

INTRODUCTION

BRCA to the future: towards best testing practice in the era of personalised healthcare

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European Journal of Human Genetics (2016) 24, S1–S2; doi:10.1038/ejhg.2016.92

The year 2014 marked the twentieth anniversary of the discovery of the breast cancer susceptibility gene, *BRCA1*.^{1,2} Since this discovery, our understanding of pathogenic *BRCA* variants and the associated increase in lifetime cancer risks has advanced significantly.^{3,4} National guidelines have been developed to help clinicians identify patients with an increased risk of pathogenic *BRCA* mutations, and genetic counselling for risk assessment is now a routine practice.^{5,6} The evolution of genetic counselling models has placed increasing importance on the use of multidisciplinary health-care teams, including medical geneticists, genetic counsellors, gynaecologists, surgeons, radiotherapists, medical oncologists and all other professionals involved in a patient's management. There has also been a surge of technological advances and new strategies allowing for the rapid turnaround of *BRCA1* and *BRCA2* mutation testing. In addition, evaluation of *BRCA* and other *BRCA*-like deleterious mutations as potentially actionable tumour targets are underway, such as the development of poly(ADP)-ribose polymerase (PARP) inhibitors in classes four or five *BRCA* mutation-positive cancers.^{7–9} The present supplement contains review articles based on presentations from a scientific symposium focused on the evolving *BRCA* testing landscape (*BRCA to the future: towards best testing practice in the era of personalised healthcare*) held at the European Human Genetics Conference in Milan, June 2014.

In the first review, Professor Dominique Stoppa-Lyonnet discusses the biological effects and clinical implications of pathogenic *BRCA* mutations. *BRCA1* and *BRCA2* have key roles in the repair of DNA double-strand breaks via the mechanism of homologous recombination (HR) and ensure genome stability.^{10–13} Germline deleterious mutations in *BRCA1* or *BRCA2* are associated with elevated risk of developing breast and/or ovarian cancer, sometimes with a genotype–phenotype correlation.^{3,4,14} *BRCA* pathogenic mutation-positive ovarian or breast cancers are susceptible to inhibitors of additional DNA damage repair pathways, such as PARP inhibitors.^{7–9} As the presence of actionable *BRCA* mutations in patients with breast and/or ovarian cancer becomes increasingly important in clinical management decisions, there is increasing support to expand the screening criteria for *BRCA* mutation testing, including testing for pathogenic mutations in other HR-deficiency genes.

In his review on *BRCA* testing, Dr Andrew Wallace covers the challenges of *BRCA* testing from the perspective of a diagnostic laboratory. Demand for *BRCA* testing is steadily increasing, placing a strain on diagnostics laboratories, particularly in those offering rapid genetic testing at the point of diagnosis. Increasingly, diagnostic

laboratories are adopting next-generation sequencing (NGS) technology for *BRCA* testing, which offers the potential of fast, scalable, cost-efficient and comprehensive sequencing.^{15–19} However, the choice and complexity surrounding NGS means that adopting this technology into the diagnostic laboratory requires many considerations, such as selection of an NGS platform, enrichment methods, sequencing chemistries, analytical procedures and techniques for somatic and germline mutation discovery. Interpreting *BRCA* test results can also be a complex process, involving the assessment of variants of unknown clinical significance, ensuring laboratory results are consistent and clearly reported, and understanding the role of passenger somatic mutations. *BRCA* testing of formalin-fixed paraffin-embedded tumour samples also presents a number of additional challenges to the diagnostics laboratory, including limited quantity and quality of extracted DNA, and artefactual sequence alterations.^{20–23}

Genetic counselling is integral to the *BRCA* testing process. In the final review, Professor Nicoline Hoogerbrugge and Dr Marjolijn Jongmans examine best practices for genetic counselling and *BRCA* testing, exploring the challenges facing current practice and looking at adapted models of genetic counselling. Current practice in many countries requires face-to-face counselling with a qualified genetics counsellor both prior to, and following, *BRCA* testing.^{5,6} As demand for genetic testing increases, new approaches for delivering genetic counselling may be necessary. It is possible that future *BRCA* testing and counselling models will incorporate different elements from these new approaches in order to optimise workflow in the era of personalised care. Examples include conserving germline *BRCA* testing and associated genetic counselling to those patients with a positive tumour pathogenic *BRCA* mutation result, enabling trained oncologists and medical specialists/staff, in addition to genetic counsellors, to manage pre-test genetic consultations with patients²⁴ and accounting for patient preferences regarding counselling and information such as telephone consultations with additional educational resources for selected patients.²⁵

I greatly appreciate the collaboration of Professor Stoppa-Lyonnet, Dr Andrew Wallace, Professor Nicoline Hoogerbrugge and Dr Marjolijn Jongmans for an engaging symposium and subsequent review articles. We hope that this supplement provides you with a comprehensive update on the practical, prognostic and clinical implications of the evolving *BRCA* testing landscape.

CONFLICT OF INTEREST

EC has received consulting fees, lecture fees and grant support from AstraZeneca.

ACKNOWLEDGEMENTS

The supplement was sponsored by AstraZeneca. Medical writing services were provided by Tom Hudson of iMed Comms, Macclesfield, UK and were funded by AstraZeneca.

