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## Infection resistance and tolerance in *Peromyscus* spp., natural reservoirs of microbes that are virulent for humans

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### Abstract

The widely-distributed North American species *Peromyscus leucopus* and *P. maniculatus* of cricetine rodents are, between them, important natural reservoirs for several zoonotic diseases of humans: Lyme disease, human granulocytic anaplasmosis, babesiosis, ehrlichiosis, hard tickborne relapsing fever, Powassan virus encephalitis, hantavirus pulmonary syndrome, and plague. While these infections are frequently disabling and sometimes fatal for humans, the peromyscines display little pathology and apparently suffer few consequences, even when prevalence of persistent infection in a population is high. While these *Peromyscus* spp. are unable to clear some of the infections, they appear to have partial resistance, which limits the burden of the pathogen. In addition, they display traits of infection tolerance, which reduces the damage of the infection. Research on these complementary resistance and tolerance phenomena in *Peromyscus* has relevance both for disease control measures targeting natural reservoirs and for understanding the mechanisms of the comparatively greater sickness of many humans with these and other infections.

### Keywords

vectorborne disease; Borrelia; zoonoses; spirochete; ecoimmunology; ecological immunology

### 1. Introduction

In September 2014 the United States' Institute of Medicine held a forum on “Vector-Borne Diseases: Exploring the Environment, Ecological, and Health Connections” [1]. The workshop highlighted the importance of animal reservoirs and arthropod vectors of pathogens for public health. Changes in climate, landscapes, transportation, and global commerce have brought the animal reservoirs of infectious diseases in ever closer contact with human populations. The majority of the emerging infectious diseases on the National Institute of Allergy and Infectious Diseases' high priority list, are zoonoses [2], which are infections acquired from a vertebrate reservoir, either by direct exposure to, or through an

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arthropod vector, and not another human. The reservoirs are usually mammals, and the vectors are usually either ticks or insects. For the following five high-priority zoonoses a *Peromyscus* species, the white-footed mouse, *P. leucopus*, is a major reservoir: Lyme disease, babesiosis, Powassan virus encephalitis, human granulocytic anaplasmosis, and hard tick-borne relapsing fever (Table 1). In North America another species, *P. maniculatus*, the deer mouse, is an important reservoir for a hantavirus [3] and *Yersinia pestis*, the agent of plague [4].

The familiar names of *Peromyscus* spp. include “mouse”, but the genus belongs to the taxon Cricetidae, together with hamsters, voles, and wood rats [5], not to the more distant clade Muridae, which includes the house mouse *Mus musculus* and, the black rat, *Rattus rattus*, by wide margins the most frequently used animal models in biomedical research. Peromyscines are distributed across North America and in a variety of habitats [6]. The *Peromyscus* species in an area is usually its most abundant mammal [7].

In North America the distribution of the *P. leucopus* largely overlaps with that of the blacklegged tick, *Ixodes scapularis*, which is commonly called the “deer tick”, because of the association of its adult stage with white-tailed deer [8]. *I. scapularis* is the vector for the agents of Lyme disease (*Borrelia burgdorferi*), babesiosis (*Babesia microti*), Powassan virus encephalitis, human granulocytic anaplasmosis (*Anaplasma phagocytophilum*), and hard tick-borne relapsing fever (*Borrelia miyamotoi*) in their various distributions in the northeastern and north-central United States and adjoining regions of Canada. Two newly-described tick-borne agents, *Ehrlichia muris* subsp. *auclairensis* and *Borrelia mayonii* are also carried by *I. scapularis* [9–11], and the ehrlichiosis agent uses *P. leucopus* as reservoir [12]. In the far-western U.S., the vector for *B. burgdorferi*, *A. phagocytophilum*, *B. miyamotoi*, and another *Babesia* species is the western blacklegged tick, *I. pacificus*.

Table 1 lists these tick-borne infections, as well as those transmitted by insects, like fleas, through an intermediate host, or by direct exposure to the rodents or their excrement, as for hantaviruses. The table also includes two other borrelias, a trypanosome species, and a schistosome that infect *P. leucopus* and/or *P. maniculatus* in nature, but for which there are no documented human infections. This list is undoubtedly incomplete. A variety of metazoan parasites, including cestodes (tapeworms), trematodes (flukes), nematodes (round worms), and other exoparasites, besides ticks and fleas, including mites, chiggers, lice, and botfly larvae are known to infect or infest *Peromyscus* spp. [13]. Botfly parasitism of *Peromyscus* spp. is not a subject of this article but has been studied by others, e.g. [14].

Here, I review natural and experimental infections of *Peromyscus* spp. with vector-borne zoonotic pathogens, and highlight the comparative health of these animals during infections with microbes that otherwise cause extensive disease in humans. The emphasis is on *B. burgdorferi* and *P. leucopus*, because that host-pathogen relationship has been most studied. But investigations of the *Peromyscus* model are far out-numbered by literature reports on experiments with *B. burgdorferi* in the house mouse, which is not the natural reservoir of a Lyme disease agent, nor any of the other infections listed in the table. So, there is scope in the review for considering other infections in the white-footed mouse and the deer mouse.

There is also literature on *Peromyscus* spp. and hantaviruses, a pathogen-host relationship that is subject of the article by Vandegrift et al. of this issue [3].

