

The Effect of Sterile Acellular Dermal Matrix Use on Complication Rates in Implant-Based Immediate Breast Reconstructions

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Background The use of acellular dermal matrix (ADM) in implant-based immediate breast reconstruction has been increasing. The current ADMs available for breast reconstruction are offered as aseptic or sterile. No published studies have compared aseptic and sterile ADM in implant-based immediate breast reconstruction. The authors performed a retrospective study to evaluate the outcomes of aseptic versus sterile ADM in implant-based immediate breast reconstruction.

Methods Implant-based immediate breast reconstructions with ADM conducted between April 2013 and January 2016 were included. The patients were divided into 2 groups: the aseptic ADM (AlloDerm) group and the sterile ADM (MegaDerm) group. Archived records were reviewed for demographic data and postoperative complication types and frequencies. The complications included were infection, flap necrosis, capsular contracture, seroma, hematoma, and explantation for any cause.

Results Twenty patients were reconstructed with aseptic ADM, and 68 patients with sterile ADM. Rates of infection (15.0% vs. 10.3%), flap necrosis (5.0% vs. 7.4%), capsular contracture (20.0% vs. 14.7%), seroma (10.0% vs. 14.7%), hematoma (0% vs. 1.5%), and explantation (10.0% vs. 8.8%) were not significantly different in the 2 groups.

Conclusions Sterile ADM did not provide better results regarding infectious complications than aseptic ADM in implant-based immediate breast reconstruction.

Keywords Breast implants / Acellular dermis / Mammoplasty

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INTRODUCTION

The use of acellular dermal matrix (ADM) in implant-based immediate breast reconstruction is increasing in popularity due to its proposed benefits [1], which include superior aesthetic outcomes, reduced capsular contracture, and reduced postoperative pain [2-4]. Since the pectoralis major muscle frequently provides inadequate inferolateral coverage for the implant, ADM

can be a good substitute to allow complete subpectoral pocket creation.

The current ADMs available for breast reconstruction are offered as aseptic or sterile. "Aseptic" refers to the processing of tissue using methods to prevent, restrict, or minimize contamination with microorganisms from the environment, processing personnel, or equipment [5]. There is no sterility assurance level associated with aseptic processing. In contrast, "sterile" means

that the tissue has completed a terminal sterilization process validated to a sterility assurance level of 10^{-6} [6].

AlloDerm (LifeCell Corp., Branchburg, NJ, USA), the first ADM used in breast reconstruction, is prepared by freeze-drying under aseptic conditions. It is the most recognized aseptic ADM by most surgeons, remains the most popular choice for postmastectomy breast reconstruction, and has been found to be very reliable and predictable [7]. MegaDerm (L&C BIO, Seongnam, Korea), which has recently been introduced as a sterile and nonimmunogenic ADM, is made using electron-beam sterilization to eliminate viruses, bacteria, and spores, achieving a 10^{-6} sterility level.

The use of a sterile ADM would theoretically be associated with a reduced risk of infection in breast reconstruction. Recently, several publications have described various correlations between the use of sterile ADM and postoperative complications in breast reconstruction [8-10]. However, no published studies have compared aseptic and sterile ADM in implant-based immediate breast reconstruction. The authors performed a retrospective study to evaluate whether sterile ADM offers a potential advantage over aseptic ADM with respect to postoperative complications, and specifically, rates of infection in implant-based immediate breast reconstruction. The authors have no vested interest of any kind in the materials or services referred to in this article.

METHODS

A retrospective review of the medical records was performed among patients who underwent implant-based immediate breast reconstruction with the application of aseptic ADM (Al-

loDerm; size, 4×12 – 4×16 cm²; thickness, 2.31–3.30 mm) or sterile ADM (MegaDerm; size, 4×12 – 6×16 cm²; thickness, 1.5–2.3 mm) in our institution. Exclusion criteria included the following: (1) patients whose breast reconstructions were performed in a delayed fashion or involved either tissue expander placement or autologous reconstruction; (2) patients who received radiation therapy at any time; (3) patients with significant comorbidities that could affect wound healing; (4) breast reconstructions performed before April 2013, the first use of sterile ADM in our institution; and (5) patients who had not yet completed a 6-month follow-up visit.

General information on patient demographics, clinical course, and postoperative complications was collated. Patients were allocated to either the aseptic ADM group (AlloDerm) or the sterile ADM group (MegaDerm).