- 1 Friedman LS, Ostermeyer EA, Szabo CI *et al*: Confirmation of BRCA1 by analysis of germline mutations linked to breast and ovarian cancer in ten families. *Nat Genet* 1994; **8**: 399–404.
- 2 Miki Y, Swensen J, Shattuck-Eidens D *et al*: A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. *Science* 1994; **266**: 66–71.
- 3 Antoniou A, Pharoah PD, Narod S *et al*: Average risks of breast and ovarian cancer associated with BRCA1 or BRCA2 mutations detected in case series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet* 2003; **72**: 1117–1130.
- 4 Chen S, Parmigiani G: Meta-analysis of BRCA1 and BRCA2 penetrance. *J Clin Oncol* 2007; **25**: 1329–1333.
- 5 Gadzicki D, Evans DG, Harris H *et al*: Genetic testing for familial/hereditary breast cancer-comparison of guidelines and recommendations from the UK, France, the Netherlands and Germany. *J Community Genet* 2011; **2**: 53–69.
- 6 NICE Clinical Guidelines: NICE Clinical Guidance 164: Familial Breast Cancer. *NICE Clinical Guidelines* 2013. Available at <https://www.nice.org.uk/guidance/cg164>.
- 7 Audeh MW, Carmichael J, Penson RT *et al*: Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet* 2010; **376**: 245–251.
- 8 Ledermann J, Harter P, Gourley C *et al*: Olaparib maintenance therapy in patients with platinum-sensitive relapsed serous ovarian cancer: a preplanned retrospective analysis of outcomes by BRCA status in a randomised phase 2 trial. *Lancet Oncol* 2014; **15**: 852–861.
- 9 Tutt A, Robson M, Garber JE *et al*: Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *Lancet* 2010; **376**: 235–244.
- 10 Collins N, McManus R, Wooster R *et al*: Consistent loss of the wild type allele in breast cancers from a family linked to the BRCA2 gene on chromosome 13q12-13. *Oncogene* 1995; **10**: 1673–1675.
- 11 Gudmundsdottir K, Ashworth A: The roles of BRCA1 and BRCA2 and associated proteins in the maintenance of genomic stability. *Oncogene* 2006; **25**: 5864–5874.
- 12 Mao Z, Bozzella M, Seluanov A, Gorbunova V: DNA repair by nonhomologous end joining and homologous recombination during cell cycle in human cells. *Cell Cycle* 2008; **7**: 2902–2906.
- 13 Venkitaraman AR: Functions of BRCA1 and BRCA2 in the biological response to DNA damage. *J Cell Sci* 2001; **114**: 3591–3598.
- 14 Rebbeck TR, Mitra N, Wan F *et al*: Association of type and location of BRCA1 and BRCA2 mutations with risk of breast and ovarian cancer. *JAMA* 2015; **313**: 1347–1361.
- 15 Grada A, Weinbrecht K: Next-generation sequencing: methodology and application. *J Invest Dermatol* 2013; **133**: e11.
- 16 Idris SF, Ahmad SS, Scott MA, Vassiliou GS, Hadfield J: The role of high-throughput technologies in clinical cancer genomics. *Expert Rev Mol Diagn* 2013; **13**: 167–181.
- 17 Loman NJ, Misra RV, Dallman TJ *et al*: Performance comparison of benchtop high-throughput sequencing platforms. *Nat Biotechnol* 2012; **30**: 434–439.
- 18 Mardis ER: Next-generation DNA sequencing methods. *Annu Rev Genomics Hum Genet* 2008; **9**: 387–402.
- 19 Walsh T, Lee MK, Casadei S *et al*: Detection of inherited mutations for breast and ovarian cancer using genomic capture and massively parallel sequencing. *Proc Natl Acad Sci USA* 2010; **107**: 12629–12633.
- 20 Do H, Dobrovic A: Dramatic reduction of sequence artefacts from DNA isolated from formalin-fixed cancer biopsies by treatment with uracil- DNA glycosylase. *Oncotarget* 2012; **3**: 546–558.
- 21 Do H, Wong SQ, Li J, Dobrovic A: Reducing sequence artifacts in amplicon-based massively parallel sequencing of formalin-fixed paraffin-embedded DNA by enzymatic depletion of uracil-containing templates. *Clin Chem* 2013; **59**: 1376–1383.
- 22 Wong SQ, Li J, Tan AY *et al*: Sequence artefacts in a prospective series of formalin-fixed tumours tested for mutations in hotspot regions by massively parallel sequencing. *BMC Med Genomics* 2014; **7**: 23.
- 23 Minucci A, Scambia G, Santonocito C *et al*: Clinical impact on ovarian cancer patients of massive parallel sequencing for BRCA mutation detection: the experience at Gemelli hospital and a literature review. *Expert Rev Mol Diagn* 2015; **15**: 1383–1403.
- 24 George A, Smith F, Cloke V *et al*: Implementation of routine BRCA gene testing of ovarian cancer (OC) patients at Royal Marsden Hospital. *Ann Oncol* 2014; **25** (Suppl 4): iv307.
- 25 Sie AS, van Zelst-Stams WA, Spruijt L *et al*: More breast cancer patients prefer BRCA-mutation testing without prior face-to-face genetic counseling. *Fam Cancer* 2014; **13**: 143–151.