## 2. Resistance to and tolerance of infection in disease reservoirs

Four key terms for what follows are “reservoir”, “competence”, “resistance”, and “tolerance”. These terms, each of which has multiple meanings in wider usage, are defined here for the present context. A *reservoir* in a life-cycle of a vector-borne virus, bacterium, or parasite is a definitive host, upon which the pathogen depends for its long-term maintenance in an environment and which also serves as a blood-meal host for the arthropod vector for the pathogen. For human zoonoses, the reservoir is almost always a vertebrate, usually a mammal. The *competence* of a reservoir in the life cycle is the capacity of that host for transmission of the pathogen to another vertebrate host, either directly or through a vector. In the example of *B. burgdorferi* and its transmission by ticks, reservoir competence of the host mammal can be defined as the proportion of the nymphs bearing infectious levels of the microbe after having fed as larvae upon an infected animal. So, it is not just the capacity of the animal to become infected and to house the pathogen transiently or persistently. The reservoir would also need to be sufficiently infectious for the feeding tick for that tick to subsequently pass through to the next stage, e.g. from larva to nymph, and then retain enough the organism for the tick to transmit the microbe to another vertebrate host at its next blood meal, as a nymph in the example.

In a simple unrestrained system, which does not take into account about the host population’s aggregate fitness, a pathogen proliferates in the vertebrate to numbers high enough for successful transmission before it kills or otherwise disables the host. The *resistance* to the pathogen’s exploitation is the degree to which the host reduces the frequency of infection, increases the rate of pathogen clearance among the infected, or both. Resistance mechanisms act on the invading pathogen. When resistance measures fail or are only partially successful, a fall-back position for the host is to minimize the amount of damage or disease caused by the infection. The mechanisms serving this trait of *tolerance* reduce the net detrimental effects of the microbe. They act on the host and mitigate effects of the infection without altering its development [15]. (This is different from a definition in the field of immunity for “tolerance”, i.e. lack of responsiveness to an antigen.) The damage to be limited could be the consequence of unmoderated responses of the host, e.g. excessive inflammation, or of virulence properties of the microbe, e.g. a toxin.

For long-term sustainable maintenance of a tick-borne pathogen in an environment, a pathogen’s main reservoir presumably should remain reproductively fit—better to provide for a new generation of competent hosts—and active enough in its home range to be exposed to ticks bearing the pathogen. What this effectively means is usually a combination of resistance to and tolerance of infection in reservoir hosts of long-standing [16]. As for the implications of the tolerance phenomenon for human health, Ayres and Schneider put it this way, “What mechanisms keep us healthy while we are fighting infections?” [15]. Experimental and field studies of natural reservoirs of infection may provide insights not only about the pathogenesis of disease but processes that keep up us comparatively healthy in the face of infection.

### 3. *B. burgdorferi* infections of *Peromyscus* spp. in nature

*B. burgdorferi*'s life-cycle comprises its tick vector, small mammals and birds that serve as both reservoirs for the pathogen and blood-meal hosts for larval and nymphal stages, and a larger mammal, usually a deer, that provides the final blood meal of the adult female for reproduction [17, 18]. This life cycle in North America has existed since at least the Last Glacial Maximum [19, 20]. In most regions of the northeastern and north-central U.S., as well as adjoining regions of Canada, *P. leucopus* is usually the predominant one, in terms of its cumulative contribution to infection of larval *I. scapularis* ticks with *B. burgdorferi* [21, 22]. This status is attributable to both its reservoir competency for *B. burgdorferi* and its greater numbers over other small mammals in most endemic locations [23]. Figure 1 illustrates the white-footed mouse's greater competency in comparison with some other mammals existing in the same environment. The figure shows the ranges for *B. burgdorferi* numbers in nymphal ticks that had been feeding as larvae on captured *P. leucopus* or different medium-sized mammals [24, 25].

In the northeastern and north-central U.S. white-footed mice become infected during the Spring and Summer, as ticks which had become infected as larvae feed again as nymphs on naïve animals (reviewed in [26]). Since the majority of *P. leucopus* born in a calendar year do not survive to the following Spring, there are few immune animals in the population as it expands with the new year. When antibody-mediated immunity is elicited, it is strain-specific, leaving an animal at risk of infection by other strains [26], as tick infestations continue throughout the season. In areas as small as a few hectares, there may be 10–15 different strains of *B. burgdorferi* present in the ticks [27]. Although white-tailed deer, as providers of the last blood meal for adult female ticks, are important for reproduction of *I. scapularis*, deer are not competent as reservoirs and, as is true for humans, are dead-ends for the pathogen.

(*P. maniculatus* in the laboratory is also a competent reservoir for *B. burgdorferi* [28, 29]. But with the exception of a few areas, such as Ontario, Canada, where both *P. leucopus* and *P. maniculatus* exist and contribute to *B. burgdorferi*'s maintenance [30], it has been difficult to assess their relative competencies under natural conditions.)