All breast reconstructions were performed according to the same surgical protocol by a single surgeon (J.H.L.). After completion of skin-sparing mastectomy, a subpectoral dissection was performed and the inferior and inferomedial attachments of the pectoralis major were released. An implant was inserted into a newly created submuscular pocket. The pectoralis muscle was used to cover the implant superiorly. An ADM sling was used to cover the implant inferolaterally. The ADM was sutured with 3-0 synthetic monofilament absorbable polyester (BIOSYN, Covidien, Mansfield, MA, USA) to the inframammary fold along the entire lower border of the breast pocket with interrupted sutures. Laterally, the ADM was sutured to the serratus fascia to define the lateral boundary of the breast pocket along the anterior axillary line (Fig. 1).

Postoperatively, two closed-suction drains were placed into the supra-ADM plane (between the ADM and the skin) and the

Fig. 1. ADM sling supporting the breast implant

The superior border of the ADM was sutured to the inferior border of the pectoralis major with an interrupted 3-0 synthetic monofilament absorbable polyester. Total implant coverage was obtained. ADM, acellular dermal matrix.



Fig. 2. Infection after implant-based immediate breast reconstruction

Intravenous antibiotic treatment or implant removal was required for infectious complications in implant-based immediate breast reconstructions.

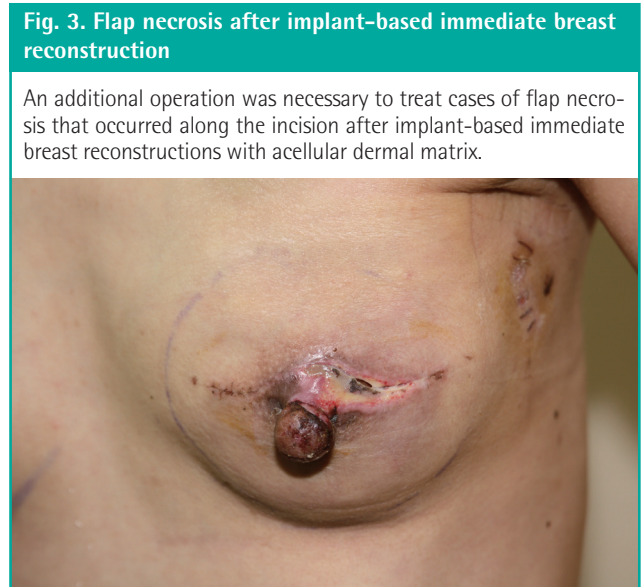


sub-ADM plane (between the implant and the ADM), until output decreased to < 20 mL over 24 hours. Postoperative antibiotics were continued until the drains were removed. Complications were analyzed, including infection, flap necrosis, capsular contracture, seroma, and hematoma. Infection was defined as erythematous changes on the affected breast with general symptoms of infection, including a sensation of heat on the breast, a generalized febrile sense, and fever (Fig. 2). Infectious complications were divided into 2 types: (1) infections requiring hospital readmission and intravenous antibiotics and (2) infections requiring surgery, including implant change or explantation. Any infectious complications encountered during the follow-up period were regarded as significant. Cultured microorganism data were collected in cases of infection requiring surgery. Flap necrosis was defined as skin necrosis only along the incision, without any erythematous change in the breast (Fig. 3). Seroma was defined as fluid collection on the affected breast that required ultrasonogram-guided aspiration after the removal of both drains. Overall explantation rates for any cause were also analyzed.

The Fisher’s exact test and chi-square test were performed to compare the outcomes between the aseptic ADM group and the sterile ADM group. P-values < 0.05 were considered to indicate statistical significance.

RESULTS

Eighty-eight breast reconstructions from April 2013 to January 2016 were included in this study. All 88 patients underwent unilateral implant-based immediate breast reconstruction using a textured silicone gel implant (Allergan Corp., Irvine, CA, USA).



Twenty patients were reconstructed with aseptic ADM and 68 patients with sterile ADM. The mean follow-up periods were 14 months in the aseptic ADM group and 12 months in the sterile ADM group.

Patients in the 2 groups had similar characteristics in terms of age, body mass index, medical comorbidities, implant size, breast cancer stage, and the need for adjuvant antineoplastic treatment (Table 1).

