

# An Evolutionarily Conserved Transcriptional Activator-Repressor Module Controls Expression of Genes for D-Galacturonic Acid Utilization in *Aspergillus niger*

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**ABSTRACT** The expression of genes encoding extracellular polymer-degrading enzymes and the metabolic pathways required for carbon utilization in fungi are tightly controlled. The control is mediated by transcription factors that are activated by the presence of specific inducers, which are often monomers or monomeric derivatives of the polymers. A D-galacturonic acid-specific transcription factor named GaaR was recently identified and shown to be an activator for the expression of genes involved in galacturonic acid utilization in *Botrytis cinerea* and *Aspergillus niger*. Using a forward genetic screen, we isolated *A. niger* mutants that constitutively express GaaR-controlled genes. Reasoning that mutations in the *gaaR* gene would lead to a constitutively activated transcription factor, the *gaaR* gene in 11 of the constitutive mutants was sequenced, but no mutations in *gaaR* were found. Full genome sequencing of five constitutive mutants revealed allelic mutations in one particular gene encoding a previously uncharacterized protein (NRRL3\_08194). The protein encoded by NRRL3\_08194 shows homology to the repressor of the quinate utilization pathway identified previously in *Neurospora crassa* (*qa-1S*) and *Aspergillus nidulans* (QutR). Deletion of NRRL3\_08194 in combination with RNA-seq analysis showed that the NRRL3\_08194 deletion mutant constitutively expresses genes involved in galacturonic acid utilization. Interestingly, NRRL3\_08194 is located next to *gaaR* (NRRL3\_08195) in the genome. The homology to the quinate repressor, the chromosomal clustering, and the constitutive phenotype of the isolated mutants suggest that NRRL3\_08194 is likely to encode a repressor, which we name GaaX. The GaaR–GaaX module and its chromosomal organization is conserved among ascomycetes filamentous fungi, resembling the quinate utilization activator-repressor module in amino acid sequence and chromosomal organization.

**KEYWORDS** gene regulation; galacturonic acid; repressor protein; genomics; transcriptomics; pectin

**T**HE filamentous fungus *Aspergillus niger* is an important producer of pectin-degrading enzymes that are used in industrial applications including in food and feed processing (Kashyap *et al.* 2001; Khan *et al.* 2013). In nature, *A. niger* is a saprotrophic fungus that feeds on organic matter from decaying plants. The major carbon sources in plant cells are the

storage polysaccharides starch, and less frequently inulin, as well as the cell wall polymers cellulose, hemicelluloses, and pectin. Of the different plant polysaccharides, pectin has the most complex structure. Pectin is made up of four substructures, including homogalacturonan, xylogalacturonan, rhamnogalacturonan I, and rhamnogalacturonan II. The abundance of each substructure varies with plant species, but typically homogalacturonan is the most abundant polysaccharide in pectin (65%), followed by rhamnogalacturonan I (25–30%). Xylogalacturonan and rhamnogalacturonan II comprise <10% of the total pectin (Mohnen 2008).

Utilization of plant polysaccharides by fungi, including *A. niger*, is accomplished by tightly controlled secretion of extracellular enzymes that degrade the polymers into

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monosaccharides or oligosaccharides that are taken up and catabolised by the fungus. The controlled regulation is not only confined to the expression of genes encoding extracellular proteins. It also includes the controlled expression of genes encoding specific sugar transporters to guarantee efficient uptake of the liberated sugars and the intracellular catabolic pathway enzymes. The precise induction of the network of genes encoding substrate-specific enzymes, transporters, and catabolic pathway enzymes has so far been shown to be mediated via Zn(II)<sub>2</sub>Cys<sub>6</sub> transcription factors. Specific transcription factors in *A. niger* have been characterized that regulate the utilization of the major polysaccharides. They include AmyR, the regulator for starch utilization (Petersen *et al.* 1999; Yuan *et al.* 2008a; vanKuyk *et al.* 2012); InuR for inulin (Yuan *et al.* 2008b); ManR, ClrA, and ClrB for cellulose (Raulo *et al.* 2016); XlnR for xylan (Van Peij *et al.* 1998; Battaglia *et al.* 2014); AraR for arabinan (Battaglia *et al.* 2014); RhaR for rhamnose (Gruben *et al.* 2014); and GaaR for polygalacturonic acid (PGA) (Alazi *et al.* 2016). These transcription factors exert coordinated regulation of the target genes by interacting with conserved binding sites that are located upstream of the target genes. Computational analysis has been used to identify the galacturonic acid (GA) responsive element (GARE) of GA-induced genes (Martens-Uzunova and Schaap 2008). The predicted sequence (CCNCCAA) was shown to be required for the induction of GA-responsive genes in *A. niger* (Niu *et al.* 2015) and *Botrytis cinerea* (Zhang *et al.* 2016). Furthermore, using the yeast one-hybrid method, it was shown in *B. cinerea* that the GaaR transcription factor interacts specifically with the GARE (Zhang *et al.* 2016).

Phenotypic characterization of mutants lacking the GA-regulator in both *B. cinerea* and *A. niger* has shown that GaaR is required for growth on GA (Alazi *et al.* 2016; Zhang *et al.* 2016). Expression analysis in both fungi confirmed that GaaR is required for the induced expression of GA-responsive genes. On complex pectins, growth of *B. cinerea* and *A. niger gaaR* deletion mutants was severely reduced and genome-wide expression analysis in *A. niger* revealed that the residual growth on pectin is likely due to the GaaR-independent expression of pectinases acting on arabinofuranosyl- and galactopyranosyl-containing side chains in rhamnogalacturonan (Alazi *et al.* 2016).

In addition to the transcription regulation via GaaR, GA-responsive genes are also under carbon catabolite repression (CCR) control (de Vries *et al.* 1999; de Vries *et al.* 2002). In filamentous fungi, CreA mediates CCR (Dowzer *et al.* 1991; Ruijter and Visser 1997). In *A. niger*, CreA also exerts CCR control on GA-responsive genes (de Vries *et al.* 1999; Niu *et al.* 2015). Using an *in vivo* reporter construct consisting of the promoter of the GA-inducible *pgaX* gene (*PpgaX*) and the acetamidase (*amdS*) gene as a reporter, both the specific induction of *pgaX* on GA as well as the carbon repression of *pgaX* via CreA had been demonstrated (Niu *et al.* 2015). In this study, we have used the *PpgaX-amdS* reporter strain to isolate mutants displaying constitutive expression of GA-responsive genes. Analysis of the mutants resulted in the identification of a protein

that likely acts as a repressor that specifically inhibits GaaR transcription activation activity under noninducing conditions.

## Materials and Methods

### *Strains, media, and growth conditions*

All strains in this study are listed in Table 1. Strains were grown in liquid or solidified (1.5% agar) minimal medium (MM) containing 7 mM KCl, 8 mM KH<sub>2</sub>PO<sub>4</sub>, 70 mM NaNO<sub>3</sub>, and 2 mM MgSO<sub>4</sub> (pH adjusted to 5.5) as described (Bennett and Lasure 1991). MM was supplemented with 50 mM glucose, 50 mM D-galacturonic acid, 50 mM fructose, or 50 mM sorbitol as carbon source. Complete medium (CM) was also used and consists of MM supplemented with 0.1% casamino acids and 0.5% w/v yeast extract and 50 mM glucose. MM agar plates containing acetamide as sole nitrogen source were made as described previously (Arentshorst *et al.* 2012).

