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## Insights from Mendelian Interferonopathies: Comparison of CANDLE, SAVI with AGS, Monogenic Lupus

Hanna Kim<sup>1</sup>, Gina A. Montealegre Sanchez<sup>2</sup>, and Raphaela Goldbach-Mansky<sup>2</sup>

Hanna Kim: hanna.kim@nih.gov; Gina A. Montealegre Sanchez: gina.montealegre@nih.gov; Raphaela Goldbach-Mansky: goldbacr@mail.nih.gov

<sup>1</sup>Lawrence Shulman Scholar in Translational Research, Pediatric Translational Research Branch, National Institute of Arthritis Musculoskeletal and Skin Diseases (NIAMS), National Institutes of Health (NIH), Bethesda, MD, USA

<sup>2</sup>Translational Autoinflammatory Disease Studies, National Institute of Allergy and Infectious Diseases (NIAID), and NIAMS, National Institutes of Health (NIH), Bethesda, MD, USA

### Abstract

Autoinflammatory disorders are sterile inflammatory conditions characterized by episodes of early-onset fever and disease-specific patterns of organ inflammation. Recently, the discoveries of monogenic disorders with strong type I interferon (IFN) signatures caused by mutations in proteasome degradation and cytoplasmic RNA and DNA sensing pathways suggest a pathogenic role of IFNs in causing autoinflammatory phenotypes. The IFN response gene signature (IGS) has been associated with systemic lupus erythematosus (SLE) and other autoimmune diseases. In this review, we compare the clinical presentations and pathogenesis of two IFN-mediated autoinflammatory diseases, CANDLE and SAVI, with Aicardi Goutières syndrome (AGS) and monogenic forms of SLE (monoSLE) caused by loss-of-function mutations in complement 1 (C1q) or the DNA nucleases, *DNASE1* and *DNASE1L3*. We outline differences in intracellular signaling pathways that fuel a pathologic type I IFN amplification cycle. While IFN amplification is caused by predominantly innate immune cell dysfunction in SAVI, CANDLE, and AGS, autoantibodies to modified RNA and DNA antigens interact with tissues and immune cells including neutrophils and contribute to IFN upregulation in some SLE patients including monoSLE, thus justifying a grouping of “autoinflammatory” and “autoimmune” interferonopathies. Understanding of the differences in the cellular sources and signaling pathways will guide new drug development and the use of emerging targeted therapies.

### Keywords

Type I IFN; Autoinflammatory; Autoimmune; Candle; SAVI; Interferonopathies

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Correspondence to: Hanna Kim, hanna.kim@nih.gov; Raphaela Goldbach-Mansky, goldbacr@mail.nih.gov.

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## Introduction

Systemic autoinflammatory diseases were introduced as a concept by Dan Kastner in 1999 to distinguish periodic fever syndromes that lack autoantibodies or antigen-specific T cells from autoimmune diseases such as systemic lupus erythematosus (SLE) and rheumatoid arthritis (RA) [1]. The characterization of patients with “autoinflammatory phenotypes” led to identification of pathogenic genetic defects in innate immune sensors and pattern recognition receptors (PRRs) pathways, pioneered by mutations in the IL-1 activating inflammasome genes *MEFV* encoding pyrin (familial Mediterranean fever), *NLRP3* (cryopyrinopathies) and, recently *NLR4* (NLRC4-associated macrophage activation syndrome), linking increased cytokine and chemokine production and recruitment of innate immune cells to the development of tissue-specific sterile organ inflammation and damage [2]. A novel group of monogenic disorders presenting with early-onset systemic and organ-specific inflammation and a prominent IFN response gene signature (IGS) including Chronic Atypical Neutrophilic Dermatitis with Lipodystrophy and Elevated temperatures (CANDLE) [3] and STING-Associated Vasculopathy with onset in Infancy (SAVI) [4] and Aicardi Goutières syndrome (AGS) [5] suggested a key role for type I IFN in disease pathogenesis. In 2003, the IGS was described in SLE [6] followed by other autoimmune diseases, such as juvenile dermatomyositis (JDM) [7]. In 2011, Yanick Crow suggested the term “interferonopathies” to indicate that IFN drives shared immunopathology [8]. The discovery of genetic causes of interferonopathies has propelled research and provided insights into innate (autoinflammatory) and autoantibody or antigen-specific T cell-mediated (autoimmune) mediated pathology in diseases with IGS. This review compares the clinical disease manifestations and pathogenesis of CANDLE and SAVI with AGS and monogenic forms of SLE caused by mutations in the complement system (*C1Q*) [9] and DNA nucleases (*DNASE1* and *DNASE1L3*) [10, 11]. Based on differences in immunopathogenesis, we suggest grouping these conditions as “autoinflammatory” and “autoimmune” interferonopathies.

## Clinical and pathogenic description of the Mendelian (monogenic) autoinflammatory interferonopathies CANDLE and SAVI

### CANDLE/PRAAS (MIM#256040)

In 2010 and 2011, several studies found that autosomal recessive (AR) loss-of-function (LOF) mutations in the proteasome subunit beta type 8 (*PSMB8*) cause conditions referred to as JMP (joint contractures, muscular atrophy, microcytic anemia, and panniculitis-induced lipodystrophy) [12], Nakajo-Nishimura syndrome (NNS) with nodular erythema, elongated and thickened fingers and emaciation [13], Japanese autoinflammatory syndrome with lipodystrophy (JASL) [14], and CANDLE [3], demonstrating a phenotypic disease spectrum with variable disease severity [3, 15], also referred to as proteasome-associated autoinflammatory syndromes (PRAAS). In addition to mutations in *PSMB8* [3, 12–14, 16] in CANDLE/PRAAS, we described loss-of-function mutations in other proteasome subunits (*PSMB9*, *PSMB4*, *PSMA3*) and the proteasome assembly protein, *POMP*, with expansion of the inheritance to compound heterozygous (mutations in *PSMB4*), digenic recessive

(combinations of *PSMA3*, *PSMB4* and/or *PSMB9*), and autosomal dominant (AD) (*POMP*) [17]. CANDLE is a rare disease with less than 100 cases described worldwide.

**Clinical manifestations**—Clinical manifestations of PRAAS/CANDLE are summarized in Tables 1 and 2. Long-standing inflammation causes organ damage (progressive lipodystrophy, joint contractures, and basal ganglia calcifications in the CNS). While inflammatory features are more prominent early in life, damage features manifest later in life. Clinical manifestations include the following:

**Systemic inflammation:** This includes fevers and elevated acute phase reactants [3, 13–21].

**Skin manifestations:** Nodular or plaque-like violaceous skin rashes and violaceous periorbital rash with or without swelling are thought to be manifestations of panniculitis that evolve to lipodystrophy [3, 13–22]. Skin biopsies show immature neutrophils (Leder stain positive) and myeloid precursors (myeloperoxidase positive) as well as atypical mononuclear cells, which are likely activated macrophages (positive for CD68 and CD163, negative for Leder stain) [3].

**Musculoskeletal manifestations:** Musculoskeletal findings include myositis and arthritis [3, 13–22]. Joint contractures are common and develop even early in childhood [3, 13–15, 17, 18, 21, 22].

**Metabolic manifestations:** Prominent metabolic features include truncal obesity, dyslipidemia, insulin resistance, and acanthosis nigricans consistent with metabolic syndrome [3, 13, 15–18, 20–22], which are exaggerated by steroids [3].

**Cardiovascular manifestations:** Recently, primary pulmonary hypertension has been observed in 2 young patients with CANDLE/PRAAS [23].

**Mortality:** Early mortality due to sudden death in the context of infections, cardiomyopathy, and cardiac arrhythmias has been reported [3, 15, 18]. See Table 1 for details.

**Treatment:** Methotrexate, azathioprine, cyclosporine, tacrolimus, rituximab, IL-1 blockade, and tumor necrosis factor antagonist (infliximab, etanercept, adalimumab) are ineffective; partial response to anti IL-6 therapy and/or high doses of steroids is seen [3, 13–15, 18]. Janus kinase or JAK inhibitors are being used with promising results (see below).

