Human pancreatic carcinoma (MIA PaCa-2)	
Down-regulation of p38	Diosgenin	Human esophageal cancer (Eca109)	[117]
		Human erythroleukemia (HEL)	[44]
Down-regulation of p52	Costunolide	Breast cancer (MDA-MB-231)	[107]
Down-regulation of p65	Costunolide	Breast cancer (MDA-MB-231)	
Down-regulation of p70S6K	Lupeol	human osteosarcoma cells (MG-63)	[31]
		human pancreatic cancer (PCNA-1)	[32]
Down-regulation of p100	Costunolide	Breast cancer (MDA-MB-231)	[107]

Table 12. Down-regulation of some anti-apoptotic molecules by *C. speciosus* active ingredients.

Mechanism	Ingredients	Cell	References
Down-regulation of actin	Diosgenin	Breast cancer (MDA-MB-231)	[98]
Down-regulation of $\beta$ -catenin	Diosgenin	Human colon carcinoma (HCT-116)	[30]
	Lupeol	Prostate cancer (CaP)	[115]
Down-regulation of FADD-like IL-1 $\beta$ -converting enzyme)-inhibitory protein (c-FLIP)	Lupeol	Pancreatic cancer (PaCa)	[55]
Down-regulation of c-Src	Diosgenin	Hepatocellular carcinoma (HCC)	[113]
Down-regulation of epidermal growth factor receptor (EGFR)	Lupeol	Gallbladder carcinoma (GBC-SD)	[95]
Down-regulation of extracellular signal-regulated kinase (ERK)	Diosgenin	Human prostate cancer (PC-3)	[87]
		Human erythroleukemia (HEL)	[118]
Down-regulation of GLI	Diosgenin	Human erythroleukemia (HEL)	[118]
Down-regulation of Gli-1	Zerumbone	Human renal carcinoma (786-0)	[51]
		Human renal carcinoma (769-P)	
Down-regulation of glycogen synthase kinase-3 (GSK3beta)	Diosgenin	Mouse melanoma (B16)	[89]
Down-regulation of Hepatocyte growth factor (HGF)	Diosgenin	Human prostate cancer (DU145)	[90]
Down-regulation of hypoxia-inducible factor 1 (HIF-1 $\alpha$ )	Diosgenin	Human gastric cancer (BGC-823)	[68]
		Gastric carcinoma (NCI-N87)	
		Human gastric adenocarcinoma (MGC80-3)	
Down-regulation of human telomerase reverse transcriptase (hTERT)	Diosgenin	Human gastric cancer (SGC-7901)	[119]
		Human lung carcinoma (A549)	
Down-regulation of myeloid leukemia cell differentiation protein (Mcl-1)	Zerumbone	Human lung carcinoma (A549)	[120]
		Human prostate cancer (PC-3)	[60]
		Human prostate cancer (DU-145)	

Table 12. contd....

Down-regulation of mouse double minute 2 homolog (Mdm2)	Diosgenin	Human prostate cancer (DU145)	[90]
Down-regulation of MAPKs	Zerumbone	Human colon carcinoma (CaCo-2)	[121]
		Human colon carcinoma (Colo320DM)	
		Human colon adenocarcinoma (HT-29)	
Down-regulation of mammalian target of rapamycin (mTOR)	Diosgenin	Human prostate cancer (DU145)	[90]
		Breast cancer (HER2)	[91]
Down-regulation of Polo-like kinase 1 (PLK-1)	Lupeol	Human prostate cancer (PC-3)	[96]
Down-regulation of Smoothed (SMO)	Diosgenin	Human erythroleukemia (HEL)	[118]
Down-regulation of survivin	Costunolide	Human bladder carcinoma (T24)	[6]
	Diosgenin	Human breast carcinoma (BCa)	[92]
	Lupeol	Prostate cancer (CaP)	[99]
	Zerumbone	Human colon carcinoma (HCT116)	[93]
Down-regulation of tumor necrosis factor-alpha (TNF- $\alpha$ )	Costunolide	Breast cancer (MDA-MB-231)	[122]
Down-regulation of vascular endothelial growth factor (VEGF)	Zerumbone	Human gastric carcinoma (AGS)	[112]
Down-regulation of Vav2	Diosgenin	Breast cancer (MDA-MB-231)	[98]
Down-regulation of vimentin	Diosgenin	Human prostate cancer (DU145)	[90]
Down-regulation of Wnt	Lupeol	Human Melanoma (Mel 928)	[123]
Down-regulation of X-linked inhibitor of apoptosis	Diosgenin	Human breast carcinoma (BCa)	[92]
	Zerumbone	Human colon carcinoma (HCT116)	[93]

regulation of caspase, Bax, antioxidant potential and PTEN. In contrary, they induce down-regulation of cyclin B, G2/M phase, Bcl-2, NF- $\kappa$ B and surviving as presented in Table 13.