### *Isolation of mutants with constitutive expression of genes involved in PGA utilization*

*A. niger* strain JN29.2 (Table 1) was used for the selection of mutants with constitutive expression of genes involved in PGA utilization. Spontaneous mutants were obtained by plating out freshly harvested and myracloth-filtered conidia (1 × 10<sup>4</sup> conidia per plate) on MM glucose/acetamide plates and incubated at 30° for 5 days. In addition, mutants were obtained after mild UV mutagenesis (80% survival) as described (Damveld *et al.* 2008). Individual mutants growing on the primary MM-glucose/acetamide selection plates were purified twice on the MM-glucose/acetamide agar plates. In total, 14 spontaneous mutants and 59 UV-mutants were isolated that grew well on MM-glucose/acetamide agar plates and they were potential mutants with constitutive expression of genes involved in PGA utilization. To identify mutants constitutively producing PGA-degrading enzymes, all 73 mutants were grown by inoculating 5 × 10<sup>7</sup> spores in 50 ml MM-glucose medium for 36 hr at 30° with shaking at 150 rpm. Supernatant of each culture was harvested by filtration. The extracellular culture fluid and the mycelia were stored at –80° for enzymatic assays and RNA extraction, respectively. A total of 10 μl supernatant of each sample was spotted on PGA plates made by dissolving 0.2% PGA (Sigma) in NaAc buffer (pH 4.2) with 1% agarose (Sphaero). The PGA plate assay was modified from the protocol used to detect cellulase activity (Teather and Wood 1982). Plates were incubated at 37° for 17 hr after spotting. PGA was stained by flooding the plates with a filter-sterile 0.05% solution of Congo Red (Sigma) dissolved in Milli-Q water for 15 min. The Congo Red solution was then poured off and the plates were washed with Milli-Q water and further treated by flooding with 1 M NaCl for 15 min. The formation of a clear zone of hydrolysis indicated PGA degradation.

The constitutive expression of genes involved in PGA degradation was further determined by Northern blot analysis. Total RNA was isolated from eleven UV-mutants and two spontaneous mutants from frozen mycelia using TRIzol

**Table 1** *Aspergillus niger* strains used in this study

Name	Genotype/description	Reference/source
N402	<i>cspA1</i> , derivative of N402	Bos <i>et al.</i> (1988)
AB4.1	<i>pyrG</i> <sup>-</sup> , derivative of N402	van Hartingsveldt <i>et al.</i> (1987)
MA234.1	$\Delta ku70::DR\_amdS\_DR$ in MA169.4	Alazi <i>et al.</i> (2016)
MA70.15	$\Delta ku70::amdS$ in AB4.1	Meyer <i>et al.</i> (2007)
MA299.2	$\Delta ku70$ in MA70.15	Niu <i>et al.</i> (2015)
MA323.1	$\Delta ku70::amdS$ , $\Delta nicB$ <sup>-</sup> , <i>pyrG</i> <sup>-</sup>	Niu <i>et al.</i> (2016)
JC1.5	<i>pgaX-amdS</i> in MA299.2, <i>pyrG</i> <sup>+</sup>	Niu <i>et al.</i> (2015)
JN29.2	$\Delta creA::hygB$ in JC1.5	Niu <i>et al.</i> (2015)
JN38	Spontaneous mutation S1 in JN29.2	This study
JN39	Spontaneous mutation S2 in JN29.2	This study
JN42	Spontaneous mutation S5 in JN29.2	This study
JN44	Spontaneous mutation S7 in JN29.2	This study
JN52	UV1 in JN29.2	This study
JN53	UV2 in JN29.2	This study
JN54	UV3 in JN29.2	This study
JN55	UV4 in JN29.2	This study
JN56	UV5 in JN29.2	This study
JN57	UV6 in JN29.2	This study
JN58	UV7 in JN29.2	This study
JN59	UV8 in JN29.2	This study
JN60	UV9 in JN29.2	This study
JN61	UV10 in JN29.2	This study
JN62	UV11 in JN29.2	This study
JN63	UV12 in JN29.2	This study
JN64	UV13 in JN29.2	This study
JN122.1, JN122.2 and JN122.3	$\Delta gaaX::phleo$ in JN29.2	This study
JN123.1, JN123.2, JN123.3	$\Delta gaaX::hygB$ in JC1.5	This study
JN125.1	$\Delta gaaX::nicB$ in MA323.1	This study
JN126.2, JN126.5, JN126.6	<i>PgaaX-gaaX::GFP-TgaaX</i> in JN125.1	This study
JN127.1, JN127.2, JN127.3	<i>PgaaX-GFP::GaaX-TgaaX</i> in JN125.1	This study

reagent (Invitrogen, Carlsbad, CA). Quantification and purity assessment of total RNA was done by spectrophotometric method (NanoDrop 2000; Thermo Scientific). Total RNA (3.5  $\mu$ g) was loaded per sample and blotted to a Hybond-N<sup>+</sup> nylon membrane (Amersham, GE Healthcare) followed by hybridization with [ $\alpha$ -<sup>32</sup>P]-dCTP-labeled probes (Rediprime II kit; Amersham, GE Healthcare). Probes were PCR-amplified using the N402 genomic DNA and the primer pairs are listed in Supplemental Material, Table S1. Standard molecular techniques were applied as described (Sambrook and Russell 2001).

#### DNA sequencing and data analysis

Sequencing of the *gaaR* gene from 11 constitutive mutants was performed by PCR amplification of the *gaaR* gene, including 137 bp upstream and 152 bp downstream sequences, using genomic DNA of the mutants as template and primers

*gaaRP7f* and *gaaRP8r* (Table S1). Genomic DNA was isolated as described (Arentshorst *et al.* 2012). The PCR fragment (2765 bp in size) was sequenced in both directions using *gaaR* sequencing primers (Table S1). Sequencing was performed by MacroGen Europe (Amsterdam, The Netherlands).

Genomic DNA of three spontaneous mutants and two UV mutants was isolated as described (Arentshorst *et al.* 2012), and was further purified with DNA Isolation Kit (MO BIO Laboratories) for whole-genome DNA sequencing. The mutant genomes were sequenced at the McGill University Génome Québec Innovation Centre (QC, Canada) using the Illumina HiSeq platform to ~50-fold coverage. The DNA reads were aligned to the NRRL3 genome with Bowtie2 (Langmead and Salzberg 2012) and sequence differences were detected with FreeBayes (Garrison and Marth 2012).

#### Deletion of *gaaX* gene

Deletion of the *gaaX* gene (NRRL3\_08194) in the JC1.5, JN29.2, and MA323.1 backgrounds (Table 1) was carried out using the split marker approach (Arentshorst *et al.* 2015). The 869 bp 5'-flank and 870 bp 3'-flank regions were PCR amplified with the primers listed in Table S1, using N402 genomic DNA as template. These PCR fragments were used in fusion PCRs with hygromycin, phleomycin resistance genes, or the *nicB* gene (Niu *et al.* 2016) to generate the split marker fragments. After amplification, the 5'-flank-*hyg* and 3'-flank-*hyg* fragments were transformed to the recipient strain JC1.5, the 5'-flank-*phleo* and 3'-flank-*phleo* fragments were transformed to the recipient strain JN29.2, and the 5'-flank-*nicB* and 3'-flank-*nicB* fragments were transformed to the recipient strain MA323.1. Putative *gaaX* disruption strains were purified by two consecutive single colony streaks. Genomic DNA was isolated as described (Arentshorst *et al.* 2012) and Southern blot hybridizations, using PCR-amplified fragments generated with primers listed in Table S1 as probes, were performed to confirm proper deletion and to exclude additional integrations.

