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## Manipulating the epigenome for the treatment of urological malignancies<sup>☆,☆☆</sup>

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### Abstract

Urological malignancies (cancers of the prostate, bladder, kidney and testes) account for 15% of all human cancers and more than 500,000 deaths worldwide each year. This group of malignancies is spread across multiple generations, affecting the young (testicular) through middle and old-age (kidney, prostate and bladder). Like most human cancers, urological cancers are characterized by widespread epigenetic insult, causing changes in DNA hypermethylation and histone modifications leading to silencing of tumor suppressor genes and genomic instability. The inherent stability yet dynamic plasticity of the epigenome lends itself well to therapeutic manipulation. Epigenetic changes are amongst the earliest lesions to occur during carcinogenesis and are essentially reversible (unlike mutations). For this reason, much attention has been placed over the past two decades on deriving pharmacological compounds that can specifically target and reverse such epi-mutations, either halting cancer on its developmental trajectory or reverting fully formed cancers to a more clinically manageable state. This review discusses DNA methyltransferase and histone deacetylase inhibitors that have been extensively studied in preclinical models and clinical trials for advanced and metastatic urological cancers.

### Keywords

Epigenome; Prostate cancer; Bladder cancer; Kidney cancer; Testicular cancer; Therapeutics

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### Conflict of interest statement

The authors declare no conflict of interest.

## 1. Introduction

Worldwide, urological malignancies account for ~15% of all non-cutaneous cancers and 12% of cancer related deaths. Urological malignancies include cancers of the prostate, bladder kidney and testes. Of all urological malignancies, prostate cancer is by far the most common with nearly one million men diagnosed annually and almost 300,000 deaths from this disease (Ferlay et al., 2010; Center et al., 2012) (Table 1, Fig. 2). Wide global variation exists for prostate cancer; incidence rates vary >25-fold worldwide, with ~75% of cases occurring in developed countries (Draisma et al., 2003). Epidemiological studies support a strong role for dietary constituents in modifying prostate cancer risk, with diets rich in lycopenes and cruciferous vegetables afforded a protective effect, while large intake of dairy, saturated fat and burnt meat is associated with increased risk. Age is the strongest risk factor for prostate cancer, with the majority of tumors occurring in men over the age of 60.

Cancers of the kidney and bladder may develop in either sex but are significantly more common in men. Bladder cancer is the 2nd most common urological malignancy and the 9th most common malignancy worldwide, with ~330,000 new cases diagnosed and ~130,000 deaths per annum (Ferlay et al., 2010). Risk factors for the development of bladder cancer include age (50–70 years), molecular abnormalities (most notably *TP63*, *EGFR*, *TP53* and *RBI* genes and Ras and p21 proteins), chemical and environmental exposures (most notably cigarette smoking) and chronic irritation (Kaufman et al., 2009).

Tumors of the kidney are considerably less common than malignancies of the bladder and prostate, although the incidence has been slowly rising over the past 3 decades (Ferlay et al., 2010). Renal cell carcinoma (RCC) has the highest mortality rate among all the genitourinary tract tumors, with a third of patients dying from their disease. Risk factors for RCC include age (50–70 years), cigarette smoking, obesity, hypertension and silencing of the *VHL* tumor suppressor gene, which drives tumor angiogenesis through VEGF over-expression (Cohen & McGovern, 2005).

Testicular cancer is the least common of all the urological cancers, accounting for <2% of male cancers and with an estimated life-time risk of ~1 in 250. It is however, the most common solid cancer in Caucasian males aged between 15 and 35 years (Houldsworth et al., 2006; Rosen et al., 2011). Various risk factors have been identified, including a range of testicular abnormalities, however in the majority of cases there is no identifiable etiology. Genetic abnormalities including Klinefelter's syndrome (47XXY), XY dysgenesis and Down's syndrome (trisomy of chromosome 21) are all associated with increased risk (Horwich et al., 2006).

The pattern of disease presentation for each of these malignancies varies widely as does their natural history, clinical characteristics and prognosis. This review discusses the evidence to date for the potential role of epigenetic modalities in novel therapeutic strategies for urological cancers.

### 1.1. Prostate cancer therapies

The last two decades have seen a dramatic rise in the incidence of prostate cancer, largely due to widespread serum prostate specific antigen (PSA) testing in developed countries (Fig. 1A). Prostate cancer is an extremely heterogeneous condition ranging from microscopic, well-differentiated tumors that may be clinically indolent to aggressive cancers that have a high likelihood of invasion and metastasis. The greatest challenge arising from PSA-detected prostate cancers is the substantial over-detection and hence over-treatment of clinically indolent disease. Nevertheless, the European Randomized Study of Screening for Prostate Cancer has shown a reduction in prostate cancer specific mortality due to PSA screening after 11 years of follow-up (Schroder et al., 2012).

Treatment options for localized prostate cancer are potentially curative by surgery (open, laparoscopic or robotic assisted radical prostatectomy) or radiation therapy (external-beam or brachytherapy) and have comparable long term survival rates. Conservative management through active surveillance or watchful waiting is now offered to patients who meet certain criteria for low-risk disease (Heidenreich et al., 2011; Tosoian et al., 2011). In recent years, advances have been made in minimally-invasive therapies, offering similar curative treatment rates but without the side-effects of conventional invasive procedures. Imaging techniques can provide precise tumor localization, increasing interest in “focal” therapies selectively targeting cancer foci, such as high-intensity focused ultrasound (HIFU), cryotherapy and photodynamic therapy (Eggerer et al., 2010; Cordeiro et al., 2012).

Advanced prostate cancer is managed by androgen-deprivation therapy. However the therapeutic response is short-lived and the disease recurs typically within 18–24 months in a castration-resistant form. Docetaxel-based chemotherapy plus prednisone is the standard of care for metastatic castration-resistant prostate cancer (CRPC), giving a median survival benefit of ~3 months (Petrylak et al., 2004; Tannock et al., 2004). In recent years, several new agents have been developed targeting different mechanistic disease pathways fundamentally altering the treatment landscape in CRPC. These treatment options include a new cytotoxic agent (Cabazitaxel), immunotherapy (sipuleucel-T) and androgen receptor-signaling inhibitors (abiraterone acetate and MDV3100) (Sartor & Fitzpatrick, 2012). These advances highlight the significant role that targeted molecular based therapeutics can play in alleviating disease burden.