## CONCLUSION AND RECOMENDATIONS

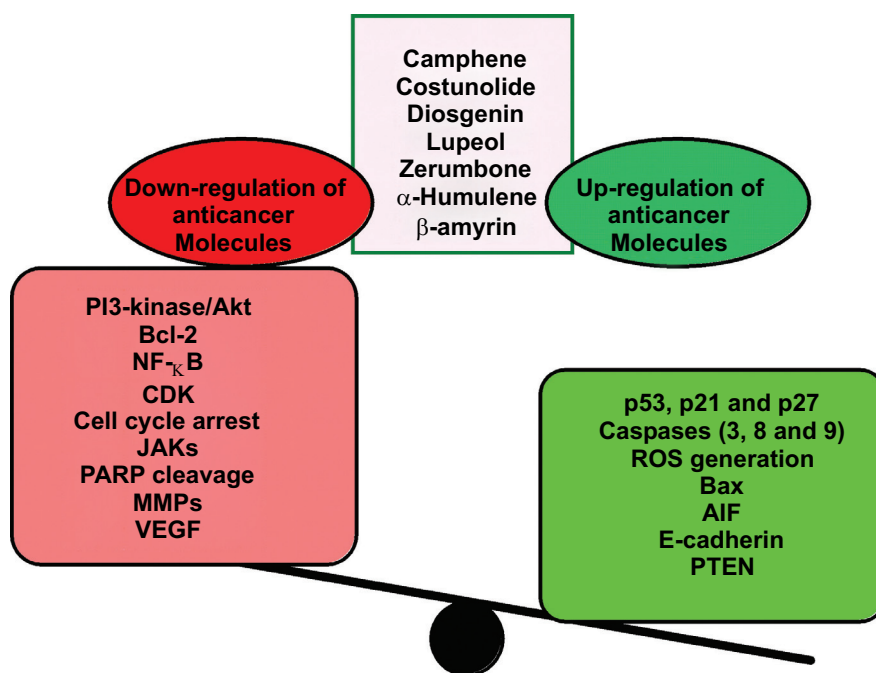
Chemical ingredients of *C. speciosus* have been described as potent anticancer therapy through induction of cancer cell apoptosis and weaken the cell survival through various mechanisms as illustrated in Fig. 2. From the data of this review we can propose research suggestions as a future plan outline that include:

- Study the preclinical novel angiogenesis inhibitor of *C. speciosus*
- Study the critical component of multiple signalling pathways that regulate proliferation, survival, metastasis, and angiogenesis especially Myeloid leukemia.
- Study the possible use of some *C. speciosus* ingredients as adjuvant therapy in chemoresistant cancer; especially hepatocellular carcinoma, breast cancer, and colorectal cancer.
- Study the possible use of some *C. speciosus* ingredients as anti-RAGE, the receptor for advanced glycation end products therapy.
- Study the change of the common regimen for treatment of cancer on a basis to improve the efficacy and reduce the common serious side effects.



Table 13. *In vivo* anticancer activity of *C. speciosus* active ingredients.

Mechanism	Ingredients	Animal	References
Up-regulation of caspase-3	Lupeol	Hamster buccal pouch carcinogenesis	[124]
		Skin of Swiss albino mice	[100]
Up-regulation of caspase-9	Lupeol	Hamster buccal pouch carcinogenesis	[124]
Up-regulation of Bax	Lupeol	Hamster buccal pouch carcinogenesis	[124]
	Zerumbone	Skin of Swiss albino mice	[100]
Antioxidant activity	Diosgenin	Hepatocarcinogenesis in rat	[125]
		Breast carcinoma in female rats	[126]
		Mouse colon carcinogenesis	[127]
	Lupeol	Hamster buccal pouch carcinogenesis	[128]
	Zerumbone	Oral carcinogenesis	[128]
	Zerumbone	Hepatocarcinogenesis in rat	[125]
Up-regulation of PTEN	Lupeol	Bladder carcinogenesis in rats	[129]
Down-regulation of cyclin B	Lupeol	Skin of Swiss albino mice	[100]
G2/M phase arrest	Lupeol	Skin of Swiss albino mice	[100]
Down-regulation of Bcl-2	Lupeol	Hamster buccal pouch carcinogenesis	[124]
		Skin of Swiss albino mice	[100]
	Zerumbone	Hepatocarcinogenesis in rat	[125]
Down-regulation of NF- $\kappa$ B	Lupeol	Skin cancer in CD-1 mice	[130]
		Colonic adenocarcinomas in mice	[131]
	Zerumbone	Lung adenomas in mice	[131]
Down-regulation of survivin	Lupeol	Skin of Swiss albino mice	[100]

Fig. (2). Summarizes the anticancer effect of *Costus speciosus* active ingredients.

- Studies the dose-dependent antiproliferative activity in the human breast cancer MCF-7 cells as a microtubule-interacting agent. These

studies demonstrated that costunolide can be related to an interaction with microtubules and inhibits the proliferation of breast cancer cells.

**CONFLICT OF INTEREST**

The authors confirm that this article content has no conflict of interest.

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Declared none.