#### Bioreactor cultivation

Controlled bioreactor cultivations for *A. niger* strains MA234.1 and JN123.1 were performed in 6.6-L BioFlo3000 bioreactors (New Brunswick Scientific) as previously described (Jørgensen *et al.* 2010). Briefly, autoclaved bioreactor vessels were filled with 5 liter of sterile MM with 0.75% fructose. During cultivation at 30°, the controller was set to maintain pH 3 by addition of titrants (2 M NaOH or 1 M HCl). Sterile air was supplied at a rate of 1 liter/min. Prior to inoculation, 1.5 ml of 10% (w/v) filter-sterilized yeast extract was added to enhance conidial germination. Cultures were inoculated with freshly harvested spores at a concentration of  $7.0 \times 10^8$  conidia per liter. To reduce the loss of hydrophobic conidia during germination, the stirrer speed was set to 250 rpm and the culture was aerated via the headspace during the first 6 hr after inoculation. Subsequently, the stirrer speed

was increased to 750 rpm, 0.5 ml of polypropyleneglycol P2000 was added as an antifoam agent, and air was supplied via the sparger. Cultures broth was harvested at regular intervals from batch cultures and mycelial biomass was retained by vacuum filtration using glass microfiber filters (Whatman). Both biomass and filtrate were quickly frozen in liquid nitrogen and subsequently stored at  $-80^{\circ}$ . Dry biomass concentrations were gravimetrically determined from lyophilized mycelia originating from a known mass of culture broth.

### Transcriptome analysis

Mycelia grown in bioreactors to midexponential phase were used to isolate RNA using TRIzol reagent (Invitrogen) and purified with NucleoSpin RNA Clean-up kit (Macherey-Nagel) with DNase treatment. Quantity and quality of the RNA samples were determined with a NanoDrop 2000 spectrophotometer and by RNA gel electrophoresis, respectively. RNA sequencing was conducted by Genome scan (Leiden, the Netherlands). Briefly, messenger RNA was isolated from the total RNA using NEBNext Ultra Directional RNA Library Prep Kit for Illumina, according to the manufacturer's protocol. After fragmentation of the messenger RNA, complementary DNA was synthesized using random primers; after a second-strand complementary DNA synthesis reaction, fragments were ligated to the sequencing adapters. Clustering and DNA sequencing was performed using the Illumina NextSeq 500 SR75. We refer to *A. niger* gene IDs based on the most up-to-date and accurate annotation of the *A. niger* NRRL3 genome (<http://genome.fungalgenomics.ca/>). The RNA-seq reads were cleaned by correcting sequencing errors with Rcorrector (Song and Florea 2015), trimming sequencing adapters and low quality sequences with Skewer (Jiang *et al.* 2014), and removing ribosomal RNA with SortMeRNA (Kopylova *et al.* 2012). The cleaned reads were mapped to NRRL3 transcripts and counted with Salmon (Patro *et al.* 2016), and the read counts were analyzed for differences in transcript expression between genotypes with DESeq2 (Love *et al.* 2014).

### Construction of strains expressing GaaX-GFP or GFP-GaaX fusion proteins

To construct fusions of GFP to the N-terminus or C-terminus of GaaX, P<sub>gaaX</sub>\_GFP::gaaX\_GaaX::et al. 2008), were used as template DNA. For constructing the P<sub>gaaX</sub>\_GFP::gaaX was PCR amplified using primers P<sub>gaaX</sub>\_P7f-NotI and P<sub>gaaX</sub>\_P11r, GFP was PCR amplified from plasmid PagsA\_eGFP\_TrpC using primers GFP\_P1f and GFP\_P3r, *gaaX* and the terminator region of *gaaX* was PCR amplified using primers *gaaX*\_P12f and TgaaX\_P10r-NotI, and the three fragments were combined together in a two-step fusion PCR. Two amino acids (Gly-Ala) were introduced as spacer between GFP and GaaX. Subsequently, the fusion fragment was cloned into

vector pJet1.2 to give plasmid pJN34. For the P<sub>gaaX</sub>\_GaaX::gaaX with the promoter region of *gaaX* was PCR amplified using primers P<sub>gaaX</sub>\_P7f-NotI and *gaaX*\_P8r, GFP was PCR amplified using primers GFP\_P1f and GFP\_P2r, the terminator region of *gaaX* was PCR amplified using primers *gaaX*\_P9f and TgaaX\_P10r-NotI, and the three fragments were combined together in a two-step fusion PCR. Again, a Gly-Ala spacer was introduced between GaaX and GFP. Subsequently, the fusion fragment was cloned into vector pJet1.2 to give plasmid pJN35.

Plasmids pJN34 and pJN35 were digested by NotI, and the fragments containing P<sub>gaaX</sub>\_GFP::gaaX\_GaaX::et al. 2015) to generate pJN36 and pJN37, respectively. The pMA334 plasmid was designed such that the reporter constructs are targeted to the *pyrG* locus. Plasmids pJN36 and pJN37 were linearized by AscI digestion and purified from gel before transformation to *A. niger* strain JN125.1. Proper integration of the P<sub>gaaX</sub>\_GFP::gaaX::GaaX\_GFP\_TgaaX fragments at the *pyrG* locus was confirmed by Southern blot analysis using PCR-amplified fragments generated with primers listed in Table S1 as probes.

### Microscopy

For microscopic analysis, conidia of strains MA323.1, JN125.1, JN126.2, and JN127.3 were inoculated on coverslips in Petri dishes. Liquid MM supplemented with 50 mM GA or 50 mM fructose as the carbon source was used. After incubation at  $30^{\circ}$  for 16 hr, the coverslips with adherent germlings were mounted upside down on glass slides and observed under a confocal laser scanning microscope (Zeiss Imager; Zeiss, Jena, Germany), equipped with an LSM 5 exciter using 63 $\times$  objectives. Images were processed by ImageJ with the exact same brightness and contrast adjustments and the median filter (radius 1.0).

### Data availability

Strains are listed in Table 1 and are available upon request. Table S2 contains SNPs and indels detected in genomes of mutants. Table S3A contains transcript per million (TPM) values of NRRL3 gene models in wild type and the *gaaX* mutant, and Table S3B contains their DESeq2 analysis. The DNA reads described in this study are deposited in the Short Read Archive under accession number SRP078415. The RNA reads described in this study are deposited in the Short Read Archive under accession number SRP078485.

