



REVIEW

The interplay between plasticity and evolution in response to human-induced environmental change [version 1; referees: 2 approved]

Sarah E. Diamond, Ryan A. Martin

Department of Biology, Case Western Reserve University, Cleveland, OH, USA

V1 **First published:** 08 Dec 2016, 5(F1000 Faculty Rev):2835 (doi: 10.12688/f1000research.9731.1)
Latest published: 08 Dec 2016, 5(F1000 Faculty Rev):2835 (doi: 10.12688/f1000research.9731.1)

Abstract

Some populations will cope with human-induced environmental change, and others will undergo extirpation; understanding the mechanisms that underlie these responses is key to forecasting responses to environmental change. In cases where organisms cannot disperse to track suitable habitats, plastic and evolved responses to environmental change will determine whether populations persist or perish. However, the majority of studies consider plasticity and evolution in isolation when in fact plasticity can shape evolution and plasticity itself can evolve. In particular, whether cryptic genetic variation exposed by environmental novelty can facilitate adaptive evolution has been a source of controversy and debate in the literature and has received even less attention in the context of human-induced environmental change. However, given that many studies indicate organisms will be unable to keep pace with environmental change, we need to understand how often and the degree to which plasticity can facilitate adaptive evolutionary change under novel environmental conditions.

Open Peer Review

Referee Status: ✓ ✓

| | Invited Referees | |
|--|------------------|---|
| | 1 | 2 |
| version 1 published 08 Dec 2016 | ✓ | ✓ |

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty. In order to make these reviews as comprehensive and accessible as possible, peer review takes place before publication; the referees are listed below, but their reports are not formally published.

1 Lauren Buckley, University of Washington USA

2 Robby Stoks, KU Leuven Belgium

Discuss this article

Comments (0)

Corresponding authors: Sarah E. Diamond (sarah.diamond@case.edu), Ryan A. Martin (ram225@case.edu)

How to cite this article: Diamond SE and Martin RA. **The interplay between plasticity and evolution in response to human-induced environmental change [version 1; referees: 2 approved]** *F1000Research* 2016, 5(F1000 Faculty Rev):2835 (doi: [10.12688/f1000research.9731.1](https://doi.org/10.12688/f1000research.9731.1))

Copyright: © 2016 Diamond SE and Martin RA. This is an open access article distributed under the terms of the [Creative Commons Attribution Licence](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Grant information: The author(s) declared that no grants were involved in supporting this work.

Competing interests: The authors declare that they have no competing interests.

First published: 08 Dec 2016, 5(F1000 Faculty Rev):2835 (doi: [10.12688/f1000research.9731.1](https://doi.org/10.12688/f1000research.9731.1))

Introduction and context

In the absence of sufficient compensatory mechanisms to cope with human-induced environmental changes, local extirpation of populations and possibly extinction of entire lineages will almost certainly occur¹. Quantifying organismal capacities for such compensatory mechanisms is therefore paramount to forecasting responses to ongoing and future human-induced changes to the environment². Of particular importance is quantifying the relative contributions of phenotypic plasticity, where a single genotype produces a range of different phenotypes in response to environmental variation, and evolution, where additive genetic differences underlie phenotypic change across generations or among populations. This distinction is critical, as plasticity and evolution often occur over different timescales and can operate under different constraints³.

Certainly much progress has been made over the last few decades in disentangling plastic from evolved changes in phenotypes under human-induced environmental change^{3–12}. Yet the distinction of plastic versus evolutionary contributions in a given population at a given time may be an oversimplification, as these mechanisms can interact to shape phenotypic changes in populations over time⁶. Basic research on evolution has long debated whether plasticity facilitates or constrains adaptive evolution¹³. However, this idea has only recently come into focus for theory^{14,15} and empirical work^{16,17} involving responses to human-induced environmental change. Here, we briefly review the interplay between plastic and evolutionary responses to human-induced environmental change and highlight new research areas for future development on the role of plasticity in shaping evolutionary responses to changes in climate, land-use, and environmental toxins. While we consider these environmental stressors separately for organizational purposes in this review, we acknowledge that many of these factors are not mutually exclusive, e.g. land-use changes can often include changes in environmental toxicity.

Phenotypic plasticity as a facilitator versus constraint on evolution

Evolutionary theory predicts that natural selection on environmentally insensitive (canalized) traits imposed by novel environments will drive populations toward their new local fitness optima⁴, yet evolution in this sense might not achieve large enough shifts in trait values or be fast enough to keep pace with the scale and rapidity of human-induced changes to the environment before populations undergo extirpation. Limits on the amount of standing genetic variation in populations, mutation rates, fitness tradeoffs, the strength and consistency of selection, and the genetic correlations among traits can all serve to slow the rate of evolutionary change⁵.

Likewise, there are limits on shifts in trait values owing to existing phenotypic plasticity. Most traits exhibit phenotypic plasticity. However, plasticity may simply fail to produce shifts in phenotypes of a great enough magnitude to cope with rapid environmental change¹⁸. There is also no guarantee that plasticity will be adaptive for the novel conditions generated by human-induced environmental change¹⁹. Whether plasticity is adaptive or not depends on whether the environmentally induced phenotypes are closer or farther from the local optimum. Because novel environments are, by definition, environments that organisms have

never experienced previously, it is entirely plausible that adaptive plasticity in current environments will be maladaptive in the novel environment^{20,21}. This prediction should, however, be tempered to some degree, as novelty can arise not only through environments not previously experienced but also through alterations to the frequency of environmental conditions experienced (e.g. 22); in the latter case, organisms may be more likely to evolve adaptive plasticity, as they are responding to frequency differences rather than absolute differences in their environment.