**REFERENCES**

- [1] Rajashree R, Gangolli D, Patil S, Ingawale K. Amla, Ashwagandha and Shatavari Formulations as Herbal Medicines and Nutraceuticals. *Res J Pharm Sci* 2012; 1: 10–15.
- [2] Gupta AK, Tondon N, Sharma M. Quality Standards of Indian Medicinal Plants, Medicinal Plants Unit. Publ by Indian Council Med Res 2008; Vol VII.
- [3] Saraf A. Phytochemical and Antimicrobial Studies of Medicinal Plant *Costus Speciosus* (Koen.). E-Journal Chem Hindawi Publishing Corporation 2010; 7: S405–S413.
- [4] Nair SVG, Hettihewa M, Rupasinghe HPV, Nair SVG, Hettihewa M, Rupasinghe HPV. Apoptotic and Inhibitory Effects on Cell Proliferation of Hepatocellular Carcinoma HepG2 Cells by Methanol Leaf Extract of *Costus speciosus*. *Biomed Res Int* 2014; 2014: 1–10.
- [5] Qiao C, Li Q, Dong H, Xu L, Wang Z. [Studies on chemical constituents of two plants from *Costus*]. *Zhongguo Zhong Yao Za Zhi* 2002; 27: 123–5.
- [6] Rasul A, Bao R, Malhi M, *et al.* Induction of apoptosis by costunolide in bladder cancer cells is mediated through ROS generation and mitochondrial dysfunction. *Molecules* 2013; 18: 1418–33.
- [7] Santos F, Frota J, Arruda B, *et al.* Antihyperglycemic and hypolipidemic effects of  $\alpha$ ,  $\beta$ -amyryn, a triterpenoid mixture from *Protium heptaphyllum* in mice. *Lipids Health Dis* 2012; 11: 98.
- [8] Lyle S, Hoover K, Colpan C, Zhu Z, Matijasevic Z, Jones SN. Dicer cooperates with p53 to suppress DNA damage and skin carcinogenesis in mice. *PLoS One* 2014; 9: e100920.
- [9] Masgras I, Carrera S, de Verdier PJ, *et al.* Reactive oxygen species and mitochondrial sensitivity to oxidative stress determine induction of cancer cell death by p21. *J Biol Chem* 2012; 287: 9845–54.
- [10] Polyak K, Xia Y, Zweier JL, Kinzler KW, Vogelstein B. A model for p53-induced apoptosis. *Nature* 1997; 389: 300–5.
- [11] Møller MB. P27 in cell cycle control and cancer. *Leuk Lymphoma* 2000; 39: 19–27.
- [12] McIlwain DR, Berger T, Mak TW. Caspase functions in cell death and disease. *Cold Spring Harb Perspect Biol* 2013; 5: a008656.
- [13] Zou H, Henzel WJ, Liu X, Lutschg A, Wang X. Apaf-1, a human protein homologous to *C. elegans* CED-4, participates in cytochrome c-dependent activation of caspase-3. *Cell* 1997; 90: 405–13.
- [14] Wajant H. The Fas signaling pathway: more than a paradigm. *Science* 2002; 296: 1635–6.
- [15] Berridge MJ, Bootman MD, Roderick HL. Calcium signalling: dynamics, homeostasis and remodelling. *Nat Rev Mol Cell Biol* 2003; 4: 517–29.
- [16] Mattson MP, Chan SL. Calcium orchestrates apoptosis. *Nat Cell Biol Nature* 2003; 5: 1041–3.
- [17] Nikolic D, van Breemen RB. DNA Oxidation Induced by Cyclooxygenase-2. *Chem Res Toxicol* 2001; 14: 351–4.
- [18] Candé C, Cohen I, Daugas E, *et al.* Apoptosis-inducing factor (AIF): a novel caspase-independent death effector released from mitochondria. *Biochimie* 2002; 84: 215–22.
- [19] Berx G, Cleton-Jansen AM, Nollet F, *et al.* E-cadherin is a tumour/invasion suppressor gene mutated in human lobular breast cancers. *EMBO J* 1995; 14: 6107–15.
- [20] Haas-Kogan D, Shalev N, Wong M, Mills G, Yount G, Stokoe D. Protein kinase B (PKB/Akt) activity is elevated in glioblastoma cells due to mutation of the tumor suppressor PTEN/MMAC. *Curr Biol* 1998; 8: 1195–8.
- [21] Corbiere C, Liagre B, Terro F, Beneytout JL. Induction of antiproliferative effect by diosgenin through activation of p53, release of apoptosis-inducing factor (AIF) and modulation of caspase-3 activity in different human cancer cells. *Cell Res* 2004; 14: 188–96.
- [22] Corbiere C, Liagre B, Bianchi A, *et al.* Different contribution of apoptosis to the antiproliferative effects of diosgenin and other plant steroids, hecogenin and tigogenin, on human 1547 osteosarcoma cells. *Int J Oncol* 2003; 22: 899–905.
- [23] Moalic S, Liagre B, Corbière C, *et al.* A plant steroid, diosgenin, induces apoptosis, cell cycle arrest and COX activity in osteosarcoma cells. *FEBS Lett* 2001; 506: 225–30.
- [24] Hu Z, Zeng Q, Zhang B, Liu H, Wang W. Promotion of p53 expression and reactive oxidative stress production is involved in zerubone-induced cisplatin sensitization of non-small cell lung cancer cells. *Biochimie* 2014; 107 Pt B: 257–62.
- [25] Zhang S, Liu Q, Liu Y, Qiao H, Liu Y. Zerubone, a Southeast Asian Ginger Sesquiterpene, Induced Apoptosis of Pancreatic Carcinoma Cells through p53 Signaling Pathway. *Evid Based Complement Alternat Med* 2012; 2012: 936030.
- [26] Hsu JL, Pan SL, Ho YF, Hwang TL, Kung FL, Guh JH. Costunolide induces apoptosis through nuclear calcium<sup>2+</sup> overload and DNA damage response in human prostate cancer. *J Urol* 2011; 185: 1967–74.
- [27] Choi YK, Seo HS, Choi HS, *et al.* Induction of Fas-mediated extrinsic apoptosis, p21WAF1-related G2/M cell cycle arrest and ROS generation by costunolide in estrogen receptor-negative breast cancer cells, MDA-MB-231. *Mol Cell Biochem* 2012; 363: 119–28.
- [28] Li Y, Wang X, Cheng S, *et al.* Diosgenin induces G2/M cell cycle arrest and apoptosis in human hepatocellular carcinoma cells. *Oncol Rep* 2015; 33: 693–8.
- [29] Liu MJ, Wang Z, Ju Y, Wong RNS, Wu QY. Diosgenin induces cell cycle arrest and apoptosis in human leukemia K562 cells with the disruption of Ca<sup>2+</sup> homeostasis. *Cancer Chemother Pharmacol* 2005; 55: 79–90.
- [30] Raju J, Bird RP. Diosgenin, a naturally occurring steroid [corrected] saponin suppresses 3-hydroxy-3-methylglutaryl CoA reductase expression and induces apoptosis in HCT-116 human colon carcinoma cells. *Cancer Lett* 2007; 255: 194–204.
- [31] Liu Y, Bi T, Dai W, *et al.* Lupeol Induces Apoptosis and Cell Cycle Arrest of Human Osteosarcoma Cells Through PI3K/AKT/mTOR Pathway. *Technol Cancer Res Treat* 2015; pii: 1533034615609014.
- [32] Liu Y, Bi T, Wang G, *et al.* Lupeol inhibits proliferation and induces apoptosis of human pancreatic cancer PCNA-1 cells through AKT/ERK pathways. *Naunyn Schmiedebergs Arch Pharmacol* 2015; 388: 295–304.
- [33] Saleem M, Maddodi N, Abu Zaid M, *et al.* Lupeol inhibits growth of highly aggressive human metastatic melanoma cells in vitro and in vivo by inducing apoptosis. *Clin Cancer Res* 2008; 14: 2119–27.
- [34] Girola N, Figueiredo CR, Farias CF, *et al.* Camphene isolated from essential oil of *Piper cernuum* (Piperaceae) induces intrinsic apoptosis in melanoma cells and displays antitumor activity in vivo. *Biochem Biophys Res Commun* 2015; 467: 928–34.
- [35] Mulyaningsih S, Youns M, El-Readi MZ, *et al.* Biological activity of the essential oil of *Kadsura longipedunculata* (Schisandraceae) and its major components. *J Pharm Pharmacol* 2010; 62: 1037–44.
- [36] Lee MG, Lee KT, Chi SG, Park JH. Costunolide induces apoptosis by ROS-mediated mitochondrial permeability transition and cytochrome C release. *Biol Pharm Bull* 2001; 24: 303–6.
- [37] Yang YI, Kim JH, Lee KT, Choi JH. Costunolide induces apoptosis in platinum-resistant human ovarian cancer cells by generating reactive oxygen species. *Gynecol Oncol* 2011; 123: 588–96.
- [38] Roy A, Manikkam R. Cytotoxic Impact of Costunolide Isolated from *Costus speciosus* on Breast Cancer via Differential Regulation of Cell Cycle-An In-vitro and In-silico Approach. *Phytother Res* 2015; 29: 1532–9.
- [39] Liu MJ, Wang Z, Ju Y, Zhou J, Wang Y, Wong RNS. The mitotic-arresting and apoptosis-inducing effects of diosgenyl saponins on human leukemia cell lines. *Biol Pharm Bull* 2004; 27: 1059–65.
- [40] He Y, Wang JS, Zhang P, Zhang WJ, Huang QL, Hua ZC. [Synergistic apoptotic effect of the combination of diosgenin and TRAIL on non-small-cell lung cancer cell line A549 evaluated with the Chou-Talalay method]. *Yao Xue Xue Bao* 2013; 48: 45–51.
- [41] Cailleteau C, Liagre B, Beneytout JL. A proteomic approach to the identification of molecular targets in subsequent apoptosis of HEL