Infectious complications were encountered in 3 patients (15.0%) from the aseptic ADM group, including 1 case of infection requiring conservative treatment with intravenous antibiotics (5.0%) and 2 cases of infection requiring surgery (10.0%). In the sterile ADM group, the overall incidence of infectious complications was 7 (10.3%), including 2 cases of infection requiring IV antibiotics (2.9%) and 5 cases of infection requiring surgery (7.4%). There were fewer infectious complications in the sterile ADM group, but the difference failed to reach statistical significance. Flap necrosis was encountered in 1 case (5.0%) in

Table 1. Characteristics of the breast reconstruction patients

Characteristic	AlloDerm (n = 20)	MegaDerm (n = 68)	P-value
Age			
Mean	46.1	45.8	0.568
Range	32–58	31–62	
Body mass index (kg/m ²)			
Mean	22.3	23.1	0.529
Range	17.5–32.3	17.7–32.1	
Comorbid conditions			
Smoking	0	0	-
Diabetes	0	0	-
Obese ^a	2 (10.0)	7 (10.3)	0.814
Specimen weight, mean ± SD (g)	349.6 ± 158.2	358.1 ± 170.1	0.795
Implant size (mL)			
< 150	3 (15.0)	9 (13.2)	0.783
150–250	11 (55.0)	39 (57.4)	0.711
250–350	4 (20.0)	13 (19.1)	0.802
> 350	2 (10.0)	7 (10.3)	0.357
Breast cancer stage			
0	7 (35.0)	21 (30.9)	0.746
1	8 (40.0)	28 (41.2)	0.692
2	3 (15.0)	12 (17.6)	0.688
3	2 (10.0)	7 (10.3)	0.412
4	0	0	-
Adjuvant CTx	7 (35.0)	20 (29.4)	0.529
Neoadjuvant CTx	1 (5.0)	5 (7.4)	0.716
ALND	7 (35.0)	22 (32.4)	0.789
NAS	11 (55.0)	40 (58.9)	0.39
Bilateral reconstruction	0	0	-

Values are presented as number (%) unless otherwise indicated. CTx, chemotherapy; ALND, axillary lymph node biopsy; NAS, nipple-areola sparing. ^aBody mass index ≥ 25 kg/m².

Table 2. Complications of breast reconstruction patients

Variable	AlloDerm (n = 20)	MegaDerm (n = 68)	P-value
Infection	3 (15.0)	7 (10.3)	0.144
Infection requiring IV antibiotics	1 (5.0)	2 (2.9)	0.283
Infection requiring surgery	2 (10.0)	5 (7.4)	0.407
Flap necrosis	1 (5.0)	5 (7.4)	0.182
Capsular contracture	4 (20.0)	10 (14.7)	0.498
Baker grade I, II	3 (15.0)	7 (10.3)	0.144
Baker grade III, IV	1 (5.0)	3 (4.4)	0.605
Seroma	2 (10.0)	10 (14.7)	0.163
Hematoma	0	1 (1.5)	0.823
Explantation by any cause	2 (10.0)	6 (8.8)	0.235

Values are presented as number (%).

the aseptic ADM group, and in 5 cases (7.4%) in the sterile ADM group. The incidence of Baker grade I or II capsular contracture was 3 (15.0%) in the aseptic ADM group and 7 (10.3%) in the sterile ADM group. The incidence of Baker grade III or IV was 1 (5.0%) in the aseptic ADM group and 3 (4.4%) in the sterile ADM group. Seroma was encountered in 2 breasts (10.0%) in the aseptic ADM group, and 10 breasts (14.7%) in the sterile ADM group. The sterile ADM group showed a higher rate of seroma than the aseptic ADM group; however, this discrepancy was not statistically significant. Overall, the explantation rate and incidence of hematomas were not significantly different between the 2 groups (Table 2). Cultured microorganisms in infectious cases from the aseptic ADM group were identified as methicillin-susceptible *Staphylococcus aureus* (MSSA) in 1 case and methicillin-resistant *Staphylococcus aureus* (MRSA) in 1 case. In the 5 infectious complication cases in the sterile ADM group, we found 2 cases of MSSA, 2 cases of MRSA, and 1 case of *Serratia marcescens* (Table 3).

DISCUSSION

ADM is a structurally intact tissue matrix that can act as a biological scaffold to induce tissue ingrowth, angiogenesis, and tissue regeneration. Since Breuing and Warren [11] reported a successful outcome of breast reconstruction with ADM in 2005, the use of ADM with breast reconstruction has been a well-accepted surgical option. After skin-sparing mastectomy, ADM can be applied to form inferolateral coverage of the pocket of the tissue expander (two-stage breast reconstruction) or the pocket of the implant (direct-to-implant immediate breast reconstruction).

AlloDerm, the first ADM used in breast reconstruction, has become the most recognized aseptic ADM worldwide. Aseptic ADM undergoes an aseptic process, including cryopreservation,

Table 3. Identified microorganisms in infectious complication cases requiring surgery

Variable	AlloDerm (n = 2)	MegaDerm (n = 5)
MSSA	1	2
MRSA	1	2
<i>Serratia marcescens</i>	-	1

MSSA, methicillin-susceptible *Staphylococcus aureus*; MRSA, methicillin-resistant *Staphylococcus aureus*.

and is freeze-dried. As the benefits of using AlloDerm in implant-based breast reconstruction have become obvious, AlloDerm remains one of the most popular, reliable, and predictable choices for postmastectomy breast reconstruction.