### 1.2. Bladder cancer therapies

More than 90% of bladder cancers are transitional cell carcinomas (TCC), 5% are squamous cell carcinomas and are largely associated with chronic urinary infection with *Schistosoma haematobium*, and less than 2% are adenocarcinomas (Table 2) (Kaufman et al., 2009). Of newly diagnosed TCC, ~70% are non-invasive superficial tumors that are potentially curative by local endoscopic resection and adjuvant chemotherapy. However, up to 70% of noninvasive TCC recur and between 10 and 20% of these become muscle-invasive. Muscle-invasive bladder cancer is treated by radical cystectomy to remove the entire bladder and its associated draining lymph nodes; a procedure associated with significant morbidity and a 5-year survival rate of ~50% (Stein & Skinner, 2006). The use of neo-adjuvant cisplatin-containing combination chemotherapy has been shown to modestly improve survival to 56%

(Winqvist et al., 2004). Alternatively, radical radiotherapy can be administered to surgically unfit candidates however outcomes are less favorable than with surgery.

It is clear that given the limitations of current clinical treatments both in preventing recurrence of superficial bladder cancer and ensuring survival from muscle-invasive disease, novel therapeutic targets and agents are urgently required. Targeted molecular therapies for localized bladder cancer have been relatively slow in forthcoming. Overexpression of ERBB2 has been found to be an independent predictor of survival in a proportion of muscle-invasive TCC (Kruger et al., 2002). This resulted in phase I and II clinical trials investigating paclitaxel and daily radiation therapy with trastuzumab (anti-ERBB2 directed therapy) for patients with muscle-invasive TCC that over-expressed ERBB2, and who are not fit for cystectomy (Hussain et al., 2007).

### 1.3. Kidney cancer therapies

RCC is the most common form of kidney cancer and represents a spectrum of histological subtypes, signifying varied biology, genetics and behavior (Table 1) (Cohen & McGovern, 2005). Due to the paucity of early clinical manifestations, detection of RCC is often delayed, with one-third of patients diagnosed with advanced or metastatic disease (Castellano et al., 2012). However, earlier and incidental detection due to abdominal imaging for other medical indications has facilitated identification of tumors at more treatable and less advanced stages.

Historically, treatment has been predominately surgical, regardless of stage; RCC are inherently resistant to radiotherapy and chemotherapy. Increased understanding of the pathobiology of RCC has led to the development of targeted immunotherapies, which have revolutionized the treatment of metastatic RCC. Targeted agents, such as the oral, multi-targeted receptor tyrosine kinase inhibitors sunitinib and sorafenib, the mammalian target of rapamycin (mTOR) inhibitors temsirolimus and everolimus (Motzer et al., 2008) and the VEGF-targeted monoclonal antibody bevacizumab (given in combination with interferon- $\alpha$ ) and pazopanib (Sternberg et al., 2010) are all well established treatments (Escudier et al., 2007a, 2007b; Rini et al., 2008; Motzer & Molina, 2009; Castellano et al., 2012).

### 1.4. Testicular cancer therapies

Testicular neoplasms denote an incongruent collection of tumors, which are classified according to their histological patterns (Table 2). The vast majority are classified as germ-cell tumors, derived from transformation of primordial germ cells (Houldsworth et al., 2006). Advances in the treatment of testicular cancer over the past number of decades, particularly in chemotherapeutic agents, have ensured that this disease has one of the highest cure rates of all cancers. The estimated 5 year mortality rate in the United States is 4%, ranking the multimodal management of testicular cancer the gold-standard model for the treatment of all solid tumors (Horwich et al., 2006).

Testicular tumors often progress silently until presentation with a painless or slightly aching scrotal lump. Serum tumor markers including alpha-fetoprotein (AFP), human chorionic gonadotrophin (hCG), and lactate dehydrogenase (LDH) are used for prognostication and treatment monitoring (Albers et al., 2011). The mainstay of treatment for testicular cancer is

radical inguinal orchidectomy followed by adjuvant surveillance, adjuvant chemotherapy and adjuvant radiotherapy, depending on tumor histology and stage (Krege et al., 2008; Tandstad et al., 2011).

## 2. Epigenetic modalities

Following the conclusive unveiling of the human genome in 2001, revealing a surprisingly low constitution of ~25,000 genes, it became clear that genetics alone was not capable of explaining the vast complexity displayed by eukaryotes. Epigenetics, defined as heritable alterations in gene expression that are not accompanied by changes in our genetic code, is recognized as a key regulator of our DNA — controlling its packaging and transcription to RNA, and as a surrogate mechanism to mutation in yielding phenotypic diversity (Fig. 2). The crux upon which this epigenetic regulatory system is based lies with the manner in which DNA is compacted within the nucleus. Epigenetic “readers, writers and erasers” catalyze a variety of biochemical modifications to either the DNA (methylation) or the histone proteins (acetylation, methylation, ubiquitylation, sumoylation, phosphorylation, citrullination, ADP-ribosylation), to establish co-operative regulation of chromatin conformation (Dawson et al., 2012). The dynamic plasticity that epigenetic modifications display is paramount to enabling the complex temporal and spatial alterations in gene expression that are required for normal eukaryotic development and adult homeostasis (Gu et al., 2010).

### 2.1. DNA methylation and the methyltransferase family

The best characterized epigenetic mark — DNA methylation involves addition of a methyl group from the universal methyl donor S-adenosylmethionine to a nucleotide residue, which in mammals is primarily the fifth carbon position of cytosine in cytosine–phosphate–guanine (CpG) dinucleotides, converting cytosine to 5-methylcytosine or the ‘fifth base of the genome’ (Bird, 1986). However, recent studies of human embryonic stem cells have shown methylation to exist in certain restricted non-CpG contexts (Lister et al., 2009).

DNA methylation is typically associated with transcriptional silencing. This is achieved directly (by prohibiting transcription factor binding) (Mancini et al., 1999) and indirectly (by attracting methyl-binding domain proteins that interact with histone deacetylases and nucleosome remodeling factors to compact the chromatin, thus rendering the promoter inaccessible for transcription) (Jones et al., 1998; Nan et al., 1998). As such, intergenic regions and repeat elements are heavily methylated (Eden et al., 2003; Roman-Gomez et al., 2005), whereas actively transcribed genes are characterized by unmethylated promoters. De novo hypermethylation of promoter CpG islands (short 200–2000 bp stretches of DNA, heavily enriched in CpG sites) is a hallmark of most human cancers, acting as a transcriptional gene silencing mechanism (Baylin, 2005).