In a study at a forested field site in Connecticut, a New England state, wild *P. leucopus*, were captured, tagged, released, and re-captured [31]. The prevalence of *B. burgdorferi* in nymphal *I. scapularis* ticks ranged between 25–50% at this location. The incidence of infection was 0.2 infections per mouse each week during the period of tick activity in this study. About 10% of the mice had *B. burgdorferi* bacteria circulating in their blood, as determined by PCR, at the time of capture. Prevalence of antibodies to *B. burgdorferi* increased with the age of the animal and, overall, from 57% in late May-early June to 95% by the end of August (Figure 2). Most of the mice that seroconverted between first and second captures had antibody to the strain-specific OspC proteins, which is a commonly recognized antigen among patients with Lyme disease [32].

Additional sera collected from captured *P. leucopus* at the Connecticut site were compared with sera from Lyme disease patients for their antibody reactivities on an microarray of 1292

proteins representing the majority of open reading frames (ORF) of the genome of *B. burgdorferi* [33]. The naturally infected white-footed mice generally recognized as antigens the same subset of proteins as the infected humans. Of the 103 most commonly recognized ORFs with the human sera, 70 were also high-ranking immunogens for the naturally-infected *P. leucopus*. This included the OspC proteins, which elicit antibodies that protect *M. musculus* against re-infection with the same strain (reviewed in [32]). This was further evidence that white-footed mice are capable of responding to infection with a variety of antibodies.

In a subsequent investigation of samples collected from *P. leucopus* at the Connecticut site, both *B. burgdorferi* and *B. miyamotoi* were studied [34]. Of 556 blood specimens, 69 (12%) contained *B. burgdorferi* DNA and 36 (6%) had *B. miyamotoi* DNA at the time of the rodent's capture. On average there were ~100 *B. burgdorferi* and ~1000 *B. miyamotoi* cells per milliliter of the blood of infected animals. A quarter of the bacteremic mice were still bacteremic with *B. burgdorferi* upon recapture a few weeks later. The prevalence of *B. burgdorferi* in ear biopsies from the captured white-footed mice was even higher: 65 (76%) of 86. The ear tissue tends to remain infected long after the bacteria have been cleared from the blood [35].

At a study site in Maryland, a mid-Atlantic state, captured and re-captured *P. leucopus* were studied for active infection with PCR and culture of ear tissue and for previous exposure by immunoassays for antibodies to *B. burgdorferi* [36]. The overall prevalence of *B. burgdorferi* in the biopsies of the captured rodents was ~30%, the prevalence rising with age of the animals. Of 77 animals who were infected on first capture, all remained infected on follow-up sampling an average of 160 days later. The authors reported "no measurable effect on the survival of infected mice."

Voordouw et al. carried out a capture-mark-capture study over a 4 year period on an island off the Connecticut coast [37]. On the basis of cultures of ear biopsies, the prevalence of active *B. burgdorferi* infection in the captured white-footed mice at the different trapping sites on the island ranged from 40–70%. As found in other studies, the prevalence of infection increased with age. The authors modeled mouse survival as a function of *B. burgdorferi* prevalence. Overall, monthly survival rates for the 4 year duration ranged from 0.55 to 0.88. For this variable, stratified by month of the year, there was no difference between infected and non-infected animals. While individual survival is an important component of fitness, neither this study nor the other longitudinal field studies to date reported on the effects of *B. burgdorferi* infection on another aspect of fitness: reproduction.

#### **4. *B. burgdorferi* infections of *Peromyscus* spp. in the laboratory**

The first experimental animal model for the newly-discovered Lyme disease agent was the golden or Syrian hamster, *Mesocricetus auratus*, a cricetine rodent like *Peromyscus* [38]. The golden hamster may remain persistently infected after inoculation [39]. Histopathologic study of chronically infected animals showed mild follicular lymphoid hyperplasia in the spleen but no signs of inflammation of kidney, eye, liver, or heart [40].

Given the minimal pathology and disability in the hamster, investigators interested in disease models turned to the laboratory mouse, *M. musculus*, which not only manifested more pathology than hamsters but came in a variety of inbred strains (reviewed in [35]). Most of these studies with inbred mouse strains focused on pathogenesis and immune response to infection, so investigators often chose strains and experimental conditions that provided for the most noticeable pathologic changes in the mice. However, while inbred strains differed in the severity of pathologic changes in tissues like the joints and heart, there was not a strain that was free of inflammation [41].

Reports of experimental infections of a *Peromyscus* sp. generally confirm the conclusions from the field studies: white-footed mice are highly susceptible to being infected with *B. burgdorferi*. In response to that infection, they produce many kinds of antibodies, which is the arm of the immune system that is most important for controlling Borrelia infection [32]. Yet, in spite of that immune response, *P. leucopus* remain persistently infected, sufficiently so for transmission to ticks to be possible for weeks to months after infection's onset. This stand-off between pathogen and host is not accompanied by much if any pathologic change or disability. The white-footed mouse has little or no defense against infection to begin with, unless it has become specifically immune to particular strain. But failing to eliminate it entirely, these rodents appear to limit the amount of growth of the organism, i.e. they are resistant to infection to some degree. At the same time, *P. leucopus*, to a greater extent than *M. musculus*, also limits or suppresses the amount of consequent damage, i.e. they are tolerant of the infection. What follows is a largely chronological summary of the studies that provided the evidence in support of this view.