Given that existing plasticity and evolution may on their own be insufficient to cope with environmental change, there has been renewed interest in the case where novel environments reveal cryptic genetic variation on which selection can act to refine the form and regulation of this novel phenotypic variation (Figure 1)²³. Plasticity may be a key source of evolutionary innovation, allowing organisms to cope with the large magnitude and rapid rate of human-induced environmental changes (the plasticity-first mechanism of evolutionary change; see 13). Indeed, plasticity may be an especially efficient source of variation, as the effects of environmental novelty can reach each individual of the population, unlike mutations, which arise in a single individual and are then likely to disappear²⁴. Despite the potential importance of the plasticity-first mechanism in the context of human-induced environmental change, there are relatively few empirical data that address its components, coupled with abundant controversy in the literature over plasticity's role in shaping evolutionary responses to environmental novelty.

Evidence is accumulating in support of the hypothesis that novel environments reveal cryptic genetic variation on which selection can act²⁵. Recent meta-analyses suggest that novel environments considered broadly tend to reveal cryptic genetic variation²⁶, as do those that focus specifically on human-induced novel environments²⁷. However, most of this human-induced environmental novelty involves variation in resource quality and quantity, representing a small subset of the environmental novelty induced by climate change, land-use change, and environmental toxins that are the focus of this review. As a consequence, while there is evidence that human-induced environmental change can reveal cryptic genetic variation, a broader range of environmental changes needs to be examined to establish the generality of this pattern. Nevertheless, laboratory selection experiments have shown that this uncovered plastic variation can facilitate evolutionary change via genetic accommodation (Box 1), which describes the evolution of environmentally induced phenotypes, including the evolution of canalized traits from initially plastic traits through the process of genetic assimilation^{23,28,29}. Despite this, considerably fewer studies have tested the relationship between initial plasticity and subsequent evolution in natural populations (but see for example 30,31).

Human-induced novel environments are excellent sources to look for the exposure of cryptic genetic variation and to explore plasticity's role in shaping evolution, but comparatively few studies have taken advantage of these opportunities³¹. On the one hand, human-induced changes to the environment yield an imperative to understand plastic and evolutionary responses to new environments for the goal of biodiversity forecasting and conservation

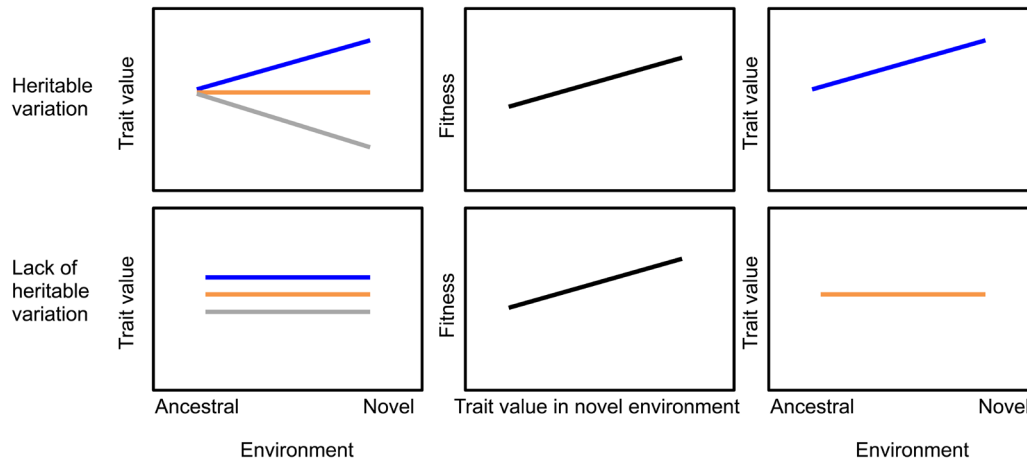


Figure 1. Adaptive trait evolution in a novel environment. When a population encounters a novel environment (for example, novel urban and ancestral rural habitats), it can either express hidden, habitat-dependent heritable variation in a given trait (top row) or not (bottom row). In the case where this variation is expressed, selection (middle column panels) can refine this phenotypically expressed genetic variation into an optimal canalized trait. In the absence of habitat-dependent genetic variation (and assuming a lack of novel mutation), the population cannot evolve towards the new trait optimum.

planning, but on the other hand, these often-rapid human-induced environmental changes may help overcome historical challenges in assessing plasticity's role in evolutionary change. Specifically, many of the comparisons between ancestral and novel populations represent divergence processes that are relatively old³². Rapid human-induced environmental changes provide access to the early stages of plastic and evolved responses to new environments. Human-induced environmental novelty may also be more straightforward for attribution—that is, determining the proximate drivers of changes in the environment—as compared with historical comparisons between ancestral and novel populations where attribution is perhaps less clear and there is more time for suites of different environmental changes to accumulate.

In this review, we discuss recent work on the interplay between plastic and evolved responses to human-induced environmental change. Specifically, we consider whether plasticity creates novel opportunities for selection to act, thereby facilitating evolution, or whether plasticity buffers environmental variation, dampening selection and constraining evolution (Box 1). Because we are covering a diverse array of changes from global climate change to environmental toxins with the aim of stimulating discussion on plastic and evolutionary responses to these stressors, we take a case study approach rather than an exhaustive review, highlighting recent and transformative work in the field.

Existing plasticity, evolutionary potential, and the role of plasticity in shaping evolutionary responses to human-induced environmental change

Climate change

Humans have made lasting impacts on the environment for thousands of years, from megafaunal extinctions during the late Quaternary period³³ to rapid industrialization and global temperature rise within the last century³⁴. The magnitude of recent human-induced changes, particularly to climate, has already led to the global redistribution of plants and animals in space and time^{35,36}.

Box 1. Plasticity's role in shaping evolution.

How plasticity can constrain evolution:

- If plasticity in novel environments results in high mean fitness (i.e. adaptive plasticity), then plasticity can weaken subsequent selection in the novel environment and hide genotypic variation from selection⁸⁶. The occurrence of adaptive plasticity in novel environments is likely to occur only when the novel environment is similar to native or ancestral environments and therefore a product of past selection⁸⁷.
- Plasticity may drive a population to extirpation in novel environments before adaptive evolution can occur if plasticity results in very low mean fitness (i.e. maladaptive plasticity), for example through developmental instability or a breakdown in homeostasis^{86,87}.

