



Complete Genome Sequence of NEB 5-alpha, a Derivative of Escherichia coli K-12 DH5 α

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Escherichia coli K-12 DH5 α is one of the most popular and widely available laboratory strains, but, surprisingly, no complete genome sequence has been publicly available. Here, we report the complete, finished sequence of NEB 5-alpha (DH5 α *fhuA2*). It should serve as a useful reference for researchers working with DH5 α .

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H5 α , constructed by Douglas Hanahan, is one of the most commonly used Escherichia coli K-12 laboratory strains. It is widely commercially available and has several properties that make it well suited for cloning applications: it can be transformed with high efficiency; yields high-quality plasmid DNA due to the absence of nonspecific endonuclease I (*endA1*) (1); maintains plasmids stably due to low levels of homologous recombination (recA1); can be transformed efficiently with unmethylated DNA due to the disruption of endonuclease EcoKI (hsdR17); supports blue/white selection due to the alphacomplementable $\Delta lacZ58(M15)$ allele; and is deficient in periplasmic alkaline phosphatase (PhoA⁻), making it useful for studying membrane proteins expressed as *phoA* fusions (2). Finally, it is the only commercially available M.EcoKI⁺ strain, so DNA passaged through it will efficiently transform classical EcoKI+ E. coli strains.

Several DH5 α markers have been characterized previously: *deoR*, originally thought to be defective and therefore responsible for the high transformability of DH5 α (3), is in fact a wild type (4); *luxS* is defective (5); and the *rfbC1* allele is actually a frameshift in *rfbD* (6). Numerous other features can be deduced from the genome sequence of its ancestor, DH1 (7). A whole-genome shotgun assembly (WGA) of DH5 α , based on short reads and comprising 89 contigs, revealed additional variants (6). Although the characteristic deletion $\Delta(argF-lac)169$ was originally determined to be 97 kb in length (8), the WGA study (erroneously, we find) inferred a shorter length of 85 kb, with the *cynS-mhpC* region not part of the deletion (6).

We have sequenced the genome of NEB 5-alpha (New England Biolabs), an immediate *fhuA2* derivative of DH5 α , using the Pacific Biosciences RSII platform with P6 chemistry. A 10-kb SMRTbell library was prepared from total DNA using the manufacturer's instructions, size-selected (4 to 50 kb) using the BluePippin instrument (Sage Science), and sequenced on 1 SMRT cell with a 360-min movie, obtaining 314× mean coverage. The genome was assembled using RS_HGAP_Assembly.3 followed by manual refinement and reassembly using RS_BridgeMapper.

The closed and finished genome of NEB 5-alpha is 4,583,637 bp in length. Our assembly shows the deletion at $\Delta(argF-lac)169$ to be

97,240 bp in length, extending from *mmuP* to *mhpD*, which is consistent with the original report (8) and contrary to the WGA study (6). The *cynS-mhpC* region, including the $\Delta lacZ58(M15)$ allele, is present on the 47,357-bp ϕ 80d[$\Delta lacZ58(M15)$] insertion at *att\phi80* (as in DH10 β [9]) but is deleted as part of Δ (*argF-lac*)169, which we confirmed by PCR. The *phoA8* allele is a 723-bp internal, in-frame deletion.

In addition to those genes disrupted in DH1, *fhuA*, *crl*, and *phoE* are disrupted in NEB 5-alpha, and *ylbE* contains a frameshift. We confirmed all of the nonsynonymous changes identified previously (6), except for *abgR* and *yicJ*, which appear to be wild type. We identified four additional nonsynonymous mutations: *flgJ* (P254S), *insH20* (W140stop), *msbB* (M33I), and *ppsA* (A50T). We suggest the following genotype: *fhuA2::IS2* Δ (*mmuP-mhpD*)169 Δ *phoA8 glnX44* ϕ 80d[Δ *lacZ58*(*M15*)] *rfbD1 gyrA96 luxS11 recA1 endA1 rph*^{WT} *thiE1 hsdR17*.

Accession number(s). This sequence has been deposited at DDBJ/ENA/GenBank under the accession number CP017100 and is also available at New England Biolabs.

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REFERENCES

- Taylor RG, Walker DC, McInnes RR. 1993. E. coli host strains significantly affect the quality of small scale plasmid DNA preparations used for sequencing. Nucleic Acids Res 21:1677–1678. http://dx.doi.org/10.1093/nar/21.7.1677.
- Rodriguez-Quinones F, Benedi VJ. 1993. *Escherichia coli* strain DH5α is a suitable host for the study of *phoA* insertions. Focus 15:110–112.
- 3. Hanahan D, Jessee J, Bloom FR. 1991. Plasmid transformation of *Escherichia coli* and other bacteria. Methods Enzymol **204**:63–113.
- Xia XX, Qian ZG, Lee SY. 2011. Comparative proteomic and genetic analyses reveal unidentified mutations in *Escherichia coli* XL1-blue and DH5α. FEMS Microbiol Lett 314:119–124. http://dx.doi.org/10.1111/ j.1574-6968.2010.02157.x.
- Surette MG, Miller MB, Bassler BL. 1999. Quorum sensing in *Escherichia coli, Salmonella typhimurium*, and *Vibrio harveyi*: a new family of genes responsible for autoinducer production. Proc Natl Acad Sci U S A 96: 1639–1644. http://dx.doi.org/10.1073/pnas.96.4.1639.
- 6. Song Y, Lee BR, Cho S, Cho YB, Kim SW, Kang TJ, Kim SC, Cho BK.

2015. Determination of single nucleotide variants in *Escherichia coli* DH5 α by using short-read sequencing. FEMS Microbiol Lett 362:fnv073. http:// dx.doi.org/10.1093/femsle/fnv073.

- 7. Suzuki S, Ono N, Furusawa C, Ying BW, Yomo T. 2011. Comparison of sequence reads obtained from three next-generation sequencing platforms. PLoS One 6:e19534. http://dx.doi.org/10.1371/journal.pone.0019534. 8. Peters JE, Thate TE, Craig NL. 2003. Definition of the *Escherichia coli*

MC4100 genome by use of a DNA array. J Bacteriol 185:2017-2021. http:// dx.doi.org/10.1128/JB.185.6.2017-2021.2003. 9. Durfee T, Nelson R, Baldwin S, Plunkett G, III, Burland V, Mau B,

Petrosino JF, Qin X, Muzny DM, Ayele M, Gibbs RA, Csörgo B, Pósfai G, Weinstock GM, Blattner FR. 2008. The complete genome sequence of *Escherichia coli* DH10B: insights into the biology of a laboratory workhorse. J Bacteriol **190:**2597–2606. http://dx.doi.org/10.1128/JB.01695-07.