Using both needle inoculation and tick bite routes, Schwan et al. infected laboratory-reared *P. leucopus* with *B. burgdorferi* and then examined the animals after 2 or 3 weeks [42]. Similarly to what has been observed with samples collected from the field, ~10% of the animals had *B. burgdorferi* in the blood at the time of sampling. Recovery of the organisms was highest from the bladders (94%), kidneys (75%), and spleens (61%) of the animals. Despite its presence in the kidney and bladder wall, the pathogen was not recovered in culture from the urine. In a study the subsequent year, Schwan et al. noted that *P. leucopus* developed persistent infections in spite of IgM and IgG antibody responses to several different antigens [43].

Wright and Nielson infected *P. leucopus* with *B. burgdorferi* by needle and tick transmission routes [44]. They observed the presence of the spirochetes in the kidney, liver, and spleen of the rodents, and the elicitation of anti-*B. burgdorferi* antibodies and their persistence in the blood over 4 months of observation. They reported that "regardless of the source of infection, no mice developed clinical signs or had any pathologic change resulting from infection." There was also no evidence of an effect on reproduction or of transplacental transmission of *B. burgdorferi* in their limited study. Mather et al. also concluded that transplacental transmission of *B. burgdorferi* to the offspring of *P. leucopus* does not occur [45].

Moody et al. reported that adult mice who became infected from needle injections of large inocula showed no gross or microscopic lesions on pathological examination [46]. On the

other hand, white-footed mice inoculated as infants became infected and also developed carditis and multifocal arthritis, similar to what was observed in adult *M. musculus*.

There have been few studies of *P. maniculatus* with experimental infections by *B. burgdorferi*, but in a small study Brown et al. found that while dusky-footed woodrats (*Neotoma fuscipes*) with infection had inflammatory infiltrates in the joints, muscles, and heart, the infected deer mice showed no pathologic changes [47]. Over an observation period of 40 days, the authors did not observe clinical signs of disease, such as reduced appetite, lethargy, lameness, or neurologic disorder, in the deer mice.

Other evidence about the duration of the transmissibility of *B. burgdorferi* from *P. leucopus* comes from a xenodiagnosis procedure, that is, the placement of unfed, uninfected larval ticks on the rodents and then assessing the proportion of the ticks on each animal that become infected. Lindsay et al. reported that transmission to ticks continued out to at least 49 days after inoculation, albeit at a lower frequency than after 7 days [48]. The pathogen was isolated from tissues of most of the mice 2 months after inoculation.

The functional effects of tick-transmitted *B. burgdorferi* infection on *P. leucopus* were studied by Schwanz et al. [49]. Infection was confirmed by the serology. After 5 weeks, infected rodents did not significantly differ from uninfected controls in blood cell counts of erythrocytes, neutrophils, eosinophils, basophils, lymphocytes, or monocytes. By the parameter of wheel running behavior, there was no measureable effect of infection after 1, 2, 3, or 6 weeks. On the basis of these findings, the authors suggested that “infection with the spirochete *B. burgdorferi* has little impact on the field activity of white-footed mice.”

In our own studies of *P. leucopus* with laboratory infections, we confirmed the findings from sera from field studies of robust antibody responses to the pathogen. The immunodominant antigens for white-footed mice largely corresponded to those that elicited antibodies during Lyme disease [31]. The specificities of the anti-OspC antibody responses among sera from experimentally-infected *P. leucopus* were similar to those exhibited by antibodies of patients with Lyme disease [32]. White-footed mice infected with different strains of *B. burgdorferi* had quantitatively similar antibody responses against whole cells, as well as purified antigens, though some strains achieved higher densities in ear, joint, tail, and heart tissues than other strains [50].

The latter study also revealed differences of an order of magnitude or greater between the individual outbred *P. leucopus* in their burdens of the bacteria [50]. Diversity in the capacities of the white-footed mice to limit pathogen burdens during infection was similarly noted in our study of experimental infections with the relapsing fever agent *B. hermsii* [51]. White-footed mice did not differ between sexes matched for age in any of the examined parameters, including change in weight over 3 weeks and burdens of bacteria in the blood or spleen. But among 30 adult male *P. leucopus*, there was a wide variation between animals in the counts of spirochetes in blood and spleen and in their spleen masses after 7 days of infection. There was also no discernible association between spirochete burdens in blood or spleen and either pathology in the liver or change in weight.

Figure 3 shows the result of a similar experiment of infection with a single strain of *B. burgdorferi* of 16 laboratory-reared *P. leucopus* and measurements of body weight before infection and then after 3 and 5 weeks; controls were 8 animals injected with PBS alone (V.J. Cook and A.G. Barbour, unpublished study). At sacrifice at 5 weeks, ear and ankle joint tissue were subjected to quantitative PCR, as previously described [50, 51]. The infected animals tended to gain less weight over the 5 weeks than uninfected animals, but the association was weak (Mann Whitney U;  $p = 0.12$ ). While the burdens of *B. burgdorferi* in the two tissues varied between animals over 2–3 orders of magnitude, there was no discernible association between pathogen burden and weight change over the 5 week course of the study (Figure 3).

## 5. Other infectious agents in *Peromyscus* in the field or laboratory

In a study of the agents of human granulocytic anaplasmosis and Lyme disease in *I. scapularis* and mammalian hosts in the north-central state of Minnesota, Johnson et al. found that prevalences of infection with *A. phagocytophilum* and *B. burgdorferi* were 20% and 42%, respectively, in white-footed mice [52]. These values corresponded to prevalences of antibodies to each agent of 29% and 48%, respectively, in the animals. By examining recaptured mice that had been infected on a previous capture, the authors noted that the rodents were more effective at eliminating *A. phagocytophilum* (~50% of the animals cleared) than *B. burgdorferi* (only 20% of the animals cleared).