How plasticity can facilitate evolution:

- Plasticity may facilitate evolution in novel environments when it acts to buffer populations from extirpation long enough for selection to act on standing or cryptic genetic variation^{86,87}.
- Gene by environment interactions may reveal **cryptic genetic variation** in novel environments when environmental variation falls outside the range generally experienced, exposing heritable phenotypes that were not expressed in the ancestral environment to selection. Since the speed of evolution is in part dependent on the degree of genetic variation underlying phenotypic traits under selection, evolution may proceed more rapidly when novel plasticity is revealed^{24,88,89}.

How plasticity itself can evolve:

- Plasticity can evolve when selection acts on genetic variation in the phenotypic expression of individual genotypes across environments (i.e. reaction norms). This process has been named **genetic accommodation**²⁴. Under this broad umbrella, selection may act on genetic variation to change the slope of the reaction norm or the mean of a trait expressed across environments³⁰. Selection to canalize the expression of a novel environmentally induced phenotype has been specifically termed **genetic assimilation**^{38,91,92}, and its reverse, the restoration of the ancestral phenotype in a novel environment, has been termed **genetic compensation**²⁰.

For those organisms that cannot get out of the way of climate change, they must rely instead on plastic and evolved responses. A major focus has been on whether current amounts of phenotypic plasticity and evolutionary potential are sufficient to keep pace with projected climate velocities^{37,38}. Of course, there are many effects of contemporary global climate change—from atmospheric deposition of elemental compounds to shifts in precipitation patterns—but temperature is perhaps the best studied and one that impacts organisms from biochemical rate processes up through species interactions, community dynamics, and ecosystem function³⁹.

By the end of the century, global temperature is expected to rise by over 4°C under the highest greenhouse gas emissions scenarios, which we are currently tracking³⁴. Studies of organismal capacities to cope with this change, either through plasticity or evolution, yield mixed evidence for whether organismal changes will be large enough and proceed fast enough to keep pace with climate change^{18,38,40,41}. In general, there has been a strong research emphasis on existing plasticity in phenology, morphology, physiology, and behavior, perhaps because of the assumption that plasticity will operate over a timetable commensurate with rapid global climate change and perhaps also stemming from the added challenges of assessing evolutionary responses⁴². This emphasis is not without merit, as plasticity has been broadly shown to be a critical component of the response to climate change⁴³. However, existing plasticity is not always sufficient to cope with climate change. For example, Anderson and colleagues found a substantial role for phenological plasticity in plant flowering time under recent climate change but concluded that plasticity alone is insufficient and evolutionary change is needed for these populations to keep pace⁴⁴. While such variance partitioning studies of responses to climate change among plastic and evolved components is growing, there remain comparatively few studies assessing how this plasticity shapes evolutionary responses³.

Those studies that have explored the role of plasticity in facilitating adaptive evolutionary responses to climate change have found little evidence in support of this mechanism, though this may be as much a product of the organisms and systems for which these data are available as it is a general biological pattern. For example, research on lizard responses to climate change from Buckley and colleagues indicates that plasticity in thermoregulatory behavior constrains lizard evolutionary responses to warming by shielding variation in thermal performance from selection⁴⁵. As a notable contrast, Logan and colleagues found evidence of selection on thermal performance traits when lizards were transplanted to a novel thermal environment⁴⁶, but plasticity's role in facilitating or constraining evolutionary change was not reported. It is worthwhile to consider why there are so few studies on genetic accommodation under climate change. One important limitation may be how the ancestral and novel populations are assessed under a temporal change in climate. Resurrection studies or studies which compare the leading edge of a rapid range expansion to the center range seem to be strong candidates for such an analysis⁴⁷. Next, we consider two environmental changes—land-use and pollution—where the ancestral and novel populations are readily accessible.

Land-use change

Although there are many types of land-use changes occurring, including deforestation and habitat fragmentation, to which organisms respond through plasticity and evolutionary change⁴⁸, urbanization is an increasingly important source of land-use change. Rates of urbanization are accelerating globally, with current levels of urbanization at three percent of the Earth's landmass, excluding Greenland and Antarctica⁴⁹. This may not seem like an exceptionally large number, but over half of the world's population lives within these urbanized areas, and pockets of urbanization dot almost every corner of the globe⁵⁰. Although studies have begun to quantify community composition and phenotypic changes in populations across urban and nearby rural habitats, few have explored the mechanisms that contribute to these changes. Because the urbanization process offers both novel urban habitats in close spatiotemporal proximity to ancestral rural habitats and the potential for replication across independent ancestral-novel comparisons^{51,52}, urbanization gradients are excellent, but underused, sources of ancestral-novel comparisons needed for assessing plasticity's role in shaping evolutionary change. Of particular interest is the rapid change in environmental temperature over space and time under the urbanization process, especially as urban warming can serve as a space-for-time proxy to understand responses to global climate change. Urban heat island effects are relatively consistent across many regions, with temperature increases in excess of several °C in the air column and over 10°C at the surface being possible (though there are exceptions, especially urbanization in desert habitats⁵³). Though, of course, many other changes accompany the urbanization process, including changes in habitat structure, nutrient availability, and pollution⁵⁴.

In general, studies that explore plastic and evolutionary responses to urbanization have lagged behind climate change studies. This is surprising, since one of the best-known and earliest examples of rapid evolutionary change, industrial melanism in peppered moths, was itself driven by the fast progression of urbanization (and environmental pollution) associated with the industrial revolution^{55,56}. More recently, Donihue and Lambert⁵⁴ provided an excellent overview of how to assess and disentangle plastic and evolved responses to urban land-use change in addition to an exhaustive review of the studies that have done so in urban environments. They defined three criteria for demonstrating adaptive evolutionary change in response to urbanization, including quantifying phenotypic shifts in traits, measuring the fitness effects and genetic basis of those traits, and identifying drivers of trait changes. Only two studies met these criteria at the time of the review, including one study on killifish responses to elevated polychlorinated biphenyl pollution in urban environments⁵⁷ and another on the reduction of dispersing versus non-dispersing plant seeds in urban environments⁵⁸. Notably, the killifish study highlights the fact that the environmental stressors we consider in our review—climate change, land-use change, and environmental toxins—are not mutually exclusive, as, for example, killifish responses to polychlorinated biphenyl compounds may be considered in context of both land-use change and environmental toxicity. Relatedly, structural changes in urban environments, specifically replacing grassland and forest with impervious surfaces like roads,

sidewalks, and buildings, contributes to the urban heat island effect; as the study on plant seed production demonstrated, these structural changes can also serve as an agent of selection themselves. Dispersing seed pods are selected against in urban environments because there are large gaps in suitable habitat owing to intervening sidewalks and roads⁵⁸.

