

Considerations for successful cancer immunotherapy in aged hosts

OTHER ARTICLES PUBLISHED IN THIS REVIEW SERIES

- Immunosenescence: the importance of considering age in health and disease. Clinical and Experimental Immunology 2017, 187: 1–3.*
The convergence of senescence and nutrient sensing during lymphocyte ageing. Clinical and Experimental Immunology 2017, 187: 4–5.
Immune senescence: significance of the stromal microenvironment. Clinical and Experimental Immunology 2017, 187: 6–15.
Innate immune responses in the ageing lung. Clinical and Experimental Immunology 2017, 187: 16–25.
Age-related alterations in immune responses to West Nile virus infection. Clinical and Experimental Immunology 2017, 187: 26–34.
Intracellular signalling pathways: targets to reverse immunosenescence. Clinical and Experimental Immunology 2017, 187: 35–43.
Ageing and inflammation in patients with HIV infection. Clinical and Experimental Immunology 2017, 187: 44–52.
Ageing and obesity similarly impair antibody responses. Clinical and Experimental Immunology 2017, 187: 64–70.
The life cycle of a T cell after vaccination – where does immune ageing strike? Clinical and Experimental Immunology 2017, 187: 71–81.
Herpes zoster and the search for an effective vaccine. Clinical and Experimental Immunology 2017, 187: 82–92.
Adult vaccination against tetanus and diphtheria: the European perspective. Clinical and Experimental Immunology 2017, 187: 93–99.

V. Hurez,* Á. S. Padrón,*

R. S. Svatek^{†‡} and T. J. Curiel^{*‡§¶}

*Department of Medicine, [†]Department of Urology, [‡]Cancer Therapy and Research Center, [§]Department of Microbiology and Immunology, and [¶]The Barshop Institute for Ageing and Longevity Studies, University of Texas Health Science Center, San Antonio, TX, USA

Summary

Immunotherapy is now experiencing unprecedented successes in treating various cancers based on new understandings of cancer immunopathogenesis. Nonetheless, although ageing is the biggest risk factor for cancer, the majority of cancer immunotherapy preclinical studies are conducted in young hosts. This review will explore age-related changes in immunity as they relate to cancer immune surveillance, immunopathogenesis and responses to immunotherapy. Although it is recognized that declining T cell function with age poses a great challenge to developing effective age-related cancer immunotherapies, examples of successful approaches to overcome this hurdle have been developed. Further, it is now recognized that immune functions do not simply decline with age, but rather change in ways than can be detrimental. For example, with age, specific immune cell populations with detrimental functions can become predominant (such as cells producing proinflammatory cytokines), suppressive cells can become more numerous or more suppressive (such as myeloid-derived suppressor cells), drugs can affect aged immune cells distinctly and the aged microenvironment is becoming recognized as a significant barrier to address. Key developments in these and other areas will be surveyed as they relate to cancer immunotherapy in aged hosts, and areas in need of more study will be assessed with some speculations for the future. We propose the term ‘age-related immune dysfunction’ (ARID) as best representative of age-associated changes in immunity.

Keywords: aging, cancer, immunity, immunotherapy

Accepted for publication 13 September 2016
Correspondence: T. Curiel, Department of Medicine, University of Texas Health Science Center at San Antonio, STRF MC 8252, 8403 Floyd Curl Drive, San Antonio, TX 78229-3900, USA.
E-mail: curielt@uthscsa.edu

Introduction

The immune system is exquisitely able to identify specific antigens and eliminate cells expressing them. Tumours are quintessentially antigenic tissues as the result of their many genetic mutations. This antigenicity (expression of antigens), however, does not usually translate into clinically meaningful immunogenicity (the ability of these antigens to elicit useful immunity), as spontaneous rejection of clinically apparent tumours occurs rarely. Tumour-specific and tumour-associated antigens were identified more than 60 years ago, prompting strategies to attempt to boost anti-tumour immunity paralleling successful approaches to boost anti-pathogen immunity. It is now

clear that the inability of endogenous immunity to eradicate clinically evident cancers derives from various factors, including tumour-driven immune dysfunction, co-evolution of anti-tumour immunity as tumours mutate to escape immune elimination (termed ‘immunoediting’, discussed below) and because anti-tumour immunity is autoimmunity [1,2]. Recent discoveries have helped thinking advance to develop more effective anti-cancer immunotherapies. With age, some of these impediments are compounded, and new barriers to successful cancer immunotherapy emerge [3–6].

Tumour immune surveillance is part of a more comprehensive process termed ‘immunoediting’ [7], associated

with the 'three Es' [8]. The first E stands for elimination of nascent cancer cells. The second E is equilibrium, in which growth of malignant cells escaping immunity is balanced by immunity that induces immune elimination. This selective pressure results in antigenic evolution of tumours that leads ultimately to immune escape, the third E is where the tumour has evaded the immune defences and becomes clinically evident in mouse models [9–11] and humans [12,13].

Characterization of cancer properties led to the original six fundamental cancer hallmarks, but these did not include immunity [14]. The updated eight fundamental hallmarks include lack of immune rejection [15]. Chronic generalized inflammation is another newly appreciated cancer hallmark [16,17], suggested as another fundamental hallmark [18], along with genomic instability [19], abnormal vasculature [20] and stem cell features [21].

As we will discuss, these cancer hallmarks (especially immune rejection and chronic inflammation) bear directly upon the challenges in developing age-specific immunotherapy. Age effects on immunity extend far beyond simple declines in functions or reductions in cell numbers. We propose the term 'age-related immune dysfunction' (ARID) to encompass the full range of age-related alterations in immunity with advancing age. The following sections address major topics relating to age effects on cancer immunotherapy.

Tumour-specific T cells

With ageing, a general reduction in T cell immunity results from various processes affecting T cell numbers, diversity, phenotype and function [22]. For example, phenotypically naive T cells (CD45RA⁺CD62L⁺CD27⁺CD28⁺) are produced throughout life in the thymus but thymic production wanes with age [23–25], reducing global T cell repertoire diversity even in healthy individuals, as overall peripheral T cell numbers remain relatively constant [26]. Reduced haematopoietic stem cell production of T cell precursors [27] also contributes. Even as naive T cell numbers decrease, T cells with a memory phenotype, including cytokine-producing cytotoxic CD8⁺ T cells, increase with age. Dysfunctional, terminally differentiated effector cells also increase, especially virus-reactive cells, with highly reduced T cell receptor repertoire diversity and with limited proliferative ability [28]. Thus, most T cells are memory/effector cells and low-level chronic inflammation is characteristic of an aged immune system. In addition, T cell signalling declines with age [29].

Therapeutic strategies have been developed to lessen these age-related defects and help to elicit effective T cell immunity. For example, CD4⁺ T cell functions decline with age, but function loss is mitigated by giving cytokines, including tumour necrosis factor (TNF)- α or

interleukin (IL)-6 [30]. Defective age-related T cell priming can be rescued using agonist α CD137 antibodies [31]. Tumour-specific immunity can also be enhanced. For example, OX40-enhanced tumour rejection and effector T cell differentiation decreases with age [32,33], but aged mice developed protective anti-tumour immunity to a lymphoid tumour with an agonist α CD40 antibody [34]. We showed that aged mice develop significant anti-tumour immunity to aggressive and poorly immunogenic B16 melanoma, comparable to young mice, with similar clinical effect by simultaneous depletion of suppressive myeloid and T cells [5], discussed in detail below.