**Pathogenesis**—The proteasome is a tubular protein degradation system for misfolded or damaged intracellular proteins, particularly those marked for degradation by polyubiquitination, which are proteolytically cleaved by the caspase, chymotrypsin, and trypsin-like activities [24]. The proteasome mutations decrease the proteolytic activities [12–14] and might impair adipocyte differentiation based on *PSMB8* knockdown experiments [14]. Clues to pathogenesis came from observations of high serum levels of IFN  $\gamma$  inducible protein 10 (IP-10) or CXCL10 and a prominent IGS [3, 12–14]. Knockdown experiments in healthy fibroblasts identified a critical threshold of proteasome dysfunction that leads to induction of type I IFN gene transcription. Proteasome dysfunction induced in wild-type

cells by proteasome inhibitors results in dose-dependent increases in *IFNA* and *IFNB* transcription [17]. Although the exact mechanism leading to IFN transcription in CANDLE/PRAAS remains unknown, the type I IFN induction is independent of STING (stimulator of IFN genes protein) and MAVS (mitochondrial antiviral signaling protein), adaptors of RNA and DNA viral sensors, respectively [17]. Patient cells are hyper-responsive to IFN  $\gamma$  [3] and IFN $\alpha$  (unpublished) stimulation and respond with increased STAT-1 phosphorylation; environmental stressors such as infections can trigger clinical flares [25].

### **SAVI (MIM #615934)**

We recently described dominant gain-of-function mutations in *TMEM173*, encoding stimulator of interferon genes (STING), as the cause for the autoinflammatory syndrome STING-associated vasculopathy with onset in infancy (SAVI) with disease-causing mutations thus far limited to exon 5 in *TMEM173* (residues V147, N154, V155) [4, 26–29], with clinical features summarized in Tables 1 and 2. A report of a family with 4 mutation positive (V155 M) members in 3 generations with variable clinical phenotype and onset (ranging from onset at 1 yo with febrile attack and rash with lung disease and failure to thrive in one individual to adult-onset arthralgia and systemic inflammation), all with overexpression of interferon-regulated genes [26], suggests modifiers may influence the severity of SAVI.

**Clinical manifestations**—Clinical manifestations of SAVI are summarized in Tables 1 and 2.

**Systemic manifestations:** Rash with systemic inflammation (fever, elevated acute phase reactants) is typically seen in the first months of life [4, 26–29].

**Skin manifestations:** Vasculopathic lesions are most prominent in cold-sensitive acral areas and present as violaceous plaques and/or nodules on the face, nose, and ears, and distal ulcerations. Features of vascular and/or tissue damage include nail dystrophy, gangrene/infarcts of fingers/toes with tissue loss, and nasal septal perforation [4, 26–28]. In lesional skin biopsy endothelial cells, there is upregulation of endothelial inflammation (inducible nitric oxide synthase), coagulation (tissue factor), and endothelial cell adhesion and activation (E-selectin). Although immune complexes (ICs) are seen in partially destroyed vessels, many vessels did not have immune complex (IC) deposition and affected cutaneous small-vessels were surrounded by neutrophils and leukocytoclasia [4].

**Lung manifestations:** Paratracheal adenopathy, abnormal pulmonary function tests, and variable interstitial lung disease on CT are seen with or without symptoms; the extent of lung fibrosis varies between patients [4, 26–29]. No vasculitis is observed in the lung of SAVI patients. STING is expressed in alveolar macrophages, type 2 pneumocytes, and bronchial epithelium. SAVI patients present with increased alveolar macrophages, activation of type 2 pneumocytes, follicular hyperplasia, and prominent fibrosis [4].

**Musculoskeletal manifestations:** Myositis, arthritis, and arthralgia are variable [4, 26, 29].

**Mortality:** Early mortality due to respiratory failure likely related to underlying SAVI lung disease often in the context of infections is seen [4, 26, 27]. See Table 1 for details.

**Treatment:** Immunosuppressive agents including high-dose steroids, azathioprine, colchicine, methotrexate, mycophenolate mofetil, cyclophosphamide, tumor necrosis factor antagonist, IL-1 blockade, IL-6 blockade, and anti-CD 20 therapy have not resulted in sustained responses [4, 26–29].

**Pathogenesis—**STING is a key dimeric adaptor protein at the endoplasmic reticulum that is essential for interferon beta induction. Viral or self double-stranded DNAs (dsDNAs) are sensed in the cytosol by cyclic GMP-AMP synthase or cGAS ligand [30, 31]. Upon binding cGAS, cyclic guanosine monophosphate-adenosine monophosphate (cGAMP) is released as a second messenger, which binds STING, leading to phosphorylation of TANK-binding kinase 1 (TBK1) and interferon regulatory factor 3 (IRF-3); IRF3 then translocates into the nucleus leading to *IFNB1* (interferon  $\beta$ ) transcription. The disease-causing STING mutations constitutively activate the pathway resulting in *IFNB1* transcription, prominent IGS, elevated serum levels of interferon-inducible protein 10 (IP-10) and constitutively elevated STAT-1 phosphorylation in patient Tand B lymphocytes [4]. Activation of the STING pathways with cGAMP in endothelial cells in vitro leads to transcription of activation markers and clotting factors as well as cell death, suggesting a role for STING in tissue homeostasis [4].

**Treatment considerations for CANDLE and SAVI—**Inhibition with Janus kinase (JAK) inhibitor tofacitinib decreased the STAT-1 phosphorylation and IP-10 in mononuclear cells (PBMCs) from CANDLE patients stimulated with IFN $\gamma$  [3]. In vitro experiments in PBMCs from SAVI patients showed that constitutive phosphorylation of STAT1 in CD4 T and CD19 B cells was reduced in cells co-cultured with JAK inhibitors (tofacitinib, ruxolitinib, and baricitinib). This phenomenon was also observed in fibroblasts. cGAMP stimulated interferon-regulated genes (*CXCL10*, *MX1*, and *OAS3*) were suppressed when cells were co-cultured with tofacitinib [4]. These observations and the absence of other suitable treatment options led to the use of JAK inhibition as therapy in CANDLE and SAVI (NCT01724580). Preliminary data show clinical improvement with significant decrease in symptom scores and daily steroid requirement in CANDLE patients [32].

## Comparison of Aicardi-Goutières syndrome (AGS) with CANDLE and SAVI

Mutations in 7 genes cause the AGS phenotype: AGS1 (*TREX1*) (MIM#225750), AGS2 (*RNASEH2B*) (MIM#610181), AGS3 (*RNASEH2C*) (MIM#610329), AGS4 (*RNASEH2A*) (MIM#610333), AGS5 (*SAMHD1*) (MIM#612952), AGS6 (*ADAR1*) (MIM#615010), AGS7 (*IFIH1*) (MIM#615846).

### Clinical presentation in AGS compared to CANDLE and SAVI

AGS is characterized by an early-onset, often monophasic, congenital infection-like syndrome resulting in encephalopathy with microcephaly, dystonia, basal ganglia calcifications, white matter abnormalities, and cerebral atrophy [33]. Cerebrospinal fluid analysis shows chronic lymphocytosis and elevated IFN $\alpha$  levels [34]. Expression of

interferon-regulated genes (IGS) in peripheral blood is also upregulated, which is sustained over time. [5].

**Systemic manifestations**—Unexplained fevers and hepatosplenomegaly are noted with AGS and are present in the context of the early/initial disease. Systemic inflammation is typically not persistent; acute phase reactants are not consistently reported [33, 35].

**CNS manifestations**—The encephalopathy and white matter disease of AGS are not seen in CANDLE or SAVI; however, CANDLE patients present with headaches and mild lymphocytosis in CSF suggestive of intermittent lymphocytic meningitis. Basal ganglia calcifications are most prominent in AGS (AGS > CANDLE > SAVI), are present in most CANDLE patients and less common in SAVI. In AGS, the onset and severity of encephalopathy correlates with CNS damage (i.e., severe brain atrophy, white matter disease, and resulting clinical sequelae), but AGS may present later in life (i.e., patients with *SAMHDI* mutations can present with later-onset vasculopathy and moyamoya disease later in childhood) [33]. Autoantibodies that trigger the CNS disease have not been described and the early-onset of AGS makes an innate immune dysregulation resulting in organ (CNS) damage most likely.

**Skin manifestations**—“Chilblains” or cold-induced acral dermatosis is present in up to 40 % of patients with AGS, but the face is typically spared. Out of 7 chilblain cases assessed in AGS, 4 were noted to have IgM deposition in the basement membrane (positive in 1 with mutated *TREX1* and 1 with mutated *RNASEH2B*, and 2 not genetically analyzed, one of whom was noted to have anti-C1q antibodies which raises the possibility of cofactors [36]; negative in 2 with mutated *SAMHDI* (at least 1 AR) and 1 with AR *TREX1* mutations) [37]. Chilblain-like vasculopathy is also seen in SAVI, but not CANDLE. Lobar neutrophilic panniculitis leading to lipodystrophy is seen in CANDLE, but is not present in SAVI and is rare in AGS (<1 %) [33].

