## Poor Evidence for C<sub>4</sub> Photosynthesis in the Wheat Grain

Research on  $C_4$  photosynthesis has seen a revival in recent years, leading to a *Nature Scientific Reports* article announcing the discovery of  $C_4$  photosynthesis in grains of the  $C_3$  plant wheat (*Triticum aestivum*; Rangan et al., 2016). The authors indicate the presence of  $C_4$  cycle enzymes, which they interpret as evidence for the existence of a  $C_4$ -type carbon-concentrating mechanism in the wheat pericarp. "Extraordinary claims require extraordinary evidence," however, and the evidence presented in this case is based entirely on transcriptome data of the entire wheat grain, with a notable absence of biochemical or physiological support, or the exact location of expression. One needs to consider the wider context of  $C_4$  photosynthesis, and this challenges the interpretation provided by Rangan et al. (2016).

It is a common misconception that in C<sub>4</sub> photosynthesis C<sub>3</sub>'s primary CO<sub>2</sub>-fixing enzyme Rubisco is simply replaced by phosphoenolpyruvate carboxylase (PEPC). PEPC indeed initially incorporates CO<sub>2</sub> as HCO<sub>3</sub><sup>-</sup> into the C<sub>4</sub> acid oxaloacetate, but this molecule is difficult to chemically reduce beyond the level of malate to that required for sugar, cellulose, and other carbohydrates, while still recycling the substrate PEP. So the CO<sub>2</sub> is later released to increase the CO<sub>2</sub> concentration in a specialized compartment, such as the bundle sheath (Leegood, 2002). There, the photosynthetic assimilation of CO<sub>2</sub> into carbohydrates occurs, via the same pathway as in C<sub>3</sub> plants, by Rubisco, ATP, and NADPH. This C<sub>4</sub> carbon-concentrating mechanism thus requires a PEPC:Rubisco activity ratio of roughly 1:1; a larger ratio means that ATP is spent on running a futile C₄ cycle. Without Rubisco, PEPC is only able to fix CO<sub>2</sub> via anaplerotic reactions. Two observations contradict the conclusions of Rangan et al. (2016) in this regard.

The authors show that the expression level of rbcS, the small subunit of Rubisco, is decreased by 99% relative to the expression in the leaf, and conclude that, together with an increase in PEPC expression and other  $C_4$  cycle enzymes, this indicates a shift from  $C_3$  to  $C_4$ photosynthesis. Without providing any measurements of the activity of Rubisco or PEPC to substantiate their claim, they refer to previous work showing that PEPC is 100 times as active in incorporating CO<sub>2</sub> into organic compounds as Rubisco (Duffus and Rosie, 1973). Assuming this to be the case here, the low concentration of Rubisco would leave insufficient capacity for photosynthetic fixation of the vast majority of CO<sub>2</sub> supplied by PEPC via a C<sub>4</sub> pump. It implies that the uptake of CO<sub>2</sub> by PEPC is to supply the large demand in amino and fatty acids. Support for the idea that PEPC facilitates the production of these compounds, and not carbohydrates, was earlier provided by pulse-chase experiments (Bort et al., 1995; Rolletschek et al., 2004).

The  $C_4$  cycle constitutes an energy-driven  $CO_2$  pump from a compartment of relatively low  $CO_2$  concentration to a largely gas-tight compartment in which the  $CO_2$  concentration is increased to reduce photorespiration (Leegood, 2002). Immunolabeling studies in wheat grains have shown that PEPC is not localized in the cross-cells of the pericarp, as suggested by Rangan et al. (2016), but in the aleurone layer and endosperm (Araus et al., 1993; González et al., 1998), the sites where one would expect the highest concentration of respiratory  $CO_2$ . With Rubisco located in the chloroplasts of the pericarp (Tambussi et al., 2005), this would render a  $C_4$ -like  $CO_2$  pump ineffective.

In our view, the claim of a  $C_4$  carbon-concentrating mechanism in wheat grains therefore cannot be upheld with the supplied data. In any such analysis it is important to distinguish production of  $C_4$  acids from the  $C_4$  photosynthetic pathway.

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## LITERATURE CITED

Araus JL, Bort J, Brown RH, Bassett CL, Cortadellas N (1993) Immunocytochemical localization of phosphoenolpyruvate carboxylase and photosynthetic gas-exchange characteristics in ears of *Triticum durum* Desf. Planta 191: 507–514

Bort J, Brown RH, Araus JL (1995) Lack of  $C_4$  photosynthetic metabolism in ears of  $C_3$  cereals. Plant Cell Environ 18: 697–702

Duffus CM, Rosie R (1973) Some enzyme activities associated with the chlorophyll containing layers of the immature barley pericarp. Planta 114: 219–226

González MC, Osuna L, Echevarría C, Vidal J, Cejudo FJ (1998) Expression and localization of phosphoenolpyruvate carboxylase in developing and germinating wheat grains. Plant Physiol 116: 1249–1258

**Leegood RC** (2002)  $C(_4)$  photosynthesis: principles of  $CO(_2)$  concentration and prospects for its introduction into  $C(_3)$  plants. J Exp Bot 53: 581–590

**Rangan P, Furtado A, Henry RJ** (2016) New evidence for grain specific  $C_4$  photosynthesis in wheat. Sci Rep **6:** 31721

Rolletschek H, Borisjuk L, Radchuk R, Miranda M, Heim U, Wobus U, Weber H (2004) Seed-specific expression of a bacterial phosphoenol-pyruvate carboxylase in *Vicia narbonensis* increases protein content and improves carbon economy. Plant Biotechnol J 2: 211–219

Tambussi EA, Nogués S, Araus JL (2005) Ear of durum wheat under water stress: water relations and photosynthetic metabolism. Planta 221: 446– 458

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