Regulatory T cells (T_{regs})

As T_{regs} are key mediators of tumour immune dysfunction, reducing T_{reg} function or numbers is a rational cancer immunotherapy strategy [35–38]. Reports of T_{reg} contributions to age-related decline in immune responses are contradictory, with some studies showing increases in T_{reg} prevalence and/or function with age in humans and mice [39–42], whereas others show no changes or reduced T_{reg} contributions [43,44].

Increased T_{reg} prevalence in lymphoid organs but not blood or thymus have been shown in aged mice [5,45,46]. The effect of age on T_{reg} functional properties is complex, depending on the experimental setting and the function assessed. Some functions appear to be reduced [47], such as suppression of delayed-type hypersensitivity responses *in vivo* [41] or inhibition of T helper type 17 (Th17) function [45]. In other studies, T_{regs} from aged mice appear to have similar or greater suppressive function *versus* young mice [5,48]. Few studies have assessed the changes in T_{reg} prevalence in elderly humans [49], but they could increase in circulation with age [50]. T_{regs} from young and elderly individuals similarly inhibited the proliferation of responder cells, whereas the production of the anti-inflammatory cytokine IL-10 was reduced in cells from aged subjects [51].

Although T_{reg} depletion is an effective approach to improving anti-tumour immunity and responses to immunotherapy [36,38], conflicting studies report the effect of T_{reg} depletion as cancer immunotherapy in aged hosts [52,53]. One early study correlated defective tumour clearance to increased T_{reg} prevalence and used α CD25 to deplete T_{regs} and improve anti-cancer immunity [53]. We used denileukin diftitox to deplete T_{regs} in mice bearing B16 melanoma [5]. While denileukin diftitox depleted T_{regs} similarly in young and aged hosts, slowed tumour growth and improved tumour-specific immunity was observed only in young mice. Denileukin diftitox-mediated T_{reg} depletion affected interferon (IFN)- γ - and IL-17 producing T cells differentially in young *versus* aged mice. Tumour-bearing aged mice had

more CD11b⁺Gr-1^{hi} myeloid-derived suppressor cells (MDSC) that were more suppressive. T_{reg} depletion resulted in a further increase in MDSC numbers. When MDSC depletion using anti-Gr-1 antibody was added to

T_{reg} depletion, anti-tumour immunity was restored in the aged mice resulting in slowed tumour growth similar to young hosts. This strategy did not improve treatment further in young mice as their MDSC did not increase with

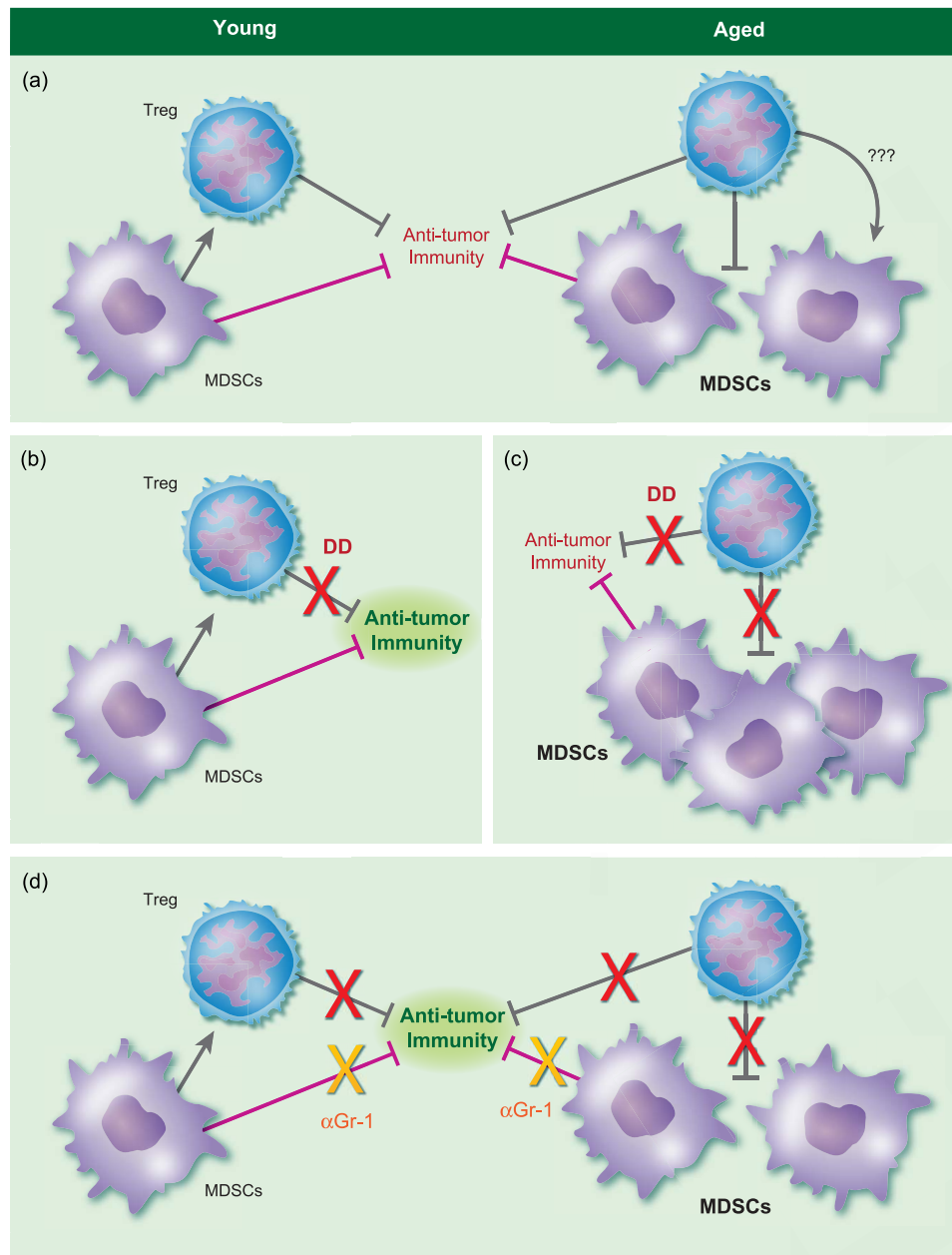


Fig. 1. Example of an age-specific immune effect that can be ameliorated to allow effective anti-melanoma immunity. (a) Young mice challenged with B16 melanoma experience increased regulatory T cells (T_{regs}), but not myeloid derived suppressor cells (MDSC), whereas aged mice experience increased T_{regs} and MDSC. In the aged, the increased MDSC could be due to poor T_{reg} control of them directly, or through indirect mechanisms (denoted by question marks). (b) In young mice, denileukin difitox (DD) reduces T_{regs} with little MDSC effect, improving anti-tumour immunity [increased interferon (IFN)- γ ⁺ T cells]. By contrast, in aged mice (c), DD-mediated T_{reg} reduction reduces T_{regs} but without significant increase in IFN- γ ⁺ T cells. Interleukin (IL)-17⁺, potentially detrimental T cells, increase, as do deleterious MDSC, thereby inhibiting beneficial anti-tumour immunity. Green anti-tumour immunity means good immunity, whereas red means less effective immunity. (d) By adding anti-granulocyte-differentiation antigen-1 (α Gr-1) antibody to DD in aged mice, the MDSC increase is blunted and aged mice now mount anti-tumour immunity comparable to young hosts receiving DD alone, with comparable clinical efficacy. Adding α Gr-1 to DD in young hosts does not improve immune or clinical effects further, as MDSC were not increased further by DD. The red 'X' denotes DD effects to reduce T_{reg} inhibition.

denileukin diftitox, representing the first cancer immunotherapy to work in aged but, to our knowledge, not young hosts [5]. An illustration to represent events is shown in Fig. 1. Thus, mitigation of aged-specific immune suppressive mechanisms allows successful cancer immunotherapy even in aged hosts.

