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IMPACT OF PLATELETS AND PLATELET-DERIVED MICROPARTICLES ON HYPERCOAGULABILITY FOLLOWING BURN INJURY

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Abstract

An acute burn induced coagulopathy develops after scald injury, which evolves into a subacute, hypercoagulable state. Microparticles, specifically platelet-derived MPs (PMPs), have been suggested as possible contributors. We first developed a model of burn-induced coagulopathy and then sought to investigate the role of platelets and PMPs in coagulation after burn. We hypothesized that changes in circulating platelet and PMP populations after injury would contribute to the post-burn, hypercoagulable state. A murine scald model with 28% TBSA full thickness burn injury was utilized and blood samples were collected at intervals after injury. Circulating MP populations, platelet counts, overall coagulation, and platelet function were determined. Burn injury led to hypercoagulability on post-burn day one (PBD1), which persisted 6 days after injury (PBD6). On PBD1, there was a significant decrease in platelet numbers and a decline in platelet contribution to clot formation with a concomitant increase in circulating procoagulant PMPs. On PBD6, there was a significant increase in platelet numbers and in platelet activation with no change in PMPs compared with sham. Further, on PBD1 decreased ADPinduced platelet activation was observed with a contrasting increase in ADP-induced platelet activation on PBD6. We therefore concluded that there was a temporal change in the mechanisms leading to a hypercoagulable state after scald injury, that PMPs are responsible for changes seen on PBD1, and finally that ADP-induced platelet activation was key to the augmented clotting mechanisms 6 days after burn.

Keywords

Burn injury; coagulation; hypercoagulability; microparticles; platelet function; thermal injury; thromboelastometry

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INTRODUCTION

Thermal injury leads to the disruption of coagulation homeostasis by inducing changes in the balanced processes of clot formation and lysis. An acute burn induced coagulopathy, characterized by endothelial injury, release of acute phase reactants, and impaired coagulation and fibrinolytic pathways (1), may also be accompanied by the development of a transient consumptive coagulopathy. The severity of these early coagulation changes correlates with age, total body surface area (TBSA) burned, presence of inhalation injury, obesity, and increased number of invasive procedures (1-5). A subsequent, subacute, and persistent hypercoagulable state follows, independently contributing to increased mortality (6, 7). Although many burn patients have been shown to develop persistent hypercoagulability after injury, the rates of venous thromboembolism (VTE), pulmonary embolism (PE), and disseminated intravascular coagulation (DIC) are highly variable, leading to much debate regarding optimal mechanical and chemoprophylaxis in these patients (8–11). Although it is clear that endothelial injury from the acute insult leads to the release of pro-inflammatory and procoagulant factors that initiate these changes, the mechanisms behind the development and persistence of this hypercoagulability remain poorly understood.

Human studies have demonstrated alterations to coagulation parameters temporally after burn using conventional prothrombin time (PT) and international normalized ratio (INR), and more recently with viscoelastic coagulation testing using thrombelastography (TEG) and rotational thromboelastometry (ROTEM) (3, 5, 12). These changes are often accompanied by increased circulating procoagulant factors and decreased anticoagulant factors (3, 4, 13). In addition to impacting overall coagulation, thermal injury is also associated with an initial transient thrombocytopenia followed by normalization of platelet counts and eventual reactive thrombocytosis in both humans (14, 15) and animal models (16). Further, lower platelet counts are associated with increased morbidity and mortality (15, 17). Proposed pathways for these coagulation state changes include tissue factor release, consumptive coagulopathy from development of DIC, and platelet dysfunction. Although microparticles (MPs), and more specifically platelet-derived MPs (PMPs), have been shown to contribute to other hypercoagulable trauma models (18–20), their role in hypercoagulability and altered platelet function after scald injury have not been explored.

The goal of the current study was first to define hypercoagulability after murine scald injury and then to investigate the roles of platelets and PMPs in coagulation after burn. We hypothesized that a murine model of major full-thickness scald injury would result in a hypercoagulable state, and that changes in circulating MP populations would contribute to the coagulation changes observed.

MATERIALS AND METHODS

Scald injury model

Male CF-1 outbred mice aged 6–8 weeks from Charles River (Wilmington, Massachusetts) were subjected to a 28% full-thickness scald injury burn as previously described (21). A 28% scald injury model was used as coagulation changes can been seen with this severity of

injury with a low risk of development of DIC (5). Briefly, mice were anesthetized with 3% isoflurane in oxygen, shaved over the dorsal surface, placed in a 28% total-body surface area template, immersed in a 90°C bath for 9 s, and resuscitated with 2mL sterile saline intraperitoneally. Sham-treated mice were treated similarly without water immersion. All murine experiments were performed between 8 AM and 10 AM using protocols approved by the Institutional Animal Care and Use Committee of the University of Cincinnati.

