

Draft Genome Sequences of *Campylobacter jejuni* Strains That Cause Abortion in Livestock

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***Campylobacter jejuni* is an intestinal bacterium that can cause abortion in livestock. This publication announces the public release of 15 *Campylobacter jejuni* genome sequences from isolates linked to abortion in livestock. These isolates are part of the 100K Pathogen Genome Project and are from clinical cases at the University of California (UC) Davis.**

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Campylobacter jejuni is transmissible between wildlife, livestock, and humans, often leading to foodborne illness in humans and disease burden among livestock (1–4). Globally, *C. jejuni* is a common foodborne pathogen (5) that infects over 1.3 million people each year in the United States (6), causing gastroenteritis; in rare cases, it may induce Guillain-Barré syndrome, an autoimmune disease (7–10). In addition to human infection, *C. jejuni* infects domesticated livestock, including sheep, cattle, goats, and pigs, most often leading to gastroenteritis in these species (11, 12).

Campylobacter spp., specifically, *C. fetus* subsp. *fetus*, are one of the leading causes of abortion in ungulates, characterized by late term abortion, stillbirths, and occasional ewe deaths (13). In recent years, many cases have recovered *C. jejuni* from aborted fetuses with similar disease pathologies (14, 15). Emergence of abortive hypervirulent *C. jejuni* isolates have been observed in various regions of the United States (14, 15). We sequenced 15 *C. jejuni* isolates associated with abortion in sheep, cows, and goats in northern California at an average coverage of 91×, assembled, and annotated (Table 1).

TABLE 1 Coverage and accession numbers of 15 abortive *C. jejuni* genomes

GenBank accession no.	SRA accession no.	Isolate name	Coverage (×)
MJZI00000000	SRR3619957	BCW_6919	93
MKAC00000000	SRR3619958	BCW_6920	91
MKAD00000000	SRR3619959	BCW_6921	79
MKAE00000000	SRR3619960	BCW_6922	100
MKAF00000000	SRR3619963	BCW_6924	90
MKAG00000000	SRR3619964	BCW_6925	84
MKAR00000000	SRR3619965	BCW_6926	84
MKAS00000000	SRR3619966	BCW_6927	80
MKAT00000000	SRR3619967	BCW_6928	50
MKAU00000000	SRR3619968	BCW_6929	76
MKAV00000000	SRR3619969	BCW_6930	101
MKIC00000000	SRR4020196	BCW_6931	111
MKID00000000	SRR4020197	BCW_6932	70
MKIB00000000	SRR4020198	BCW_6933	81
MKIA00000000	SRR4020199	BCW_6934	182

All *C. jejuni* isolates were cases from the University of California (UC) Davis California Animal Health and Food Safety Laboratory System (CAHFS) and sequenced by the 100K Pathogen Genome Project (<http://www.100kgenomes.org>) in the laboratory of Bart Weimer (UC Davis, Davis, CA). As described (16), isolates were checked for purity (17), genomic DNA (gDNA) was extracted from cultures grown on 5% blood agar plates (UC Davis, VetMed Biological Services) for 1 to 2 days, lysed (18), purified with Qiagen QIAamp DNA minikit, and analyzed on Agilent 2200 TapeStation system using the Genomic DNA ScreenTape assay for integrity of gDNA (19). Isolated gDNA was used to construct libraries using the Kapa HyperPlus kit (KR1145 version 3.16; Kapa Biosystems, Wilmington, MA, USA) with dual-SPRI size selection (20). Libraries were constructed using the PerkinElmer Sciclone next-generation sequencing (NGS) Workstation (PerkinElmer, Hopkinton, MA). Library quantitation was done using the Kapa SYBR Fast quantitative PCR (qPCR) kits (Kapa Biosystems) to ensure the starting concentration of 400 ng and a fragment insert size between 350 and 450 bp (20). Libraries were indexed using Integrated DNA Technologies Weimer 384 TS-LT DNA barcodes, which allowed multiplexing up to 384 isolates. Sequencing was done at the UC Davis Genome Center (Davis, CA, USA) on the HiSeq 3000 instrument using a paired-end 150 protocol (Illumina, Inc., San Diego, CA, USA) (21, 22). Paired-end reads were assembled using ABySS 1.5.2 using $k = 64$ (23).