**Hematologic manifestations**—Hematologic manifestations including thrombocytopenia, lymphopenia, and leukopenia may be autoantibody-mediated or the result of IFN-mediated bone marrow suppression. Cytopenias with IFN treatment are reported [38, 39] and IFN-mediated transient bone marrow suppression (mostly lymphopenia) is the likely mechanism of cytopenias in patients with SAVI and CANDLE. Thrombocytosis is common in SAVI. In AGS, thrombocytopenia and leukopenia have been reported, but their association with antibodies or flare-induced bone marrow suppression remains unclear [33, 40].

**Cardiovascular manifestations**—Systemic hypertension and pulmonary hypertension are recently observed morbidities in CANDLE and SAVI, which suggest a shared role of interferon in vasculopathy, but have not been reported in AGS.

**Autoantibodies**—Features of complex SLE were systematically assessed in 18 mutation positive AGS patients such as oral ulcers (3/18, 16.7 %) and detectable autoantibody titers including ANA (3/18, 16.7 %), anti-dsDNA (1/18, 5.6 %), and anticardiolipin (ACA) (2/18, 11.1 %) antibodies [40]. Some SLE clinical criteria [41] thought to be triggered by



respective immune complex deposition include classic photosensitive butterfly rash (Fig. 1) [42], pleuritis [43], SLE nephritis and presence of antibodies to chromatin (anti-dsDNA) and to spliceosome components (anti-Sm) and these criteria are rarely positive in AGS patients (Table 4). Autoantibodies are also detected with mostly low titers in CANDLE (12 ANA, 3 anti-dsDNA, 2 antiphospholipid (anti-cardiolipin, beta 2 glycoprotein) antibodies, 2 anti-SSA, 1 anti-smRNP, 1 anti-SmB, 5 ANCA), and SAVI (9 ANA, 2 anti-dsDNA, 7 antiphospholipid antibodies, 2 anti-RNP, 1 anti-Ro, 6 ANCA). Variable lupus anticoagulant positivity was found in 7/15 (46.7 %) CANDLE and in 4/9 (44.4 %) SAVI. Data are based on patients who had at least one positive result over time. Autoantibody titers fluctuate and do not correlate with disease activity in CANDLE and SAVI and are thus likely secondary to persistent IFN upregulation rather than driving pathology. Metabolic and pulmonary manifestations seen in CANDLE or SAVI have not been described in AGS.

### Genetics and pathogenesis of AGS compared to SAVI and CANDLE

The pathogenesis of most forms of AGS is thought to result from accumulation of nucleic acids that cannot be sufficiently metabolized, thus raising the pool of cytoplasmic nucleic acids that signal through the DNA sensor cGAS which activates adaptor STING and/or RNA sensors MDA-5 and RIG-I which activate adaptor MAVS [45]. In 2006, AR LOF mutations in 3' repair exonuclease *TREX1* became the first monogenic cause of AGS [46]. Since then, AR LOF mutations in components of the RNASEH2 endonuclease complex (*RNASEH2A*, *RNASEH2B*, and *RNASEH2C*) [47], a deoxynucleotide triphosphohydrolase *SAMHD1* [48], and an adenosine deaminase, *ADARI* or *ADAR* [49] were found to cause AGS. Most recently, GOF AD mutations in *IFIH1* (interferon-induced with helicase C domain 1 or AGS7), also known as *MDA5* or melanoma-differentiation-associated protein 5 [50, 51] have been reported to cause an AGS phenotype with more variable disease severity.

As the major cytosolic 3'-to-5' exonuclease, *TREX1* deficiency can lead to the accumulation of interferogenic DNA-RNA triggers. *TREX1* can act in concert with a granzyme A-activated endonuclease, NM23-H1, to degrade DNA during granzyme A-dependent apoptosis. A defect in *TREX1* would lead to decreased degradation of damaged DNA that is generated with granzyme A-dependent apoptosis [52]. Furthermore, the absence of TREX1 in *Trex1*<sup>-/-</sup> mouse embryo fibroblasts upon gamma radiation destabilizes *Chk2*, a central DNA damage checkpoint [53], and results in the accumulation of aberrant extranuclear single-stranded DNA (ssDNA) that result from uncleared DNA replication intermediates and leads to chronic ATM-mediated Chk2 activation [54]. Lastly, the absence of *TREX1* has also been associated with the accumulation of reverse-transcribed DNA derived from transcription of endogenous retroelements [55]. The accumulation of cytoplasmic DNA leads to STING-dependent but mostly endosomal TLR-independent upregulation of type I IFN transcription in the cell types examined [55, 56]. LOF mutations in RNASEH2 endonuclease complex are associated with increased formation of RNA:DNA hybrids [57] that are involved in the regulation of a wide range of functions including immunoglobulin class switching, regulation of gene expression and replication of mitochondrial DNA [58]. Furthermore, mutations in the RNASEH2 complex result in defective removal of misincorporated ribonucleotides from DNA due to deficient ribonucleotide excision repair which causes DNA damage and triggers a cGAS and STING-

dependent type I IFN response [59, 60]. LOF mutations in *SAMHD1* reduce DNA damage repair and increase the dNTP pools [61], similarly resulting in STING-dependent type I IFN signaling [60, 62, 63]. Adenosine deaminase ADAR has dsRNA-editing function through deaminating adenosine bases to inosines [49, 64]. When mutated, there is accumulation of dsRNA stimulating MAVS-dependent type I IFN transcription [65]. Finally, GOF mutations in *IFIH1* lead to MAVS-dependent IFN production without nucleic acid accumulation with incomplete penetrance and variable clinical phenotypes suggesting a possible role for modifier genes [50, 51].

The actual pathomechanisms that lead to CNS damage in AGS patients are unknown. In a mouse model transgenic for a glial fibrillary acidic protein-interferon-alpha fusion protein where IFN $\alpha$  is expressed in an astrocyte-restricted fashion, transgenic animals develop a progressive inflammatory encephalopathy with neuropathologic features similar to AGS [66] suggesting a neurotoxic role of IFN $\alpha$  in the developing brain. In contrast, the disease-causing mutations in CANDLE and SAVI are not known nor expected to alter nucleic acid metabolism. The type I IFN production in CANDLE is MAVS and STING independent [17]. In SAVI, GOF STING mutations directly increase *IFNB1* transcription.

## Comparison of clinical features in IFN and IL-1 mediated autoinflammatory diseases

Clinical features that are overlapping in at least 2 of the 3 disease groups (AGS, CANDLE, SAVI) are summarized in Table 3 and contrasted with clinical features that are seen in patients with IL-1 mediated diseases [44]. The features more commonly seen in patients with known IFN dysregulation may suggest a role for IFN in perpetuating organ inflammation and damage and may help with defining a pathogenic role of IFN in yet uncharacterized autoinflammatory diseases.

## Familial chilblain lupus (FCL), retinal vasculopathy with cerebral leukodystrophy (RVCL), and Singleton Merten syndrome (SMS)

FCL phenotype has been associated with AD mutations in *TREX1* [67, 68], *SAMHD1* [69] and, more recently, *TMEM173/STING* [70]. FCL is characterized by painful bluish-red papular or nodular lesions of skin in acral locations precipitated by cold or wet exposure, some with ulceration and generally without photosensitivity, consistent with cutaneous chilblain lupus. Four different AD *TREX1* mutations have been reported in 4 kindreds with FCL. The most common FCL mutation (D18N) is also associated with heterozygous AGS, usually de novo, but has also been seen with both phenotypes (AGS, FCL) in the same family; this same mutation has been seen in an asymptomatic mother who has a child with a FCL phenotype [71, 72]. An AD *SAMHD1* mutation, which has also been observed with AGS when inherited as recessive mutation, has been reported in 1 family with chilblain lupus and photosensitivity but a negative autoantibody panel [69]. In most patients with FCL the positive ANA antibody titers fluctuate over time, and anti-dsDNA Abs or immune-complex mediated kidney disease have not been described [67, 71]. Upregulation of IFN-stimulated genes in peripheral blood from FCL patients and increased expression of IFN-



inducible IFN myxovirus resistance protein A (MxA) and CXCL10 in lesional skin biopsies from FCL patients have been seen [71, 73]. Cold-induced skin features of FCL resemble the cutaneous features in SAVI in some [74] and a distinct *TMEM173* mutation identified in a single kindred has the phenotype of FCL [70], but no lung disease in FCL has been reported as seen in SAVI. The pathophysiology of FCL remains unclear. The fact that mutations in 3 different genes cause a FCL phenotype suggests additional modifiers that lead to the disease. Whether the vasculitis in cold-exposed areas is mediated by direct endothelial cell damage or by cold-induced autoantibodies needs further investigation.