Thromboelastometry

Rotational thromboelastometry (ROTEM, TEM Systems Inc, Durham, North Carolina) analyses were performed to determine alterations in coagulation per manufacturer instructions. Whole blood collected via cardiac puncture from sham and scald-injured mice was anticoagulated with 10% citrate. Overall coagulation was determined using the NATEM test, extrinsic pathway coagulation using the EXTEM test, and fibrin contribution to clot using the FIBTEM test. For EXTEM and FIBTEM tests, 20μ L of thromboplastin was added to 300μ L of citrated blood to initiate clot formation. In addition, cytochalasin D was added to the FIBTEM samples to inhibit platelet activation. Clotting time (CT), clot formation time (CFT), clot lysis (LI30), α -angle (AA), and maximum clot firmness (MCF) were determined for each test. Platelet contribution to clot strength (%MCFPlatelet) was calculated by the equation: (EXTEM_{MCF} – FIBTEM_{MCF})/EXTEM_{MCF}, similar to the methods previously described (20, 22). All analyses were initiated within 10 min of whole blood collection.

Platelet count determination

Whole blood collected via cardiac puncture from injured mice was anticoagulated with heparin. Coulter AcT 10 Hematology Analyzer (Beckman Coulter, Brea, California) was used to determine complete blood count, including white blood cell count, hemoglobin, hematocrit, and platelet count.

Multiplate analysis

Whole blood was collected via cardiac puncture, anticoagulated with 10% hirudin, and analyzed by Multiplate software (Roche Diagnostics, Rotkreuz, Switzerland). ADPtest was used to determine ADP-induced platelet activation. Platelet aggregation velocity, total platelet aggregation (AU), and area under the curve (AUC) were measured.

Microparticle isolation, enumeration, and characterization

The MP isolation protocol used was adapted from those previously published by our laboratory (23). In short, whole blood collected via cardiac and was anticoagulated with heparin. It was then centrifuged at 450 *g* for 10 min, and the supernatant collected and centrifuged at 10,000 *g* for 10 min to remove platelets. The platelet-free supernatant containing the MPs was then diluted 1:1,000 with Roswell Park Memorial Institute media (RPMI) and labeled with 10 μ L/sample CD41 antibody (BD Pharmingen, Clone MWReg30, San Jose, California). Nanoparticle Tracking Analysis (Nanosight, Malvern Instruments Ltd, Worcestershire, UK) was then used to enumerate total and CD41 + MP (PMP) concentrations.

Microparticle procoagulant activity

Microparticle procoagulant activity was determined using a Zymuphen MP-Activity thrombin generation assay (Aniara, West Chester, Ohio). Whole blood was collected via cardiac puncture from sham and TBI mice 24 h after injury and anticoagulated with 10% citrate. Microparticles were isolated per protocol above, diluted 1:20, and the assay was performed per manufacturer protocol. The lower limit of detection was 0.2 nm.

Coagulation factor measurement by ELISA

Whole blood collected via cardiac puncture was placed in serum separator tubes (BD Bioscience, San Diego, California), centrifuged at 1,000 *g* for 10 min, and serum collected. Serum levels of fibrinogen and von Willebrand factor (vWF) were measured by ELISA according to the manufacturer's protocol (MyBioSource, San Diego, California). The lower limit of detection for fibrinogen was 5 ng/mL and for vWF was 156 pg/mL.

Statistical analysis

Student's *t* tests were used when comparisons were made between two treatment groups. One-way ANOVA with Tukey post-hoc test was used to compare multiple populations. Prism 6 (GraphPad Software, La Jolla, California) was used for all statistical analyses. Experiments containing multiple data points were used to calculate means and standard errors of the mean. A *P*-value of 0.05 was considered significant.

RESULTS

Increased clot strength is seen after scald injury

The development of a hypercoagulable state after burn injury is well documented in humans (2, 4). This hypercoagulable state has been demonstrated in various animal models (24); however, it has not yet been shown in a murine model. We therefore sought to develop a murine model of scald injury that resulted in a hypercoagulable state. We performed 28% TBSA scald injury to mice and analyzed whole blood collected temporally after injury. At both 1 and 6 days after injury, we found a significant increase in maximum clot strength on NATEM analysis in burn mice when compared with sham mice (Fig. 1).

Platelet counts and platelet function are altered 1 day after scald injury

We next sought to focus on the coagulation changes seen on post-burn day one (PBD1) and to determine whether circulating platelets were impacted by scald injury. Whole blood platelet counts were significantly decreased in PBD1 mice compared with sham (Fig. 2A). Hematocrit and white blood cell counts were similar between scald and sham mice (data not shown). Further, when ADP-induced platelet function of sham and scald-injured mice was compared, there was a significant decrease in platelet activation following stimulation with this agonist (Fig. 2B). There was also a complementary decrease in the calculated platelet contribution to clot formation at this time point (Fig. 2C).