Innate immunity

Declines in T cell function and chronic low-level inflammation with age can promote expansion of suppressive myeloid cells. Thus, as anti-tumour T cells may not be optimally functional in elderly cancer patients, targeting innate cells could represent a better strategy [54].

Dendritic cells (DCs)

DCs are antigen-presenting cells that play a key role in mediating T cell immunity [55]. The tumour environment promotes expansion of dysfunctional DC subsets that hamper effective anti-tumour immune responses and immunotherapy [56,57].

Circulating DC precursors and skin Langerhans cells decrease with age. Age effects on DC function depend on the function assessed and are reported variably as decreased or unchanged [58]. As DC function declines, any related impaired T cell function or increased inflammation could contribute to age-related cancer risk. Augmenting DC antigen-presenting abilities, which is reported inconsistently to decline with age, induces strong tumour-specific cytotoxic T cell immunity. For example, CD40L or agonist anti-CD40 antibodies can boost DC activation in animal and human studies [59]. In older cancer patients, a vaccine using CD40L linked to specific antigens has potential [60].

Macrophages

In contrast to reduced lymphopoiesis in age, myelopoiesis increases [61], increasing myeloid cell numbers. Macrophages are an important component of tumour stroma and tumour-associated immune dysfunction [62]. Tumour-resident macrophages are characterized as proinflammatory M1 or anti-inflammatory M2 macrophages and can switch phenotypes. M1 macrophages secrete proinflammatory cytokines, such as TNF- α and IL-12, that boost anti-tumour immunity. M2 macrophages produce anti-inflammatory cytokines (e.g. IL-10, TGF- β) that promote tumorigenesis [62].

Age effects on macrophage differentiation and function are complex. Macrophage prevalence in lymphoid organs increases in aged mice [63]. M1 macrophage function could increase through production of age-related reactive oxygen species (ROS), but other studies show reduced function with age in M1 macrophages [64], which could be from increased IL-10-producing M2 macrophages [62].

Thus, macrophages from ageing hosts could promote tumour growth.

Tumour-associated macrophages from elderly, but not young, mice produce high levels of immune-suppressive TGF- β , consistent with an M2-type phenotype [63]. Detrimental M2 macrophages can be converted to beneficial M1 using IL-12 or poly-(cysteine 5' to guanine) (CpG) plus an α IL-10 receptor [65]. Thus, targeting tumour-associated age-related M2 macrophages could be useful to treat cancers in aged hosts.

MDSC

MDSC are immature, immunosuppressive, myeloid cells that increase in inflammatory diseases, particularly tumours [66–69], and suppress anti-tumour immunity [70]. MDSCs produce inhibitory factors (e.g. IL-10, arginase) that inhibit T cells and promote T_{regs} and detrimental M2 macrophages [71]. MDSCs increase during ageing in human blood [72] and in lymphoid organs in mice [5,63]. MDSC contribute to immunopathology in aged hosts, including in cancer [73,74]. T_{reg} depletion with denileukin diftitox increased MDSCs in aged mice, suggesting T_{reg} control over MDSC [5], which could include indirect effects of T_{reg}-controlled cytokines that alter MDSC mobilization, proliferation or differentiation. In a CT26 colon cancer model, a *Lentinula edodes* mycelia extract reduced MDSC infiltration with a whole tumour cell vaccine in aged mice by suppressing the inflammatory MDSC-promoting cytokines IL-6 and TNF- α [75]. This extract also improved vaccine-induced *in-vivo* priming of tumour-specific cytotoxic T cells. Targeting MDSC is another potentially effective approach to reverse cancer-associated immune dysfunction in aged hosts.

Immune check-point inhibitors

Immune check-point receptor blockade has become one of the most successful immunotherapy strategies for various cancers [76,77]. None the less, the impact of those novel approaches in treating elderly patients has been little reported.

Immune check-point molecules are those that control the degree of immune responses either positively (activating immunity) or negatively (dampening immunity), which is often the case in cancers. Immune check-point inhibitor antibodies block these negative signals to improve anti-cancer immunity [76,77]. The expression of regulatory immune check-points on T cells increases with age in humans and mice [5,78,79], consistent with accumulation of hyporesponsive memory-like T cells that express these molecules. The immune check-point molecules identifying exhausted (poorly functional) T cells, such as programmed death 1 (PD-1), lymphocyte activation gene 3 (Lag-3) and T-cell immunoglobulin and mucin-domain containing 3

(Tim-3) also increase with age. These and other related molecules on T cells are targets for immune check-point blockade anti-cancer immunotherapies. Other immune check-point receptors, such as PD-L1, are more prominent on myeloid cells or B cells in young hosts, but can be expressed at high levels on tumour cells and on CD8⁺ T cells in aged mice [80]. Defining age-specific strategies to reduce those inhibitory signals and reverse the hyporesponsiveness or exhaustion of aged T cells while reducing potential cytotoxic effects is important in developing these promising immunotherapies.

PD-1

PD-1 is immunopathogenic in cancers by impeding anti-tumour PD-1⁺ T cells. Monoclonal α PD-1 antibodies (α PD-1) have demonstrated remarkable clinical efficacy against a variety of cancers and two distinct α PD-1 antibodies are US Food and Drug Administration (FDA)-approved to treat melanoma, renal cell carcinoma, non-small-cell lung cancer, lymphoma and head and neck cancer.

PD-1 is expressed preferentially on the surface of effector-memory (CD44^{hi}CD62L^{lo}) T cells that increase with ageing. CD4⁺PD-1⁺ T cells from old mice exhibit proliferative hyporesponsiveness, suggesting that the up-regulation of surface-expressed PD-1 could contribute to the age-dependent functional decline in effector-memory T cells [81]. PD-1 increases on T cells with age [82]. α PD-1 improves T cell functions in aged mice [83], although this strategy has not been reported in aged hosts with cancer to our knowledge. We showed that rapamycin reduces age-related T cell PD-1 expression, and PD-1⁺ cells in rapamycin-treated mice were more functional *versus* PD-1⁺ T cells in untreated, aged mice [82], suggesting that rapamycin or another mammalian target of rapamycin (mTOR) inhibitor could improve anti-tumour immunity in aged hosts, as was shown for the ability of the related molecule, everolimus, to improve B cell immunity to influenza vaccine in aged humans [84].

PD-ligand 1 (PD-L1)

PD-L1 is an immune co-signalling molecule that signals through PD-1. Like α PD-1, α PD-L1 is thought to work as cancer immunotherapy by protecting PD-1⁺ anti-tumour T cells from inhibition by tumour PD-L1 expression [77,85–88].

Most old naive CD8⁺ T cells in mice are reportedly PD-L1⁺ *versus* 25% in young mice. Aged CD8⁺ T cells showed reduced proliferation, but α PD-L1 improved their proliferation to a level similar to young CD8⁺ T cells *in vitro*. α PD-L1 improved anti-tumour immunity in aged hosts to young host levels in a mouse lymphoma model [80].