RVCL is an adult-onset autosomal dominant neurological disease caused by frameshift mutations in the C-terminus of *TREX1* that are distinct from the AGS causing mutations. Prominent features include vascular retinopathy leading to blindness, neuropsychiatric symptoms, and progressive neurologic disease with characteristic neuroimaging abnormalities [72, 75]. Though there is a shared gene, relationship to IFN signaling in RVCL is not yet established [72, 76].

SMS is associated with AD presumed gain-of-function mutations in *IFIH1* and *DDX58* [77, 78], characterized by dental dysplasia, early-onset aortic and valvular calcification, and acro-osteolysis. One *IFIH1* mutation, which had not previously been associated with AGS phenotype, was identified in 3 families with SMS phenotype, with upregulation of interferon-regulated genes [78]. Interestingly, a distinct mutation in *IFIH1* in a family characterized by acral lentiginos, scarring and ulceration of distal ears, erythematous cheeks, loss of secondary dentition, contractures, variable neurologic involvement, skeletal abnormalities, and glaucoma was associated with overlapping features of AGS, FCL, and SMS and upregulated interferon-stimulated genes [79]. Two *DDX58* mutations in 2 respective families were associated with atypical SMS presenting with glaucoma, aortic calcifications, and skeletal abnormalities and notably normal dentition [77]. The more variable clinical features in the context of vascular calcifications suggest that other genetic and/or environmental factors may influence the disease phenotype.

## **Comparison of the “autoimmune” interferonopathies (complement deficient SLE, and DNase deficient SLE) with the “autoinflammatory” interferonopathies SAVI and CANDLE**

The IGS was initially described in SLE in 2003 [6] and in other polygenic autoimmune conditions. The discovery of “monogenic interferonopathies” raises the question of similarities and differences of monogenic versus complex autoimmune diseases. The finding of monogenic mutations in a complement factor (*C1Q*) and the DNA nucleases (*DNASE1* and *DNASE1L3*), which encode proteins involved in the rapid clearance of apoptotic cells, provide monogenic disease models of SLE (monoSLE) that are invaluable in comparison with CANDLE, SAVI, and AGS.

### **MonoSLE**

AR LOF mutations in complement component 1 (C1q) that lead to C1q deficiency (C1QMIM#613652), and AD LOF mutations in *DNASE1* (MIM# 152700), and AR LOF

mutations in *DNASE1L3* (MIM#614420) predispose to development of SLE with characteristic clinical features, including positive anti-dsDNA antibodies, positive anti-spliceosome antibodies (i.e., anti-Sm), immune-complex-mediated glomerulonephritis (GN), malar photosensitivity rashes, and LE (lupus erythematosus) band seen on skin biopsy indicating complement or IC deposition in the dermis-epidermis interface [41, 42]. More than 90 % of individuals with C1q deficiency develop SLE features with equal incidence among males and females with childhood-onset of disease. The high incidence and defined clinical presentation of SLE associated with mutations in these genes led to their designation as a “monogenic form of SLE.” LOF mutations in other complement components (i.e., C1r/C1s, C2, C3, C4) are associated with a lower risk of developing SLE and other autoimmune conditions, with a SLE risk of about 65 % with C1r/C1s deficiency, 10–20 % for patients with C2 and C3 deficiencies, and about 80 % for patients with C4 deficiencies respectively [9, 80]. C1r and/or C1s deficiency is associated with a prominent immunodeficiency phenotype with recurrent bacterial, viral, or fungal infections in 85% and many dying at a young age due to severe infection [80]. C4 deficiencies have also been associated with increased infections, autoimmune hepatitis, and type I diabetes [81, 82]. This suggests that these mutations are strong modifiers of autoimmunity but may not cause a “monogenic form of SLE.”

### Clinical features in monoSLE compared to CANDLE, SAVI, and AGS

**C1q deficiency**—Kidney disease reported is primarily mesangial proliferative glomerulonephritis, but IgA nephropathy has been reported and IC deposition has been demonstrated [83, 84]. Variable rashes have been reported including a photosensitive rash with a positive LE band on skin biopsy reported in one case [84, 86, 87]. Antibodies are not uniformly assessed or reported. In one family, ANA and anti-Ro antibodies are reported in 2/2 [86] and in another kindred, 6/6 were ANA positive, 1/7 had positive anti-Sm antibody, and 2/6 had positive anti-Ro antibody titers [87]. Elevated IP-10 and IFN $\alpha$  in peripheral serum have been reported [85].

**DNASE1 deficiency**—AD mutation in *DNASE1* resulting in low serum DNASE1 activity was identified in 2 unrelated patients with juvenile-onset SLE (jSLE) (13yo, 17yo) who presented with fever and generalized erythematoid rash; 1 patient also had parotitis and renal disease with mesangial IgG deposition. Both patients had high-titer ANA, and positive anti-dsDNA, anti-RNP, and anti-SSA antibody titers [10].

**DNASE1L3 deficiency**—AR LOF mutations in *DNASE1L3* resulting in absent DNase activity in vitro, cause jSLE as reported in 17 individuals (11 male) from 6 Arab families. In 11 individuals (from 4 families) positive ANCA antibodies were measured. 15/17 patients had positive anti-dsDNA antibodies and all 17 had a positive ANA with low C3 and C4. Positive anti-cardiolipin antibody (ACA) titers were seen in 5 patients from 2 families and 11 patients from 5 families had nephritis [11].

SLE-specific diagnostic criteria are listed in Table 4 and were compared in monoSLE, CANDLE, SAVI, and AGS. Photosensitive rashes, positive LE band on skin biopsy, glomerulonephritis, and positive anti-dsDNA and anti-Sm antibodies are seen in patients

with monoSLE, but rarely in patients with CANDLE ( $n = 3$  with anti-dsDNA,  $n = 1$  with anti-SmB antibody), SAVI ( $n = 2$  with anti-dsDNA, none with anti-Sm antibody reported), and AGS ( $n = 1$  with anti-dsDNA and none with anti-Sm antibody reported). Interestingly, IgM deposition at the basement membrane or dermal-epidermal junction consistent with a LE band has been described with chilblain lesions in AGS patients [37, 46] as noted above. Other disease manifestations are as follows:

**Systemic manifestations:** Systemic inflammation with fever similar to CANDLE and SAVI is seen in early-onset of monoSLE [10].

**Skin manifestations:** The differences in the distribution of the facial rashes in SAVI and genetically undefined SLE are depicted in Fig. 1 with representative images [88, 89]. In SAVI, the erythematous rash is localized to acral areas such as the tip of the nose, cheeks and the ears and is aggravated by cold. In contrast, the characteristic malar rash in SLE is distributed in a butterfly pattern and photosensitive [42]. Skin biopsies from SAVI show small vessel vasculopathy and a predominantly neutrophilic infiltrate but absence of an LE band; however, a LE band at the dermal-epidermal junction has been described in monoSLE (C1q deficiency). In SAVI, ICs are seen in some vessels but typically later in the course when significant destruction has occurred, thus suggesting IC deposition in the context of clearance of tissue damage rather than as early triggers of the inflammatory response. In CANDLE, neutrophilic lobar panniculitis on biopsy is thought to lead to adipocyte destruction and lipoatrophy [14]. Panniculitis has not been reported in SAVI with rare reports in AGS [33] and so far no reports in monoSLE. In contrast to CANDLE, histologic features of SLE panniculitis (SLE profundus) include lobular lymphocytic infiltrates with increased interstitial mucin and hyaline necrosis in subcutaneous tissue [90], suggesting a distinct innate pathogenesis in CANDLE.

**Lung manifestations:** Interstitial lung disease in SAVI often with significant fibrosis at the time of biopsy is very rare in CANDLE, not reported in AGS, and not characteristic of SLE. Although not reported in monoSLE, pleuritis is a common lung manifestation of not yet genetically defined SLE. The presence of ICs in pleural fluid has been reported but their contribution to pathology is unclear [91].

**Autoantibodies:** Although autoantibodies are more frequently seen in CANDLE and SAVI than in IL-1 mediated autoinflammatory disease, the titers are typically low and the presence or absence of an autoantibody does not correlate with disease severity. Importantly, IC-mediated glomerulonephritis is a feature not yet described with CANDLE, SAVI, or AGS in contrast to monoSLE.