Evidence consistent with an interpretation of urban evolution continues to accumulate. For example, Winchell and colleagues⁵⁹ have found phenotypic shifts in lizard morphology in urban areas where lizards use artificial surfaces as perches as opposed to vegetation in unaltered habitats (an interesting parallel with the structural changes in urban habitats altering plant seed dispersal); a common garden experiment suggested a genetic basis for these changes in morphology, though the adaptive nature of these changes has yet to be demonstrated, and maternal effects cannot be fully excluded with their use of first-generation offspring from field-collected parents. Interestingly, evolutionary responses to urban heat islands appear to be one of the least-studied aspects of the urbanization process despite the ubiquity of the urban heat island signal among cities. Angilletta and colleagues⁶⁰ demonstrated an increase in upper thermal tolerance in city ants compared with rural ants, though it is unclear whether this shift is adaptive and has a genetic basis. McLean and colleagues used a common garden experiment to demonstrate a genetic basis for shifts in growth rate of a chitinolytic fungus along an urbanization gradient, though, again, the adaptive nature of these changes is unclear, and only one urban-rural comparison was made⁶¹. Although urban evolution is gaining traction in the literature, the use of urban environments to assess the role of plasticity in shaping evolutionary responses has been used only to a marginal extent and remains open for future development. In particular, replicated urban-rural comparisons with multigenerational common garden studies are needed to explore the potential for urban evolution and its interplay with plasticity.

Environmental toxins

In conjunction with increasing CO₂ emissions and changes in land-use, more than a century of industrialization has led to the accumulation of toxins in the environment, notably heavy metal and pesticide contaminants⁶². Environmental toxins can have severe negative consequences for organismal fitness and population survival through a diverse range of effects and mechanisms of action⁶³. Despite this diversity and in contrast to urbanization and climate change, the evolution of resistance to environmental toxins is well established⁶⁴⁻⁶⁶: for instance, the classic example of divergent evolution and speciation of plant populations living in and adjacent to toxic mine tailings⁶⁷⁻⁷⁰.

Phenotypic plasticity also contributes to organismal responses to environmental toxins, as prior exposure to heavy metals⁷¹ and pesticides⁷² can reduce the negative effects of later exposure. Interestingly, exposure to one toxin may induce a plastic response with positive fitness effects (i.e. an adaptation) to other toxins^{73,74}, but the generality of this inducible “cross-tolerance” is currently unknown. This shared adaptive plasticity does not appear to be restricted to toxins with common mechanisms of action, contrary to expectations⁷³. Indeed, this result suggests an important role of plasticity in the evolution of toxin tolerance, as cross-tolerance

would induce initially adaptive rather than maladaptive plasticity in response to novel toxins, possibly facilitating subsequent evolutionary refinement.

Like urbanization, the spatiotemporal heterogeneity of environmental toxins provides an opportunity to evaluate the role of plasticity in shaping evolution. Indeed, comparisons among populations differing in their exposure to pesticides provide some of our best evidence that genetic assimilation plays a role in evolutionary responses to anthropogenic change. Historically, studies focused on the evolution of tolerance in pest species directly targeted by pesticide application. A recent emphasis on non-pest species incidentally affected by pesticides has provided novel insights into the mechanisms underlying pesticide tolerance. By comparing pesticide tolerance among naïve populations to populations exposed to pesticides in contaminated environments (e.g. populations close to agriculture), numerous studies have now found evidence for the evolution of greater resistance to pesticides in affected populations of anurans and aquatic invertebrates compared to naïve populations^{73,75-83}.

Using an agricultural land-use gradient in a space-for-time substitution, Hua and colleagues recently extended these findings to test key predictions of genetic assimilation (**Box 1**): the canalization of inducible phenotypic variation by the action of natural selection. By quantifying both evolved and inducible (i.e. plastic) tolerances of wood frog populations varying in their distance from agriculture to an insecticide, Hua and colleagues found that evolved tolerance to a potentially lethal exposure of insecticide decreased with distance from agriculture^{76,79}. In contrast, the degree of plastic tolerance induced from an initial sublethal exposure increased with distance from agriculture⁷⁶. Together, these results are consistent with the predictions of genetic assimilation; exposure to novel environments induced the expression of phenotypic variation in ancestral populations, and this variation was then canalized in populations with a history of exposure to the novel environment^{28,29}. We emphasize that while these studies provide indirect evidence of genetic assimilation, more direct tests are needed. To accomplish this goal, resurrection studies and the study of extant populations varying in their time of exposure to novel environments may provide powerful means of testing the role of plasticity and evolution for responses to human-induced environmental change.

Ways forward: improving our understanding of plastic and evolutionary responses to a rapidly changing world

That plasticity is a powerful mechanism for allowing organisms to cope with rapid human-induced environmental change has widespread empirical support. Evidence in support of rapid evolution in response to environmental change is less abundant³ but accumulating. Although quantifying organismal capacities to respond to environmental change through plasticity and evolution is and should continue to be a major research priority, a key shortcoming in this area is a limited quantity of empirical data on the interaction of these mechanisms, specifically the role of plasticity in shaping evolutionary responses to environmental change³¹. Climate change, land-use change, and environmental toxins provide conservation and biodiversity forecasting challenges; however, these agents of environmental change also provide opportunities

to explore the potential for novel environments to release cryptic genetic variation and how selection might act on that variation to enact evolutionary change. To move forward, we suggest researchers should especially focus on (i) quantifying the expression of cryptic genetic variation in the wild, (ii) assessing patterns of selection on cryptic genetic variation in natural populations, and (iii) quantifying the strength and direction of evolutionary versus plastic responses to novel environments. An especially powerful approach may be assessing evolutionary and plastic responses across a gradient of time rather than space. By identifying and studying populations exposed to novel selective environments for different lengths of time using multi-generational common garden designs, we can directly assess the mechanisms underlying these responses. Human-mediated changes via urbanization and environmental toxins may be especially amenable to such an approach where historical records can be harnessed to calibrate temporal variation in the selective environment.