- cells after diosgenin-induced megakaryocytic differentiation. *J Cell Biochem* 2009; 107: 785–96.
- [42] Selim S, Al Jaouni S. Anticancer and apoptotic effects on cell proliferation of diosgenin isolated from *Costus speciosus* (Koen.) Sm *BMC Complement Altern Med* 2015; 15: 301.
- [43] Das S, Dey KK, Dey G, *et al.* Antineoplastic and apoptotic potential of traditional medicines thymoquinone and diosgenin in squamous cell carcinoma. *PLoS One* 2012; 7: e46641.
- [44] Leger DY, Liagre B, Beneytout JL. Role of MAPKs and NF-kappaB in diosgenin-induced megakaryocytic differentiation and subsequent apoptosis in HEL cells. *Int J Oncol* 2006; 28: 201–7.
- [45] Raju J, Patlolla JMR, Swamy MV, Rao CV. Diosgenin, a steroid saponin of *Trigonella foenum graecum* (Fenugreek), inhibits azoxymethane-induced aberrant crypt foci formation in F344 rats and induces apoptosis in HT-29 human colon cancer cells. *Cancer Epidemiol Biomarkers Prev* 2004; 13: 1392–8.
- [46] Prasad S, Kalra N, Shukla Y. Hepatoprotective effects of lupeol and mango pulp extract of carcinogen induced alteration in Swiss albino mice. *Mol Nutr Food Res* 2007; 51: 352–9.
- [47] He Y, Liu F, Zhang L, *et al.* Growth inhibition and apoptosis induced by lupeol, a dietary triterpene, in human hepatocellular carcinoma cells. *Biol Pharm Bull* 2011; 34: 517–22.
- [48] Xian M, Ito K, Nakazato T, *et al.* Zerumbone, a bioactive sesquiterpene, induces G2/M cell cycle arrest and apoptosis in leukemia cells via a Fas- and mitochondria-mediated pathway. *Cancer Sci* 2007; 98: 118–26.
- [49] Rajan I, Jayasree PR, Kumar PRM. Zerumbone induces mitochondria-mediated apoptosis via increased calcium, generation of reactive oxygen species and upregulation of soluble histone H2AX in K562 chronic myelogenous leukemia cells. *Tumor Biol* 2015; 36: 8479–89.
- [50] Rahman HS, Rasedee A, Abdul AB, *et al.* Zerumbone-loaded nanostructured lipid carrier induces G2/M cell cycle arrest and apoptosis via mitochondrial pathway in a human lymphoblastic leukemia cell line. *Int J Nanomedicine* 2014; 9: 527–38.
- [51] Sun Y, Sheng Q, Cheng Y, *et al.* Zerumbone induces apoptosis in human renal cell carcinoma via Gli-1/Bcl-2 pathway. *Pharmazie* 2013; 68: 141–5.
- [52] Weng HY, Hsu MJ, Wang CC, *et al.* Zerumbone suppresses IKK $\alpha$ , Akt, and FOXO1 activation, resulting in apoptosis of GBM 8401 cells. *J Biomed Sci* 2012; 19: 86.
- [53] Abdel Wahab SI, Abdul AB, Alzubairi AS, Mohamed Elhassan M, Mohan S. In vitro ultramorphological assessment of apoptosis induced by Zerumbone on (HeLa). *J Biomed Biotechnol* 2009; 2009: 769568.
- [54] Tabata K, Nishimura Y, Takeda T, Kurita M, Uchiyama T, Suzuki T. Sesquiterpene lactones derived from *Saussurea lappa* induce apoptosis and inhibit invasion and migration in neuroblastoma cells. *J Pharmacol Sci* 2015; 127: 397–403.
- [55] Murtaza I, Saleem M, Adhami VM, Hafeez BB, Mukhtar H. Suppression of cFLIP by lupeol, a dietary triterpene, is sufficient to overcome resistance to TRAIL-mediated apoptosis in chemoresistant human pancreatic cancer cells. *Cancer Res* 2009; 69: 1156–65.
- [56] Siveen KS, Nguyen AH, Lee JH, *et al.* Negative regulation of signal transducer and activator of transcription-3 signalling cascade by lupeol inhibits growth and induces apoptosis in hepatocellular carcinoma cells. *Br J Cancer* 2014; 111: 1327–37.
- [57] Léger DY, Liagre B, Cardot PJP, Beneytout JL, Battu S. Diosgenin dose-dependent apoptosis and differentiation induction in human erythroleukemia cell line and sedimentation field-flow fractionation monitoring. *Anal Biochem* 2004; 335: 267–78.
- [58] Prasad S, Madan E, Nigam N, Roy P, George J, Shukla Y. Induction of apoptosis by lupeol in human epidermoid carcinoma A431 cells through regulation of mitochondrial, Akt/PKB and NFkappaB signaling pathways. *Cancer Biol Ther* 2009; 8: 1632–9.
- [59] Sakinah SAS, Handayani ST, Hawariah LPA. Zerumbone induced apoptosis in liver cancer cells via modulation of Bax/Bcl-2 ratio. *Cancer Cell Int* 2007; 7: 4.
- [60] Chan ML, Liang JW, Hsu LC, Chang WL, Lee SS, Guh JH. Zerumbone, a ginger sesquiterpene, induces apoptosis and autophagy in human hormone-refractory prostate cancers through tubulin binding and crosstalk between endoplasmic reticulum stress and mitochondrial insult. *Naunyn Schmiedebergs Arch Pharmacol* 2015; 388: 1223–36.