**Hematologic and CNS manifestations:** Hematologic features are part of the diagnostic clinical criteria of SLE [41] (Table 4) but have not systematically been assessed or described in reports of monoSLE. CNS disease has not yet been reported in monoSLE.

## Pathogenesis of “monogenic SLE” compared to SAVI, CANDLER, and AGS

The development of pathogenic ICs against nucleosomes including chromatin, ribonucleoproteins, and histones that are generated in the context of delayed clearance of apoptotic debris are thought to be the hallmark in SLE pathogenesis [92, 93]. The characterization of patients and their respective murine models with C1q-deficiency [94] after DNASE1 [95, 96] and DNASE1L3 [97] deficiency have provided insights into the pathogenic role of immune complexes and pathomechanisms that lead to the autoimmune-mediated organ damage that has significant overlap with the pathogenesis of respective clinical features in not yet genetically defined SLE.

1. Clearance of apoptotic debris: C1q is a strong opsonin for phagocytosis of late apoptotic and necrotic cells; it is essential for effective uptake of degraded chromatin by monocyte-derived phagocytes [94, 98]. A role of the two major serum DNases (Deoxyribonuclease I (DNASE1) and Deoxyribonuclease I-like 3 (DNASE1L3)) in degrading chromatin has been described [96, 97]. DNASE1L3 has a specific role in removing chromatin from the surface of microparticles that are released from apoptotic cells thus indicating a direct mechanism in controlling circulating levels of immunogenic chromatin [97]. Impaired clearance of apoptotic tissue and hematopoietic cells including neutrophil extracellular traps (NETs) induces secondary necrosis, which leads to presentation of dsDNA-protein including chromatin and RNA-protein complexes (ribonuclear proteins) on the cell surface (i.e., in blebs on apoptotic keratinocytes [99], on glomerular basement membrane (GBM) in kidneys [96], or released during neutrophil death by NETosis [100]). Secondary necrotic cell-derived material (SNECs) is more immunogenic and associated with exposure of autoantigens that are potent stimulators of autoantibodies [101].
2. Development of autoantibodies and break of tolerance: The cellular/tissue locations of the break in tolerance remain unclear and are likely variable and specific to autoantibodies present. Dendritic cells containing apoptotic bodies migrate from peripheral sites to draining lymph nodes (LN). Chromatin taken up in apoptotic blebs can induce myeloid dendritic cell (myDC) maturation [122], upregulation of proinflammatory cytokines, expression of MHCs and co-stimulatory molecules [102–104], and effective stimulation of autoreactive T-helper cells and autoantibody production by autoreactive B cells. Chromatin-Ig complexes are found in non-genetically defined SLE patients and can activate B cells by dual engagement of IgM through the B-cell receptor, and through the endosomal Toll-like receptors (TLRs) [106, 107]. Furthermore, DNASE-1 and DNASE1L3 deficiency affects tissue susceptibility to organ injury and damage. In a *Dnase1* deficient murine model, anti-chromatin antibodies selectively bind epitopes in renal tissue where Dnase-1 deficiency leads to necrotic death of kidney cells and the generation of chromatin fragments. Evidence of in situ immune complex deposition by binding to uncleared

cellular debris at tissue sites or blood have been supported in murine models of GN [108] and reflect a bidirectional interaction between the immune response and the target tissues (recently reviewed in detail in [92]).

3. Complement activation in tissues and tissue injury: Circulating autoantibodies/ICs can bind to tissue antigens exposed during cell stress, such as injury [92]. The clearance of antinuclear immune-complexes via Fc-gamma receptors by blood phagocytes, macrophages, and dendritic cells that get recruited to tissues may lead to the secretion of proinflammatory cytokines (IL-8, IL-1 $\beta$ , TNF $\beta$ , IFN $\alpha$ , IL-18) and tissue inflammation [105]. In the kidney, an organ sensitive to IC-mediated injury, anti-chromatin containing immunoglobulin (Ig) binds to the cell basal membranes, stimulating a local inflammatory response [109] and activating the complement cascade; The attack complex causes damage to the GBM for example and perpetuates an abnormal immune response [110].
4. Amplification of a vicious cycle of dysregulated immune responses and the role of IFN: The clearance of NETs is facilitated by DNASE1 and C1q-assisted extracellular pre-processing [98, 111]. After phagocytosis, NETs are normally degraded in an immunologically “silent” process [111]. Delay of NET clearance (i.e., in the context of anti-HMGB1, anti-LL37 and/or anti-C1q antibody and possibly anti-chromatin autoantibody interference that prevents access of DNASE1) can extend the survival of the NETs that are immunogenic and interferogenic [109, 112]. Type I interferon aggravates the abnormal tissue responses by accelerating autoreactive B cell maturation, precipitating cell death and organ damage [113], and by increased NETosis induction [114]. Uncleared NETs serve as a source of interferogenic autoantigens by exposure of chromatin, myeloperoxidase, proteinase 3, and release of oxidized mitochondrial DNA, thus fueling a type I IFN amplification loop that perpetuates the abnormal immune responses in SLE [100, 115]. IFN upregulation has been reported in C1q deficiency; pathogenic mechanisms in DNASE1 and DNASE1L3 deficiency suggest similar upregulation of IFN.

### **Distinction between autoinflammatory and autoimmune interferonopathies**

SAVI and CANDLE patients do not have genetic defects that affect clearance of apoptotic debris. In SAVI, constitutive transcription of IFN $\beta$  and the tissue-specific effects of STING signaling on cell homeostasis can lead to immune-complex and/or autoantibody independent organ inflammation and damage [4]. In SAVI, the lungs and small vessels in the skin are affected, but not the kidneys. The specific interferogenic pathomechanism in CANDLE remains incompletely defined, but the absence of significant autoimmunity despite significant tissue damage (high LDH, and development of lipodystrophy, myositis and other organ manifestations) and our data on independence of MAVS and STING signaling suggest

“independence” from the endosomal and RNA/DNA sensing pathways as well [17]. Although the persistent abnormal and chronic IFN response can and might stimulate autoimmunity [116] and may lead to “secondary autoimmune manifestations,” the CANDLE and SAVI-specific disease manifestations seen early in life are independent of autoantibody production and immune complex deposition. In that context, the presence of positive anti-citrullinated peptide antibody (ACPA) erosive RA in a 7 year old SAVI patient [4] may have been facilitated by the abnormal type I IFN production on the background of other genetically predisposing variants or modifiers (i.e., presence of shared epitope in the patient).

The overlap with autoimmunity, however, may occur more frequently in patients with AGS and SPENCDI (Fig. 2). Although the accumulation of cytoplasmic nucleic acids in AGS seems to mainly trigger the disease manifestations through stimulation of STING and MAVS dependent nucleic acid receptors, it cannot be excluded that stimulation through endosomal TLRs in B cells also occurs and may trigger autoimmune manifestations. Similarly, SPENCDI is caused by recessive mutations in *ACPS5*, which encodes tartrate-resistant phosphatase (TRAP) and presents with some features similar to lupus including anemia, nephritis, autoantibody positivity, thrombocytopenia, and myositis [117, 118] with 6/10 cases with dsDNA antibodies and 3/10 with lupus nephritis in one report [117]. Interestingly, increased levels of intracellularly phosphorylated osteopontin (OPN) are seen in the context of impaired TRAP function, which mediates signaling through TLR9 in predominantly plasmacytoid dendritic cells (pDCs), resulting in increased IFN $\alpha$  production, and thus activating a signaling pathway more likely to induce autoimmunity [66, 117, 118].

Scleroderma (SSc) [119, 120] and juvenile dermatomyositis (JDM) [7, 121] are genetically unsolved autoimmune diseases with prominent blood and tissue IGS that fluctuate with disease severity [7, 119, 120] and have overlapping clinical features with CANDLE and SAVI (JDM: myositis, lipodystrophy, vasculopathy, and lung disease; SSc: vasculopathy and lung fibrosis). A comparison of these conditions with CANDLE, SAVI, and the monogenic forms of SLE will likely provide clues to the pathogenesis of these diseases and to pathways leading to the IFN signature.