The Sin Nombre hantavirus of *Peromyscus* spp. is considered at length in an accompanying article of this issue [3]. But for the present context, I note that in an experimental model with *P. maniculatus*, Botten et al. detected viral RNA throughout many of the organs and tissues of the rodents, but noted “no consistent histopathologic changes associated with infection, even when RNA load was high” [53].

*Bartonella* spp. in North America are usually transmitted by fleas and not ticks, but these blood-borne bacteria are common in many mammalian species in the wild. Bai et al. observed a high prevalence of persistent infection with *B. vinsonii* subsp. *arupensis* among in *P. maniculatus* in Colorado, but also no correlation between *Bartonella* prevalence and deer mouse weight [54]. This sub-species of *Bartonella* was isolated from a case of human illness [55].

While metazoan parasites of *Peromyscus* spp. do not appear to pose the same cross-over threat to humans as the foregoing microbes do, there is a considerable literature on parasitic worms in wildlife, either in the field or as an experimental model. Of relevance here is a report of laboratory infection of *P. maniculatus* with its natural parasite, the schistosome trematode, *Schistosomatium douthitti* [56]. While the parasitized deer mice had increased liver and spleen masses, the animals did not consume more food, decrease in body weight, or reduce their activity. The authors noted reduced thermoregulation during short-term cold exposure, which might be a fitness cost of the parasitism, but otherwise no changes in basal metabolic rate and cold-induced maximal metabolic rate.



On the other hand, *Peromyscus* is not resistant and/or tolerant of all potential pathogens, especially if it is not one for which it serves as a natural reservoir. Although *Peromyscus* spp. host their own vector-borne trypanosomes [57, 58], the agent of African trypanosomiasis, *Trypanosoma brucei*, and the closely-related apicomplexan parasite *T. equiperdum* are not among these. Following the Packchanian's description of the *T. brucei*-*P. maniculatus* model, in which average survival of the animals after inoculation of the parasite was 80 days [59], Moulton and colleagues carried out a series of studies [60]. They observed predictable mortality between 60–100 days after inoculation, pathologic changes in a number of organs, including the brain, a proliferative glomerulonephritis with antigen-antibody complexes, and suppression of cell-mediated immunity. While this certainly is a worse outcome for this rodent than we have seen up to now, the deer mouse was attractive as an experimental model because it did not undergo an “acute parasitemic death” [60], unlike the usual laboratory models of mice and rats.

## 6. *Peromyscus* spp. as target for disease control measures

*P. leucopus* and *P. maniculatus* are of importance for translational research or disease prevention applications, in particular for strategies to lower risk of human disease by targeting the infection reservoirs. This includes attracting the rodents with baits for application of pesticides to kill the attached ticks [61], and administering antibiotics in the field to kill bacterial pathogens the animals may be carrying [62]. There is also the controversial concept of increasing the proportion of animals that are less competent as reservoirs than *P. leucopus* to eventually “dilute out” *B. burgdorferi* in the population of ticks [63–65].

But more relevant to this review is the candidacy of *P. leucopus* as targets for vaccines to interrupt the life cycle for *B. burgdorferi*. The feasibility of this was demonstrated by Tsao et al. in the laboratory with an antigen, the OspA protein, that is expressed in the tick but not in mammals with natural infections [66]. A proof-of-principle for such a vaccine was provided by a field trial [67]. Subsequent studies of an orally-delivered OspA protein or a recombinant viral vector for OspA confirmed the efficacy of this approach [68–70]. *P. leucopus* would also be the prime target for an anti-tick vaccine [71], which would aim to reduce transmission to this reservoir of not only *B. burgdorferi* but also other Ixodes tick-transmitted pathogens. A transmission-blocking vaccine is the thinking behind a Sin Nombre hantavirus vaccine directed at *P. maniculatus* [72].

## 7. Conclusions

As some of the cited articles indicate, there is increasing interest in traits that allow an organism—be it a plant, nematode, fruitfly, mouse, or human—to minimize the fitness or health consequences of an infection, even if it fails to eliminate it [73–75]. This tolerance of infection is usually in partnership with resistance to infection in a mixed strategy of defense. While none of the field and laboratory studies of *Peromyscus* spp. discussed here were intentionally designed to tease out the relative contributions of resistance and tolerance, as they are defined here, one can see in the cumulative results different phenotypes that might align with either the resistance or tolerance trait. The status of these *Peromyscus* spp. as

widely-distributed, abundant, major reservoirs for several disease agents for humans in North America (Table 1) suggests a more-or-less stable, probably long-standing and mutual accommodation between parasitic microbes and mammalian species that appear to prosper in many parts of their distributions.

Further *Peromyscus* spp. studies should have testable hypotheses about resistance and tolerance and specify quantitative phenotypes. Whether they will provide insights of eventual benefit for human health remains to be seen [15, 76]. But I can think of no other vertebrate that would as well fit the bill for a model organism for these sorts of questions, including the causes of the disabilities of Lyme disease.