Humans contain three catalytically-active DNA methyltransferase enzymes (DNMT1, DNMT3a, DNMT3b) which regulate DNA methylation, in addition to an associated regulatory protein (DNMT3L) (Denis et al., 2011). DNMT1 is essential for maintaining patterns of methylation over replicative time by copying the pattern of the parent strand onto the new strand during DNA replication (Song et al., 2012). DNMT3a and DNMT3b possess

the capacity to target unmethylated CpG sites and therefore function in establishing de novo methylation patterns (Rhee et al., 2002; Chedin, 2011). DNMT3L does not possess innate catalytic capabilities but does play an important role in orchestrating DNA methylation by regulating activity of the other DNMT3 members (Jurkowska et al., 2011; Van Emburgh & Robertson, 2011). Related family member DNMT2 shows poor in vitro DNA methyltransferase activity acting instead on RNA methylation (Schaefer et al., 2010; Thiagarajan et al., 2011).

DNA methylation patterns become altered during the very early transition from a normal cell phenotype to the pre-neoplastic state, with promoter hypermethylation of tumor suppressor genes (Brooks et al., 1998; Kang et al., 2004; Christoph et al., 2006b, 2007). This is followed by hypomethylation of intergenic sequences (Aporntewan et al., 2011), retroviral oncogenes (Goering et al., 2011), proto-oncogene promoters (Wang et al., 2007) and pericentromeric regions (Narayan et al., 1998), all facilitating epigenetic instability. Overall, there is significant reduction in the 5-methylcytosine content of the genome.

There is strong evidence to show that methylation patterns are clonally inherited during prostate tumor progression (Yegnasubramanian et al., 2004). However, recent advances in whole-genome methylation analysis have identified distinct subsets of genes that become hypermethylated in late stage prostate cancer and during metastatic progression, during which epigenetic changes become markedly more heterogeneous (Kim et al., 2011; Mahapatra et al., 2012). LINE1 hypomethylation is significantly enhanced in metastatic prostate cancer (Yegnasubramanian et al., 2008) and it has recently been shown that ETS-negative prostate cancers show reduced LINE1 methylation compared to ETS-positive tumors (Kim et al., 2011). It also appears that non-invasive and invasive bladder cancers may arise via distinct epigenetic pathways, in which non-invasive tumors display a distinct hypomethylated phenotype whereas invasive tumors exhibit widespread promoter hypermethylation (Wolff et al., 2010).

## 2.2. Histone modifications

Histone acetylation is the best studied post-translational modification of the histone octamer. Histone acetyltransferases catalyze the addition of an acetyl group from acetyl coenzyme A to a lysine typically at the histone N-terminus, which negates the positive histone charge, resulting in chromatin expansion and facilitating transcription (Marmorstein, 2001). This process is counter-balanced by the action of the histone deacetylase (HDAC) family. In removing the acetyl group, the HDACs restore the positive charge of the histone proteins, promoting chromatin compaction and transcriptional repression (Ng & Bird, 2000). Hence, acetylation is a core modulator of regulating the transition between eu- and heterochromatin.

Histone methylation is less well characterized and can have both activating (e.g. H3K4me2, H3k36me2) and repressing (e.g. H3k27me2/3 and H3K9me2/3) effects on gene expression, depending on the amino acid position and the number of methylated residues (Klose & Zhang, 2007; Crea et al., 2012b). Histone methylation is catalyzed by the histone methyltransferase (HMT) family of enzymes, with lysine undergoing mono-, di- or tri-methylation and arginine becoming di-methylated either symmetrically or non-symmetrically (Bedford & Richard, 2005; Martin & Zhang, 2005). While acetylation

negates the positive histone charge, methylation alters the basicity and hydrophobicity of histones, changing the nucleosome's affinity for a multitude of transcription factors.

The histone lysine demethylase (KDM) class of epigenetic enzymes removes both activating and repressive methyl groups from specific histone lysine residues causing chromatin reconfiguration. Of particular interest in this review are lysine-specific demethylase 1 (LSD1 or KDM1A) and members of the Jumonji domain containing family (JMJD2A, JMJD2C/GASC1, JMJD2D) that also function as p53 and nuclear receptor co-regulators to cooperatively mediate sex steroid hormone-induced transcription through their KDM activity (Metzger et al., 2005; Huang et al., 2007; Perillo et al., 2008).

During tumorigenesis, the histone methylome has been reported to undergo insult. Aberrant HMT activity has been implicated in survival and proliferation and epithelial–mesenchymal transition in prostate cancer (Ezponda et al., 2012), in bladder cancer (Kottakis et al., 2011) and in RCC (Duns et al., 2010). KDM expression is also dysregulated in human cancers, whereby KDMs may act as either tumor suppressors or oncogenes, depending on their gene regulatory function. For example, aberrant expression of LSD1 is reported throughout different stages of prostate and bladder cancer (Kauffman et al., 2011; Crea et al., 2012b). As a component of co-repressor complexes, LSD1 demethylates activating methyl marks at H3K4. In the presence of hormone-activated AR however, the LSD1–JMJD2 complex cooperate to remove the repressive H3K9 tri-methylation mark, thus facilitating AR-dependent gene expression and prostate tumor cell proliferation (Metzger et al., 2005; Wissmann et al., 2007).

### 2.3. On the road — epimutations as drivers of cancer

In spite of widespread heterogeneity between tumor types, it is believed that all cancers strive towards the acquisition of eight key traits: self-sufficiency in growth signals, resistance to anti-proliferative signals, apoptotic evasion, limitless replicative potential, angiogenesis, metastasis and, most recently, bioenergetic reprogramming and immune evasion (Hanahan & Weinberg, 2011). Add to this the 'epigenetic progenitor model of cancer' (Feinberg et al., 2006), and we can begin to understand how epigenetic alterations in precursor cells can create genomic instability, which fosters additional genetic 'hits', such as point mutations and deletions, unwanted transcription of proviral and retrotransposon repeats, leading to disruption of their surrounding genes, further epigenetic catastrophe and rapid acquisition of the cancerous phenotype (Eden et al., 2003; Chalitchagorn et al., 2004). Virtually all tiers of the epigenetic hierarchy undergo subversion during tumor evolution.