We showed that α PD-L1 treats B16 melanoma in young but not aged mice, but that treatment efficacy was partially

restored by combining α PD-L1 with anti-cytotoxic T lymphocyte antigen 4 (α CTLA-4) [89]. The above study [80] tested α PD-L1 in a lymphoma model in BALB/c mice, whereas we studied a carcinoma in BL6 mice. Thus, either lymphoma *versus* carcinoma or host genetic background, among other considerations, could help to explain different results with α PD-L1.

CTLA-4

The anti-CTLA-4 antibody ipilimumab was the first immune check-point inhibitor agent that clearly benefited human cancer by improving overall and disease-free survival in metastatic melanoma [90]. There are no reports, to our knowledge, of published studies of age effects of anti-CTLA-4 in human cancer. We showed that α CTLA-4 was the only single-agent immunotherapy effective in aged mice in the B16 melanoma model [89].

Immune check-point agonists

OX40 agonists and CD137 agonists are discussed above under T cells [31,34].

Adoptive cell transfers

Older adoptive cell strategies (passive vaccination) using DC or tumour-infiltrating lymphocytes to treat cancer were generally poorly effective. Chimeric antigen receptor (CAR) cell transfers have shown great promise more recently in haematological malignancies. To make a CAR-expressing cell, an artificial antigen recognition receptor (the CAR) is engineered into a lymphocyte that is then transfused into the patient [91]. To our knowledge, there are no published reports on the effects of aged T cells on generating CAR T cells, aside from the recognition that *in vitro* culture of T cells from older patients generally yields fewer CAR T cells *versus* younger patients. To the extent that aged T cells have reduced cytolytic or homing potential, they could generate less effective CAR T cells for cancer immunotherapy. Means to boost aged T cell functions discussed above could potentially be used to increase efficacy of CAR T cells made from aged cells. As natural killer (NK) cells and $\gamma\delta$ T cells do not require major histocompatibility matching to kill their targets, CAR engineered into such cells could potentially be useful to treat aged cancer patients with CAR NK or CAR $\gamma\delta$ T cells, as these strategies are under investigation in younger hosts. Similarly, to the extent that aged DC have reduced T cell-activating or antigen-presenting functions, their utility in DC adoptive cell transfers could be limited, although means to improve aged DC functions discussed above could be tested in this regard.

mTOR inhibitors

mTOR inhibitors at doses lower than used usually for immunosuppression can improve antigen-specific immunity, including against cancers [92]. We showed that rapamycin can reduce T cell PD-1 expression and improves PD-1⁺ T cell function in aged mice [47] and improves $\gamma\delta$ T cell-mediated anti-cancer immunity [93]. Rapamycin prolongs life in mice even when given late in life [94]. Thus, mTOR inhibitors could be used as adjuncts to various cancer immunotherapy agents to improve aged immune function, as we showed is possible [82], with potentially additional life or health extension effects.

mTOR signals are critical to T cell differentiation, and it is well known that mTOR suppression can promote T_{reg} cell function differentiation [82,95]. Thus, there is some concern that small molecule inhibitors could reduce anti-tumour immunity by promoting T_{regs}, as these can be detrimental to anti-tumour immunity [36]. However, in appropriate settings and at sufficiently low doses, small molecule mTOR inhibitors produce net benefits to anti-tumour immunity [47,82,92,93] and do not appear to promote increased T_{reg} numbers or function [82]. We showed recently that low-dose rapamycin could be beneficial to, whereas typical therapeutic rapamycin doses could impair, anti-cancer immunity (Yang *et al.*, in revision). More work on this important issue is needed.

Caloric restriction

Caloric restriction suppresses mTOR similar to rapamycin. When used during 6–8 months, caloric restriction improved α CD40 treatment responses in models of sarcoma and breast cancer when mice were aged 12 months. Tumour antigen-specific CD4⁺ but not CD8⁺ T cell priming was similar to young controls [96]. Certain types of caloric restriction are tolerable in aged patients, and in young subjects can improve anti-tumour immunity [97]. However, mTOR suppression can reduce immunity in the aged [24] and caloric restriction at too late an age or at too great a caloric reduction can harm aged hosts. This interesting intervention merits additional studies in aged hosts to mitigate cancer treatment symptoms, or prevent or treat cancer.

Tumour microenvironment

The tumour microenvironment consists of tumour, immune and stromal cells. Until recently, most studies of ageing and cancer have focused upon the tumour and the immune system, but tumour microenvironmental/stromal effects are now also recognized as important drug discovery areas. The aged tumour microenvironment could be more immunosuppressive than in young hosts. Factors include increased M2 macrophages and MDSC that result from chronic, low-grade age-related inflammation. This aged, protumorigenic tissue

microenvironment could be compromised further by tumour cells that can attract detrimental MDSC, T_{regs} or neutrophils that not only inhibit anti-tumour immunity but also produce protumorigenic factors [63,98]. Primary prostate stromal fibroblasts from older *versus* younger patients produce more proinflammatory factors [99].

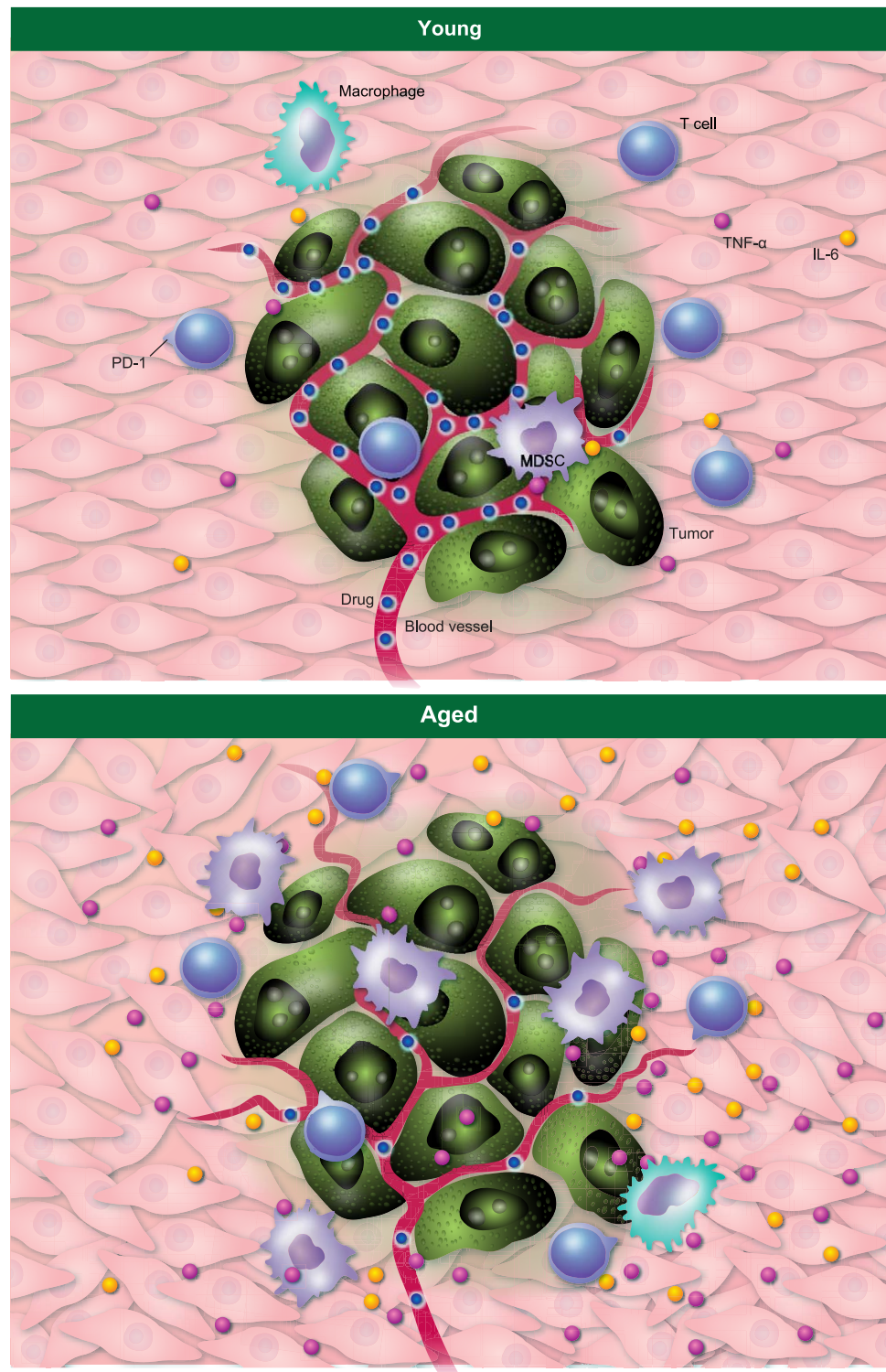
A transgenic murine model of prostate cancer and benign prostate hyperplasia found an age-related increase in TGF- β 1 that resulted in altered tissue architecture consistent with local inflammation [100], a result corroborated in prostates of aged normal mice [101]. Aged primary fibroblasts attract CD4⁺ T cells that promote prostate epithelial cell proliferation. Macrophages, neutrophils and CD8⁺ T cells promote prostate cancer cell proliferation [90].