Accession number(s). These sequences can be found in the 100K Project BioProject at the NCBI SRA BioProject PRJNA186441 and in the NCBI GenBank. Individual GenBank and SRA accession numbers are presented in Table 1.

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REFERENCES

- Weis AM, Miller WA, Byrne BA, Chouicha N, Boyce WM, Townsend AK. 2014. Prevalence and pathogenic potential of *Campylobacter* isolates from free-living, human-commensal American crows. *Appl Environ Microbiol* 80:1639–1644. <http://dx.doi.org/10.1128/AEM.03393-13>.
- Conor C, Taff AMW, Wheeler S, Hinton MG, Weimer BC, Barker CM, Jones M, Logsdon R, Smith WA, Boyce WM, Townsend AK. 2016. Influence of host ecology and behavior on *Campylobacter jejuni* prevalence and environmental contamination risk in a synanthropic wild bird. *Appl Environ Microbiol* 82. <http://dx.doi.org/10.1128/AEM.01456-16>.
- Gardner TJ, Fitzgerald C, Xavier C, Klein R, Pruckler J, Stroika S, McLaughlin JB. 2011. Outbreak of campylobacteriosis associated with consumption of raw peas. *Clin Infect Dis* 53:26–32. <http://dx.doi.org/10.1093/cid/cir249>.
- Straw BE. 1990. Effect of *Campylobacter* spp.-induced enteritis on growth rate and feed efficiency in pigs. *J Am Vet Med Assoc* 197:355–357.
- Coker AO, Isokpehi RD, Thomas BN, Amisu KO, Obi CL. 2002. Human campylobacteriosis in developing countries. *Emerg Infect Dis* 8:237–244. <http://dx.doi.org/10.3201/eid0803.010233>.
- CDC. 2012. Foodborne Diseases Active Surveillance Network: FoodNet surveillance report for 2011 (final report). U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, Atlanta, GA. http://www.cdc.gov/foodnet/pdfs/2011_annual_report_508c.pdf.
- Nachamkin I. 2003. *Campylobacter* and *Arcobacter*. In Murray PE, Baron EJ, Jorgensen JH, Phaller MA, Tenover FC, Manual of clinical microbiology, ASM Press, Washington, DC.
- Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, Jones JL, Griffin PM. 2011. Foodborne illness acquired in the United States—major pathogens. *Emerg Infect Dis* 17:7–15. <http://dx.doi.org/10.3201/eid1701.091101p1>.
- Altekruse SF, Stern NJ, Fields PI, Swerdlow DL. 1999. *Campylobacter jejuni*—an emerging foodborne pathogen. *Emerg Infect Dis* 5:28–35. <http://dx.doi.org/10.3201/eid0501.990104>.
- Young KT, Davis LM, DiRita VJ. 2007. *Campylobacter jejuni*: molecular biology and pathogenesis. *Nat Rev Microbiol* 5:665–679. <http://dx.doi.org/10.1038/nrmicro1718>.
- Nielsen EM. 2002. Occurrence and strain diversity of thermophilic *campylobacters* in cattle of different age groups in dairy herds. *Lett Appl Microbiol* 35:85–89. <http://dx.doi.org/10.1046/j.1472-765X.2002.01143.x>.
- Jacobs-Reitsma W, Lyhs U, Wagenaar J. 2008. *Campylobacter* in the food supply, p 627–644. In Nachamkin I, Szymanski C, Blaser M (ed), *Campylobacter*, 3rd ed. ASM Press, Washington, DC.
- Hedstrom OR, Sonn RJ, Lassen ED, Hultgren BD, Crisman RO, Smith BB, Snyder SP. 1987. Pathology of *Campylobacter jejuni* abortion in sheep. *Vet Pathol* 24:419–426.