## Results

### Mutants constitutively expressing genes related to galacturonic acid utilization

To identify mutants that constitutively express genes related to PGA degradation and GA utilization in *A. niger*, we designed a forward screening procedure using a reporter strain containing a P<sub>ppgaX</sub>-*amdS* reporter construct for positive selection of the desired mutants. We recently showed that the

*pgaX* gene is specifically induced by GA, PGA, and pectin, allowing the reporter *PpgaX-amdS* strain to grow on acetamide as a nitrogen source when GA, PGA, or pectin is present as a carbon source (Niu *et al.* 2015). We also showed that deletion of the CCR protein (CreA) did not result in growth of the *PpgaX*-reporter strain on glucose, indicating that derepression via *creA* deletion was not sufficient to drive *PpgaX-amdS* expression to sustain growth. For the mutant screen, we used the *PpgaX-amdS* reporter strain in the  $\Delta creA$  background (JN29.2) to prevent interference with possible CreA pathway-related repression mechanisms. Spores of *A. niger* strain JN29.2 were UV-mutagenized and surviving spores (80%) were plated on MM-glucose-acetamide plates. After mutagenesis, 59 mutants were isolated based on growth on acetamide. In addition to UV-generated mutants, 14 spontaneous mutants were isolated, resulting in a total of 73 mutants that could grow on glucose/acetamide plates.

To determine whether mutations were *cis*- or *trans*-acting, mutants were cultured in glucose medium for 36 hr and the medium was analyzed for polygalacturonase activity. Initial experiments showed that cultivation of JN29.2 in glucose medium resulted in very low polygalacturonase levels and no halo was formed on Congo Red-stained PGA plates when culture medium was spotted on a PGA plate (Figure 1A). We reasoned that if the mutation is *trans*-acting, the medium should contain increased levels of both exo- and endo-polygalacturonases, leading to the formation of a halo. On the other hand, if the mutation is *cis*-acting, thereby only affecting the *PpgaX-amdS* reporter construct, it would not result in a halo on a PGA plate. Based on this assay, we concluded that the mutations of 65 out of 73 mutants are *trans*-acting, while the remaining eight mutants carry presumed *cis*-acting mutations.

To further demonstrate that the presumed *trans*-acting mutations indeed affected expression of multiple genes related to GA utilization and belonging to the GA-induced genes (Martens-Uzunova and Schaap 2008; Niu *et al.* 2015; Alazi *et al.* 2016), the expression of three GA-induced genes (*pgaX*, *gatA*, and *gaaB*) was examined by Northern blot analysis in a subset of mutants after growth on glucose. The *pgaX*, *gatA*, and *gaaB* genes encode an exo-polygalacturonase, a GA-specific transporter, and the L-galactonic acid dehydratase involved in the GA release from PGA, the uptake of GA, and subsequent metabolism of GA, respectively. As shown in Figure 1B, expression of these genes was not detected in the wild type (N402) and the parent JN29.2 ( $\Delta creA$ , *PpgaX-amdS*) strains, whereas these genes were expressed in the constitutive mutants that displayed increased polygalacturonase activity. The presumed *cis*-acting mutants from the plate assay (UV2, UV7, UV9, UV10, UV11, and S2) did not constitutively express *pgaX*, *gatA*, and *gaaB*, and showed a small halo on PGA plates, indicating the halo assay can be used to discriminate between *cis*- and *trans*-acting mutants. To determine whether a *cis*-acting mutation in the *pgaX* promoter in front of the *amdS* gene was responsible for the ability of this class of mutants to grow on acetamide, the *pgaX* promoter in front of the *amdS* gene of all eight *cis*-acting

mutants was PCR amplified using *pyrG*- and *amdS*-specific primers (Table S1). This analysis revealed no mutations in the *pgaX* promoter region of any of the eight presumed *cis*-acting mutants. Hence, the nature of the mutation(s) in these strains that allows growth on acetamide remains unknown. A possibility could be the activation of expression of endogenous *amdS* genes, as at least four *amdS*-like genes are present in the genome of *A. niger*.

#### **Identification of mutations responsible for the constitutive expression of the galacturonic acid utilization genes**

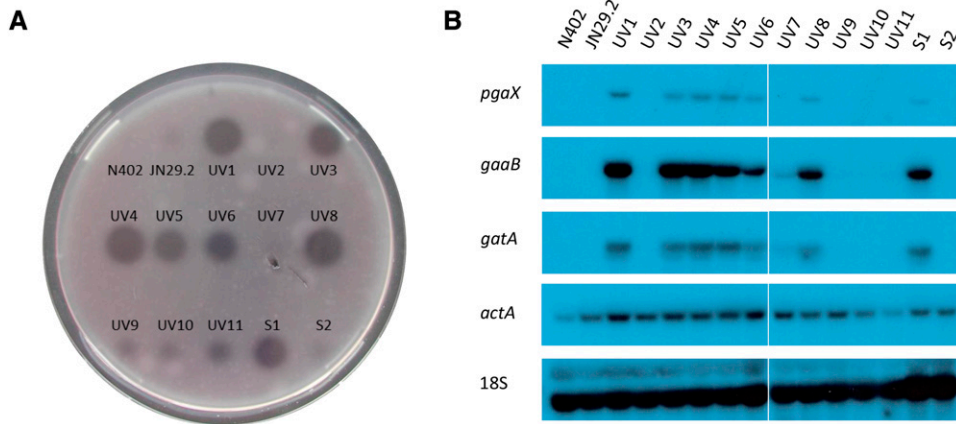
A possible explanation for the constitutive expression of GA utilization genes in the mutants is that they carry mutations in the recently identified GaaR transcriptional activator (Alazi *et al.* 2016). We therefore PCR amplified and sequenced the *gaaR* locus of eight constitutive mutants obtained after UV mutagenesis (UV1, UV3, UV4, UV5, UV6, UV8, UV12, and UV13) and three spontaneous mutants (S1, S5, and S7). The *gaaR* coding regions as well as 300 bp flanking regions were sequenced, but no mutation in the *gaaR* gene in any of these eleven mutants was found (data not shown).

To determine whether the constitutive expression of GA-induced genes in these mutants involves a functioning GaaR transcription factor, we deleted the *gaaR* gene in seven of the constitutive mutants (UV1, UV3, UV4, UV5, UV6, UV8, and S1) and analyzed constitutive expression using the *PpgaX-amdS* reporter. All seven mutants were unable to grow on glucose/acetamide plates (data not shown), indicating that the constitutive expression of the GA-induced genes requires a functional *gaaR* gene.

To identify mutation(s) in the gene(s) responsible for the constitutive phenotype, the genomes of five mutants (UV1, UV8, S1, S5, and S7) and the parental strain JN29.2 were sequenced. Table 2 summarizes the number of SNPs and indels detected in the five mutant strains and Table S2 lists positions and type of all SNPs and indels detected. Spontaneous mutant S7 contains only 11 SNPs or indels, of which 10 are located in intergenic regions and only one SNP mutated a gene (NRRL3\_08194). Remarkably, in all four other mutants a mutation was found in the same gene. Two of the mutants carry nonsense mutations (UV1 and S5) and one carries a frame-shift mutation (S3), all leading to premature stop codons and predicted to result in truncated proteins (Table 2). Mutants S7 and UV8 have missense mutations in the C-terminal part of the protein. These results strongly suggest that the constitutive expression of genes encoding pectin-degrading enzymes in the five mutants is caused by a loss of function of the protein encoded by NRRL3\_08194.