## 3. DNA methylation inhibitors (DNMTi)

Endogenously, DNA methylation provides a means to rapidly and stably alter eukaryotic gene expression in response to stimuli. DNA methylation can be considered as an attractive therapeutic target because it is reversible and relatively simple to manipulate biochemically. By globally inducing hypomethylation, genes which have become aberrantly silenced, in particular tumor suppressors, may be reactivated. To date, numerous DNA methylation inhibitors (DNMTi) have been developed with varying pre-clinical results and the most promising clinical translatability for hematological malignancies with FDA approval granted

for patients with myelodysplastic syndrome (Tsai et al., 2012). Notably, the efficacy of hypomethylating agents is dependent on the tumor genotype (Pulukuri & Rao, 2005; Richter et al., 2007).

### 3.1. Re-achieving the healthy genome with nucleoside analogs

The first class of DNMTi comprises simple nucleoside analogs: azacytidine (5-Aza-CR; trade name Vidaza), a simple cytidine derivative and 5-aza-2'-deoxycytidine (5-Aza-CdR; decitabine; trade name Dacogen), a deoxycytidine derivative. Via active demethylation of tumor suppressor genes and cytotoxicity arising from incorporation into nucleic acids, these DNMTi exert potent anti-tumorigenic activity (Jones & Liang, 2009). In vitro evidence suggests that DNMTi may also passively induce gradual demethylation by competitively binding DNMTs over replicative time (Kuo et al., 2007). A significant drawback of these agents, however, is their relative toxicity and limited potency at tolerable levels. Therefore, efforts have been made to develop novel azacytidine derivatives (Matousova et al., 2011). Zebularine, although initially developed in 1961, has recently attracted attention as a ribonucleoside DNMTi (Champion et al., 2010). It exhibits greater intrinsic stability and lower toxicity than its counterparts, thus enabling longer oral administration, facilitating maintenance of a demethylated state (Billam et al., 2010).

Multiple preclinical studies of nucleoside analogs have validated their potential use in restoring expression of a myriad of genes in in vitro models of prostate, bladder, renal and testicular cancer at a consistent rate over time (Christoph et al., 2006a). Over a decade ago, treatment of the androgen-insensitive DU145 cell line with 5,6-dihydro-5-azacytidine was shown to restore androgen receptor (AR) reactivation (Nakayama et al., 2000) as well as responsiveness to dihydrotestosterone (DHT) (Izbicka et al., 1999). Treatment with histone deacetylase inhibitor (HDACi) trichostatin A (TSA) invariably shows substantial synergistic gene reactivation in vitro when used in combination with a DNMTi (Cameron et al., 1999). In renal cancer, TGF $\beta$  receptor type II expression, in addition to TGF $\beta$  responsiveness, was restored by 5-Aza-CdR (Zhang et al., 2005). Furthermore, combined treatment with 5-Aza-CdR and genistein, a non-toxic isoflavone, significantly induced cell cycle arrest and halted proliferation across three renal carcinoma cell lines, over genistein alone (Majid et al., 2009). More recently, it was shown in T24 bladder cancer cell line that 5-Aza-CdR is capable of concomitant reactivation of host mRNA and intragenic intronic microRNA expression (Saito et al., 2009). Nucleoside analogs have also been revealed as capable of potentially targeting epithelial-to-mesenchymal transition and meta-static spread in bladder cancer with 5-Aza-CdR restoring prostatic expression in cell lines, with concomitant reactivation of E-cadherin and recovery of epithelial morphology (Chen et al., 2009).

Bladder cancer was the first malignancy to show epigenetic reactivation of a tumor suppressor gene after oral delivery of a hypomethylating agent. Following promising results by demethylating the p16 gene promoter and globally decreasing hypermethylated regions in T24 human bladder carcinoma cells, studies on zebularine were escalated to in vivo models. Using the xenograft murine model of EJ6 human bladder carcinoma culture in nude mice, zebularine was shown clinically efficient when delivered either intravenously or orally,



producing an 11% average maximum weight change between test cases and controls at 1000 mg/kg dosing (Cheng et al., 2003, 2004).

### 3.2. Restoration of radio- and chemosensitivity

A promising avenue for the clinical utility of hypomethylating agents is their potential to promote radio- and/or chemosensitivity through reactivation of specific enzymes involved in drug metabolism (Shen et al., 2007; Zhang et al., 2010). Combined treatment of prostate cancer cell line DU145 with docetaxel (Taxotere<sup>R</sup>) and either 5-Aza-CdR or 5-Aza-CR enhanced docetaxel sensitivity through demethylation and reactivation of DNA damage response gene *GADD45A* (Ramachandran et al., 2009) and tumor suppressors E-cadherin and maspin and significantly reduced proliferation over either agent alone (Hurtubise & Momparler, 2004). Similarly, 5-Aza-CdR was found to enhance the apoptotic effect of cisplatin (trade name: cisplatin) in DU145 cells (Fang et al., 2004). In RCC, cytotoxicity of vinblastine was increased both in vitro and in a Caki-1 xenograft model by co-treatment with 5-Aza-CdR (Iwata et al., 2011). Hypermethylation of the pro-apoptotic *XAF1* gene has been associated with the apoptotic resistant phenotype of RCC that emerges post-immunotherapy. Rescuing *XAF1* expression by either 5-Aza-CdR or a DNMT antisense oligonucleotide has been shown to restore susceptibility to interferon-induced apoptosis (Reu et al., 2006).

Cisplatin resistance poses a key problem in the treatment of bladder cancer. 5-Aza-CdR has been shown to sensitize TCC to the effects of cisplatin (Shang et al., 2008).

Hypermethylation of homeobox gene *TLX3* has been associated with chemoresistance in 21% of patients and overexpression of *TLX3* restored chemosensitivity (Tada et al., 2011). Similarly, it has been shown that 5-Aza-C reactivation of proapoptotic gene *TMS1* restores cisplatin sensitivity in the UMUC3 bladder cancer cell line (Ramachandran et al., 2011).

Testicular germ cell tumors are characterized by hypersensitivity to cisplatin and DNMTi, which probably mechanistically relates to their high endogenous levels of DNMT3b (Beyrouthy et al., 2009). In comparison to other solid tumors, very low nanomolar concentrations of 5-Aza-CdR were capable of recovering expression of 3 hypermethylated genes (*RASSF1A*, *MGMT*, *HOXA9*), and activating the p21 and ATM pathways, resulting in decreased proliferation and cell survival. This study went on to demonstrate that in testicular germ cell tumors that develop cisplatin resistance, sensitivity can be restored by 5-Aza-CdR (Beyrouthy et al., 2009).