Circulating platelet-derived microparticles and microparticle procoagulant activity is increased 1 day after scald injury

As previous studies have shown microparticles to be hypercoagulable (18–20), we next sought to determine whether MPs, and specifically PMPs, were contributing to coagulation changes seen after burn. We first enumerated circulating PMPs in whole blood. In contrast to the decrease in total serum microparticles, there was a significant increase in the concentration of PMPs in the blood of PBD1 mice compared with sham mice (Fig. 3A). Further, a significant increase in procoagulant activity of these MPs was observed at this timepoint (Fig. 3B).

Plasma fibrinogen and von Willebrand factor levels are unchanged after scald injury

We next sought to determine whether other circulating coagulation factors could be contributing to the hypercoagulability seen after injury. As the independent contributions of fibrinogen and platelets to clot strength can alter NATEM tests, we measured plasma levels of both fibrinogen and von Willebrand factor (vWF). On PBD1, there were no changes in circulating fibrinogen (71.4 ng/mL sham vs. 46.3 ng/mL PBD1, P = 0.13) or vWF (406.5 pg/mL sham vs. 526.4 pg/mL PBD1, P = 0.54) in scald-injured mice when compared with sham mice.

Platelet counts and function are increased 6 days after scald injury

Six days after burn (PBD6), mice remained hypercoagulable as demonstrated in Figure 1. We therefore next investigated whether these coagulation changes were because of similar mechanisms as those seen on PBD1. In contrast to PBD1, when thrombocytopenia was observed, on PBD6 there was a significant increase in circulating platelet counts (Fig. 4A) with similar hematocrit and white blood cell counts compared with sham (data not shown). In addition, there was no change in PBD6 circulating PMP levels compared with sham (Fig. 4B). Further, on PBD6, a significant increase in ADP-induced platelet activation was demonstrated in scald-injured mice compared with sham mice (Fig. 4C). Finally, there was an increase in clot strength via the extrinsic pathway with no change in the relative platelet contribution to clot formation (Fig. 4D).

DISCUSSION

In the current study, we demonstrated sustained posttraumatic hypercoagulability in a murine model of full thickness scald injury (Fig. 1). The contributing factors to this hypercoagulable state evolved over time. On examination of circulating platelet populations and platelet function 1 day after injury, we found decreased platelet counts, decreased platelet activation, and reduced platelet contribution to clot formation, suggesting that platelets were not responsible for the hypercoagulability seen at this time point (Fig. 2). There were also increased procoagulant PMPs in burn mice (Fig. 3) with no changes in circulating fibrinogen or vWF, supporting the hypothesis that PMPs are responsible for PBD1 hypercoagulability. By contrast, 6 days after injury, increased circulating platelet populations were observed with a similar number of circulating PMPs as sham mice (Fig. 4, A and B). Further, ADP-induced platelet activation was increased with a parallel increase in clot strength via the extrinsic pathway compared with sham mice. Altogether, this suggests

that platelets, rather than PMPs, contribute to the hypercoagulable state on PBD6 (Fig. 4, C and D).

Our findings of hypercoagulability after scald injury in a severe burn model on PBD1 and PBD6 are consistent with previous human studies (3-5, 12) and animal models with similar trauma injuries (24, 25). Further, alterations in circulating platelet counts after injury, with an acute thrombocytopenia and subsequent recovery with thrombocytosis, are also well supported in both human (14, 15) and murine studies (16, 26, 27). Thromboelastometry has not been previously utilized in murine models of thermal injury; however, our results are consistent with increased clot strength seen in human studies (5, 12). Platelet function has also been shown to be altered after scald injury (26, 28); however, demonstration of changes in platelet aggregation over time in a murine scald injury is novel. Eurenius et al. (26) saw decreased platelet aggregation early after scald injury in rats, and hypothesized that this change in platelet function was secondary to factors extrinsic to platelets rather that because of the platelets themselves. They were unable to identify a specific factor; however, our study clearly shows PMPs are at least one factor contributing to these changes. The development of DIC has also been described in select burn populations (29); however, it is likely found in more severe burn injuries as human studies suggest that DIC mainly occurs in patients with greater than 40% TBSA burns, and may be in part because of aggressive crystalloid resuscitation (5). As our model was only a 28% TBSA burn without ongoing volume resuscitation, our finding of normal circulating fibrinogen levels would be expected.