In addition to the issues with plastic and evolutionary responses to environmental change that we considered here, several related areas are also seeing further developments. One of these areas concerns the covariation between selection and genetic variation when both are affected by the same environmental factor^{27,84,85}. A major assumption underlying the genetic accommodation mechanism is that there will be a positive relationship between the degree of genetic variation revealed and the strength of natural selection in novel environments. However, this outcome is not guaranteed, and it is possible that genetic variance could be lower for the targets of selection when selection is stronger; in practical terms, this means that evolution could be slower, as trait heritability (the ratio of additive genetic variance to total phenotypic variance) would be lower in the novel environment despite stronger selection. There have been very few empirical examinations of the nature of the covariation between the amount of variation exposed

and the strength of selection in novel environments. Wood and Brodie have examined this relationship with several agents of selection, including variation in environmental temperature²⁷. Interestingly, while the authors found the strength of selection to vary with temperature, this variation was not correlated with genetic variance, as greater and lesser values of genetic variance were detected with increasing strength of selection. In contrast, environmental novelty, as defined by the study authors, consistently revealed increased genetic variance but the studies synthesized by Wood and Brodie did not concurrently measure selection in these novel environments. Going forward, we need additional empirical data on both the strength and targets of selection and the degree of heritable variation available for selection to act to determine when and how the assumption of a positive correlation between strength of selection and revealed genetic variance in novel or changing environments is upheld or broken. Evolution on contemporary timescales in response to human-induced environmental change provides at least as many opportunities to explore fundamental questions on interactions among plasticity, organismal ecologies, and the magnitude and direction of evolutionary change as it does challenges in predicting complex responses to environmental change for conservation planning.

Competing interests

The authors declare that they have no competing interests.

Grant information

The author(s) declared that no grants were involved in supporting this work.

Acknowledgements

We thank Joel Kingsolver, Mike Moore, Lauren Buckley, and Robby Stoks for comments that improved this manuscript.

References



1. **F** Bellard C, Bertelsmeier C, Leadley P, *et al.*: **Impacts of climate change on the future of biodiversity.** *Ecol Lett.* 2012; **15**(4): 365–77.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | **F1000 Recommendation**
2. Huey RB, Kearney MR, Krockenberger A, *et al.*: **Predicting organismal vulnerability to climate warming: roles of behaviour, physiology and adaptation.** *Philos Trans R Soc Lond B Biol Sci.* 2012; **367**(1596): 1665–79.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
3. Merilä J, Hendry AP: **Climate change, adaptation, and phenotypic plasticity: the problem and the evidence.** *Evol Appl.* 2014; **7**(1): 1–14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
4. **F** Hoffmann AA, Sgrò CM: **Climate change and evolutionary adaptation.** *Nature.* 2011; **470**(7335): 479–85.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
5. Franks SJ, Hoffmann AA: **Genetics of climate change adaptation.** *Annu Rev Genet.* 2012; **46**: 185–208.
[PubMed Abstract](#) | [Publisher Full Text](#)
6. **F** Sgrò CM, Terblanche JS, Hoffmann AA: **What Can Plasticity Contribute to Insect Responses to Climate Change?** *Annu Rev Entomol.* 2016; **61**: 433–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
7. Chown SL, Hoffmann AA, Kristensen TN, *et al.*: **Adapting to climate change: a perspective from evolutionary physiology.** *Clim Res.* 2010; **43**(1/2): 3–15.
[Publisher Full Text](#)
8. Hill JK, Griffiths HM, Thomas CD: **Climate change and evolutionary adaptations at species' range margins.** *Annu Rev Entomol.* 2011; **56**: 143–59.
[PubMed Abstract](#) | [Publisher Full Text](#)
9. Anderson JT, Panetta AM, Mitchell-Olds T: **Evolutionary and ecological responses to anthropogenic climate change: update on anthropogenic climate change.** *Plant Physiol.* 2012; **160**(4): 1728–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Somero GN: **The physiology of climate change: how potentials for acclimatization and genetic adaptation will determine 'winners' and 'losers'.** *J Exp Biol.* 2010; **213**(6): 912–20.
[PubMed Abstract](#) | [Publisher Full Text](#)
11. **F** Gienapp P, Teplitsky C, Alho JS, *et al.*: **Climate change and evolution: disentangling environmental and genetic responses.** *Mol Ecol.* 2008; **17**(1): 167–78.
[PubMed Abstract](#) | [Publisher Full Text](#) | **F1000 Recommendation**
12. Walters RJ, Blanckenhorn WU, Berger D: **Forecasting extinction risk of ectotherms under climate warming: an evolutionary perspective.** *Funct Ecol.* 2012; **26**(6): 1324–38.
[Publisher Full Text](#)

