

Genome Sequencing and Analysis of the Filamentous Fungus *Penicillium sclerotiorum* 113, Isolated after Hurricane Sandy

Guohua Yin,^{a,b} Yuliang Zhang,^b Kayla K. Pennerman,^a Sui Sheng T. Hua,^c Qixing Huang,^b Anping Guo,^b Zhixin Liu,^b Joan W. Bennett^a

Department of Plant Biology and Pathology, Rutgers, The State University of New Jersey, New Brunswick, New Jersey, USA^a; Key Laboratory of Biology and Genetic Resources of Tropical Crops, Ministry of Agriculture, Institute of Tropical Bioscience and Biotechnology, Chinese Academy of Tropical Agricultural Sciences, Haikou, Hainan, China^b; U.S. Department of Agriculture, Western Regional Research Center, ARS, Albany, California, USA^c

G.Y. and Y.Z. contributed equally to this work.

***Penicillium sclerotiorum* is a distinctive species within the genus *Penicillium* that usually produces vivid orange to red colonies, sometimes with colorful sclerotia. Here, we report the first draft genome sequence of *P. sclerotiorum* strain 113, isolated in 2013 in the aftermath of Hurricane Sandy from a flooded home in New Jersey.**

Received 19 September 2016 Accepted 5 October 2016 Published 23 November 2016

Citation Yin G, Zhang Y, Pennerman KK, Hua SST, Huang Q, Guo A, Liu Z, Bennett JW. 2016. Genome sequencing and analysis of the filamentous fungus *Penicillium sclerotiorum* 113, isolated after Hurricane Sandy. *Genome Announc* 4(6):e01153-16. doi:10.1128/genomeA.01153-16.

Copyright © 2016 Yin et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Guohua Yin, guohuayin1997@gmail.com.

Penicillium sclerotiorum was first isolated from an air sample in Java, Indonesia, and described in 1937 (1). It has subsequently been isolated from Africa, Asia, and North America (2, 3). The fungus usually produces distinctive bright orange pigments, some of which have been identified as carotenoids (4). It also makes several xylanases (5, 6), as well as sclerotiorin, a secondary metabolite that has been used in anti-acne creams and as an inhibitor of aldose reductase (7–10). Furthermore, it has biotechnological potential for producing calcium malate from glucose (11). A recent report showed that *P. sclerotiorum* is the etiological agent of postharvest fungal diseases of pomegranate in Spain (12). Here, we report and characterize the genome of *P. sclerotiorum* 113, a strain isolated from a home in Manasquan, New Jersey, that was flooded with marine water in 2013 during Hurricane Sandy (13).

For genomic sequencing, we grew the fungus in potato dextrose broth with shaking at 200 rpm, 25°C for 7 days. We used an OMEGA Bio-Tek E.Z.N.A fungal DNA midi kit to extract genomic DNA and prepared 500-bp, 2-kb, and 8-kb fragments using an Illumina MiSeq benchtop sequencer. The sequence depths for each library were 186×, 94×, and 67×, respectively. Genome assembly using SOAPdenovo version 2.04 (<http://soap.genomics.org.cn>) resulted in 737 contigs, with an N_{50} value of 157,787 bp and a G+C content of 48.63%. We estimated the genome size of *P. sclerotiorum* to be about 35 Mb based on a 17 *k*-mer statistical analysis.

The assembled genome was annotated using the MAKER2 program (14). *P. sclerotiorum* 113 has an estimated 12,649 putative genes averaging 1,418 bp in length and comprising 52.7% of the whole genome. Repetitive sequences were 279,139 bp in total, constituting 0.82% of the genome. We compared the orthologous genes shared by *P. sclerotiorum* and five other *Penicillium* strains available in public databases (*P. griseofulvum* PG3, *P. expansum* R19, *P. expansum* ATCC 24692, *P. solitum* RS1, and *P. glabrum*

DAOM239074) using OrthoMCL. A total of 6,249 common orthologous genes were identified. *P. sclerotiorum* 113 contained the highest number of nonorthologous genes (2,205), followed by *P. glabrum* DAOM239074 (1,960), *P. expansum* ATCC 24692 (866), *P. solitum* RS1 (672), *P. griseofulvum* PG3 (509), and *P. expansum* R19 (326). This is the first report of the genome of *P. sclerotiorum* and its analysis. These data will be a foundation for improving our understanding of the distinctness of *P. sclerotiorum* from other members of the genus, its cosmopolitan distribution, and its biotechnological potential.