Fibroblasts in the aged melanoma microenvironment can contribute to melanoma growth and progression. Aged fibroblasts undergo senescence and can drive increased tumour metastases and angiogenesis through production of secreted frizzled-related protein 2. Older fibroblasts also produced fewer ROS scavengers that can promote DNA damage [6]. Finally, in studies of an agonist α CD40 antibody, aged mice failed to respond to cancer treatment from defective CD40 signalling in tumour microenvironmental CD40 signals, not to T cell-intrinsic effects [33]. Thus, therapeutically modifying stromal elements, in particular myeloid cells or their products, could be relevant for elderly cancer patients.

Toll-like receptor (TLR) agonists

Bacille Calmette–Guérin (BCG) is an attenuated *Mycobacterium bovis* that is FDA-approved to treat non-muscle invasive bladder cancer. It is a strong TLR agonist. Although its mechanism of anti-cancer action is incompletely understood, it is thought to promote anti-cancer immunity. Observational data suggest that age could influence response to BCG therapy in bladder cancer. In a trial of BCG plus IFN- α immunotherapy in bladder cancer, aged patients had increased relapse rates following BCG compared to younger patients [102]. Of the patients who were aged 61–70 years *versus* patients older than 80 years, cancer-free survival was 39 *versus* 61%, respectively, at a median follow-up of 24 months ($P = 0.002$). In analyses of aged patients receiving BCG for non-muscle invasive bladder cancer, age was an independent risk for progression and the 2-year progression-free survival was 87% among patients aged less than 75 years compared to 65% among patients aged greater than 75 years [103]. In a large ($n = 805$) cohort of patients with bladder cancer age was found to have a small but measurable association with response to BCG therapy [104]. Nevertheless, the influence of competing morbidities and selective surgical management of elderly patients *versus* immune effects is not clear from these studies. Thus, although BCG is the sole agent to our knowledge for which age-specific immunotherapy

Fig. 2. Examples of age effects in the tumour microenvironment. The aged tumour microenvironment can have elevated levels of proinflammatory molecules that reduce anti-tumour immunity such as interleukin (IL)-6 (yellow dots) and tumour necrosis factor (TNF)- α (purple dots). Green cells = tumour. Myeloid cells, including myeloid-derived suppressor cells (MDSC) (light purple) and macrophages (light green) that inhibit anti-tumour immunity can also be increased in the aged tumour microenvironment. These cells can be a source of detrimental molecules. Local IL-6 and other factors can increase MDSC. T cells programmed death 1 (PD)-1 expression can be increased, suggesting reduced T cell functions. As the Wnt antagonist secreted frizzled-related protein 2 (sFRP2) can promote angiogenesis, we have taken the liberty of showing bigger and more numerous blood vessels in the aged microenvironment, although more work is needed in this area. Drug delivery could be reduced from abnormal vasculature.



effects in humans has been reported extensively, a clear conclusion on age-related immune effects cannot be made from these data. As BCG is standard-of-care immunotherapy and bladder cancer patients tend to be older than for

many other cancer types, these data merit follow-up. There are no reports to our knowledge of age effects on efficacy of other approved or investigational TLR agonists for cancer treatment.

Immunotherapy toxicity mitigation

Cancer immunotherapy in aged mice can generate rapid and/or lethal toxicities in distinct organs due to proinflammatory cytokine production, both systemically and locally in affected tissues. Proinflammatory effects could be driven by macrophages rather than T or NK cells. Myeloid cell depletion during therapy mitigates these toxicities without significantly compromising treatment efficacy. Aged macrophages from mice and normal humans produce higher proinflammatory cytokines, including TNF- α and IL-6 *versus* young macrophages. Blocking TNF- α in aged mice also improves survival and reduces toxicities without loss of anti-tumour effects [3,4]. As α TNF- α , α IL-6 and α IL-6R antibodies are FDA-approved, this concept merits further investigation. However, as other studies have shown that IL-6 and TNF- α administration can improve aged T cell function [30], much additional work is required to understand how such approaches could be incorporated safely and effectively into cancer immunotherapy strategies.

Microbiome effects

The gut microbiome affects systemic immunity and is affected by age [105–107]. Two recent reports showed that gut microbes affected anti-cancer immunotherapy in human subjects receiving α PD-L1 [108] or α CTLA-4 [109]. There is currently much justified interest in understanding microbial effects on anti-cancer immunity although, to our knowledge, age effects are not yet reported in cancer immunotherapy. As the gut microbiome can affect anti-cancer treatments through immune effects [110], including modulation of tumour microenvironmental myeloid cells [111] which themselves are also affected by age [82], it is reasonable for there to be age-associated microbial effects on anti-cancer immunotherapy, an area deserving of additional studies.

Conclusions

Immune therapy for cancer, including in elderly people, has a strong scientific rationale. Recent breakthroughs in understanding the basis for cancer-driven immune dysfunction have led to the development of highly successful anti-cancer immunotherapies. Nonetheless, much of our understanding of tumour immunity derives from studies of younger hosts. There is now a growing, but still relatively small, body of literature demonstrating age effects on cancer immunity and responses to immunotherapies, which have led in some instances to demonstrations of novel approaches to improving anti-cancer immunotherapy in aged hosts most at risk for cancer.

'Immune decline' is an inaccurate term, and 'immunosenescence' and 'inflammageing' describe specific attributes of an aged immune system. We propose the term 'age related immune dysfunction' (ARID) to describe the totality of age-

related immune changes to include increased or decreased numbers of certain immune cells, appearance of novel immune cell populations, increased proinflammatory or immune suppressive functions, restricted T cell receptor (TCR) repertoire and the myriad other changes that age brings to immunity. To the extent that underlying ARID can be reversed in aged cancer patients, clinically relevant anti-tumour immunity could potentially be achieved, as has now been demonstrated clearly.