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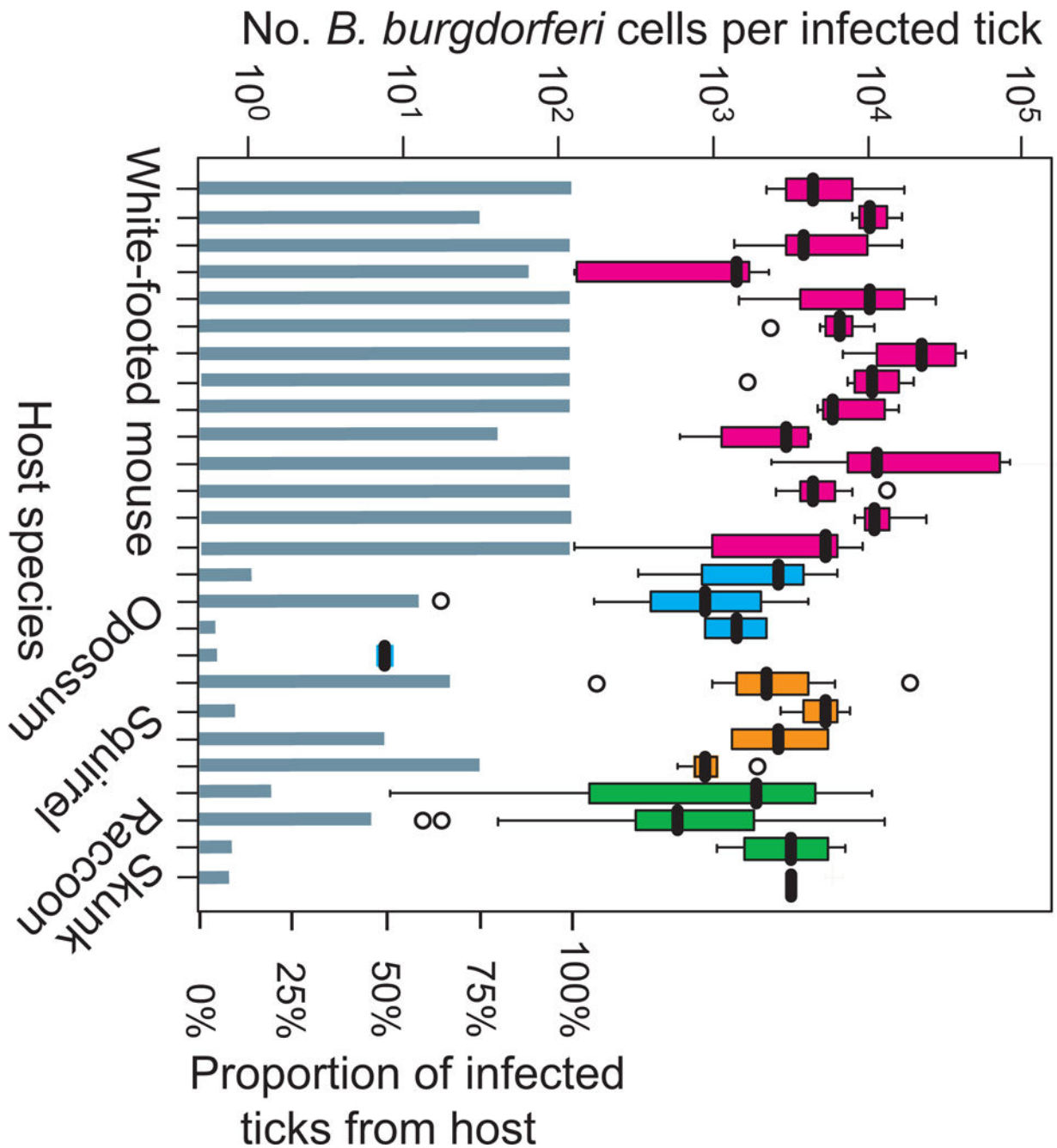
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**Figure 1.**

Greater reservoir competence of the white-footed mouse *P. leucopus* in comparison to medium-sized small mammals in a Connecticut forest.

Larval *Ixodes scapularis* ticks were collected after they had been found feeding on different mammalian species, which had been captured and temporarily held before release. The engorged larvae that fell off were allowed to molt to nymphs, and then the numbers of *B. burgdorferi* cells per individual tick were determined by quantitative PCR. For this analysis the results for 14 white-footed mice (red), 4 Virginia opossum (royal blue), 4 gray squirrel (orange), 3 common raccoon (green), and 1 striped skunk (black) are shown. Box-whisker