#### **Deletion of NRRL3\_08194 results in the constitutive expression of genes required for PGA breakdown and GA catabolism**

To determine whether the constitutive expression of the target genes of the GaaR transcriptional activator (Alazi *et al.* 2016) is caused by a loss-of-function mutation in NRRL3\_08194, we



**Figure 1** Enzymatic and RNA blot analysis of mutants with constitutive expression of genes involved in polygalacturonic acid utilization. (A) Supernatant (10  $\mu$ l) from glucose-grown cultures of reference strains N402 and JN29.2, 11 UV mutants, and two spontaneous mutants were spotted on polygalacturonic acid agarose medium to detect polygalacturonase activity. (B) Northern blot analysis of selected GA-responsive genes in the reference strains N402 and JN29.2, 11 UV mutants, and two spontaneous mutants.

deleted this gene in the *pgaX-amdS* reporter strains JC5.1 and JN29.2 ( $\Delta creA$ ), as well in a parental background without reporter constructs (MA323.1) (Table 1). The deletion mutants were purified and deletion of NRRL3\_08194 was confirmed by Southern blot analysis for JC5.1, JN29.2, and MA323.1 (Figure S1 and Figure S2). The verified deletion mutants were tested for growth on acetamide plates containing different carbon sources (Figure 2), as well as for constitutive expression of polygalacturonases (see section GaaX is induced on galacturonic acid and localized in the cytosol). Figure 2 shows that deletion of NRRL3\_08194 in the  $\Delta creA$  background (JN122) resulted in the ability to grow on glucose/acetamide, fructose/acetamide, and sorbitol/acetamide. The colony size on all three different carbon sources was similar, indicating that the *amdS* gene was expressed regardless of the carbon source used. However, deletion of NRRL3\_08194 in the JC1.5 reporter strain (JN123) resulted in similar growth on fructose/acetamide and sorbitol/acetamide plates as JN122, but reduced growth on glucose. This indicates that glucose-mediated CCR repressed *PpgaX*-driven *amdS* expression even in the absence of NRRL3\_08194. The ability of the  $\Delta NRRL3_08194$  strain to grow on acetamide plates strongly suggests that a loss of function of NRRL3\_08194 results in constitutive expression of *pgaX* and other pectinolytic genes. Furthermore, deletion of *gaaX* did not result in an altered growth behavior on GA, PGA, apple pectin, glucose, fructose, sorbitol, xylose, and arabinose (data not shown). These results are most easily explained by proposing that NRRL3\_08194 encodes a repressor protein, which we name GaaX, that represses the activity of the GaaR transcription factor in the absence of GA. Interestingly, GaaX shows sequence similarity to a previously identified repressor protein, QutR (Grant *et al.* 1988). Moreover, the transcriptional activator (GaaR, NRRL3\_08195) and the repressor (GaaX, NRRL3\_08194) are clustered in the genome, similar to the quinic acid utilization transcriptional activator (QutA/Qa-1F) and repressor (QutR/Qa-1S) in *Aspergillus nidulans* and *Neurospora crassa*, respectively (Geever *et al.* 1989; Levesley *et al.* 1996).

### The GaaR–GaaX target gene regulon

We posit that the regulation of GA-responsive genes is likely to be negatively controlled by the repressor protein GaaX. A

possible mode of action is that the repressor GaaX inhibits the activity of the transcriptional activator GaaR in the absence of an inducer. This would imply that deletion of the repressor or activation of the transcription factor by growth on GA would result in activation of the same set of genes. To show that the loss of function of the repressor activates the GA regulon and to identify the genes repressed by GaaX under noninducing conditions, RNA-seq profiles of the  $\Delta gaaX$  and its parental strain (MA234.1) were compared after growth on fructose, a nonrepressing carbon source. Controlled cultivations in bioreactors showed that the growth rates [ $\mu_{max}$  parental strain  $0.214 \pm 0.007$  g dry wt/kg/hr ( $n = 3$ );  $\mu_{max}$   $\Delta gaaX$   $0.223 \pm 0.004$  g dry wt/kg/hr ( $n = 2$ )] as well as biomass yields [ $Y_{max}$  parental strain  $4.15 \pm 0.13$  g dry wt/kg ( $n = 3$ );  $Y_{max}$   $\Delta gaaX$   $4.29 \pm 0.19$  g dry wt/kg ( $n = 2$ )] of the two strains were highly comparable, indicating the *gaaX* deletion did not result in major physiological changes affecting the growth and biomass yield.

To identify differentially expressed genes in the  $\Delta gaaX$  strain as compared to its parental strain, RNA-seq was performed on RNA isolated from exponentially growing cells at the time point at which about 75–80% of the maximum biomass yield was reached. RNA-seq reads were mapped to the NRRL3/N400 genome as this is the parent of the laboratory strain N402 and derivatives used in this study. TPM values were calculated using Salmon (Patro *et al.* 2016) (Table S3). Analysis of differential gene expression, based on a stringent false discovery rate (FDR)  $< 0.001$  and a fold change (FC)  $> 4.0$ , identified 37 upregulated genes (Table 3). Gene Ontology (GO) enrichment analysis using FetGOat (Nitsche *et al.* 2012) and manual inspection of the genes upregulated in the  $\Delta gaaX$  mutant indicated that genes involved in pectin catabolism were highly enriched. Of the 37 genes, 16 are predicted to encode extracellular enzymes acting on the GA backbone of pectin or acting on pectin side chains (Table 3). Nine genes in the group of 37 upregulated genes in the  $\Delta gaaX$  strain are predicted to encode intracellular proteins. Four of these nine genes (*gaaA–gaaD*) are required for the conversion of GA into pyruvate and glycerol (Martens-Uzunova and Schaap 2008). The exact role of the other five genes and their possible role in GA catabolism is currently unknown. The

**Table 2 Mutations in the constitutive mutants as compared to the parental strain JN29.2**

Strain	Total number of SNPs and indels	SNPs or indels in coding region	Mutation in NRRL3_08194	Position of mutation relative to ATG of NRRL3_08194	Mutation in codon (bold)	Amino acid change	Predicted protein length (full length protein is 697 amino acids)
JN29.2_UV1	40	19	G-T	1372	<b>GAG-TAG</b>	E to Stop	457 aa
JN29.2_UV8	21	4	G-A	1577	<b>GGA-GAA</b>	G to E (526)	697 aa
JN29.2_S1	68	34	Extra G	1958	GTT-G <b>GT</b>	V to G out of frame	663 aa
JN29.2_S5	48	11	C-T	1105	<b>CAA-TAA</b>	Q to Stop	368 aa
JN29.2_S7	11	1	T-C	2027	<b>CTG-CCG</b>	L-P (676)	697 aa