Challenges in developing effective age-appropriate cancer immunotherapy

Improved understanding of tumour-specific immunopathology and related age-specific effects will help to guide more effective treatment approaches. Individual immunotherapy agents are unlikely to treat most cancers effectively, and thus optimal means to combine agents and approaches need to be defined, including specific agents, doses and scheduling. We need to identify biomarkers that differentiate aged patients with sufficiently responsive tumour-specific immunity from those in whom additional adjuncts will be needed. Adjuncts for those patients with impaired immunity could include adoptive transfer of rejuvenated or young CAR-expressing cells, novel antigen-presenting cells, thymic transplants, gene therapy [including with caspase 9/clustered regularly interspaced short palindromic repeats (CRISPR) approaches] or other approaches.

Toxicity mitigation strategies that do not compromise clinical efficacy are also needed, as is identification of rational combination therapies to reduce dosage of potentially harmful individual agents. Studies of the effect of age on the human microbiome are in their infancy, and could help understanding of the heterogeneity of responses to immunotherapy observed both in humans and in preclinical models. Tumour mutational burden clearly affects the efficacy of many types of cancer immunotherapies. Mutational burden will be dictated, among other considerations, by immune editing. We thus need a clearer understanding of age effects on immune editing and tumour mutational burden, which could produce related or individual contributions to cancer immunotherapy efficacy. Finally, we need to deepen understanding of age effects on tumour stroma, including vasculature effects that can alter immune cell trafficking and drug delivery. An illustration of some challenges is shown in Fig. 2.

As we use ever more powerful but costly and potentially harmful approaches in ever-older cancer patients, economic, feasibility and ethical issues will need to be addressed.

Acknowledgements

Financial support is acknowledged for T. C. (CA170491, CA54174, CDMRP, The Holly Beach Public Library, The

Owens Foundation, The Barker Foundation and the Skinner endowment).

Disclosure

None to declare.

Author contributions

V. H., R. S. S. and T. J. C. wrote the manuscript. V. H. and A. P. performed some experiments described.

References

- Pardoll D. T cells and tumours. *Nature* 2001; **411**:1010–2.
- Pardoll D. Does the immune system see tumors as foreign or self? *Annu Rev Immunol* 2003; **21**:807–39.
- Bouchlaka MN, Murphy WJ. Impact of aging in cancer immunotherapy: the importance of using accurate preclinical models. *Oncoimmunology* 2013; **2**:e27186.
- Bouchlaka MN, Schisel GD, Chen M *et al.* Aging predisposes to acute inflammatory induced pathology after tumor immunotherapy. *J Exp Med* 2013; **210**:2223–37.
- Hurez V, Daniel BJ, Sun L *et al.* Mitigating age-related immune dysfunction heightens the efficacy of tumor immunotherapy in aged mice. *Cancer Res* 2012; **72**:2089–99.
- Kaur A, Webster MR, Marchbank K *et al.* sFRP2 in the aged microenvironment drives melanoma metastasis and therapy resistance. *Nature* 2016; **532**:250–4.
- Schreiber RD, Old LJ, Smyth MJ. Cancer immunoeediting: integrating immunity's roles in cancer suppression and promotion. *Science* 2011; **331**:1565–70.
- Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoeediting. *Annu Rev Immunol* 2004; **22**:329–60.
- Koebel CM, Vermi W, Swann JB *et al.* Adaptive immunity maintains occult cancer in an equilibrium state. *Nature* 2007; **450**:903–7.
- Matsushita H, Vesely MD, Koboldt DC *et al.* Cancer exome analysis reveals a T-cell-dependent mechanism of cancer immunoeediting. *Nature* 2012; **482**:400–4.
- DuPage M, Mazumdar C, Schmidt LM, Cheung AF, Jacks T. Expression of tumour-specific antigens underlies cancer immunoeediting. *Nature* 2012; **482**:405–9.
- Strauss DC, Thomas JM. Transmission of donor melanoma by organ transplantation. *Lancet Oncol* 2010; **11**:790–6.
- Stephens JK, Everson GT, Elliott CL *et al.* Fatal transfer of malignant melanoma from multiorgan donor to four allograft recipients. *Transplantation* 2000; **70**:232–6.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; **100**:57–70.
- Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even warburg did not anticipate. *Cancer Cell* 2012; **21**:297–308.
- Demaria S, Pikarsky E, Karin M *et al.* Cancer and inflammation: promise for biologic therapy. *J Immunother* 2010; **33**:335–51.
- Grivnickov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell* 2010; **140**:883–99.
- Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis* 2009; **30**:1073–81.
- Negrini S, Gorgoulis VG, Halazonetis TD. Genomic instability – an evolving hallmark of cancer. *Nat Rev Mol Cell Biol* 2010; **11**:220–8.
- De Bock K, Cauwenberghs S, Carmeliet P. Vessel abnormalization: another hallmark of cancer? Molecular mechanisms and therapeutic implications. *Curr Opin Genet Dev* 2010; **21**:73–9.
- Teschendorff AE, Menon U, Gentry-Maharaj A *et al.* Age-dependent DNA methylation of genes that are suppressed in stem cells is a hallmark of cancer. *Genome Res* 2010; **20**:440–6.
- Hakim FT, Flomerfelt FA, Boyiadzis M, Gress RE. Aging, immunity and cancer. *Curr Opin Immunol* 2004; **16**:151–6.
- Haynes BF, Sempowski GD, Wells AF, Hale LP. The human thymus during aging. *Immunol Rev* 2000; **22**:253–61.
- Goldberg EL, Romero-Aleshire MJ, Renkema KR *et al.* Life-span-extending caloric restriction or mTOR inhibition impair adaptive immunity of old mice by distinct mechanisms. *Aging Cell* 2015; **14**:130–8.
- Yang H, Youm YH, Vandanmagsar B *et al.* Obesity accelerates thymic aging. *Blood* 2009; **114**:3803–12.
- Surh CD, Boyman O, Purton JF, Sprent J. Homeostasis of memory T cells. *Immunol Rev* 2006; **211**:154–63.
- Anderlini P, Przepiora D, Seong C *et al.* Factors affecting mobilization of CD34+ cells in normal donors treated with filgrastim. *Transfusion* 1997; **37**:507–12.
- Fulop T, Larbi A, Pawelec G. Human T cell aging and the impact of persistent viral infections. *Front Immunol* 2013; **4**:271.
- Fulop T, Witkowski JM, Le Page A, Fortin C, Pawelec G, Larbi A. Intracellular signalling pathways: targets to reverse immunosenescence. *Clin Exp Immunol* 2016.
- Haynes L, Eaton SM, Burns EM, Rincon M, Swain SL. Inflammatory cytokines overcome age-related defects in CD4 T cell responses in vivo. *J Immunol* 2004; **172**:5194.
- Bansal-Pakala P, Croft M. Defective T cell priming associated with aging can be rescued by signaling through 4-1BB (CD137). *J Immunol* 2002; **169**:5005.
- Ruby CE, Weinberg AD. OX40-enhanced tumor rejection and effector T cell differentiation decreases with age. *J Immunol* 2009; **182**:1481–9.
- Ruby CE, Weinberg AD. The effect of aging on OX40 agonist-mediated cancer immunotherapy. *Cancer Immunol Immunother* 2009; **58**:1941–7.
- Lustgarten J, Dominguez AL, Thoman M. Aged mice develop protective antitumor immune responses with appropriate costimulation. *J Immunol* 2004; **173**:4510–5.
- Curiel TJ. Tregs and rethinking cancer immunotherapy. *J Clin Invest* 2007; **117**:1167–74.
- Curiel TJ. Regulatory T cells and treatment of cancer. *Curr Opin Immunol* 2008; **20**:241–6.
- Curiel TJ, Coukos G, Zou L *et al.* Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004; **10**:942.
- Zou W. Regulatory T cells, tumour immunity and immunotherapy. *Nat Rev Immunol* 2006; **6**:295–307.
- Zhang H, Podojil JR, Luo X, Miller SD. Intrinsic and induced regulation of the age-associated onset of spontaneous experimental autoimmune encephalomyelitis. *J Immunol* 2008; **181**:4638–47.