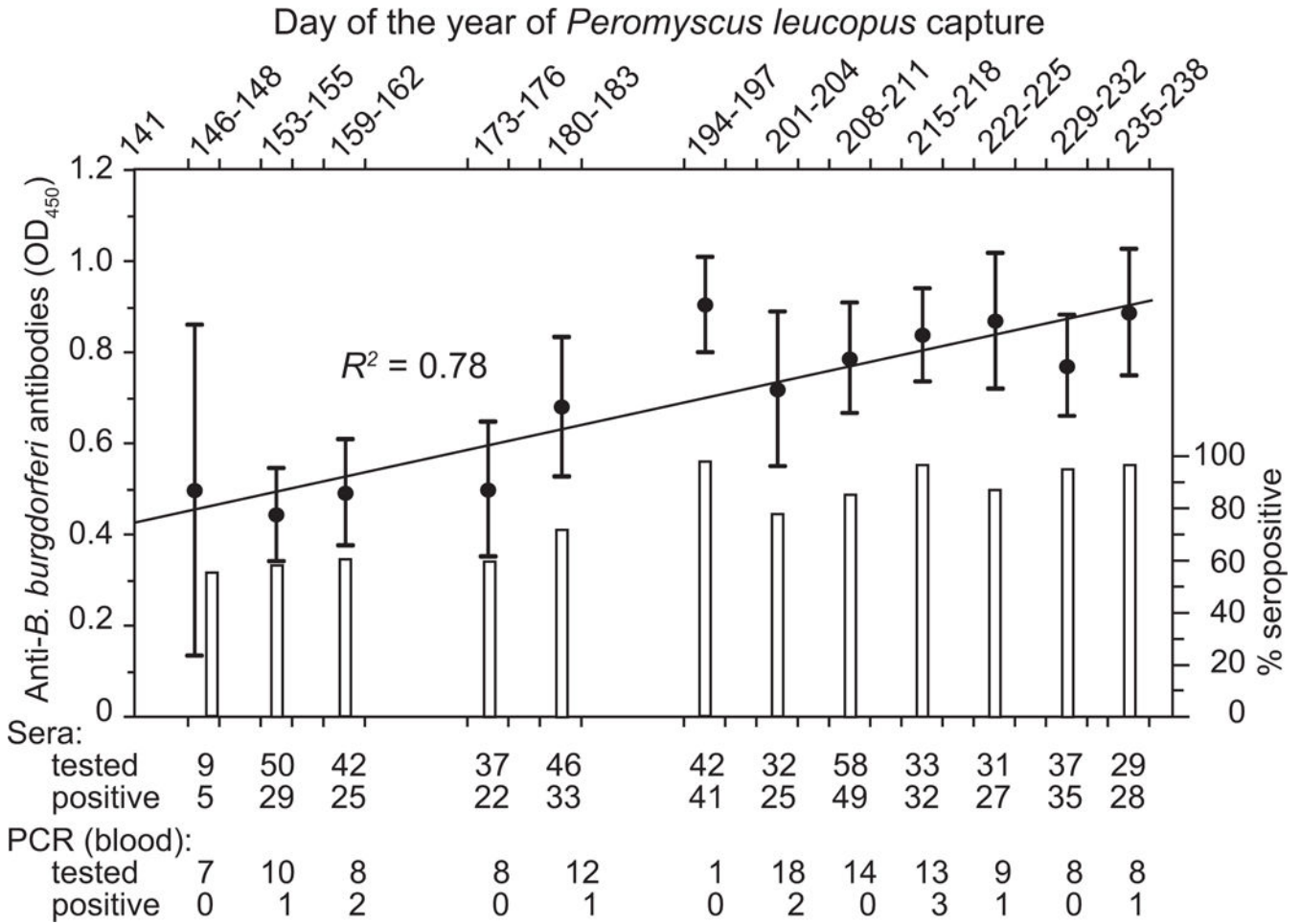
plots summarize the distributions of bacterial counts per individual tick, with the values indicated on the left  $y$ -axis. Each horizontal box indicates the first and third quartiles, and the black horizontal line inside the box is the median. The  $1.5\times$  interquartile range is indicated by the whiskers bisecting the box, and a value outside this range is indicated by an open circle. The lower half of the graph shows the proportions (right  $y$ -axis) of the collected ticks from each animal that were infected with *B. burgdorferi*. Methods and data are from the studies of references 25 and 94.