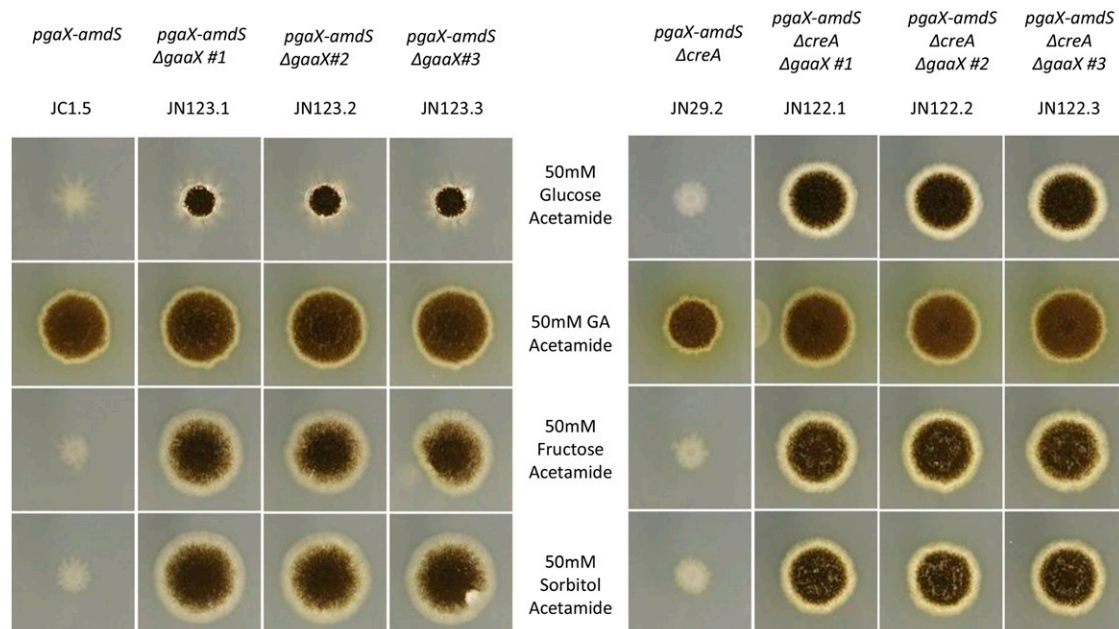
group of 37 upregulated genes also includes seven genes predicted to encode sugar transporter proteins. Of these seven transporter-encoding genes, only GatA has been studied in detail and shown to be able to transport GA (Sloothaak *et al.* 2014). Apart from the genes encoding extracellular enzymes (16), transporters (seven), and enzymes possibly involved in GA catabolism (nine), the remaining five genes in this group encode proteins with unknown functions or with similarities to known proteins that, for now, cannot be directly linked to GA metabolism. The deletion of *gaaX* has the most profound effect on the transcript levels of the genes encoding the first three steps of the GA utilization pathway (*gaaA*, *gab*, and *gaaC*) and on the expression of *gatA*. Deletion of *gaaX* resulted in a 1.24-fold ( $P = 0.000035$ ) increase in *gaaR* gene activity. Since the upregulation of *gaaR* in the  $\Delta gaaX$  mutant is modest, it is likely that the repressing activity of GaaX is mediated at the protein level (*e.g.*, by interacting with GaaR) rather than by transcriptional control of *gaaR*. Seventeen of the 37 genes upregulated in the  $\Delta gaaX$  mutant were previously identified as part of the GA regulon (Martens-Uzunova and Schaap 2008; Alazi *et al.* 2016) (Figure 3 and Table 3). Sixteen of the 17 genes found in common with previous studies are predicted or demonstrated to encode extracellular pectin-degrading enzymes. These results indicate that loss of function of *gaaX* affects the expression of the GA regulon. The other 20 genes were identified as significantly upregulated in the *gaaX* mutant, but these were not identified previously as being part of the GA regulon (Figure 3 and Table 3). A re-examination of the expression of these 20 genes in the RNA-seq data published earlier (Alazi *et al.* 2016) indicated that 10 of the genes (indicated in Table 3 by the asterisk, Table S4) were also GA-induced or GaaR-dependent for induction in this previous study. On the other hand, 15 genes identified to be GA-induced in a GaaR-dependent manner in the previous study (Alazi *et al.* 2016) were not significantly upregulated in the *gaaX* deletion strain (Figure 3). These results therefore suggest that full induction of GA-inducible genes requires more than the loss of GaaX activity, and that an additional induction mechanism plays a role.

An additional GA-induced gene identified in the study of Martens-Uzunova and Schaap (2008) but missing in the GaaR study is *gaaX* itself. Expression of *gaaX* was not examined in the Alazi *et al.* (2016) study as its function was not yet directly linked to GA utilization. However, re-evaluation of

the dataset revealed that the induced expression of *gaaX* on GA is dependent on GaaR (FC of wild type *vs.*  $\Delta gaaR$ : 18.7;  $P = 0.003$ ; Table S4). Combining the expression data of the  $\Delta gaaX$  mutant (this study), the  $\Delta gaaR$  mutant (Alazi *et al.* 2016) and the genes induced on GA (Martens-Uzunova and Schaap 2008), we propose a panregulon of 53 GaaR–GaaX controlled genes and a core GaaR–GaaX regulon of at least 27 genes (Figure 3, Table 3, and Table S4). These 27 genes include 11 genes present in the intersection of all three data sets, six genes present in the intersection of the  $\Delta gaaX$  data and the  $\Delta gaaR$  data (Alazi *et al.* 2016), nine genes identified by examining the *gaaX* dataset with supporting evidence from previous studies, and *gaaX* (Figure 3). Of these 27 genes, all except NRRL3\_00660 (carboxyesterase), NRRL3\_10865 ( $\alpha$ -N-arabinofuranosidase), NRRL3\_03342 (short-chain dehydrogenase/reductase), NRRL3\_08833 (hypothetical protein), and NRRL3\_02479 ( $\beta$ -galactosidase), have at least one predicted GARE motif in the upstream regions of the coding region (Table 3). It is interesting to note that among the genes listed in Figure 3 and Table 3 that are upregulated in the  $\Delta gaaX$ , some of them (NRRL3\_00957 and NRRL3\_00958; NRRL3\_09862 and NRRL3\_09863; NRRL3\_03291 and NRRL3\_03292) are clustered. Except for NRRL3\_00958, which encodes a GA-specific transporter (Sloothaak *et al.* 2014), the possible role of these genes in pectin degradation is currently unknown.

#### ***GaaX is induced on galacturonic acid and localized in the cytosol***

GaaX was previously identified as a GA-induced gene with unknown function (Martens-Uzunova and Schaap 2008). To monitor the induction of GaaX and to localize the GaaX protein in the cell, GaaX was fused to GFP at either the N- or C-terminal part of GaaX and expressed from the endogenous GaaX promoter. Fusion constructs were targeted to the *pyrG* locus of *A. niger* in a strain lacking endogenous *gaaX* (JN125.1) to be able to test complementation of the GFP-GaaX and GaaX-GFP fusion proteins (Figure S3). As shown in Figure 4A, JN125.1 ( $\Delta gaaX::nicB$ ) constitutively expressed pectinases indicated by the halo on PGA plates, while both the C-terminally tagged as well as the N-terminally tagged versions of GaaX (JN126.2 and JN127.3, respectively) complemented the constitutive expression phenotype, indicating that both fusion proteins are functional. Confocal fluorescent



**Figure 2** Regulation of the *pgaX* expression is controlled by GaaX and by CreA-mediated glucose repression. Growth of *pgaX-amdS* reporter strains in *gaaX* deletion and their parental strains was examined on various carbon sources. Parental strains and corresponding deletion strains were grown on MM-acetamide supplemented with 50 mM glucose, galacturonic acid (GA), fructose, or sorbitol. All strains carry the *PpgaX-amdS* reporter construct. Strain JC1.5 is the parent of three independent transformants (JN123.1, JN123.2, and JN123.3) that contain a deletion in the *gaaX* gene. Strain JN29.2 carries the  $\Delta creA$  marker and is the parent of three independent transformants (JN122.1, JN122.2, and JN122.3) that contain a deletion in the *gaaX* gene.

microscopy was performed on GFP-tagged strains to localize GaaX (Figure 4B). Spores were germinated either on GA or on fructose (a nonrepressing carbon source) and a fluorescent signal was only detectable in the GFP-labeled strains after growth on GA. This observation confirms the results from the expression data that indicate that GaaX is lowly expressed under noninducing conditions and is induced on GA. The expression of GaaX is low on fructose and no GFP signal above the background level was detected on fructose. Based on the fluorescent pictures, GaaX is likely to be localized in the cytosol.