- [61] Lepage C, Liagre B, Cook-Moreau J, Pinon A, Beneytout JL. Cyclooxygenase-2 and 5-lipoxygenase pathways in diosgenin-induced apoptosis in HT-29 and HCT-116 colon cancer cells. *Int J Oncol* 2010; 36: 1183–91.
- [62] Lepage C, Léger DY, Bertrand J, Martin F, Beneytout JL, Liagre B. Diosgenin induces death receptor-5 through activation of p38 pathway and promotes TRAIL-induced apoptosis in colon cancer cells. *Cancer Lett* 2011; 301: 193–202.
- [63] Prasad S, Kalra N, Shukla Y. Induction of apoptosis by lupeol and mango extract in mouse prostate and LNCaP cells. *Nutr Cancer* 2008; 60: 120–30.
- [64] Yodkeeree S, Sung B, Limtrakul P, Aggarwal BB. Zerumbone enhances TRAIL-induced apoptosis through the induction of death receptors in human colon cancer cells: Evidence for an essential role of reactive oxygen species. *Cancer Res* 2009; 69: 6581–9.
- [65] Ambrož M, Boušová I, Skarka A, *et al.* The Influence of Sesquiterpenes from *Myrica rubra* on the Antiproliferative and Pro-Oxidative Effects of Doxorubicin and Its Accumulation in Cancer Cells. *Molecules* 2015; 20: 15343–58.
- [66] Lin KW, Huang AM, Tu HY, *et al.* Xanthine oxidase inhibitory triterpenoid and phloroglucinol from guttiferaceous plants inhibit growth and induced apoptosis in human NTUB1 cells through a ROS-dependent mechanism. *J Agric Food Chem* 2011; 59: 407–14.
- [67] Edagawa M, Kawauchi J, Hirata M, *et al.* Role of activating transcription factor 3 (ATF3) in endoplasmic reticulum (ER) stress-induced sensitization of p53-deficient human colon cancer cells to tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL)-mediated apoptosis through up-re. *J Biol Chem* 2014; 289: 21544–61.
- [68] Mao ZJ, Tang QJ, Zhang CA, *et al.* Anti-proliferation and anti-invasion effects of diosgenin on gastric cancer BGC-823 cells with HIF-1 $\alpha$  shRNAs. *Int J Mol Sci* 2012; 13: 6521–33.
- [69] Zhang L, Zhang Y, Zhang L, Yang X, Lv Z. Lupeol, a dietary triterpene, inhibited growth, and induced apoptosis through down-regulation of DR3 in SMMC7721 cells. *Cancer Invest* 2009; 27: 163–70.
- [70] Sehrawat A, Sakao K, Singh SV. Notch2 activation is protective against anticancer effects of zerumbone in human breast cancer cells. *Breast Cancer Res Treat* 2014; 146: 543–55.
- [71] Lee TKW, Castilho A, Cheung VCH, Tang KH, Ma S, Ng IOL. Lupeol targets liver tumor-initiating cells through phosphatase and tensin homolog modulation. *Hepatology* 2011; 53: 160–70.
- [72] Ogiwara K, Hata K. Melanoma cell differentiation induced by lupeol separates into two stages: morphological and functional changes. *J Nat Med* 2009; 63: 323–6.
- [73] West KA, Castillo SS, Dennis PA. Activation of the PI3K/Akt pathway and chemotherapeutic resistance. *Drug Resist Updat* 2002; 5: 234–48.
- [74] Pisu M, Concas A, Cao G. A novel quantitative model of cell cycle progression based on cyclin-dependent kinases activity and population balances. *Comput Biol Chem* 2015; 55: 1–13.
- [75] Simon HU, Friis R. ATG5: a distinct role in the nucleus. *Autophagy* 2014; 10: 176–7.
- [76] Youle RJ, Strasser A. The BCL-2 protein family: opposing activities that mediate cell death. *Nat Rev Mol Cell Biol* 2008; 9: 47–59.
- [77] Chao DT, Korsmeyer SJ. BCL-2 family: regulators of cell death. *Annu Rev Immunol* 1998; 16: 395–419.
- [78] Cao JP, Niu HY, Wang HJ, Huang XG, Gao DS. NF- $\kappa$ B p65/p52 plays a role in GDNF up-regulating Bcl-2 and Bcl-w expression in 6-OHDA-induced apoptosis of MN9D cell. *Int J Neurosci* 2013; 123: 705–10.
- [79] Perkins ND. Integrating cell-signalling pathways with NF-kappaB and IKK function. *Nat Rev Mol Cell Biol* 2007; 8: 49–62.
- [80] Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. *Oncogene* 1999; 18: 6853–66.
- [81] Park M, Hong J. Roles of NF- $\kappa$ B in Cancer and Inflammatory Diseases and Their Therapeutic Approaches. *Cells* 2016; 5: 15.
- [82] Krishnakumar R, Kraus WL. The PARP side of the nucleus: molecular actions, physiological outcomes, and clinical targets. *Mol Cell* 2010; 39: 8–24.
- [83] O'Shea JJ, Gadina M, Schreiber RD. Cytokine signaling in 2002: new surprises in the Jak/Stat pathway. *Cell* 2002; 109 Suppl: S121–31.
- [84] Sansone P, Piazza G, Paterini P, *et al.* Cyclooxygenase-2/carbonic anhydrase-IX up-regulation promotes invasive potential and