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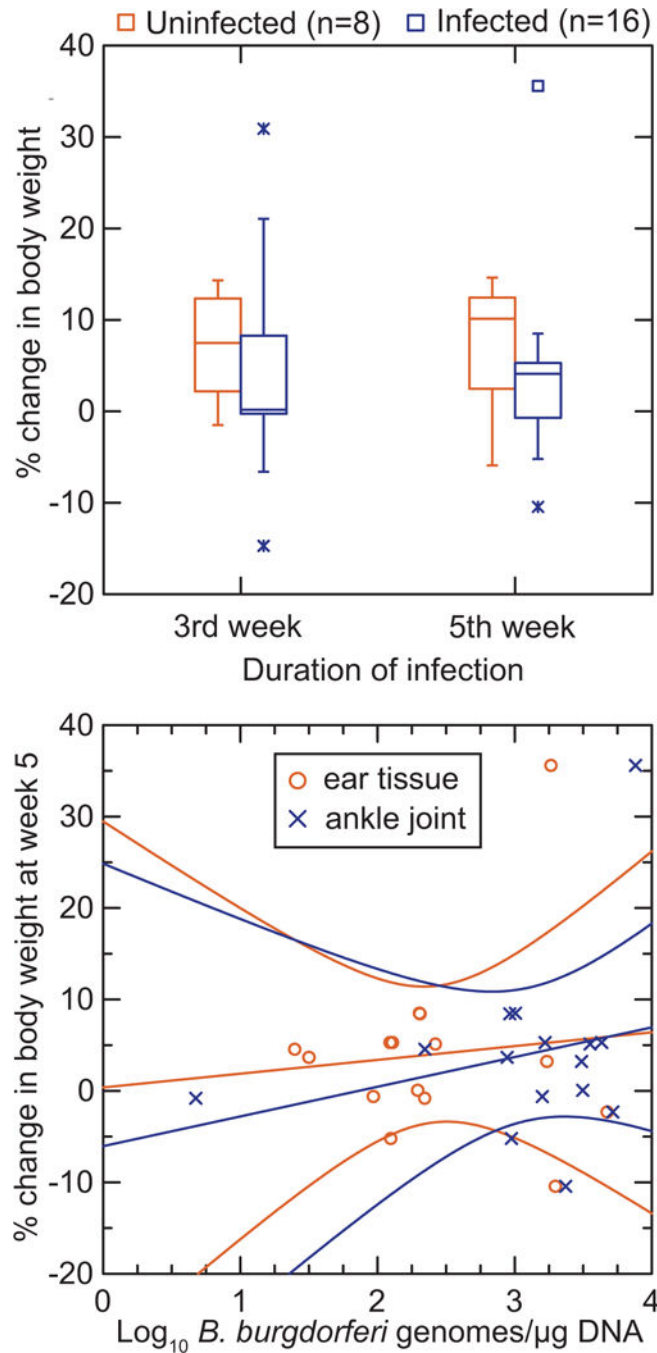
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**Figure 2.** Highly prevalent *Borrelia burgdorferi* infection of *Peromyscus leucopus* in a Connecticut forest. Infection in capture-release rodents was monitored over a late Spring-Summer tick transmission season with enzyme-linked immunoassay (EIA) of serum samples for antibodies to *B. burgdorferi* and polymerase chain reaction (PCR) assay of whole blood samples for *B. burgdorferi*. The colorimetric EIA OD<sub>405</sub> value intervals and the percentage of positive serum samples for a sampling period are shown on the left and the right y-axis, respectively. Below the graph are figures on the total number of blood samples subjected to EIA and PCR analysis, from different collection periods, as well as the number of samples that were found to be positive by each of these assays. The coefficient of determination ( $R^2$ ) value between the collection dates and the values by EIA is shown. Methods and data are from reference 31.



**Figure 3.**

Lack of effect on body weight over 5 weeks during an experimental infection of laboratory-reared *P. leucopus* with *B. burgdorferi*.

Adult male *P. leucopus* were infected by needle injection with the HB19 strain of *B. burgdorferi* or with a buffer control. The animals were weighed just before injection and then at 3 weeks and 5 weeks, at which point they were euthanized. DNA was extracted from ear tissue and ankle joints and subjected to quantitative PCR. The sources of animals and bacteria, methods, and conditions of the experiment are described in references 50 and 51.

The upper panel compares in box-whisker plots the % change in weight from day of injection at weeks 3 and 5 for uninfected (left plots in each pair) and infected mice (right plots in each pair). The lower panel is a scatter plot of % change in body weight on counts of bacteria in either ear tissue or ankle joints. The linear regression lines with 95% confidence intervals are shown.

Table 1

Zoonotic pathogens, their human diseases, vectors, and *Peromyscus* spp. Hosts

Microbe/parasite	Human disease	Vector/intermediate host	<i>P. leucopus</i> [references]	<i>P. maniculatus</i> [references]
<i>Borrelia burgdorferi</i>	Lyme disease (Lyme borreliosis)	<i>Ixodes scapularis</i> , <i>I. pacificus</i> ticks	+ <sup>a</sup> [21, 22, 77]	+ [28, 47]
<i>Borrelia miyamotoi</i>	Hard tick-borne relapsing fever	<i>I. scapularis</i> , <i>I. pacificus</i>	+ [34, 78]	?
<i>Anaplasma phagocytophilum</i>	Human granulocytic anaplasmosis	<i>I. scapularis</i> , <i>I. pacificus</i> , <i>I. spinipalpis</i>	+ [79, 80]	+ [81]
<i>Babesia microti</i>	Babesiosis	<i>I. scapularis</i>	+ [82, 83]	–
Powassan/deer tick virus	Viral encephalitis	<i>I. scapularis</i>	+ [84]	+ [85]
<i>Ehrlichia muris</i> subsp. <i>euclairensis</i>	Ehrlichiosis	<i>I. scapularis</i>	+ [9, 12]	–
Sin Nombre-like hantaviruses	Hantavirus pulmonary syndrome	None	+ [86]	+ [87]
<i>Yersinia pestis</i>	Plague	Fleas	–	+ [4, 88]
<i>Bartonella</i> spp.	Bartonellosis	Fleas	+ [89]	+ [54, 55]
<i>Borrelia bissettii</i>	None known	<i>I. pacificus</i> , <i>I. spinipalpis</i>	–	+ [90, 91]
" <i>Borrelia davisi</i> "	None known	?	+ [92]	?
<i>Trypanosoma</i> sp. <sup>a</sup>	None known	?	+ [58]	+ [57]
<i>Schistosomatium douthitti</i>	None known	Snail	?	+ [56, 93]

<sup>a</sup> +, the listed *Peromyscus* species is a reservoir; –, not a reservoir; ?, reservoir status not known

<sup>b</sup> Exclusive of agents of Chagas disease (*T. cruzi*) and African trypanosomiasis (*T. brucei*)