### 3.3. Beyond the nucleoside: alternative DNMTi

Due to the inherent toxicity and adverse effects of nucleoside analogs, efforts are ongoing to establish synthetic or dietary non-nucleoside compounds that do not become integrated into the DNA yet can still effectively exert hypomethylating activity. Procainamide, an antiarrhythmic drug with observed in vitro demethylating capacity, was shown to be capable of hypomethylating the prostate cancer stalwart, *GSTP1*, in the LNCaP cell line as well as in xenograft athymic nude mouse models (Lin et al., 2001). Disulfiram, a thiol-reactive compound used in the treatment of chronic alcoholism, was proven to possess intrinsic DNMTi properties, globally decreasing the quantity of 5-methylcytosine and capable of

reactivating *APC* and *RAR $\beta$*  through inhibition of DNMT1 activity. Treatment also resulted in decreased proliferation and increased apoptosis in cell line and xenograft prostate cancer models (Lin et al., 2011). Curcumin (diferuloylmethane), a phytochemical compound found endogenously in turmeric curry, also appears to possess demethylating properties, restoring expression of transcription factor *NEUROG1* in prostate cancer LNCaP cells through preventing MeCP2 binding to the *NEUROG1* promoter (Shu et al., 2011). It was recently shown that curcumin is also capable of demethylating and reactivating noncoding microRNA genes, namely tumor suppressor microRNA miR-203 in bladder cancer cell lines. This resulted in down-regulation of miR-203 target genes Akt2 and Src and culminated in decreased proliferation and increased apoptosis (Saini et al., 2011).

### 3.4. DNMTi in clinical trials for urological cancers

While the full mechanistic action of many potential hypomethylating agents is still being actively investigated at the preclinical stage, several notable clinical trials have been completed for urological malignancies (Table 3). Overall the outcomes have not been particularly hopeful, with perhaps the most promising results seen in metastatic CRPC patients. As early as 1998, a clinical trial took place for 5-aza-2'-deoxycytidine on men who developed metastatic prostate cancer following complete androgen blockade and flutamide withdrawal. Of the 14 men recruited, 12 were evaluable with 2 cases displaying a time to progression greater than 10 weeks. Interestingly, both men were African-American, suggesting ethnicity may be an important variable for consideration (Thibault et al., 1998). More recently, a phase II clinical trial of Vidaza in CRPC patients (n=36) revealed a significant impact upon PSA doubling time (PSA-DT), the median PSA-DT of the treated group (2.8 months) far exceeded that of the control group (1.5 months) (Sonpavde et al., 2011).

In bladder, renal and testicular cancers, few clinical trials have been carried out on these agents. Several have been terminated due to toxicity or poor uptake (Winquist et al., 2006). More recent trials are studying the synergistic benefit of combined epigenetic and immunotherapies (Table 3).

## 4. Histone deacetylase inhibitors (HDACi)

Beyond the DNA level, post-translational modifications of histone proteins also function in modulating eukaryotic gene expression and therefore may be pharmacologically targeted. Among these, histone acetylation has received most attention for its capacity to neutralize the positive charge of histones, thereby facilitating euchromatin expansion and gene transcription. Hence, a multitude of HDAC inhibitors (HDACi) have been developed, targeting various members of the HDAC family of enzymes (Khan & La Thangue, 2012).

Due to their diversity, HDACi are broadly categorized into 5 groups based on chemical composition and structure: short-chain fatty acids, hydroxamic acids, cyclic tetrapeptides, benzamides and aliphatic acids. Alternatively, HDACi may be classified based on the member(s) of the HDAC enzyme family they target, with further disparity both between and within the five structural groups (Khan & La Thangue, 2012). Some act specifically on certain HDACs, for example, entinostat (MS-275) inhibits class I HDACs, whereas others

such as trichostatin A, vorinostat and panobinostat can inhibit multiple classes of HDAC. Perhaps surprisingly, it has been shown that <10% of genes are influenced by HDACi directly (Peart et al., 2005). Therefore, it appears that HDAC inhibition serves as an initial instigator in restoring normal gene expression but that phenotypic effects are perpetuated by multiple downstream mechanisms. This notion is supported by the observation that HDACs interact with numerous non-histone targets, many of which are important for driving tumorigenesis (Johnstone & Licht, 2003), such as  $\alpha$ -tubulin (Marcus et al., 2005), important for cell motility and metastasis, bcl-6 (Dhordain et al., 1998), an oncogene involved in apoptotic regulation, and p53 (Murphy et al., 1999). Although the exact mechanisms and pathways involved in HDACi remain unclear, several recurrent anti-tumorigenic phenotypic outcomes have been observed. HDACi mediate selective tumor-cell death through several pathways, including induction of apoptosis, growth arrest and autophagy, and effects on DNA repair, mitosis, and angiogenesis (Long et al., 2009; Khan & La Thangue, 2012). Pan-HDACi belinostat potently inhibited the growth of prostate cancer cell lines by increasing expression of p21 and inducing G2/M arrest and reduced tumor growth and metastatic spread in orthotopic prostate cancer tumor models (Qian et al., 2008). Similarly it was shown in RCC cell lines, that panobinostat (LBH589) induced degradation of both Aurora A and B kinases through a proteasome-mediated pathway by targeting HDAC3 and HDAC6, resulting in G2/M arrest and apoptosis (Cha et al., 2009).

Multiple lines of evidence support a role for combining pharmacological compounds to sensitize tumors to the effects of HDACi. Interferon alpha enhances the anti-tumor effects of HDACi valproic acid (VPA) on prostate cancer growth (Hudak et al., 2012). In bladder cancer, both cisplatin and gemcitabine have been demonstrated to potentiate the antitumor effects of trichostatin A (Jeon et al., 2011). Vorinostat was found to synergistically improve the anti-cancer activity of mTOR inhibitor temsirolimus in a panel of RCC cell lines in vitro and in two xenograft models in vivo. Combined treatment significantly reduced survivin expression leading to apoptosis and a reduction in angiogenesis (Mahalingam et al., 2010). Similarly in mouse models of prostate cancer, the combination of panobinostat with the mTOR inhibitor everolimus resulted in enhanced anti-tumor activity (over either agent alone) mediated by increased p21 and p27 expression and reduced angiogenesis and tumor proliferation via androgen receptor, c-Myc and HIF-1 $\alpha$  signaling (Ellis et al., 2011).