## Conclusion

Recent studies on a number of monogenic conditions that present with an IFN signatures and overlapping clinical features have provided insights into the different molecular pathways that drive type I IFN production. The comparisons of SAVI and CANDLE with AGS and monogenic forms of SLE (C1q deficiency, DNase1 deficiency, DNase1L3 deficiency) point to differences in innate and adaptive immune dysregulation that lead to IFN production. The type I IFN production is induced by constitutive *IFNB1* transcription in SAVI, and in most forms of AGS (AGS1-6), the accumulation of nucleic acids triggers innate immune nucleic acid (RNA and DNA) sensors in a STING or MAVS-dependent fashion causing innate immune activation. In contrast, when modified, chromatin fragments are exposed in the context of clearance or apoptosis defects that underlie the pathogenesis of monoSLE and polygenic forms of SLE. These modified autoantigens can trigger adaptive immune dysregulation by stimulating autoreactive B cell differentiation and production of



interferogenic ICs (i.e., against chromatin and spliceosome antigens) which can trigger IFN responses. In the autoinflammatory (CANDLE and SAVI) and autoimmune (monoSLE, some forms of SLE) interferonopathies, type I IFN leads to an amplification loop that reinforces the underlying abnormal pathomechanisms seen in these conditions and may provide a target for treatment. Insights gained from the study of these monogenic conditions can guide our understanding of other as yet genetically undefined autoinflammatory and autoimmune interferonopathies, guide new drug development, and provide rational approach to the use of emerging therapies targeting the type I IFN pathway.

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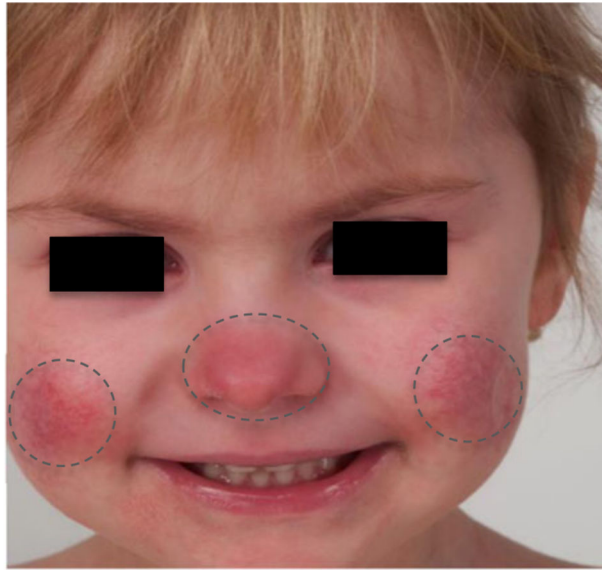
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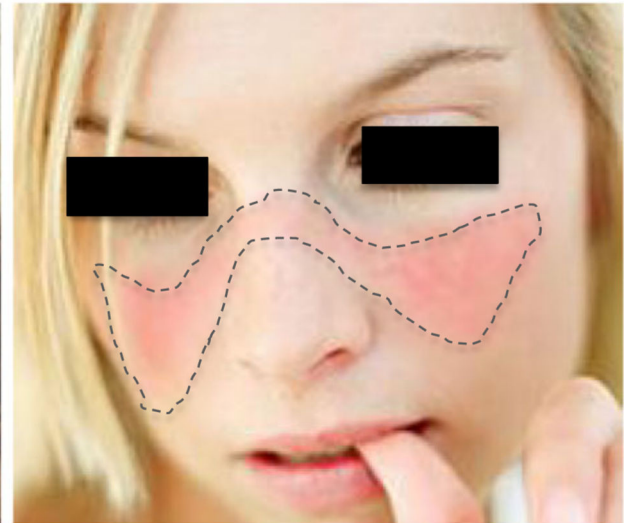
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SAVI

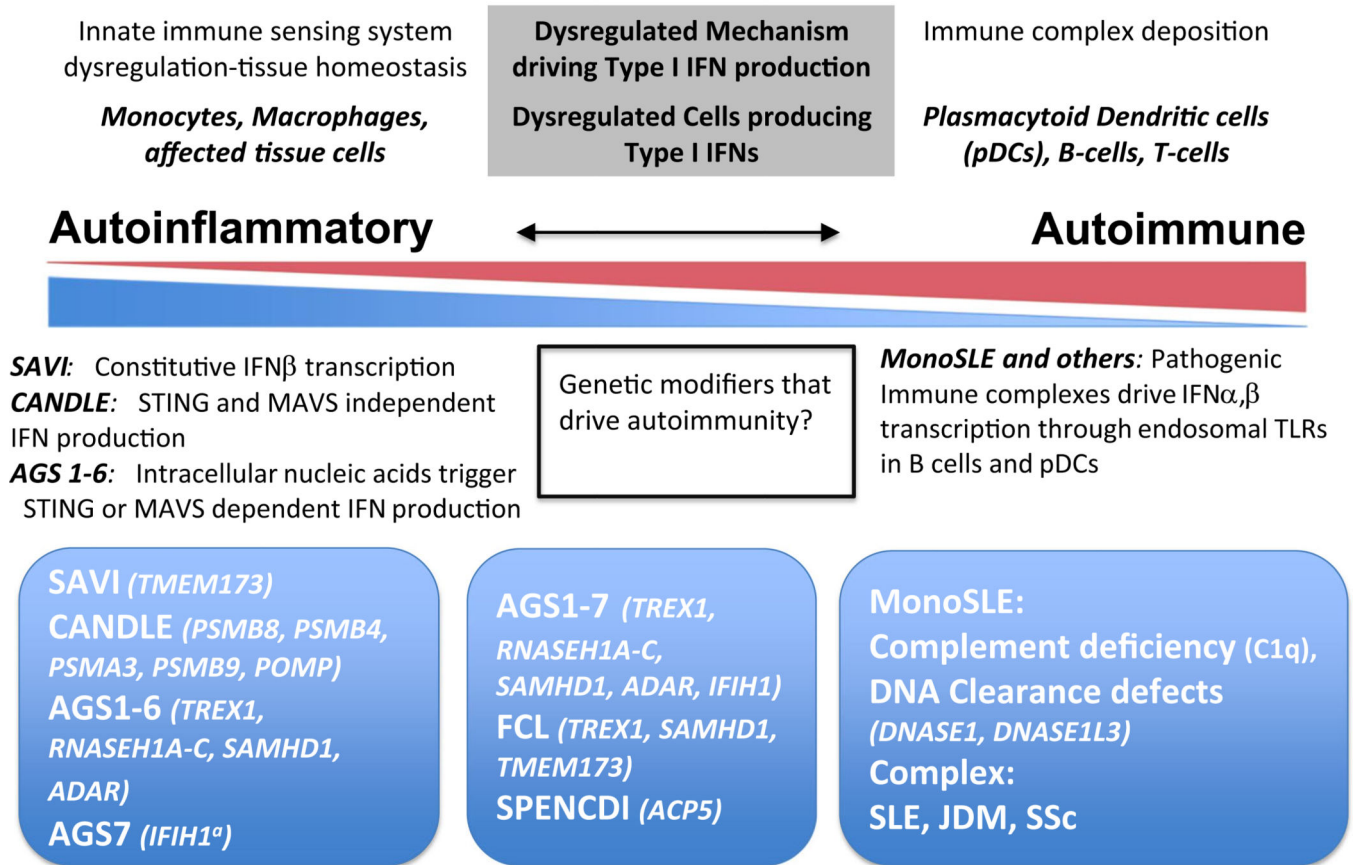


SLE

**Fig. 1.**

Different patterns in facial rash of SAVI and SLE. In SAVI patients (*upper panels*), the rashes are aggravated by cold exposure and are prominent in cold-sensitive areas including the tip of the nose and the lower part of the cheeks and the ears (not shown). The characteristic malar (facial) rash with a butterfly distribution on pictures from genetically undefined SLE patients (*lower panels*) show a photosensitive rash that is often induced by exposure to sunlight and associated with immune complex deposition at the dermal-epidermal junction (LE band) on biopsy. References: Lower left photo: © 2016 American College of Rheumatology. Used with permission. Lower right photo: [www.mollysfund.org](http://www.mollysfund.org). Used with permission.