Despite the removal during manufacture of all the cellular components in ADM that could cause rejection and inflammation, several reports have suggested that the use of ADM may be associated with elevated rates of infection and seromas [12,13]. Chun et al. [2] reported a series of 415 immediate breast reconstructions performed on 283 patients. The authors found that both seroma and infection rates were higher in the aseptic ADM group than in the non-ADM group. Similarly, Liu et al. [12] also found in their large series of 470 immediate reconstructions that the infection rate in the ADM group was significantly higher than in the non-ADM group. Antony et al. [13] reported a higher incidence of overall complications, specifically, seroma and reconstructive failures, in their ADM group versus their non-ADM group and their previously published non-ADM group. Similarly, Lanier et al. [14] found a significantly higher rate of infection, reoperation, explantation, and overall complications when they compared their aseptic ADM group to their non-ADM group.

In contrast, several studies have reported that ADM was not associated with an elevated rate of infectious complications. In his case series, Nahabedian [15] showed that the overall incidence of infection was essentially the same in the ADM and non-ADM groups. Vardanian et al. [16] also found that their aseptic ADM group had a similar safety profile to their non-ADM group.

In terms of infectious complication for the use of ADM in breast reconstruction, the use of sterile ADM would theoretically be associated with a reduced risk of infection in breast reconstruction. "Sterile" means that the ADM has completed a terminal sterilization process validated to a sterility assurance level of 10⁻⁶. In other words, a microorganism has a one in 1 million chance of surviving the sterilization process.

Buseman et al. [8] reported a comparative study of aseptic and sterile ADM. The authors found that sterile ADM has a risk of infection equivalent to that of aseptic ADM. However, the ster-

ile ADM group showed a higher incidence of seroma than the aseptic ADM group. Yuen et al. [17] reported a significantly higher incidence of infection and seroma in the sterile ADM group than in the aseptic ADM group. Previous studies comparing aseptic and sterile ADM have been performed with ready-to-use AlloDerm as the sterile ADM. MegaDerm, which has been recently introduced as a sterile ADM in South Korea, is prepared using an electron-beam sterilization process. No reports have been published on the use of MegaDerm for breast reconstruction.

Our results showed no statistically significant differences between the 2 groups in terms of infection, flap necrosis, capsular contracture, seroma, hematoma, or overall explantation rates. It should be noted that sterile ADM showed complication rates equivalent to those observed using aseptic ADM, including rates of infectious complications. However, our study has several limitations, including its retrospective nature. In practice, many potential factors could cause infectious complications in implant-based immediate breast reconstruction. To minimize the effect of all other factors except ADM, all cases before the first use of sterile ADM (MegaDerm) at our institution were excluded, and the only 20 cases were included in the aseptic group. The number of cases in the aseptic group was smaller than in recent studies of outcomes of ADM in breast reconstruction. In our study, no cases of red breast syndrome occurred. Since red breast syndrome also appears as erythema along the inferior pole of the breast, it is possible to misdiagnose a case as infection. Despite trying to render all conditions between the 2 groups as similar as possible, some factors that could influence the outcomes remained. Because the ADM was produced in a rectangular shape, trimming or removal of unnecessary parts of the ADM was performed to fit the inferolateral breast pocket. The amounts of microorganisms that could cause infectious complications were therefore different within the same group. Three breast surgeons performed skin-sparing mastectomies at our institution over the course of this study. A breast surgeon who leaves behind very well-vascularized mastectomy flaps affords the reconstructive surgeon an optimal setting to minimize complications. For example, the thickness of the skin after a skin-sparing mastectomy is important, because this skin flap can cause flap necrosis or infection. Additionally, the operating time may differ among surgeons, with longer operating time being a factor associated with infection. To minimize the influence of these biases on the outcomes of implant-based immediate breast reconstruction, a randomized controlled prospective trial would be necessary.

To summarize, sterile ADM did not provide improved results related to infectious complications, contrary to the theoretical

expectation. In other words, the sterilization of ADM for the sole purpose of minimizing infectious complications was unnecessary. It is noteworthy that infectious complications in implant-based immediate breast reconstructions can be influenced more by contamination during the intraoperative procedure or postoperative care than by contamination of the ADM product itself. Although the sterilization of ADM did not demonstrate more preferable outcomes in terms of infection, sterile ADM has some advantages, including resistance to degradation by collagenase, improvement of durability, and strength due to cross-linking [18]. These biomechanical characteristics of sterile ADM need to be investigated in further studies.

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