#### 4.1. HDACi in clinical trials for urological cancers

The main challenge in translating HDACi into cancer therapeutics is trying to establish which molecules will yield maximum efficacy while minimizing off-target toxicity; difficult due to the pleiotropic nature of the enzymes involved. Several HDACi have entered clinical trials for the treatment of prostate and renal cancer, with mixed success (Table 4).

In 2009, Bradley and colleagues released the results of their phase II clinical trial of vorinostat (Suberoylanilide Hydroxamic Acid) for metastatic CRPC (Bradley et al., 2009). In this study, 27 patients with disease progression following chemotherapy were recruited. Overall survival, PSA decline rate and safety were amongst those listed as secondary endpoints. Unfortunately, only 7% patients displayed stable disease and no PSA decline of  $\geq 50\%$  was noted. Furthermore, all participants ceased treatment before the 6 month interval

due to associated toxicities. Hence, the toxicity of vorinostat at such doses is clearly not translatable to the clinic.

Molife et al. completed a phase II clinical trial in 2010, examining the potential therapeutic benefit of romidepsin (depsipeptide) in metastatic CRPC (Molife et al., 2010). Via treatment with this HDACi, it was hoped that AR signaling could be abrogated by inducing acetylation of heat shock protein 90. The primary endpoint was the absence of radiological progression at 6 months. Only two patients were reported as displaying radiological partial response (RECIST) lasting 6 months in conjunction with PSA decline 50%. Additionally, 11 patients had to be discontinued from trial due to adverse events. Hence, romidepsin does not appear of clinical use at such concentrations and toxicities. Results of a phase I combination trial of oral panobinostat alone in conjunction with docetaxel in 16 CRPC patients showed that docetaxel did not augment panobinostat efficacy at high dose (20 mg) but in the second arm of the study in patients receiving a lower dose of 15 mg panobinostat, 63% patients showed at least a 50% decline in PSA levels (Rathkopf et al., 2010).

Similarly, the potential to treat metastatic RCC with HDACi romidepsin and panobinostat has been disappointing showing either no response or very high levels of toxicity, respectively (Stadler et al., 2006; Hainsworth et al., 2011).

#### 4.2. Inhibitors targeting the histone methylome

EZH2 belongs to the polycomb group of transcriptional repressors, and forms the catalytic subunit of the Polycomb repressive complex 2 (PRC2), which catalyzes trimethylation on H3K27, triggering gene silencing. It also acts as a molecular scaffold, attracting DNMTs thus bridging two distinct epigenetic modes of gene regulation: chromatin conformation and DNA methylation (Vire et al., 2006). EZH2 over-expression is associated with metastatic progression and poor prognosis in urological malignancies (Varambally et al., 2002; Raman et al., 2005; Wagener et al., 2008), although it does not appear to exert its characteristic oncogenic properties in testicular cancer (Hinz et al., 2010).

EZH2 inhibition may be employed as an anti-angiogenic and anti-metastatic strategy. One potential compound is 3-dezaneplanocin-A (DZNeP), a HMT inhibitor, which inhibits S-adenosyl-homocysteine, required for EZH2-dependent lysine methylation. DZNeP exerts anti-tumor activity in many cancer cell lines including prostate, where it also blocks cancer cell migration and invasion (Crea et al., 2012a). Several selective small molecule inhibitors of EZH2 have recently been described that decrease global H3K27me3 levels and reactivate silenced PRC2 target genes, such as GSK126 (McCabe et al., 2012) and EPZ005687 (Knutson et al., 2012).

The possibility of pharmacologically targeting KDMs has been demonstrated (Kruidenier et al., 2012). A range of KDM inhibitors (KDMi) have been identified (Rotili & Mai, 2011; Crea et al., 2012b). In prostate cancer, LSD1 remains the most promising target for KDMi, and several substrate analogs, polyamine analogs and inhibitors of the mono-amine-oxidase domain have been developed such as tranilcypromine and clorgyline. Tranilcypromine and its analogs effectively inhibit H3K4 demethylation and may inhibit progression to castration resistance (Crea et al., 2012b). Clorgyline exerts anti-proliferative and pro-differentiation

activity on high-grade prostate cancer cells in part mediated by inhibiting growth factor signaling and down-regulating HMT EZH2 (Zhao et al., 2009). Namoline is a more specific LSD1 inhibitor which has been shown to reduce androgen-dependent cell proliferation and xenograft tumor growth (Willmann et al., 2012).

## 5. Future directions: remaining challenges

As we continue to advance our understanding of the molecular pathobiology of urological malignancies, we move towards an improved ability to predict which patients will progress to lethal disease. This in turn will enable patient-orientated individualized treatment and specific targeted therapies for patients with advanced disease.

Without doubt the greatest obstacle in translating epigenetic drugs to clinical use for solid tumors, not just urological malignancies, remains their genome-wide mode of action and thus lack of specificity and hence severe toxicity. Unless hypermethylation of individual tumor suppressor genes can be restored without the side-effects of whole genome hypomethylation, it is difficult to see how DNMTi and HDACi will significantly advance the treatment of solid tumors in their current form. Although HDACs exhibit selective toxicity against tumor cells at nanomolar concentration, their prolonged use is associated with significant toxicity (Prince et al., 2009). A number of plant-derived polyphenols have cancer preventative properties, display structural features compatible with HDAC inhibition and may be more tolerable. For example, epigallocatechin-3-gallate (the polyphenol from green tea) has been shown to cause cell cycle arrest and induce apoptosis in prostate cancer cells by upregulating p21 and Bax through H3 acetylation (increasing promoter accessibility), causing epigenetic modifications and down-regulating HDACs through increasing their proteasomal degradation (Thakur et al., 2012).

While acetylation remains the best studied histone modification to date, it in itself is only one component of the elusive histone code. Multiple other post-translational modifications have been described and are implicated in various cancers and essentially remain largely understudied. Unlike DNMTs and HDACs, for which inhibitors have been developed and discovered, the quest to establish histone methyltransferase inhibitors lies ahead. Whether these agents can be engineered with increased specificity and hence lower toxicity, remains to be seen.