## Discussion

The forward genetic screen with a positive selection strategy for the isolation of *A. niger* mutants with constitutive expression of genes involved in PGA degradation resulted in the identification of a repressor protein (NRRL3\_08194), which we named GaaX. Both the genome sequencing of five independently obtained mutants, as well as the analysis of a targeted deletion mutant ( $\Delta gaaX$ ), showed that the loss of function of *gaaX* leads to constitutive expression of genes previously identified as GA-induced genes (Martens-Uzunova and Schaap 2008) and genes encoding pectinolytic enzymes that are activated via the transcription factor GaaR (Alazi *et al.* 2016). Deletion of *gaaX* did not result in a growth alteration on any carbon source tested (Figure 2 and data not shown). Transcriptome analysis (Table S3) strongly suggests that deletion of *gaaX* only affects the expression of genes related to the degradation and metabolism of (poly)galacturonic acid. Genes

encoding enzymes involved in the hydrolysis of nonpectin polysaccharides are not differentially regulated in  $\Delta gaaX$ . In addition, GO enrichment analysis of  $\Delta gaaX$  transcriptome shows a strong correlation only between the activity of GaaX and the expression of GA-induced genes. In agreement with these observations, the phenotype of the *gaaR* deletion mutant was specific for (poly)galacturonic acid, with no growth defect observed on other substrates tested (glucuronic acid, rhamnose, xylose, and arabinose) (Alazi *et al.* 2016). Taken together, these findings indicate that GaaR and GaaX are specifically involved in the regulation of pectin catabolism.

Interestingly, the *gaaX* gene is located next to the recently identified GA-specific transcriptional activator *gaaR* (NRRL3\_08195). The GaaR transcriptional activator is conserved in 19 out of the 20 *Aspergillus* species for which genomic sequences are available via *Aspergillus* Genome Database (AspGD), and only absent in *Aspergillus glaucus* (Alazi *et al.* 2016), which corresponds with the inability of *A. glaucus* to grow on GA (<http://www.fung-growth.org/>). In all 19 *Aspergillus* species containing GaaR, a GaaX ortholog could be identified adjacent to GaaR. Only in *Aspergillus fumigatus* (Figure 5) and *Aspergillus wentii* (data not shown) were ORFs predicted to be present in between *gaaX* and *gaaR*. The ORFs between *gaaX* and *gaaR* in *A. fumigatus* are Afu4g06430 and Afu4g06450. Afu4g06430 is predicted to encode a 128-aa long protein that has no ortholog in other aspergilli. According to available expression data (Lind *et al.* 2015), this gene is not expressed, and it is questionable whether this predicted gene actually encodes a protein. Afu4g06450 is predicted to encode a Tan1-related transposase of the DDE family. This type of transposase is



**Table 3 Comparative RNA-seq analysis between wild-type and  $\Delta$ gaaX strains: genes (37) upregulated in  $\Delta$ gaaX strain**