- hypoxia survival in colorectal cancer cells. *J Cell Mol Med* 2009; 13: 3876–87.
- [85] Nagata Y, Takahashi N, Davis RJ, Todokoro K. Activation of p38 MAP kinase and JNK but not ERK is required for erythropoietin-induced erythroid differentiation. *Blood* 1998; 92: 1859–69.
- [86] Liu F, He Y, Liang Y, *et al.* PI3-kinase inhibition synergistically promoted the anti-tumor effect of lupeol in hepatocellular carcinoma. *Cancer Cell Int* 2013; 13: 108.
- [87] Chen PS, Shih YW, Huang HC, Cheng HW. Diosgenin, a steroidal saponin, inhibits migration and invasion of human prostate cancer PC-3 cells by reducing matrix metalloproteinases expression. *PLoS One* 2011; 6: e20164.
- [88] Léger DY, Battu S, Liagre B, Beneytout JL, Cardot PJP. Megakaryocyte cell sorting from diosgenin-differentiated human erythroleukemia cells by sedimentation field-flow fractionation. *Anal Biochem* 2006; 355: 19–28.
- [89] Lee J, Jung K, Kim YS, Park D. Diosgenin inhibits melanogenesis through the activation of phosphatidylinositol-3-kinase pathway (PI3K) signaling. *Life Sci* 2007; 81: 249–54.
- [90] Chang HY, Kao MC, Way TD, Ho CT, Fu E. Diosgenin suppresses hepatocyte growth factor (HGF)-induced epithelial-mesenchymal transition by down-regulation of Mdm2 and vimentin. *J Agric Food Chem* 2011; 59: 5357–63.
- [91] Chiang CT, Way TD, Tsai SJ, Lin JK. Diosgenin, a naturally occurring steroid, suppresses fatty acid synthase expression in HER2-overexpressing breast cancer cells through modulating Akt, mTOR and JNK phosphorylation. *FEBS Lett* 2007; 581: 5735–42.
- [92] Srinivasan S, Koduru S, Kumar R, Venguswamy G, Kyprianou N, Damodaran C. Diosgenin targets Akt-mediated prosurvival signaling in human breast cancer cells. *Int J cancer* 2009; 125: 961–7.
- [93] Sobhan PK, Seervi M, Deb L, *et al.* Calpain and Reactive Oxygen Species Targets Bax for Mitochondrial Permeabilisation and Caspase Activation in Zerumbone Induced Apoptosis. *PLoS One* 2013; 8: e59350.
- [94] Kang CG, Lee HJ, Kim SH, Lee EO. Zerumbone Suppresses Osteopontin-Induced Cell Invasion Through Inhibiting the FAK/AKT/ROCK Pathway in Human Non-Small Cell Lung Cancer A549 Cells. *J Nat Prod* 2016; 79: 156–60.
- [95] Liu Y, Bi T, Shen G, *et al.* Lupeol induces apoptosis and inhibits invasion in gallbladder carcinoma GBC-SD cells by suppression of EGFR/MMP-9 signaling pathway. *Cytotechnology* 2016; 68: 123–133.
- [96] Prasad S, Nigam N, Kalra N, Shukla Y. Regulation of signaling pathways involved in lupeol induced inhibition of proliferation and induction of apoptosis in human prostate cancer cells. *Mol Carcinog* 2008; 47: 916–24.
- [97] Sehrawat A, Arlotti JA, Murakami A, Singh SV. Zerumbone causes Bax- and Bak-mediated apoptosis in human breast cancer cells and inhibits orthotopic xenograft growth in vivo. *Breast Cancer Res Treat* 2012; 136: 429–41.
- [98] He Z, Chen H, Li G, *et al.* Diosgenin inhibits the migration of human breast cancer MDA-MB-231 cells by suppressing Vav2 activity. *Phytomedicine* 2014; 21: 871–6.
- [99] Saleem M, Murtaza I, Witkowsky O, Kohl AM, Maddodi N. Lupeol triterpene, a novel diet-based microtubule targeting agent: disrupts survivin/cFLIP activation in prostate cancer cells. *Biochem Biophys Res Commun* 2009; 388: 576–82.
- [100] Nigam N, Prasad S, George J, Shukla Y. Lupeol induces p53 and cyclin-B-mediated G2/M arrest and targets apoptosis through activation of caspase in mouse skin. *Biochem Biophys Res Commun* 2009; 381: 253–8.
- [101] Jorvig JE, Chakraborty A. Zerumbone inhibits growth of hormone refractory prostate cancer cells by inhibiting JAK2/STAT3 pathway and increases paclitaxel sensitivity. *Anticancer Drugs* 2015; 26: 160–6.
- [102] Kirana C, McIntosh GH, Record IR, Jones GP. Antitumor activity of extract of *Zingiber aromaticum* and its bioactive sesquiterpenoid zerumbone. *Nutr Cancer* 2003; 45: 218–25.
- [103] Cailleteau C, Micallef L, Lepage C, *et al.* Investigating the relationship between cell cycle stage and diosgenin-induced megakaryocytic differentiation of HEL cells using sedimentation field-flow fractionation. *Anal Bioanal Chem* 2010; 398: 1273–83.
- [104] Abdelwahab SI, Abdul AB, Zain ZNM, Hadi AHA. Zerumbone inhibits interleukin-6 and induces apoptosis and cell cycle arrest in ovarian and cervical cancer cells. *Int Immunopharmacol* 2012; 12: 594–602.