- 40 Rosenkranz D, Weyer S, Tolosa E *et al.* Higher frequency of regulatory T cells in the elderly and increased suppressive activity in neurodegeneration. *J Neuroimmunol* 2007; **188**:117–27.
- 41 Zhao L, Sun L, Wang H, Ma H, Liu G, Zhao Y. Changes of CD4+CD25+Foxp3+ regulatory T cells in aged Balb/c mice. *J Leukoc Biol* 2007; **81**:1386–94.
- 42 Kryczek I, Liu R, Wang G *et al.* FOXP3 defines regulatory T cells in human tumor and autoimmune disease. *Cancer Res* 2009; **69**:3995–4000.
- 43 Kozłowska E, Biernacka M, Ciechomska M, Drela N. Age-related changes in the occurrence and characteristics of thymic CD4(+) CD25(+) T cells in mice. *Immunology* 2007; **122**:445–53.
- 44 Thomas DC, Mellanby RJ, Phillips JM, Cooke A. An early age-related increase in the frequency of CD4+ Foxp3+ cells in BDC2.5NOD mice. *Immunology* 2007; **121**:565–76.
- 45 Sun L, Hurez VJ, Thibodeaux SR *et al.* Aged regulatory T cells protect from autoimmune inflammation despite reduced STAT3 activation and decreased constraint of IL-17 producing T cells. *Aging Cell* 2012; **11**:509–19.
- 46 Lages CS, Suffia I, Velilla PA *et al.* Functional regulatory T cells accumulate in aged hosts and promote chronic infectious disease reactivation. *J Immunol* 2008; **181**:1835–48.
- 47 Tsaknaris L, Spencer L, Culbertson N *et al.* Functional assay for human CD4+CD25+ Treg cells reveals an age-dependent loss of suppressive activity. *J Neurosci Res* 2003; **74**:296–308.
- 48 Garg SK, Delaney C, Toubai T *et al.* Aging is associated with increased regulatory T-cell function. *Ageing Cell* 2014; **13**:441–8.
- 49 Fessler J, Ficjan A, Duftner C, Dejaco C. The impact of aging on regulatory T-cells. *Front Immunol* 2013; **4**:231.
- 50 Gregg R, Smith CM, Clark FJ *et al.* The number of human peripheral blood CD4+ CD25high regulatory T cells increases with age. *Clin Exp Immunol* 2005; **140**:540–6.
- 51 Hwang KA, Kim HR, Kang I. Aging and human CD4(+) regulatory T cells. *Mech Ageing Dev* 2009; **130**:509–17.
- 52 Dominguez AL, Lustgarten J. Implications of aging and self-tolerance on the generation of immune and antitumor immune responses. *Cancer Res* 2008; **68**:5423–31.
- 53 Sharma S, Dominguez AL, Lustgarten J. High accumulation of T regulatory cells prevents the activation of immune responses in aged animals. *J Immunol* 2006; **177**:8348–55.
- 54 Jackaman C, Dye DE, Nelson DJ. IL-2/CD40-activated macrophages rescue age and tumor-induced T cell dysfunction in elderly mice. *Age (Dordr)* 2014; **36**:9655.
- 55 Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. *Nat Rev Cancer* 2012; **12**:265–77.
- 56 Zou W, Machelon V, Coulomb-L'Hermin A *et al.* Stromal-derived factor-1 in human tumors recruits and alters the function of plasmacytoid precursor dendritic cells. *Nat Med* 2001; **7**:1339–46.
- 57 Curiel TJ, Wei S, Dong H *et al.* Blockade of B7-H1 improves myeloid dendritic cell-mediated antitumor immunity. *Nat Med* 2003; **9**:562–7.
- 58 Agrawal A, Agrawal S, Tay J, Gupta S. Biology of dendritic cells in aging. *J Clin Immunol* 2008; **28**:14–20.
- 59 Khong A, Nelson DJ, Nowak AK, Lake RA, Robinson BW. The use of agonistic anti-CD40 therapy in treatments for cancer. *Int Rev Immunol* 2012; **31**:246–66.
- 60 Tang YC, Thoman M, Linton PJ, Deisseroth A. Use of CD40L immunoconjugates to overcome the defective immune response to vaccines for infections and cancer in the aged. *Cancer Immunol Immunother* 2009; **58**:1949–57.
- 61 Geiger H, Rudolph KL. Aging in the lympho-hematopoietic stem cell compartment. *Trends Immunol* 2009; **30**:360–5.
- 62 Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* 2008; **454**:436–44.
- 63 Jackaman C, Radley-Crabb HG, Soffe Z, Shavlakadze T, Grounds MD, Nelson DJ. Targeting macrophages rescues age-related immune deficiencies in C57BL/6J geriatric mice. *Aging Cell* 2013; **12**:345–57.
- 64 Mahbub S, Deburghgraeve CR, Kovacs EJ. Advanced age impairs macrophage polarization. *J Interferon Cytokine Res* 2012; **32**:18–26.
- 65 Watkins SK, Egilmez NK, Suttles J, Stout RD. IL-12 rapidly alters the functional profile of tumor-associated and tumor-infiltrating macrophages *in vitro* and *in vivo*. *J Immunol* 2007; **178**:1357–62.
- 66 Ostrand-Rosenberg S, Sinha P. Myeloid-derived suppressor cells: linking inflammation and cancer. *J Immunol* 2009; **182**:4499–506.
- 67 Li H, Han Y, Guo Q, Zhang M, Cao X. Cancer-expanded myeloid-derived suppressor cells induce energy of NK cells through membrane-bound TGF-beta 1. *J Immunol* 2009; **182**:240–9.
- 68 Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumour-bearing mice. *J Immunol* 2008; **181**:5791–802.
- 69 Marigo I, Dolcetti L, Serafini P, Zanovello P, Bronte V. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunol Rev* 2008; **222**:162–79.
- 70 Huang B, Pan PY, Li Q *et al.* Gr-1+CD115+ immature myeloid suppressor cells mediate the development of tumor-induced T regulatory cells and T-cell energy in tumor-bearing host. *Cancer Res* 2006; **66**:1123–31.
- 71 Marvel D, Gabrilovich DI. Myeloid-derived suppressor cells in the tumor microenvironment: expect the unexpected. *J Clin Invest* 2015; **125**:3356–64.
- 72 Verschoor CP, Johnstone J, Millar J *et al.* Blood CD33(+)/HLA-DR(-) myeloid-derived suppressor cells are increased with age and a history of cancer. *J Leukoc Biol* 2013; **93**:633–7.]
- 73 Grizzle WE, Xu X, Zhang S *et al.* Age-related increase of tumor susceptibility is associated with myeloid-derived suppressor cell mediated suppression of T cell cytotoxicity in recombinant inbred BXD12 mice. *Mech Ageing Dev* 2007; **128**:672–80.
- 74 Enioutina EY, Bareyan D, Daynes RA. A role for immature myeloid cells in immune senescence. *J Immunol* 2011; **186**:697–707.
- 75 Ishikawa S, Matsui Y, Wachi S, Yamaguchi H, Harashima N, Harada M. Age-associated impairment of antitumor immunity in carcinoma-bearing mice and restoration by oral administration of *Lentinula edodes* mycelia extract. *Cancer Immunol Immunother* 2016; **65**:961–72.
- 76 Pardoll D, Drake C. Immunotherapy earns its spot in the ranks of cancer therapy. *J Exp Med* 2012; **209**:201–9.
- 77 Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 2012; **12**:252–64.