13. Levis NA, Pfennig DW: **Evaluating 'Plasticity-First' Evolution in Nature: Key Criteria and Empirical Approaches.** *Trends Ecol Evol.* 2016; **31**(7): 563–74.
[PubMed Abstract](#) | [Publisher Full Text](#)
14. Ashander J, Chevin LM, Baskett ML: **Predicting evolutionary rescue via evolving plasticity in stochastic environments.** *Proc Biol Sci.* 2016; **283**(1839): pii: 20161690.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Chevin LM, Lande R, Mace GM: **Adaptation, plasticity, and extinction in a changing environment: towards a predictive theory.** *PLoS Biol.* 2010; **8**(4): e1000357.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
16. Ghalambor CK, Hoke KL, Ruel EW, *et al.*: **Non-adaptive plasticity potentiates rapid adaptive evolution of gene expression in nature.** *Nature.* 2015; **525**(7569): 372–5.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
17. Stoks R, Govaert L, Pauwels K, *et al.*: **Resurrecting complexity: the interplay of plasticity and rapid evolution in the multiple trait response to strong changes in predation pressure in the water flea *Daphnia magna*.** *Ecol Lett.* 2015; **19**(2): 180–190.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
18. Gunderson AR, Stillman JH: **Plasticity in thermal tolerance has limited potential to buffer ectotherms from global warming.** *Proc Biol Sci.* 2015; **282**(1808): 20150401.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
19. Telemeco RS, Fletcher B, Levy O, *et al.*: **Lizards fail to plastically adjust nesting behavior or thermal tolerance as needed to buffer populations from climate warming.** *Glob Chang Biol.* 2016.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Grether GF: **Environmental change, phenotypic plasticity, and genetic compensation.** *Am Nat.* 2005; **166**(4): E115–23.
[PubMed Abstract](#) | [Publisher Full Text](#)
21. Schaum CE, Collins S: **Plasticity predicts evolution in a marine alga.** *Proc Biol Sci.* 2014; **281**(1793): pii: 20141486.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Williams JW, Jackson ST, Kutzbach JE: **Projected distributions of novel and disappearing climates by 2100 AD.** *Proc Natl Acad Sci U S A.* 2007; **104**(14): 5738–42.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
23. Schlichting CD, Wund MA: **Phenotypic plasticity and epigenetic marking: an assessment of evidence for genetic accommodation.** *Evolution.* 2014; **68**(3): 656–72.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. West-Eberhard MJ: **Developmental Plasticity and Evolution.** Oxford University Press, 2003; 815.
[Reference Source](#)
25. Schlichting CD: **Hidden reaction norms, cryptic genetic variation, and evolvability.** *Ann N Y Acad Sci.* 2008; **1133**: 187–203.
[PubMed Abstract](#) | [Publisher Full Text](#)
26. Murren CJ, Maclean HJ, Diamond SE, *et al.*: **Evolutionary change in continuous reaction norms.** *Am Nat.* 2014; **183**(4): 453–67.
[PubMed Abstract](#) | [Publisher Full Text](#)
27. Wood CW, Brodie ED 3rd: **Evolutionary response when selection and genetic variation covary across environments.** *Ecol Lett.* 2016; **19**(10): 1189–200.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
28. Waddington CH: **Genetic assimilation.** In: Thoday EWC and JM, editor. *Adv Genet* [Internet]. Academic Press. 1961; 10: 257–93 [cited 2016 Sep 6].
[PubMed Abstract](#) | [Publisher Full Text](#)
29. Suzuki Y, Nijhout HF: **Evolution of a polyphenism by genetic accommodation.** *Science.* 2006; **311**(5761): 650–2.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
30. Moczek AP, Sultan S, Foster S, *et al.*: **The role of developmental plasticity in evolutionary innovation.** *Proc Biol Sci.* 2011; **278**(1719): 2705–13.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
31. McGuigan K, Sgrò CM: **Evolutionary consequences of cryptic genetic variation.** *Trends Ecol Evol.* 2009; **24**(6): 305–11.
[PubMed Abstract](#) | [Publisher Full Text](#)
32. Wund MA, Baker JA, Clancy B, *et al.*: **A test of the “flexible stem” model of evolution: ancestral plasticity, genetic accommodation, and morphological divergence in the threespine stickleback radiation.** *Am Nat.* 2008; **172**(4): 449–62.
[PubMed Abstract](#) | [Publisher Full Text](#)
33. Sandom C, Faurby S, Sandel B, *et al.*: **Global late Quaternary megafauna extinctions linked to humans, not climate change.** *Proc Biol Sci.* 2014; **281**(1787): pii: 20133254.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
34. Intergovernmental Panel on Climate Change, editor: **Climate Change 2013 - The Physical Science Basis: Working Group I Contribution to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change.** Cambridge: Cambridge University Press; 2014.
[Reference Source](#)
35. Parmesan C, Yohe G: **A globally coherent fingerprint of climate change impacts across natural systems.** *Nature.* 2003; **421**(6918): 37–42.