## Abbreviations

<b>CRPC</b>	castration-resistant prostate cancer
<b>DNMT</b>	DNA methyltransferase
<b>DNMTi</b>	DNA methyltransferase inhibitors
<b>HDAC</b>	histone deacetylase
<b>HDACi</b>	histone deacetylation inhibitors
<b>PSA</b>	prostate specific antigen

<b>RCC</b>	renal cell carcinoma
<b>TCC</b>	transitional cell carcinoma

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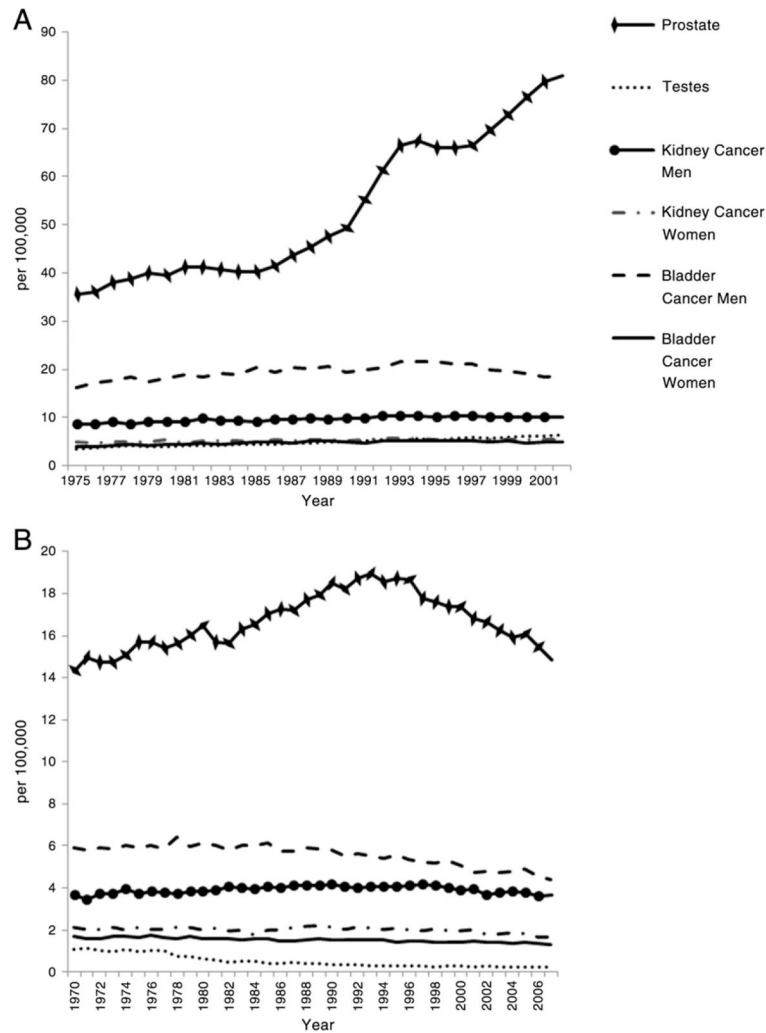
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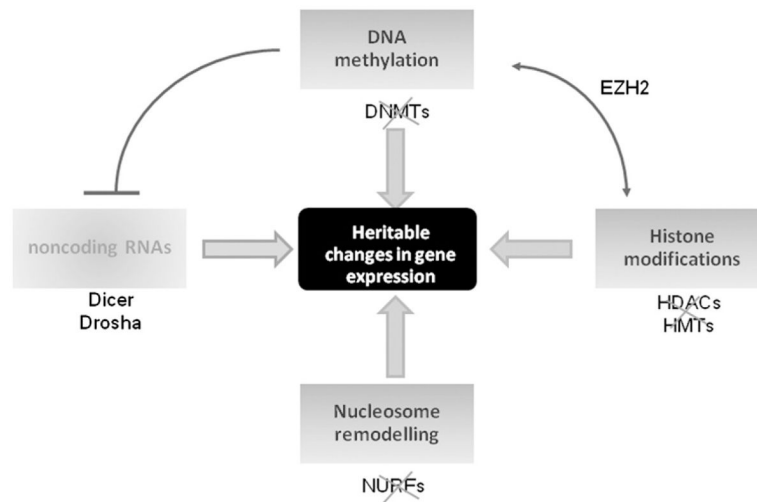
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**Fig. 1.** Global facts and figures for urological cancers. **A)** World age-standardized incidence and **B)** world age-standardized mortality. Data are shown for USA, Canada, Denmark, Finland, France, Norway, Sweden, Netherlands, UK and Australia and were sourced from the WHO International Agency for Research on cancer *CI5plus* Cancer Incidence in Five Continents Annual Dataset (<http://ci5.iarc.fr/CI5plus/ci5plus.htm>) and Cancer Mortality database (<http://www-dep.iarc.fr/WHODb/WHODb.htm>).



**Fig. 2.**

The co-operative network of epigenetic modifications. Crosstalk between DNA methylation, histone modifications, nucleosome remodeling and noncoding RNAs is the cornerstone for epigenetic gene regulation. Evidence now supports a role for small noncoding RNAs in epigenetic gene silencing; believed to be involved indirectly though post-transcriptionally inactivating key epigenetic enzymes. The microRNA class of noncoding RNAs have recently emerged as a target of DNA methylation in cancer cells.

**Table 1**

Summary of facts and figures for urological malignancies.

	<b>Prostate</b>	<b>Bladder</b>	<b>Kidney</b>	<b>Testicular</b>
Number of new cases diagnosed per year <sup>a</sup>	899,102	382,660	273,518	52,322
Number of deaths per year <sup>a</sup>	258,133	150,282	116,368	9,874
Median age at diagnosis (years) <sup>b</sup>	72	73	64	33
Median age at metastasis (years) <sup>b</sup>	82	75	69	33
Major treatments for localized disease	Prostatectomy Radiotherapy	Cystectomy	Nephrectomy	Orchidectomy
Major side effects of treatments	Impotence, incontinence	Impotence, urinary diversion	Minimal	Minimal
Cure-rate for treated localized disease	80%	Superficial=85% Muscle-invasive=50%	70%	>99%
Major treatments for metastatic disease	Hormonal therapy, chemotherapy	Chemotherapy	Targeted meds	Chemotherapy
Mean survival for metastatic disease	22 months	14 months	24 months	23 years

<sup>a</sup>Obtained from GLOBOCAN 2008.<sup>b</sup>Median age according to the US Surveillance, Epidemiology and End Results (SEER) database.



**Table 2**

Classification of urological malignancies at diagnosis.