# Pathogenesis of Interferonopathies



**Fig. 2.**

Comparative pathogenic considerations in autoinflammatory and autoimmune interferonopathies. On the *left end of the spectrum*, in monogenic conditions CANDLE/PRAAS and SAVI, the IFN signature is derived from proteasome suppression leading to *IFNA* and *IFNB1* transcription and constitutively increased *IFNB1* transcription, respectively, and are not triggered by immune complex deposition and can thus be called autoinflammatory. In AGS 1–6, sterile pyrexia and CNS damage are likely driven STING- or MAVS-dependent type I IFN production, suggesting innate (autoinflammatory) immune activation triggered by intracellular nucleic acid accumulation. *IFIH1*<sup>a</sup> (AGS7) mutations directly activate MAVS-dependent upregulation of IFN production without nucleic acid accumulation. The incomplete penetrance and variable clinical phenotype with *IFIH1* and *TREX1* mutations suggests a possible role for modifier genes. FCL and SMS have clinical features not seen in other interferonopathies. Though upregulated interferon-regulated genes are reported in SMS, little information regarding pathogenesis is available. Both autoimmune and autoinflammatory features are seen in the *middle box*. On the *right end of the spectrum*, clinical features of monoSLE (C1q deficiency, DNA clearance defects due to deficiency in DNASE1 or DNASE1L3) are likely driven by immune complex deposition (see text). The immune complexes can drive IFN production through activation of

interferogenic dendritic and B cell responses or interferogenic release of oxidized mitochondrial DNA, suggesting involvement of adaptive “autoimmune” pathways in disease pathogenesis. Pathogenic antibodies and immune complexes are seen in SLE, and likely in JDM and SSc

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**Table 1**

## CANDLE/PRAAS and SAVI demographics

Demographics	PRAAS/CANDLE (n = 37)	SAVI <sup>b</sup> (n = 14)
Age of onset <sup>a</sup>	Infancy–6 months	Infancy–6 months
Gender	13 F/24 M	5 F/9 M
Ethnicity		
Hispanic	15	1
Japanese	10	0
Caucasian	7	3
Afro-Jamaican	2	0
Palestinian	1	0
Ashkenazi Jewish	1	0
French-Canadian	0	5
Turkish	0	2
Ukrainian	0	1
Mixed Ethnicity	1	1
Unknown	0	1
Clinical outcome at last published assessment		
Alive	26 (70.3 %)	10 (71.4 %)
Deceased	11 (29.7 %)	4 (28.6 %)
Respiratory failure (6, 54, and 63 years old)/(14, 15, and 16 years old <sup>c</sup> )	3	3
Congestive heart failure (47 years old)	1	0
Presumed progression of chronic liver and kidney disease (25 years old)	1	0
Sepsis, recurrent pancreatitis (10 years old)	1	0
Presumed sepsis (8 years old)	1	0
Necrotizing fasciitis (29 years old)	0	1
Unknown (4, 6, 14, and 26 years old)	4	0

**References.** PRAAS/CANDLE: 3, 13–23, includes 7 previously unpublished patients evaluated or reviewed at NIH (3 with limited information not included in Table 2). SAVI: 4, 26–29, includes 1 previously unpublished patient evaluated at NIH

<sup>a</sup>PRAAS/CANDLE: seven with later onset (between 11 months–4 years). SAVI: three within one family with later onset, two as teenagers, one as adult

<sup>b</sup>Four cases within one family

<sup>c</sup>Respiratory failure in SAVI likely related to underlying SAVI-related lung disease. Secondary infection was noted in two (14 and 15 year old)



Table 2

## Clinical manifestations in CANDLE/PRAAS and SAVI

Organ system	Inflammatory manifestations	PRAAS/ CANDLE <sup>a</sup> fraction (%)	SAVI <sup>b</sup> fraction (%)	End organ damage	PRAAS/ CANDLE <sup>a</sup> fraction (%)	SAVI <sup>b</sup> fraction (%)
Systemic inflammation	Systemic inflammation (elevated acute phase reactants)	31/32 (96.9)	14/14 (100)	Growth retardation/failure to thrive	24/31 (77.4)	13/13 (100)
	Fever	24/24 (100)	9/13 (69.2)	Delayed puberty	6/6 (100)	3/3 (100)
	Anemia	29/32 (90.6)	9/9 (100)	Bone marrow suppression	13/20 (65)	5/10 (50)
	Thrombocytosis	5/23 (21.7)	6/7 (85.7)	Thrombocytopenia	10/20 (50)	0/7 (0)
Skin manifestations	Hepatomegaly and/or splenomegaly and/or lymphadenopathy	29/33 (87.9)	6/6 (100)	Leukopenia	2/14 (14.3)	5/10 (50)
	Panniculitis, nodular violaceous erythema	16/16 (100)		Neutropenia	2/17 (11.8)	3/7 (42.9)
	Annular plaques	15/16 (93.8)		Lymphopenia	11/17 (64.7)	4/8 (50)
	Acral violaceous plaques, nodules			Lipodystrophy	33/33 (100)	None
	Erythematous/violaceous rash at cheeks, tip of nose, tips of ears			Eschars	None	10/10 (100)
	Painful distal ulcers with purulent discharge or tissue infarcts			Hyperpigmented macules and scarring	3/3 (100)	
Vascular manifestations	Oral ulcers, pustules (including generalized), vesicles		7/7 (100)			
	Violaceous and or swollen eyelids	23/25 (92)				
	Perioral swelling	7/11 (63.6)				
	Livedo reticularis		7/7 (100)	Peripheral calcinosis	2/11 (18.2)	
	Raynaud phenomenon		4/4 (100)	Amputation of extremities	None	7/9 (77.8)
	Nailfold capillary changes		3/3 (100)	Nail dystrophy/loss	None	6/6 (100)
	Telangiectasia		8/8 (100)	Systemic hypertension	7/15 (46.7)	2/7 (28.6)
Musculoskeletal manifestations	Episodic or patchy myositis	13/19 (68.4)	3/6 (50)	Pulmonary hypertension	3/13 (23.1)	2/6 (33.3)
				Nasal septal perforation	None	7/9 (77.8)
				Muscle atrophy	13/18 (72.2)	2/2 (100)

Organ system	Inflammatory manifestations	PRAAS/ CANDLE <sup>a</sup> fraction (%)	SAVI <sup>b</sup> fraction (%)	End organ damage	PRAAS/ CANDLE <sup>a</sup> fraction (%)	SAVI <sup>b</sup> fraction (%)
	Synovitis (non-erosive), arthralgia, stiff joints	23/25 (92)	6/12 (50)	Joint contractures	25/32 (78.1)	3/4 (75)
	Finger swelling	17/20 (85)		Finger deformities, clubbing	24/25 (96)	
Metabolic manifestations	Myositis, or elevated CK/aldolase	22/30 (73.3)	2/5 (40)	Metabolic syndrome	4/5 (80)	
				Insulin intolerance, diabetes mellitus	9/22 (40.9)	1/4 (25)
				Dyslipidemia <sup>c</sup>	23/31 (74.2)	4/6 (66.7)
				Prominent abdomen and/or increased intraabdominal fat	15/20 (75)	1/3 (33.3)
				Acanthosis nigricans	8/20 (40)	
				Hepatic steatosis	5/10 (50)	1/3 (33.3)
CNS manifestations	Mild lymphocytic meningitis	4/16 (25)		Basal ganglia calcifications	20/29 (69)	3/7 (42.9)
	Headache	12/13 (92.3)	3/5 (60)	Cognitive dysfunction/ developmental delay, low IQ	10/28 (35.7)	2/8 (25)
				Seizures	3/16 (18.8)	
				Cardiac abnormalities <sup>d</sup>	6/20 (30)	1/6 (16.7)
Other organ manifestations	Interstitial lung disease/other lung disease <sup>e</sup>	3/19 (15.8)	12/14 (85.7)	Lung fibrosis/chronic interstitial lung disease		9/13 (69.2)
Immune dysregulation	Autoantibodies (variable, transient)	11/20 (55)	13/13 (100)			
	Recurrent infection <sup>f</sup>	14/19 (73.7)	7/7 (100)			
	Low complement (C3 or C4)	1/13 (7.7)	0/7 (0)			
	Hypergammaglobulinemia	9/16 (56.3)	7/7 (100)			

References. PRAAS/CANDLE: 3, 13–23, 4 previously unpublished patients evaluated or reviewed at NIH, and clinical updates from evaluation at NIH on 8 patients. SAVI: 4, 26–29 with clinical updates on 4 patients, 1 previously unpublished patient evaluated at NIH