[PubMed Abstract](#) | [Publisher Full Text](#)
36. Chen IC, Hill JK, Ohlemüller R, *et al.*: **Rapid range shifts of species associated with high levels of climate warming.** *Science.* 2011; **333**(6045): 1024–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
37. Diamond SE: **Evolutionary potential of upper thermal tolerance: biogeographic patterns and expectations under climate change.** *Ann N Y Acad Sci.* 2016.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Visser ME: **Keeping up with a warming world; assessing the rate of adaptation to climate change.** *Proc Biol Sci.* 2008; **275**(1635): 649–59.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. Kingsolver JG: **The well-temperated biologist. (American Society of Naturalists Presidential Address).** *Am Nat.* 2009; **174**(6): 755–68.
[PubMed Abstract](#) | [Publisher Full Text](#)
40. Seebacher F, White CR, Franklin CE: **Physiological plasticity increases resilience of ectothermic animals to climate change.** *Nat Clim Change.* 2015; **5**(1): 61–6.
[Publisher Full Text](#)
41. Gienapp P, Lof M, Reed TE: **Predicting demographically sustainable rates of adaptation: can great tit breeding time keep pace with climate change?** *Philos Trans R Soc Lond B Biol Sci.* 2013; **368**(1610): 20120289.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
42. Merilä J: **Evolution in response to climate change: in pursuit of the missing evidence.** *Bioessays.* 2012; **34**(9): 811–8.
[PubMed Abstract](#) | [Publisher Full Text](#)
43. Crozier LG, Hutchings JA: **Plastic and evolutionary responses to climate change in fish.** *Evol Appl.* 2014; **7**(1): 68–87.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
44. Anderson JT, Inouye DW, McKinney AM, *et al.*: **Phenotypic plasticity and adaptive evolution contribute to advancing flowering phenology in response to climate change.** *Proc Biol Sci.* 2012; **279**(1743): 3843–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
45. Buckley LB, Ehrenberger JC, Angilletta MJ Jr: **Thermoregulatory behaviour limits local adaptation of thermal niches and confers sensitivity to climate change.** *Funct Ecol.* 2015; **29**(8): 1038–47.
[Publisher Full Text](#)
46. Logan ML, Cox RM, Calsbeek R: **Natural selection on thermal performance in a novel thermal environment.** *Proc Natl Acad Sci U S A.* 2014; **111**(39): 14165–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
47. Geerts AN, Vanoverbeke J, Vanschoenwinkel B, *et al.*: **Rapid evolution of thermal tolerance in the water flea *Daphnia*.** *Nat Clim Change.* 2015; **5**(7): 665–8.
[Publisher Full Text](#)
48. Heinen-Kay JL, Noel HG, Layman CA, *et al.*: **Human-caused habitat fragmentation can drive rapid divergence of male genitalia.** *Evol Appl.* 2014; **7**(10): 1252–67.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
49. Liu Z, He C, Zhou Y, *et al.*: **How much of the world's land has been urbanized, really? A hierarchical framework for avoiding confusion.** *Landsc Ecol.* 2014; **29**(5): 763–71.
[Publisher Full Text](#)
50. Seto KC, Güneralp B, Hutyra LR: **Global forecasts of urban expansion to 2030 and direct impacts on biodiversity and carbon pools.** *Proc Natl Acad Sci U S A.* 2012; **109**(10): 16083–8.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
51. Evans KL, Gaston KJ, Frantz AC, *et al.*: **Independent colonization of multiple urban centres by a formerly forest specialist bird species.** *Proc Biol Sci.* 2009; **276**(1666): 2403–10.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
52. Diamond SE, Dunn RR, Frank SD, *et al.*: **Shared and unique responses of insects to the interaction of urbanization and background climate.** *Current Opinion in Insect Science.* 2015; **11**: 71–7.
[Publisher Full Text](#)
53. Imhoff ML, Zhang P, Wolfe RE, *et al.*: **Remote sensing of the urban heat island effect across biomes in the continental USA.** *Remote Sensing of Environment.* 2010; **114**(3): 504–13.
[Publisher Full Text](#)
54. Donihue CM, Lambert MR: **Adaptive evolution in urban ecosystems.** *Ambio.* 2015; **44**: 194–203.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
55. Kettlewell HBD: **Selection experiments on industrial melanism in the *Lepidoptera*.** *Heredity.* 1955; **9**: 323–42.
[Publisher Full Text](#)
56. Cook LM, Grant BS, Saccheri IJ, *et al.*: **Selective bird predation on the peppered moth: the last experiment of Michael Majerus.** *Biol Lett.* 2012; **8**(4): 609–12.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
57. Whitehead A, Triant DA, Champlin D, *et al.*: **Comparative transcriptomics implicates mechanisms of evolved pollution tolerance in a killifish population.** *Mol Ecol.* 2010; **19**(23): 5186–203.
[PubMed Abstract](#) | [Publisher Full Text](#)
58. Cheptou P, Carrue O, Rouifed S, *et al.*: **Rapid evolution of seed dispersal**