- Sahin O, Fitzgerald C, Stroika S, Zhao S, Sippy RJ, Kwan P, Plummer PJ, Han J, Yaeger MJ, Zhang Q. 2012. Molecular evidence for zoonotic transmission of an emergent, highly pathogenic *Campylobacter jejuni* clone in the United States. *J Clin Microbiol* 50:680–687. <http://dx.doi.org/10.1128/JCM.06167-11>.
- Sahin O, Plummer PJ, Jordan DM, Sulaj K, Pereira S, Robbe-Austerman S, Wang L, Yaeger MJ, Hoffman LJ, Zhang Q. 2008. Emergence of a tetracycline-resistant *Campylobacter jejuni* clone associated with outbreaks of ovine abortion in the United States. *J Clin Microbiol* 46:1663–1671. <http://dx.doi.org/10.1128/JCM.00031-08>.
- Lüdeke CH, Kong N, Weimer BC, Fischer M, Jones JL. 2015. Complete genome sequences of a clinical isolate and an environmental isolate of *Vibrio parahaemolyticus*. *Genome Announc* 3(2):e00216-15. <http://dx.doi.org/10.1128/genomeA.00216-15>.
- Kong N, Ng W, Lee V, Kelly L, Weimer BC. 2013. Production and analysis of high molecular weight genomic DNA for NGS pipelines using Agilent DNA extraction kit (p/n 200600). (5991-3722EN; <http://dx.doi.org/10.13140/RG.2.1.2961.4807>) Agilent Technologies, Santa Clara, CA.
- Jeannotte R, Lee E, Kong N, Ng W, Kelly L, Weimer BC. 2014. High-throughput analysis of foodborne bacterial genomic DNA using Agilent 2200 TapeStation and genomic DNA ScreenTape system. (5991-4003EN; <http://dx.doi.org/10.13140/RG.2.1.3354.6961>) Agilent Technologies, Santa Clara, CA.
- Kong N, Ng W, Cai L, Leonardo A, Kelly L, Weimer BC. 2014. Integrating the DNA integrity number (DIN) to assess genomic DNA (gDNA) quality control using the Agilent 2200 TapeStation system. (5991-5442EN; <http://dx.doi.org/10.13140/RG.2.1.3616.8409>) Agilent Technologies, Santa Clara, CA.
- Kong N, Ng W, Foutouhi A, Huang BH, Kelly L, Weimer BC. 2014. Quality control of high-throughput library construction pipeline for Kapa HTP library using an Agilent 2200 TapeStation. (5991-5141EN; <http://dx.doi.org/10.13140/RG.2.1.4927.5604>) Agilent Technologies, Santa Clara, CA.
- Kong N, Thao K, Huang C, Appel M, Lappin S, Knapp L, Kelly L, Weimer BC. 2014. Automated library construction using Kapa library preparation kits on the Agilent NGS workstation yields high-quality libraries for whole-genome sequencing on the Illumina platform. (5991-4296EN; <http://dx.doi.org/10.13140/RG.2.1.2306.1203>) Agilent Technologies, Santa Clara, CA.
- Miller B, Kets VV, Rooyen BV, Whitehorn H, Jones P, Ranik M, Geldart A, Walt EVD, Appel M, Kong N, Huang BH, Storey D, Weimer BC. 2015. A novel, single tube enzymatic fragmentation and library construction method enables fast turnaround times and improved data quality for microbial whole-genome sequencing. Kapa Biosystems, Wilmington, MA. https://www.kapabiosystems.com/assets/App_Note_HyperPlus_APP109001-v1.15_low.pdf.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJM, Birol I. 2009. ABySS: A parallel assembler for short read sequence data. *Genome Res* 19:1117–1123.