- 78 Canaday DH, Parker KE, Aung H, Chen HE, Nunez-Medina D, Burant CJ. Age-dependent changes in the expression of regulatory cell surface ligands in activated human T-cells. *BMC Immunol* 2013; **14**:45.
- 79 Channappanavar R, Twardy BS, Krishna P, Suvas S. Advancing age leads to predominance of inhibitory receptor expressing CD4 T cells. *Mech Ageing Dev* 2009; **130**:709–12.
- 80 Mirza N, Duque MA, Dominguez AL, Schrum AG, Dong H, Lustgarten J. B7-H1 expression on old CD8+ T cells negatively regulates the activation of immune responses in aged animals. *J Immunol* 2010; **184**:5466–74.
- 81 Shimada Y, Hayashi M, Nagasaka Y, Ohno-Iwashita Y, Inomata M. Age-associated up-regulation of a negative co-stimulatory receptor PD-1 in mouse CD4+ T cells. *Exp Gerontol* 2009; **44**:517–22.
- 82 Hurez V, Dao V, Liu A *et al.* Chronic mTOR inhibition in mice with rapamycin alters T, B, myeloid, and innate lymphoid cells and gut flora and prolongs life of immune-deficient mice. *Ageing Cell* 2015; **14**:945–56.
- 83 Lages CS, Lewkowich I, Sproles A, Wills-Karp M, Chougnet C. Partial restoration of T cell function in aged mice by *in vitro* blockade of the PD-1/PD-L1 pathway. *Ageing Cell* 2010; **9**:785–98.
- 84 Mannick JB, Del Giudice G, Lattanzi M *et al.* mTOR inhibition improves immune function in the elderly. *Sci Transl Med* 2014; **6**:268ra179.
- 85 Topalian SL, Hodi FS, Brahmer JR *et al.* Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* 2012; **366**:2443–54.
- 86 Dong H, Strome SE, Salomao DR *et al.* Tumor-associated B7-H1 promotes T-cell apoptosis: a potential mechanism of immune evasion. *Nat Med* 2002; **8**:793–800.
- 87 Brahmer JR, Tykodi SS, Chow LQ *et al.* Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med* 2012; **366**:2455–65.
- 88 Taube JM, Anders RA, Young GD *et al.* Colocalization of inflammatory response with B7-h1 expression in human melanocytic lesions supports an adaptive resistance mechanism of immune escape. *Sci Transl Med* 2012; **4**:127ra37.
- 89 Figueiredo ASP, Hurez V, Liu A. TJCI. Age and sex affect α CTLA-4 efficacy alone and combined with α B7-H1 or regulatory T cell depletion in a melanoma model. *J Immunol* 2016; **196**:213.4.
- 90 Hodi FS, O'Day SJ, McDermott DF *et al.* Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010; **363**:711–23.
- 91 June CH, Riddell SR, Schumacher TN. Adoptive cellular therapy: a race to the finish line. *Sci Transl Med* 2015; **7**:280ps7.
- 92 Pedicord VA, Cross JR, Montalvo-Ortiz W, Miller ML, Allison JP. Friends not foes: CTLA-4 blockade and mTOR inhibition cooperate during CD8+ T cell priming to promote memory formation and metabolic readiness. *J Immunol* 2015; **194**:2089–98.
- 93 Dao V, Liu Y, Pandeswara S *et al.* Immune stimulatory effects of rapamycin are mediated by stimulation of antitumor $\gamma\delta$ T cells. *Cancer Res*, 2016; **76**:5970–82.
- 94 Hasty P, Livi CB, Dodds SG *et al.* eRapa restores a normal life span in a FAP mouse model. *Cancer Prev Res (Phila)* 2014; **7**: 169–78.
- 95 Pollizzi KN, Patel CH, Sun IH *et al.* mTORC1 and mTORC2 selectively regulate CD8+ T cell differentiation. *J Clin Invest* 2015; **125**:2090–108.
- 96 Farazi M, Nguyen J, Goldufsky J *et al.* Caloric restriction maintains OX40 agonist-mediated tumor immunity and CD4 T cell priming during aging. *Cancer Immunol Immunother* 2014; **63**:615–26.
- 97 Di Biase S, Lee C, Brandhorst S *et al.* Fasting-mimicking diet reduces HO-1 to promote T cell-mediated tumor cytotoxicity. *Cancer Cell* 2016; **30**:136–46.
- 98 Jackaman C, Nelson DJ. Are macrophages, myeloid derived suppressor cells and neutrophils mediators of local suppression in healthy and cancerous tissues in aging hosts? *Exp Gerontol* 2014; **54**:53–7.
- 99 Begley LA, Kasina S, MacDonald J, Macoska JA. The inflammatory microenvironment of the aging prostate facilitates cellular proliferation and hypertrophy. *Cytokine* 2008; **43**:194–9.
- 100 Barron DA, Strand DW, Ressler SJ *et al.* TGF-beta1 induces an age-dependent inflammation of nerve ganglia and fibroplasia in the prostate gland stroma of a novel transgenic mouse. *PLOS ONE* 2010; **5**:e13751.
- 101 Bianchi-Frias D, Vakar-Lopez F, Coleman IM, Plymate SR, Reed MJ, Nelson PS. The effects of aging on the molecular and cellular composition of the prostate microenvironment. *PLOS ONE* 2010; **5**:e12501.
- 102 Joudi FN, Smith BJ, O'Donnell MA, Konety BR. The impact of age on the response of patients with superficial bladder cancer to intravesical immunotherapy. *J Urol* 2006; **175**:1634–9. discussion 9–40.
- 103 Margel D, Alkhateeb SS, Finelli A, Fleshner N. Diminished efficacy of bacille Calmette–Guerin among elderly patients with nonmuscle invasive bladder cancer. *Urology* 2011; **78**:848–54.
- 104 Herr HW. Age and outcome of superficial bladder cancer treated with bacille Calmette–Guerin therapy. *Urology* 2007; **70**:65–8.
- 105 Yatsunenkov T, Rey FE, Manary MJ *et al.* Human gut microbiome viewed across age and geography. *Nature* 2012; **486**: 222–7.
- 106 Pitt JM, Vetizou M, Waldschmitt N *et al.* Fine-tuning cancer immunotherapy: optimizing the gut microbiome. *Cancer Res* 2016; **76**:4602–7.
- 107 Zitvogel L, Ayyoub M, Routy B, Kroemer G. Microbiome and anticancer immunosurveillance. *Cell* 2016; **165**:276–87.
- 108 Sivan A, Corrales L, Hubert N *et al.* Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science* 2015; **350**:1084–9.
- 109 Vetizou M, Pitt JM, Daillere R *et al.* Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science* 2015; **350**:1079–84.
- 110 Viaud S, Saccheri F, Mignot G *et al.* The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. *Science* 2013; **342**:971–6.
- 111 Iida N, Dzutsev A, Stewart CA *et al.* Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science* 2013; **342**:967–70.