- [105] Deorukhkar A, Ahuja N, Mercado AL, *et al.* Zerumbone increases oxidative stress in a thiol-dependent ROS-independent manner to increase DNA damage and sensitize colorectal cancer cells to radiation. *Cancer Med* 2015; 4: 278–92.
- [106] Pitchai D, Roy A, Ignatius C. In vitro evaluation of anticancer potentials of lupeol isolated from *Elephantopus scaber* L. on MCF-7 cell line. *J Adv Pharm Technol Res* 2014; 5: 179–84.
- [107] Pitchai D, Roy A, Banu S. In vitro and in silico evaluation of NF- $\kappa$ B targeted costunolide action on estrogen receptor-negative breast cancer cells—a comparison with normal breast cells. *Phytother Res* 2014; 28: 1499–505.
- [108] Saleem M, Kweon MH, Yun JM, *et al.* A novel dietary triterpene Lupeol induces fas-mediated apoptotic death of androgen-sensitive prostate cancer cells and inhibits tumor growth in a xenograft model. *Cancer Res* 2005; 65: 11203–13.
- [109] Shamoto T, Matsuo Y, Shibata T, *et al.* Zerumbone inhibits angiogenesis by blocking NF- $\kappa$ B activity in pancreatic cancer. *Pancreas* 2014; 43: 396–404.
- [110] Sung B, Jhurani S, Ahn KS, *et al.* Zerumbone down-regulates chemokine receptor CXCR4 expression leading to inhibition of CXCL12-induced invasion of breast and pancreatic tumor cells. *Cancer Res* 2008; 68: 8938–44.
- [111] Sung B, Murakami A, Oyajobi BO, Aggarwal BB. Zerumbone abolishes RANKL-induced NF- $\kappa$ B activation, inhibits osteoclastogenesis, and suppresses human breast cancer-induced bone loss in athymic nude mice. *Cancer Res* 2009; 69: 1477–84.
- [112] Tsuboi K, Matsuo Y, Shamoto T, *et al.* Zerumbone inhibits tumor angiogenesis via NF- $\kappa$ B in gastric cancer. *Oncol Rep* 2014; 31: 57–64.
- [113] Li F, Fernandez PP, Rajendran P, Hui KM, Sethi G. Diosgenin, a steroidal saponin, inhibits STAT3 signaling pathway leading to suppression of proliferation and chemosensitization of human hepatocellular carcinoma cells. *Cancer Lett* 2010; 292: 197–207.
- [114] Kim S, Kil WH, Lee J, *et al.* Zerumbone suppresses EGF-induced CD44 expression through the inhibition of STAT3 in breast cancer cells. *Oncol Rep* 2014; 32: 2666–72.
- [115] Saleem M, Murtaza I, Tarapore RS, *et al.* Lupeol inhibits proliferation of human prostate cancer cells by targeting beta-catenin signaling. *Carcinogenesis* 2009; 30: 808–17.
- [116] Han J, Bae SY, Oh SJ, *et al.* Zerumbone suppresses IL-1 $\beta$ -induced cell migration and invasion by inhibiting IL-8 and MMP-3 expression in human triple-negative breast cancer cells. *Phytother Res* 2014; 28: 1654–60.
- [117] Han J, Bae SY, Oh SJ, *et al.* Zerumbone Suppresses IL-1 $\beta$ -induced Cell Migration and Invasion by Inhibiting IL-8 and MMP-3 Expression in Human Triple-negative Breast Cancer Cells. *Phyther Res* 2014; 28: 1654–60.
- [118] Ghezali L, Liagre B, Limami Y, Beneytout JL, Leger DY. Sonic Hedgehog activation is implicated in diosgenin-induced megakaryocytic differentiation of human erythroleukemia cells. *PLoS One* 2014; 9: e95016.
- [119] Mohammad RY, Somayyeh G, Gholamreza H, Majid M, Yousef R. Diosgenin inhibits hTERT gene expression in the A549 lung cancer cell line. *Asian Pac J Cancer Prev* 2013; 14: 6945–8.
- [120] Rahmati-Yamchi M, Ghareghomi S, Haddadchi G, Milani M, Aghazadeh M, Daroushnejad H. Fenugreek extract diosgenin and pure diosgenin inhibit the hTERT gene expression in A549 lung cancer cell line. *Mol Biol Rep* 2014; 41: 6247–52.
- [121] Murakami A, Miyamoto M, Ohigashi H. Zerumbone, an anti-inflammatory phytochemical, induces expression of proinflammatory cytokine genes in human colon adenocarcinoma cell lines. *Biofactors* 2004; 21: 95–101.
- [122] Choi YK, Cho SG, Woo SM, *et al.* Saussurea lappa Clarke-Derived Costunolide Prevents TNF  $\alpha$ -Induced Breast Cancer Cell Migration and Invasion by Inhibiting NF- $\kappa$ B Activity. *Evid Based Complement Alternat Med* 2013; 2013: 936257.
- [123] Tarapore RS, Siddiqui IA, Saleem M, Adhmi VM, Spiegelman VS, Mukhtar H. Specific targeting of Wnt/ $\beta$ -catenin signaling in human melanoma cells by a dietary triterpene lupeol. *Carcinogenesis* 2010; 31: 1844–53.
- [124] Manoharan S, Palanimuthu D, Baskaran N, Silvan S. Modulating effect of lupeol on the expression pattern of apoptotic markers in 7,