<sup>a</sup>Other features seen in PRAAS/CANDLE: transaminitis ( $n = 16$ ), diarrhea ( $n = 10$ ), conjunctivitis/episcleritis/keratitis ( $n = 10$ ), hypertrichosis likely in the context of steroid therapy ( $n = 6$ ), teeth abnormalities/loss of teeth ( $n = 4$ ), hyperhidrosis ( $n = 4$ ), epididymitis/testicular swelling ( $n = 3$ ), pancreatic abnormalities/pancreatitis ( $n = 3$ ), dysphagia ( $n = 3$ ), constipation ( $n = 2$ ), gynecomasia ( $n = 2$ ), parotitis ( $n = 2$ ), renal calculi ( $n = 1$ ), eosinophilic esophagitis ( $n = 1$ ), bacterial overgrowth syndrome ( $n = 1$ ), nephrotic syndrome ( $n = 1$ ), pericarditis ( $n = 1$ ), keratoconus ( $n = 1$ ), IgA nephropathy ( $n = 1$ ),

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portal hypertension with esophageal varices ( $n = 1$ ), nodular regenerative hyperplasia of liver ( $n = 1$ ), antiphospholipid syndrome with venous thrombosis ( $n = 1$ ), subarachnoid hemorrhage with cerebral edema ( $n = 1$ ), bowel pneumatosis ( $n = 1$ ), undetectable IgA ( $n = 1$ )

<sup>b</sup> Other features seen in SAVI: sparse/thin hair ( $n = 3$ ), tubular/asymptomatic proteinuria ( $n = 2$ ), transaminitis ( $n = 2$ ), conductive hearing loss ( $n = 2$ ), ACPA (or anti-citrullinated peptide antibody) positive erosive arthritis ( $n = 1$ ), lentiginos ( $n = 1$ ), balanitis ( $n = 1$ ), hoarse voice ( $n = 1$ ), lymphedema ( $n = 1$ ), pectus carinatum ( $n = 1$ ), gynecomastia ( $n = 1$ )

<sup>c</sup> Dyslipidemia features include high LDL, low HDL, and/or hypertriglyceridemia in CANDLE. Only low HDL seen in SAVI

<sup>d</sup> PRAAS/CANDLE cardiac abnormalities: right ventricle dilation in setting of pulmonary hypertension ( $n = 2$ ), mitral valve prolapse ( $n = 1$ ), arrhythmia, premature ventricular contraction, congestive heart failure ( $n = 1$ ), atrial fibrillation ( $n = 1$ ), right ventricular hypertrophy with secondary repolarization abnormality in setting of pulmonary hypertension ( $n = 1$ ). SAVI cardiac abnormality: right ventricle dilation in setting of pulmonary hypertension ( $n = 1$ )

<sup>e</sup> PRAAS/CANDLE lung disease: interstitial lung disease ( $n = 1$ ), bronchiolitis-obliterans organizing pneumonia (BOOP)-like lung disease ( $n = 1$ ), interstitial pneumonitis ( $n = 1$ ). SAVI lung disease: all interstitial lung disease

<sup>f</sup> PRAAS/CANDLE infections: otitis media (OM), sinusitis, upper respiratory infection (URI), urinary tract infection (UTI), flu, mouth infection, bronchiolitis, sepsis following bowel perforation, pneumonia, oral candidiasis, gingivitis, periodontitis, onychomycosis. SAVI infections: URI, UTI, pneumonia, pansinusitis, oral candidiasis/thrush, otitis externa, OM, recurrent skin infections, cellulitis

**Table 3**

Differences in clinical manifestations of IL-1 and TFN-mediated autoinflammatory diseases

	<b>IL-1 mediated diseases</b>	<b>IFN mediated disease (CANDLE, SAVI, AGS)</b>
<b>Systemic</b>	CRP closely correlates with disease activity	CRP only elevated in severe disease
	Granulocytosis with flares	Lymphopenia, leukopenia with flares
<b>CNS</b>	Aseptic neutrophilic meningitis	Mild lymphocytic meningitis
	Arachnoid adhesions (severe disease)	Basal ganglion calcifications, CNS vasculopathy, white matter disease
<b>Skin / Vessels</b>	Neutrophilic dermatitis (urticaria-like with a mature neutrophilic infiltrate)	Panniculitis (“immature” neutrophils), lipoatrophy, vasculitis (chilblain-like lesion) microthrombotic disease
<b>Lung / Heart</b>	Serositis, pericarditis	Pulmonary fibrosis/interstitial lung disease HTN, pulmonary HTN
<b>MSK</b>	Osteomyelitis, bony overgrowth, fasciitis, arthralgia	Myositis, non-erosive arthritis, arthralgia
<b>ENT</b>	Hearing loss (inflammatory)	NA
<b>Eyes</b>	Conjunctivitis, anterior uveitis	Glaucoma, episcleritis
<b>Serology</b>	40% lupus anticoagulant positive, other autoantibodies uncommon	40–50% lupus anticoagulant positive in CANDLE/SAVI, some with other variable/transient autoantibodies

**Table 4**

Presence of 1997 ACR SLE Criteria in CANDLE, SAVI, AGS, and MonoSLE

	CANDLE <sup>d</sup>	SAVI <sup>d</sup>	AGS <sup>e</sup>	MonoSLE
1. Malar rash <sup>a</sup> butterfly-shaped rash across cheeks and nose	NO	NO	NO	YES
2. Discoid (skin) rash <sup>a</sup> raised red patches	NO	NO	NO	YES
3. Photosensitivity: skin rash as result of unusual reaction to sunlight	NO	NO	NO	YES
4. Mouth or nose ulcers: usually painless <sup>a</sup>	NO	YES	RARE	YES
5. Arthritis (non-erosive) in two or more joints, along with tenderness, swelling, or effusion. <sup>b</sup>	YES	YES	RARE	YES
6. Cardio-pulmonary involvement: inflammation of the lining around the heart (pericarditis) <sup>a</sup> and/or lungs (pleuritis) <sup>a</sup>	NO	NO	NO	UNK
7. Neurologic disorder <sup>a</sup> seizures and/or psychosis	RARE	NO	YES	YES
8. Renal (kidney) disorder <sup>a</sup> excessive protein in the urine, or cellular casts in the urine	NO	NO	NO	YES
9. Hematologic (blood) disorder <sup>b</sup> :				
• hemolytic anemia,	NO	NO	UNK	UNK
• low white blood cell count	NO	YES	YES	UNK
• low platelet count	YES	YES	YES	YES
10. Immunologic disorder, antibodies to:				
• Anti-double stranded DNA antibodies <sup>c</sup> ,	RARE	RARE	RARE	YES
• Anti-Sm antibodies <sup>c</sup> ,	RARE	RARE	RARE	YES
• Antiphospholipid antibodies <sup>b</sup> (anticardiolipin IgG/IgM, lupus anticoagulant, false positive Treponemal test)	YES	YES	RARE	YES
11. Antinuclear antibodies (ANA) <sup>b</sup> positive test in absence of drugs	YES	YES	RARE	YES

References. SLE Criteria: 61. PRAAS/CANDLE: 3, 13–23, includes 4 previously unpublished patients evaluated or reviewed at NIH, and clinical updates from evaluation at NIH on 8 patients. SAVI: 4, 26–29 with clinical updates on 4 patients, 1 previously unpublished patient evaluated at NIH. MonoSLE: DNASE1 deficiency,  $n = 2$  (10), C1q deficiency,  $n = 50$ –100 (9, 82, 85–88), DNASE1L3 deficiency,  $n = 17$  (11).

Four or more of the 11 criteria are needed for diagnosis of SLE. Clinical features that are more specific for SLE and are typically not observed in SAVI are depicted in black. In SLE, these are triggered by immune complex deposition. Features in *gray* are less specific for SLE and can be triggered by mechanisms other than immune complex deposition (i.e., hematologic manifestations seen in SLE can be caused by autoantibodies directed to cell subsets or by bone marrow suppression aggravated by the release of type I IFN). Antibodies to cardiolipin can often be present in patients with other immune dysregulatory diseases and are less specific for SLE. *Rare* indicates reported in less than 20 % of cases that report on the feature

<sup>a</sup>Evidence of immune complex deposition has been reported

<sup>b</sup>Multifactorial or nonspecific findings in isolation

<sup>c</sup>Antibodies specific to SLE are detected shortly before the development of specific organ disease

<sup>d</sup>CANDLE: oral ulcers  $n = 1$ , pericarditis  $n = 1$ , seizures  $n = 2$ , nephrotic syndrome unknown etiology  $n = 1$ , Coombs positive anemia  $n = 1$ , SAVI: oral ulcers in 4/5 with 1 associated with recurrent HSV labialis. ANA titers fluctuate (inconsistently positive) and do not correlate with disease activity in CANDLE  $n = 12$ , SAVI  $n = 8$

<sup>e</sup>Seizures are a common feature in AGS as part of primary encephalopathy