- in an urban environment in the weed *Crepis sancta*. *Proc Natl Acad Sci U S A*. 2008; **105**(10): 3796–9.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
59. **F** Winchell KM, Reynolds RG, Prado-Irwin SR, *et al.*: Phenotypic shifts in urban areas in the tropical lizard *Anolis cristatellus*. *Evolution*. 2016; **70**(5): 1009–22.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
60. Angilletta MJ Jr, Wilson RS, Niehaus AC, *et al.*: Urban physiology: city ants possess high heat tolerance. *PLoS One*. 2007; **2**(2): e258.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
61. McLean MA, Angilletta MJ Jr, Williams KS: If you can't stand the heat, stay out of the city: Thermal reaction norms of chitinolytic fungi in an urban heat island. *J Therm Biol*. 2005; **30**(5): 384–91.
[Publisher Full Text](#)
62. van Straalen NM: Ecotoxicology becomes stress ecology. *Environ Sci Technol*. 2003; **37**(17): 324A–30.
[PubMed Abstract](#) | [Publisher Full Text](#)
63. Jansen M, Stoks R, Coors A, *et al.*: Collateral damage: rapid exposure-induced evolution of pesticide resistance leads to increased susceptibility to parasites. *Evolution*. 2011; **65**(9): 2681–91.
[PubMed Abstract](#) | [Publisher Full Text](#)
64. Antonovics J, Bradshaw AD, Turner RG: Heavy Metal Tolerance in Plants. In: *Advances in Ecological Research*. Elsevier. 1971; **7**: 1–85.
[Publisher Full Text](#)
65. Mallet J: The evolution of insecticide resistance: Have the insects won? *Trends Ecol Evol*. 1989; **4**(11): 336–40.
[PubMed Abstract](#) | [Publisher Full Text](#)
66. Posthuma L, van Straalen NM: Heavy-metal adaptation in terrestrial invertebrates: A review of occurrence, genetics, physiology and ecological consequences. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol*. 1993; **106**(1): 11–38.
[Publisher Full Text](#)
67. Jain SK, Bradshaw AD: Evolutionary divergence among adjacent plant populations I. The evidence and its theoretical analysis. *Heredity*. 1966; **21**: 407–41.
[Publisher Full Text](#)
68. Antonovics J: Evolution in closely adjacent plant populations V. Evolution of self-fertility. *Heredity*. 1968; **23**: 219–38.
[Publisher Full Text](#)
69. McNeilly T, Antonovics J: Evolution in closely adjacent plant populations IV. Barriers to gene flow. *Heredity*. 1968; **23**: 205–18.
[Publisher Full Text](#)
70. **F** Antonovics J: Evolution in closely adjacent plant populations X: long-term persistence of prereproductive isolation at a mine boundary. *Heredity (Edinb)*. 2006; **97**(1): 33–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
71. Kwok KW, Grist EP, Leung KM: Acclimation effect and fitness cost of copper resistance in the marine copepod *Tigriopus japonicus*. *Ecotoxicol Environ Saf*. 2009; **72**(2): 358–64.
[PubMed Abstract](#) | [Publisher Full Text](#)
72. Baker AJ, Walker PL: Physiological responses of plants to heavy metals and the quantification of tolerance and toxicity. *Chem Speciat Bioavailab*. 1989; **1**(1): 7–17.
[Reference Source](#)
73. Hua J, Jones DK, Relyea RA: Induced tolerance from a sublethal insecticide leads to cross-tolerance to other insecticides. *Environ Sci Technol*. 2014; **48**(7): 4078–85.
[PubMed Abstract](#) | [Publisher Full Text](#)
74. Poupardin R, Reynaud S, Strode C, *et al.*: Cross-induction of detoxification genes by environmental xenobiotics and insecticides in the mosquito *Aedes aegypti*: impact on larval tolerance to chemical insecticides. *Insect Biochem Mol Biol*. 2008; **38**(5): 540–51.
[PubMed Abstract](#) | [Publisher Full Text](#)
75. Hua J, Jones DK, Mattes BM, *et al.*: Evolved pesticide tolerance in amphibians: Predicting mechanisms based on pesticide novelty and mode of action. *Environ Pollut*. 2015; **206**: 56–63.
[PubMed Abstract](#) | [Publisher Full Text](#)
76. **F** Hua J, Jones DK, Mattes BM, *et al.*: The contribution of phenotypic plasticity to the evolution of insecticide tolerance in amphibian populations. *Evol Appl*. 2015; **8**(6): 586–96.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
77. Brausch JM, Smith PN: Pesticide resistance from historical agricultural chemical exposure in *Thamnocephalus platyurus* (Crustacea: Anostraca). *Environ Pollut*. 2009; **157**(2): 481–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
78. Coors A, Vanoverbeke J, De Bie T, *et al.*: Land use, genetic diversity and toxicant tolerance in natural populations of *Daphnia magna*. *Aquat Toxicol*. 2009; **95**(1): 71–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
79. Hua J, Morehouse NI, Relyea R: Pesticide tolerance in amphibians: induced tolerance in susceptible populations, constitutive tolerance in tolerant populations. *Evol Appl*. 2013; **6**(7): 1028–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
80. Cothran RD, Brown JM, Relyea RA: Proximity to agriculture is correlated with pesticide tolerance: Evidence for the evolution of amphibian resistance to modern pesticides. *Evol Appl*. 2013; **6**: 832–41.
[Publisher Full Text](#)
81. Weston DP, Poynton HC, Wellborn GA, *et al.*: Multiple origins of pyrethroid insecticide resistance across the species complex of a nontarget aquatic crustacean, *Hyalella azteca*. *Proc Natl Acad Sci U S A*. 2013; **110**(41): 16532–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
82. Bendis RJ, Relyea RA: Living on the edge: populations of two zooplankton species living closer to agricultural fields are more resistant to a common insecticide. *Environ Toxicol Chem*. 2014; **33**(12): 2835–41.
[PubMed Abstract](#) | [Publisher Full Text](#)
83. Jansen M, Coors A, Vanoverbeke J, *et al.*: Experimental evolution reveals high insecticide tolerance in *Daphnia* inhabiting farmland ponds. *Evol Appl*. 2015; **8**(5): 442–53.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
84. **F** Wilson AJ, Pemberton JM, Pilkington JG, *et al.*: Environmental coupling of selection and heritability limits evolution. *PLoS Biol*. 2006; **4**(7): e216.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
85. **F** Husby A, Visser ME, Kruuk LE: Speeding up microevolution: the effects of increasing temperature on selection and genetic variance in a wild bird population. *PLoS Biol*. 2011; **9**(2): e1000585.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
86. Price TD, Qvarnström A, Irwin DE: The role of phenotypic plasticity in driving genetic evolution. *Proc Biol Sci*. 2003; **270**(1523): 1433–40.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
87. Ghalambor CK, McKay JK, Carroll SP, *et al.*: Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Funct Ecology*. 2007; **21**: 394–407.
[Publisher Full Text](#)
88. Ledón-Rettig CC, Pfennig DW, Chunco AJ, *et al.*: Cryptic genetic variation in natural populations: a predictive framework. *Integr Comp Biol*. 2014; **54**(5): 783–93.
[PubMed Abstract](#) | [Publisher Full Text](#)
89. Hoffmann AA, Merilä J: Heritable variation and evolution under favourable and unfavourable conditions. *Trends Ecol Evol*. 1999; **14**(3): 96–101.
[PubMed Abstract](#) | [Publisher Full Text](#)
90. Via S, Gomulkiewicz R, De Jong G, *et al.*: Adaptive phenotypic plasticity: consensus and controversy. *Trends Ecol Evol*. 1995; **10**(5): 212–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
91. Waddington CH: Canalization of Development and the Inheritance of Acquired Characters. *Nature*. 1942; **150**: 563–5.
[Publisher Full Text](#)
92. Waddington CH: Genetic Assimilation of an Acquired Character. *Evolution*. 1953; **7**(2): 118–126.
[Publisher Full Text](#)

Open Peer Review

Current Referee Status:



Editorial Note on the Review Process

F1000 Faculty Reviews are commissioned from members of the prestigious F1000 Faculty and are edited as a service to readers. In order to make these reviews as comprehensive and accessible as possible, the referees provide input before publication and only the final, revised version is published. The referees who approved the final version are listed with their names and affiliations but without their reports on earlier versions (any comments will already have been addressed in the published version).

The referees who approved this article are:

Version 1

- 1 **Robby Stoks**, Department of Biology, KU Leuven, Leuven, Belgium
Competing Interests: No competing interests were disclosed.
- 2 **Lauren Buckley**, Department of Biology, University of Washington, Seattle, WA, USA
Competing Interests: No competing interests were disclosed.