- 12-dimethylbenz(a)anthracene induced oral carcinogenesis. Asian Pac J Cancer Prev 2012; 13: 5753–7.
- [125] Taha MME, Abdul AB, Abdullah R, Ibrahim TAT, Abdelwahab SI, Mohan S. Potential chemoprevention of diethylnitrosamine-initiated and 2-acetylaminofluorene-promoted hepatocarcinogenesis by zerumbone from the rhizomes of the subtropical ginger (*Zingiber zerumbet*). Chem Biol Interact 2010; 186: 295–305.
- [126] Jagadeesan J, Nandakumar N, Rengarajan T, Balasubramanian MP. Diosgenin, a steroidal saponin, exhibits anticancer activity by attenuating lipid peroxidation via enhancing antioxidant defense system during NMU-induced breast carcinoma. J Environ Pathol Toxicol Oncol 2012; 31: 121–9.
- [127] Miyoshi N, Nagasawa T, Mabuchi R, *et al.* Chemoprevention of azoxymethane/dextran sodium sulfate-induced mouse colon carcinogenesis by freeze-dried yam sanyaku and its constituent diosgenin. Cancer Prev Res (Phila) 2011; 4: 924–34.
- [128] Rajalingam K, Sugunadevi G, Arokia Vijayaanand M, Kalaimathi J, Suresh K. Anti-tumor and anti-oxidative potential of diosgenin against 7, 12-dimethylbenz(a)anthracene induced experimental oral carcinogenesis. Pathol Oncol Res 2012; 18: 405–12.
- [129] Prabhu B, Balakrishnan D, Sundaresan S. Antiproliferative and anti-inflammatory properties of diindolylmethane and lupeol against N-butyl-N-(4-hydroxybutyl) nitrosamine induced bladder carcinogenesis in experimental rats. Hum Exp Toxicol 2016; 35: 685–92.
- [130] Saleem M, Afaq F, Adhami VM, Mukhtar H. Lupeol modulates NF-kappaB and PI3K/Akt pathways and inhibits skin cancer in CD-1 mice. Oncogene 2004; 23: 5203–14.
- [131] Kim M, Miyamoto S, Yasui Y, Oyama T, Murakami A, Tanaka T. Zerumbone, a tropical ginger sesquiterpene, inhibits colon and lung carcinogenesis in mice. Int J cancer 2009; 124: 264–71.