

Adenovirus Strand Nomenclature: a Proposal^{1, 2}

Thirty-six scientists have agreed to standardize the nomenclature of DNA strands of all adenovirus serotypes according to certain criteria.

The study of the biochemistry and molecular biology of adenoviruses began soon after their identification, but the construction of a genomic map of viral mRNA sequences and proteins awaited the purification of base-sequence specific endonucleases and the separation of the two strands of viral DNA. These endonucleases are in many cases known to be a component of restriction and modification systems of bacteria and are therefore commonly designated restriction endonucleases (11). The sites of cleavage of many restriction endonucleases have been mapped on the genome of adenovirus serotypes 2, 5, and 12 and adenovirus 2-simian virus 40 hybrid viruses (8, 12). Thus, specific fragments of viral DNA can be used to map biochemical products of adenovirus infections to discrete regions of the genome.

In common with the DNA of most bacteriophages, the strands of adenoviruses 2, 5, and 12 DNA can be separated by centrifugation to equilibrium in CsCl gradients after binding of poly(U,G) (7, 9, 10, 14, 16). One strand of the viral DNA binds a greater quantity of the ribopolymer and thus comes to equilibrium at a higher density in the gradient than its complement. The strand banding at higher density has been referred to as the "heavy" strand and denoted as both *h* and H; its complement, the "light" strand, is denoted as either *l* or L. The strands of adenoviruses 2 and 5 DNA have also been separated by equilibrium sedimentation of denatured viral DNA in alkaline CsCl gradients (15). A net bias in the distribution of guanine and thymine between the two strands results in one strand binding more Cs⁺ ion and therefore banding at a higher density than its complement (17). Again, the strand sediment-

ing to the higher density in alkaline CsCl gradients has been called the "heavy" strand and denoted as the H strand; its complement has been called the "light" strand and denoted as the L strand. Unfortunately, the adenovirus DNA strand sedimenting to the higher density in alkaline CsCl gradients bands at the lower density in poly(U,G)-containing CsCl gradients (13, 16). The *h* and *l* designation for the separated strands of adenovirus is therefore ambiguous and depends on the method used for strand separation.

Electrophoresis of denatured DNA in gels has been used to separate the strands of specific restriction endonuclease fragments of adenovirus types 2 and 5 DNA (13). This procedure was originally developed for the separation of the two strands of bacteriophage DNA (5). The strands of all six *EcoRI* cleavage fragments, five of the seven *HpaI* cleavage fragments, and all four *BamHI* cleavage fragments of adenovirus 2 DNA migrate, for example, as a faster (F) and slower (S) band during electrophoresis (2). The bias in base sequence between the two strands that causes their different mobility in these gel systems is not known. However, Weingärtner et al. (18) have reported that the strand migrating more slowly during electrophoresis has a lower thymine content than its complement. Viral DNA recovered from the faster and slower migrating bands in agarose gels has been assigned as a component of either total viral strand (2, 13).

Genomic maps are now being constructed from data from many laboratories, and it is therefore necessary to develop conventions for referring to the two strands of adenovirus DNA and the left-to-right orientation of the conventional map. To avoid the cumbersome notation of designating a particular strand after the physical property employed to separate the two complements, we suggest acceptance of the nomenclature developed in bacteriophage systems; that is, on the conventional map, the strand transcribed from left to right should be denoted the *r* strand and its complement, which is transcribed from right to left, the *l* strand. By generally accepted convention, the genetic maps of adenovirus subgroup C serotypes have been drawn with the following asymmetric physical and biological features defining the

¹ Consenting scientists listed alphabetically: Steven Bachenheimer, A. J. D. Bellett, J. E. Darnell, W. Doerfler, A. J. van der Eb, S. J. Flint, Kei Fujinaga, R. F. Gesteland, Harold S. Ginsberg, Maurice Green, M. S. Horwitz, Thomas J. Kelly, Jr., A. S. Levine, A. J. Levine, A. M. Lewis, Jr., J. B. Lewis, Stanley Mak, James K. McDougall, George D. Pearson, Ulf Pettersson, Lennart Philipson, K. Raska, Jr., H. J. Raskas, Richard Roberts, James A. Rose, W. C. Russell, Joe Sambrook, R. W. Schlesinger, Phillip A. Sharp,* H. Shimojo, J. S. Sussenbach, Clark Tibbets, P. C. van der Vliet, Joseph Weber, E. L. Winnacker, and William S. M. Wold.

² Requests for reprints should be addressed to Phillip A. Sharp, Center for Cancer Research, Massachusetts Institute of Technology, Cambridge, MA 02139.

left-to-right orientation: (i) the AT-rich half of the viral DNA is positioned to the right (1, 8); (ii) a non-essential region of the genome deleted in nondefective adenovirus 2-simian virus 40 hybrid viruses is positioned to the right (6); (iii) the only segment of the viral genome necessary for transformation of cells in culture is positioned to the left (3, 4); (iv) the strand of the genome coding for the virion major structural proteins is transcribed from left to right.