This study is the first to assess platelet function, platelet contribution to clot formation, and the role of altered microparticle populations in these changes after scald injury. Although human and animal studies have consistently found that platelet populations are altered after injury (14–16, 26, 27), no previous models of scald injury have examined platelet function and shown temporal changes after scald injury. Although ROTEM changes have been previously described in other murine trauma models (5, 12), this is the first murine scald model that has demonstrated hypercoagulability on thromboelastometry. As previous human studies have shown that viscoelastic coagulation testing is a better predictor of resuscitation status and risk for VTE compared with PT/INR (30, 31), we believe this may be a more relevant parameter to study. Further, these results suggest that VTE prophylaxis may not be as straightforward as previously thought and that the anticoagulability.

Understanding the mechanisms behind coagulation changes after burn is important as variable VTE rates (2) and use of anticoagulation (32) in this patient population are prevalent. The presence of hypercoagulability after burn is well established, and both the rate of VTE (11) and the need for increased chemo-prophylactic dosing (33, 34) are correlated with increased burn size. Further, post-mortem human studies show microvascular thrombosis in multiple organs, specifically lung and renal tissues, leading to multiple organ dysfunction syndrome (MODS) and death (35, 36). As MODS is the major cause of death in burn patients (37), preventing microvascular thrombosis in these patients may be life-saving. Further, although rates of VTE are heavily debated, autopsies have shown pulmonary embolism (PE) rates up to 35% (38) and PE-related mortality rates as high as 40% (11). Microparticles are known contributors to hypercoagulability in various disease processes

(18, 19, 39), and previous research has demonstrated that PMPs are associated with increased procoagulant activity in fresh frozen plasma (40) and are associated with hypercoagulability in other injury models (22). The current study is the first to highlight the link between PMPs and coagulation in burns and future burn care may require targeted anticoagulation strategies based on the time course after injury, including use of agents that target platelets and PMPs specifically, to more effectively prevent thrombosis-related morbidity and mortality.

Although this study establishes a novel, murine model of hypercoagulability after scald injury with associated changes in platelet function, there are some limitations to our study. First, we utilized a single scald model with 28% TBSA burns and therefore these findings may not be applicable to minor burns or non-thermal burns. We also focused on 1 and 6 days after burn, and have yet to determine when the mechanistic changes driving hypercoagulability are occurring. In addition, we did not perform *in vivo* studies to further demonstrate the impact of altered microparticle populations from scald-injured mice on coagulation in naive mice. Further, whereas we were able to show alterations to platelet function, we have yet to determine whether microparticles directly impact platelet function early after burn and identify the mechanism behind increased platelet functionality seen 6 days after burn. Future studies will focus on elucidating these mechanisms, as they will allow us to better understand and manage the coagulation changes seen after scald injury.

CONCLUSION

In conclusion, acute hypercoagulability after scald injury persists 1 week after injury in a murine burn model. There is a clear temporal change in the mechanisms that lead to this hypercoagulability. Platelet-derived MPs are likely responsible for changes seen on PBD1 and platelet activation is key to the altered clotting mechanisms seen on PBD6. Better understanding of these mechanisms will allow us to target thrombosis prophylaxis to the underlying cause of hypercoagulability after injury and prevent VTE-associated morbidity and mortality.

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Midura et al.



Fig. 1. Increased clot strength is seen after scald injury

Mice were subjected to scald injury or sham and whole blood collected temporally after injury for NATEM analysis. Sample size is 4–7 in each group. Significance was determined using ANOVA analysis with Tukey's post-hoc test. *P < 0.01 compared with sham.

Midura et al.

Fig. 2. Platelet counts and platelet function are altered 1 day after scald injury Mice were subjected to scald injury or sham and whole blood collected 1 day after injury. A, Platelet counts, B, ADP-induced platelet function, and C, platelet contribution to clot formation were then determined. Sample sizes are (A) n = 12-18, (B) n = 15-16, and (C) n = 6-10. Significance was determined using Student *t* test. **P*<0.05 compared with sham.

Midura et al.

Fig. 3. Circulating platelet-derived microparticles and microparticle procoagulant activity is increased 1 day after scald injury

Mice were subjected to scald injury or sham and whole blood collected on day after injury. A, Circulating platelet-derived MP populations and B, MP procoagulant activity were then determined. Sample sizes are (A) n = 11 and (B) n = 6-10. Significance was determined using Student *t* test. **P*<0.05 compared with sham.

Midura et al.

Fig. 4. Platelet counts and function are increased 6 days after scald injury

Mice were subjected to scald injury or sham and whole blood collected 6 days after injury. A, Platelet counts, B, circulating platelet-derived MP populations, C, ADP-induced platelet function, and D, ROTEM analyses of EXTEM and FIBTEM tests with the calculated platelet contribution to clot formation were then determined. Sample sizes are (A) n = 12–18, (B) n = 20, (C) n = 5–10, and (D) n = 7–10. Significance was determined using Student *t* test. *P< 0.05 compared with sham.