Tumor type		5-year survival according to pathological stage at diagnosis
Prostate	Epithelial (>95%)	Organ confined prostate cancer (T1–T2): 100%
	Adenocarcinoma (95%)	Locally advanced prostate cancer (T3–T4): >95%
	Urothelial	Metastatic prostate cancer (N+/M+): 29%
	Squamous	
	Basal cell Other (<5%)	
Bladder	Epithelial (>95%)	Superficial bladder cancer & carcinoma in situ (Ta/Tis/T1): >98%
	Transitional cell carcinoma (>90%)	Muscle-invasive bladder cancer (T2–T4): 63%–88%
	Squamous cell carcinoma (1–7%)	Metastatic bladder cancer (N+/M+): 15%–46%
	Adenocarcinoma (2–3%)	
	Other (<5%)	
Kidney	Renal cell carcinoma (>95%)	Organ confined renal cell carcinoma (T1–T2): 81%
	Clear cell renal carcinoma (75–80%)	Locally advanced renal cell carcinoma (T3–T4): 53%–74%
	Papillary renal cell carcinoma (10–15%)	Metastatic renal cell carcinoma (N+/M+): 8%
	Chromophobe renal cell carcinoma (5%)	
	Other (<5%)	
Testicular	Germ cell tumors (90%)	Testicular cancer confined to testis and epididymis (T1–T2): 99%
	Seminoma (48%)	Locally advanced testicular cancer (T3–T4): 96%
	Non-seminomatous (42%)	Metastatic testicular cancer (N+/M+): 72%
	Mixed NSGCT (10%)	
	Sex cord stromal tumors (3%)	
	Other (7%)	

Abbreviations: is=in situ and NSGCT=non seminomatous germ cell tumor.

Survival data obtained from American and European Associations of Urology guidelines

Table 3

Clinical trials of DNMTi for urological cancers.

Drug	Clinical trial identifier	Phase	Status	Protocol	Outcome
<i>Prostate</i>					
Vidaza	NCT00384839	Phase II	Completed	CRPC cases on combined androgen blockade with PSA-DT<3 months treated with 75 mg/m <sup>2</sup> s.c. on days 1–5 of each 28 day cycle for up to 12 cycles where tolerable. n=36	PSA-DT>3 months achieved in 55.8% patients; median progression free survival=12.4 weeks
Decitabine	–	Phase II	Completed	Metastatic cases post-total blockade and flutamide withdrawal were treated with 57 mg/m <sup>2</sup> /dose i.v. as 1 h infusion every 8 h for 3 doses; cycles repeated every 5–8 weeks. n=14	17% had stable disease with progression time>10 weeks
Vidaza, docetaxel and prednisone	NCT00503984	Phase I/II	Recruiting	CRPC patients previously treated with docetaxel. Azacitidine: i.v., days 1–5 of each 3 weekly cycle; docetaxel: i.v. on day 6 of each 3 weekly cycle; prednisone: 5 mg twice daily from day 1 to 21 of each cycle. Predicted enrolment: n=42	NYA
Bladder & testes					
Decitabine	NCT00030615	Phase I	Completed	Advanced bladder and testicular cancer patients received daily escalating doses of decitabine for 4 weeks until MTT was determined.	NYA
<i>Kidney</i>					
Vidaza, bevacizumab	NCT00934440	Phase I/II	Recruiting	Advanced RCC patients receiving: bevacizumab at standard dose of 10 mg/kg IV every two weeks; vidaza: several different doses s.c. dependent on phase I/II. Predicted enrolment: n=23	NYA
Decitabine, IFN $\alpha$ -2B	NCT00561912	Phase II	Terminated	Advanced RCC patients received decitabine: 15 mg/m <sup>2</sup> i.v. daily for 5 days; IFN $\alpha$ -2B s.c. twice daily continuously as of day 1 cycle 3.	Terminated due to poor accrual (n=2)
MG98	NCT00003890	Phase I/II	Terminated	Untreated metastatic RCC patients received 360 mg/m <sup>2</sup> /dose i.v. twice weekly for 3 out of 4 weeks. n=17	No conclusive pattern of decreased DNMT1 activity in PBMCs was detected post MG98 treatment. Terminated due to toxicity.

Abbreviations: MTT: maximum tolerated dose, PBMCs: peripheral blood mononuclear cells, and NYA: not yet available.

Table 4

Clinical trials of HDACi for urological cancers.

Drug	Clinical trial identifier	Phase	Status	Protocol	Outcome
<i>Prostate</i>					
Panobinostat, docetaxel and prednisone	NCT00663832	Phase I	Completed	CRPC received panobinostat 10, 15 or 20 mg i.v. on days 1 and 8 in combination with docetaxel i.v. on day 1 and prednisone p.o. 5 mg bid every day of a 21-day cycle. n=44	Determined the MTD of i.v. Panobinostat is 15 mg in combination with docetaxel and prednisone in patients with CRPC. 63% of patients showed >50% decline in PSA levels
Panobinostat, bicalutamide	NCT00878436	Phase I/II	Recruiting	CRPC patients receiving panobinostat at either 60 or 120 mg per week for 2 consecutive weeks, with one week rest and bicalutamide 50 mg p.o. daily, continuously. Predicted enrolment: n=78	Investigating safety and efficacy of combined treatment of panobinostat at 2 dose levels combined with bicalutamide for CRPC as measured by time to PSA progression and proportion of patients that achieve a 50% PSA decline by 9 months of therapy
Vorinostat	NCT00330161	Phase II	Active, not recruiting	Metastatic CRPC patients (PSA>5 ng/ml, disease progression, adequate organ function) received 400 mg vorinostat orally each day. n=27	7% patients achieved an objective response rate; no PSA decline>50% observed; significant toxicities reported
Romidepsin	NCT00106418	Phase II	Completed	Metastatic CRPC patients received romidepsin 13 mg/m <sup>2</sup> i.v. over 4 h on days 1, 8 and 15 every 4 weeks. n=35	2 patients reached a confirmed radiological partial response of >6 months, in addition to >50% PSA decline
<i>Kidney</i>					
Panobinostat	NCT00550277	Phase II	Completed	Metastatic RCC patients received 45 mg panobinostat twice per week for 8 week cycles with re-evaluation at the end of each cycle; n=20	No objective response observed; generally well-tolerated with minimal side effects
Romidepsin	NCT00106613	Phase II	Completed	Metastatic RCC patients received 13 mg/m <sup>2</sup> romidepsin i.v. on days 1, 8 and 15 over 4 week cycles with disease progression analyzed every 8 weeks. n=29	7% experienced an objective response. Terminated due to toxicity.