Accession number(s). The whole-genome sequence of *P. sclerotiorum* 113 has been deposited at DDBJ/ENA/GenBank under the accession number [MJCA01000000](https://www.ncbi.nlm.nih.gov/nuclink/MJCA01000000). The version described in this paper is the first version, MJCA01000000.

ACKNOWLEDGMENT

Use of a company or product name by the U.S. Department of Agriculture does not imply approval or recommendation of the product to the exclusion of others that may also be suitable.

FUNDING INFORMATION

This work was funded by the Special Fund for Agro-scientific Research in the Public Interest of the People's Republic of China (grant no. 201403075).

REFERENCES

1. Van Beyma J. 1937. *Penicillium sclerotiorum* nov. spec. *Centrblatt für Bakteriologie, Parastenkunde und Infektionskrankheiten* 96:481–491.
2. Rivera KG, Seifert KA. 2011. A taxonomic and phylogenetic revision of the *Penicillium sclerotiorum* complex. *Stud Mycol* 70:139–158. <http://dx.doi.org/10.3114/sim.2011.70.03>.
3. Pitt JI. 1979. The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. Academic Press, London.
4. Mase Y, Rabourn WJ, Quackenbush FW. 1957. Carotene production by *Penicillium sclerotiorum*. *Arch Biochem Biophys* 68:150–156. [http://dx.doi.org/10.1016/0003-9861\(57\)90335-1](http://dx.doi.org/10.1016/0003-9861(57)90335-1).

5. Knob A, Carmona EC. 2008. Xylanase production by *Penicillium sclerotiorum* and its characterization. *World Appl Sci J* 4:277–283.
6. Knob A, Carmona EC. 2010. Purification and characterization of two extracellular xylanases from *Penicillium sclerotiorum*: a novel acidophilic xylanase. *Appl Biochem Biotechnol* 162:429–443. <http://dx.doi.org/10.1007/s12010-009-8731-8>.
7. Curtin T, Reilly J. 1940. Sclerotiorin, a chlorinated metabolic product of *Penicillium sclerotiorum*, Van Beyma. *Nature* 3697:335.
8. Pairet L, Wrigley SK, Chetland I, Reynolds EE, Hayes MA, Holloway J, Ainsworth AM, Katzer W, Cheng XM, Hupe DJ, et al. 1995. Azapbilones with endothelin receptor binding activity produced by *Penicillium sclerotiorum*: taxonomy, fermentation, isolation, structure elucidation and biological activity. *J Antibiot* 48:913–923. <http://dx.doi.org/10.7164/antibiotics.48.913>.
9. Chidananda C, Rao LJ, Sattur AP. 2006. Sclerotiorin, from *Penicillium frequentans*, a potent inhibitor of aldose reductase. *Biotechnol Lett* 28: 1633–1636. <http://dx.doi.org/10.1007/s10529-006-9133-4>.
10. Lucas E, Machado Y, Ferreira A, Dolabella L, Takahashi J. 2010. Improved production of pharmacologically active sclerotiorin by *Penicillium sclerotiorum*. *Trop J Pharm Res* 9:365–371. <http://dx.doi.org/10.4314/tjpr.v9i4.58930>.
11. Wang Z-P, Wang G-Y, Khan I, Chi Z-M. 2013. High-level production of calcium malate from glucose by *Penicillium sclerotiorum* K302. *Bioresour Technol* 143:674–677. <http://dx.doi.org/10.1016/j.biortech.2013.06.051>.
12. Palou L, Taberner V, Guardado A, Del Río MÁ, Montesinos-Herrero C. 2013. Incidence and etiology of postharvest fungal diseases of pomegranate (*Punica granatum* cv. Mollar de Elche) in Spain. *Phytopathologia Mediterranea* 52:478–489. http://dx.doi.org/10.14601/Phytopathol_Mediterr-11581.
13. Zhao H, Yin G, Inamdar AA, Luo J, Zhang N, Yang I, Buckley B, Bennett JW. 17 Oct 2016. Volatile organic compounds emitted by filamentous fungi isolated from flooded homes after hurricane sandy show toxicity in a *Drosophila* bioassay. *Indoor Air* [Epub ahead of print.] <http://dx.doi.org/10.1111/ina.12350>.
14. Holt C, Yandell M. 2011. MAKER2: an annotation pipeline and genome-database management tool for second-generation genome projects. *BMC Bioinformatics* 12:491. <http://dx.doi.org/10.1186/1471-2105-12-491>.