When the map has been oriented according to these parameters, we recommend that the positions on the map be defined as the percentage of the total distance of the adenovirus genome.

Maps of other adenovirus serotypes should be oriented in accord with as many of these criteria as possible to facilitate direct comparison between different serotypes.

LITERATURE CITED

- Doerfler, W., and A. K. Kleinschmidt. 1970. Denaturation pattern of the DNA of adenovirus type 2 as determined by electron microscopy. *J. Mol. Biol.* 50:579-593.
- Flint, S. J., S. M. Berget, and P. A. Sharp. 1976. Adenovirus transcription. III. Mapping of viral RNA sequences in cells productively infected by adenovirus type 5. *Virology* 72:443-455.
- Gallimore, P. H., P. A. Sharp, and J. Sambrook. 1974. Viral DNA in transformed cells. II. A study of the sequences of adenovirus 2 DNA in nine lines of transformed rat cells using specific fragments of the viral genome. *J. Mol. Biol.* 89:49-72.
- Graham, F. L., P. J. Abrahams, C. Mulder, H. L. Heijneker, S. O. Warnaar, F. A. J. de Vries, W. Fiers, and A. J. van der Eb. 1974. Studies on *in vitro* transformation by DNA and DNA fragments of human adenoviruses and simian virus 40. *Cold Spring Harbor Symp. Quant. Biol.* 39:637-650.
- Hayward, G. S. 1972. Gel electrophoretic separation of complementary strands of bacteriophage DNA. *Virology* 49:342-344.
- Kelly, T. J., Jr., and A. M. Lewis. 1973. Use of nondefective adenovirus-simian virus 40 hybrids for mapping the simian virus 40 genome. *J. Virol.* 12:643-652.
- Landgraf-Leurs, M., and M. Green. 1971. Adenovirus DNA. III. Separation of the complementary strands of adenovirus 2, 7, and 12 DNA molecules. *J. Mol. Biol.* 60:185-202.
- Mulder, C., J. R. Arrand, H. Delius, W. Keller, U. Pettersson, R. J. Roberts, and P. A. Sharp. 1974. Cleavage mapping of DNA from adenovirus types 2 and 5 by restriction endonucleases *Eco* RI and *Hpa* I. *Cold Spring Harbor Symp. Quant. Biol.* 39:397-400.
- Ortin, J., K.-H. Scheidtmann, R. Greenberg, M. Westphal, and W. Doerfler. 1976. Transcription of the genome of adenovirus type 12. III. Maps of stable RNA from productively infected human cells and abortively infected and transformed hamster cells. *J. Virol.* 20:355-372.
- Patch, C. T., A. M. Lewis, and A. S. Levine. 1972. Evidence for a transcription control region of SV40 in the adenovirus 2-SV40 hybrid Ad2+ND₁. *Proc. Natl. Acad. Sci. U.S.A.* 69:3375-3379.
- Roberts, R. J. 1976. Restriction endonuclease, Chemical Rubber Co. Reviews, in press.
- Sambrook, J., J. F. Williams, P. A. Sharp, and T. Grodzicker. 1975. Physical mapping of temperature-sensitive mutations of adenoviruses. *J. Mol. Biol.* 97:369-390.
- Sharp, P. A., P. H. Gallimore, and S. J. Flint. 1974. Mapping of adenovirus 2 RNA sequences in lytically infected cells and transformed cell lines. *Cold Spring Harbor Symp. Quant. Biol.* 39:457-474.
- Smiley, J. R., and S. Mak. 1976. Adenovirus type 12-specific RNA sequences during productive infection of KB cells. *J. Virol.* 19:36-42.
- Sussenbach, J. S., D. J. Ellens, and H. S. Jansz. 1973. Studies on the mechanism of replication of adenovirus DNA. II. The nature of single-stranded DNA in replicative intermediates. *J. Virol.* 12:1131-1138.
- Tibbetts, C., U. Pettersson, K. Johansson, and L. Philipson. 1974. Relationship of mRNA from productively infected cells to the complementary strands of adenovirus type 2 DNA. *J. Virol.* 13:370-377.
- Vinograd, J., J. Morris, N. Davidson, and W. F. Dove. 1963. The buoyant behaviour of viral and bacterial DNA in alkaline CsCl. *Proc. Natl. Acad. Sci. U.S.A.* 49:12-17.
- Weingärtner, B., E.-L. Winnacker, A. Tolun, and U. Pettersson. 1976. Two complementary strand-specific termination sites for adenovirus DNA replication. *Cell* 9:259-268.