Gene ID (NRRL3)	Gene ID (CBS513-88)	Gene name	Description	Average expression value wt <sup>a</sup>	Average expression value $\Delta$ gaaX <sup>a</sup>	FC $\Delta$ gaaX vs. wt <sup>b</sup>	False discovery rate <sup>b</sup>	Predicted localization	GARE element <sup>c</sup>
NRRL3_05649	An02g07720 <sup>d,e</sup>	<i>gaaC</i>	L-threo-3-deoxy-hexylosanate aldolase	12.54	2283.77	169.39	0.00E+00	Intracellular	(-) -292, -606
NRRL3_09863 <sup>f</sup>	An11g03500		$\alpha$ -Hydroxy acid dehydrogenase	0.82	165.84	160.07	0.00E+00	Intracellular	(+) -543, (-) -182
NRRL3_00958	An14g04280 <sup>d,e</sup>	<i>gatA</i>	MFS-type sugar/inositol transporter	3.47	524.95	140.38	0.00E+00	Membrane	(+) -360
NRRL3_06890	An16g05390 <sup>d,e</sup>	<i>gaaB</i>	L-galactonic acid dehydratase	47.77	6256.70	121.49	0.00E+00	Intracellular	(+) -326
NRRL3_03291	An12g05600		Heterokaryon incompatibility protein	0.00	6.78	82.98	6.48E-78	Intracellular	(+) -737, -325
NRRL3_05650	An02g07710 <sup>d,e</sup>	<i>gaaA</i>	D-galacturonic acid reductase	19.92	1515.44	71.84	0.00E+00	Intracellular	(+) -414, -100
NRRL3_03144	An12g07500 <sup>d,e</sup>	<i>pgaX</i>	Exo-polygalacturonase	1.36	51.63	32.66	2.60E-272	Extracellular	(+) -388
NRRL3_05252	An02g12505 <sup>e</sup>	<i>pmeC</i>	Pectin methylesterase	1.06	31.32	25.03	7.10E-189	Extracellular	(+) -275, -246, -35
NRRL3_06244 <sup>f</sup>	An02g00140		Glycoside hydrolase family 43 protein	0.81	22.19	23.46	8.13E-193	Extracellular	(-) -96, (-) -712
NRRL3_08281	An03g06740 <sup>d,e</sup>	<i>pgxB</i>	Exo-polygalacturonase Pgx28B	0.00	2.10	22.95	3.59E-43	Extracellular	(-) -298, -823
NRRL3_10559	An18g04810 <sup>e</sup>	<i>rgxC</i>	Glycoside hydrolase family 28 protein	0.08	3.11	17.32	1.33E-45	Extracellular	(+) -852, (-) -250
NRRL3_05260	An02g12450 <sup>d,e</sup>	<i>pgxC</i>	Exo-polygalacturonase Pgx28C	0.95	16.77	15.22	1.83E-144	Extracellular	(+) -268, (-) -642
NRRL3_08663	An03g01620 <sup>d,e</sup>		MFS-type sugar/inositol transporter	0.27	5.62	14.28	8.28E-46	Membrane	(+) -673
NRRL3_03342 <sup>f</sup>	An12g04990		Short-chain dehydrogenase/reductase	0.81	14.46	13.51	2.10E-62	Intracellular	None
NRRL3_10865	An08g01710 <sup>d,e</sup>	<i>abfC</i>	$\alpha$ -N-arabinofuranosidase	0.80	11.21	11.92	1.19E-84	Extracellular	None
NRRL3_09862	An11g03510		Hypothetical protein	0.00	0.93	11.05	4.32E-18	Unknown	(-) -517, -829, -843
NRRL3_10050	An11g01120 <sup>d,e</sup>	<i>gaaD/larA</i>	NADPH-dependent erythrose reductase Err1	256.41	2732.37	10.16	0.00E+00	Intracellular	(-) -538, -583, -801 -813
NRRL3_00957	An14g04260		B3/B4 domain-containing protein	1.53	18.56	10.05	2.46E-64	Unknown	None
NRRL3_00502	An09g06200		Hypothetical protein	0.93	12.38	9.36	1.08E-32	Unknown	(-) -189
NRRL3_10558 <sup>f</sup>	An18g04800		$\alpha$ -L-rhamnosidase	0.35	3.85	9.09	2.19E-61	Extracellular	(+) -365
NRRL3_11710	An06g00620		MFS-type sugar/inositol transporter	2.86	25.29	8.05	1.39E-118	Membrane	(+) -487, (-) -368
NRRL3_06053	An02g02540 <sup>d,e</sup>	<i>paeA</i>	Carbohydrate esterase family 16 protein	2.06	17.62	7.76	5.39E-107	Extracellular	(+) -1238
NRRL3_01073	An14g05840		O-methyltransferase, COMT-type	0.54	5.57	7.16	1.06E-21	Intracellular	(+) -300
NRRL3_07382	An16g00540		$\alpha$ -L-fucosidase	0.04	0.67	7.07	5.92E-14	Extracellular	(-) -606
NRRL3_00660 <sup>f</sup>	An14g00860		Carboxylesterase	0.21	1.87	6.98	1.47E-31	Extracellular	None
NRRL3_08499	An03g03960		Uncharacterized protein	0.48	4.58	6.71	3.84E-21	Unknown	None
NRRL3_07470	An04g09690 <sup>e</sup>	<i>pmeB</i>	Pectin methylesterase	0.75	5.72	6.36	3.59E-43	Extracellular	(+) -389
NRRL3_02479	An01g10350 <sup>e</sup>	<i>lacB</i>	Exo- $\beta$ -1,4-galactanase	3.48	22.29	6.03	1.37E-179	Extracellular	None
NRRL3_03292	An12g05590		Carboxylesterase	0.18	1.86	5.26	1.75E-09	Extracellular	(-) -1000
NRRL3_01237	An19g00270 <sup>e</sup>	<i>pelD</i>	Pectin lyase	0.17	1.47	5.25	6.14E-14	Extracellular	(-) -409, -465
NRRL3_03467	An12g03550		MFS-type transporter	1.13	6.73	5.05	4.03E-28	Membrane	None
NRRL3_01127 <sup>f</sup>	An14g06500	DHAK	Dihydroxyacetone kinase	18.90	100.64	4.98	1.61E-102	Intracellular	(+) -69
NRRL3_11054 <sup>f</sup>	An08g04040		MFS-type sugar/inositol transporter	6.62	31.89	4.57	2.57E-126	Membrane	(+) -778, -726, (-) -646
NRRL3_08833 <sup>f</sup>	N.a.		Hypothetical protein	0.31	2.02	4.52	1.58E-12	Unknown	None
NRRL3_01398	An13g02090 <sup>f</sup>		MFS-type transporter	2.81	13.10	4.16	5.62E-30	Membrane	(-) -270
NRRL3_08325	An03g06310 <sup>e</sup>	<i>pmeA</i>	Pectin methylesterase Pme8A	0.04	0.56	4.10	1.49E-07	Extracellular	(+) -983, (-) -308
NRRL3_02770	An01g13880		MFS-type transporter	0.87	4.07	4.06	1.57E-20	Membrane	(+) -578

Gray boxes indicate genes proposed to be part of the GaaR/GaaX regulon.

<sup>a</sup> Expression values (TPM) are averages of biological triplicates for the wild type (wt) and duplicates for the  $\Delta$ gaaX strain.

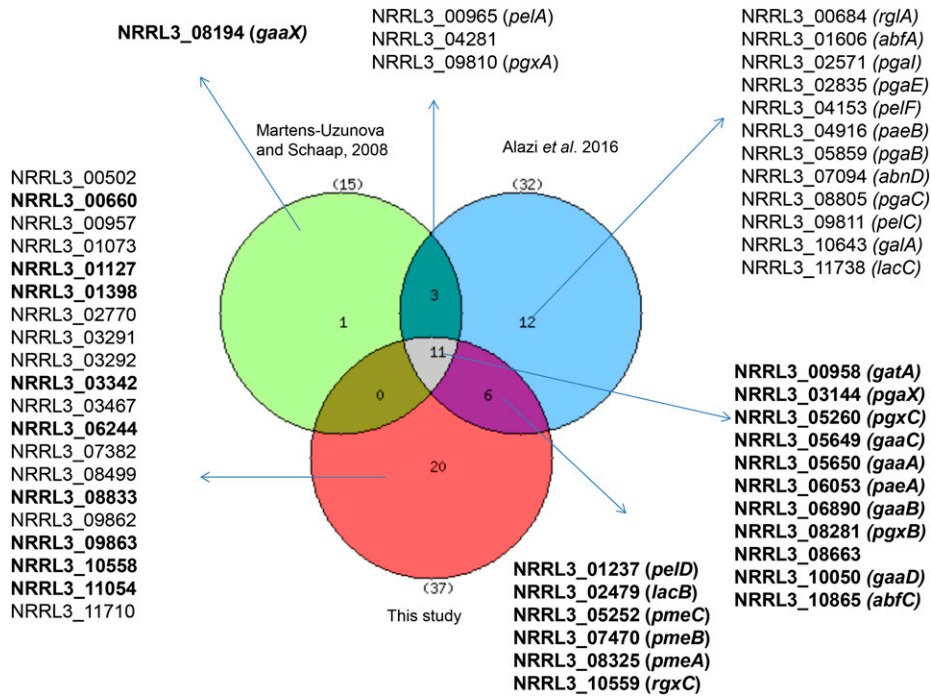
<sup>b</sup> Fold changes and FDR-values are calculated with DESeq2, and genes with fold changes  $\geq 4$  and FDR-values  $\leq 0.001$  are listed.

<sup>c</sup> GA-responsive element (GARE) position (CCNCAA) is given with respect to the translational start site.

<sup>d</sup> Genes identified as the GA-regulon by Martens-Uzunova and Schaap (2008).

<sup>e</sup> Genes identified as the GA-regulon by Alazi et al. (2016).

<sup>f</sup> Gene not identified as components of the GA regulons in the studies by Martens-Uzunova and Schaap (2008) and Alazi et al. (2016), but are likely to be under the regulation of GaaR (see text for details).



**Figure 3** Venn diagram showing the overlaps between upregulated genes in the wt\_fructose vs. wt\_GA study (Martens-Uzunova and Schaap 2008), the upregulated genes between  $\Delta gaaR$ -GA vs. wt-GA (Alazi et al. 2016), and the upregulated genes in wt\_fructose vs.  $\Delta gaaX$ -fructose (this study) to identify the GA regulon. The 27 genes defining the GaaR-GaaX core regulon are indicated in bold.

found in both *A. nidulans* and *A. niger*, as well as in many other organisms. This transposase is also lowly expressed in *A. fumigatus* (Lind et al. 2015).

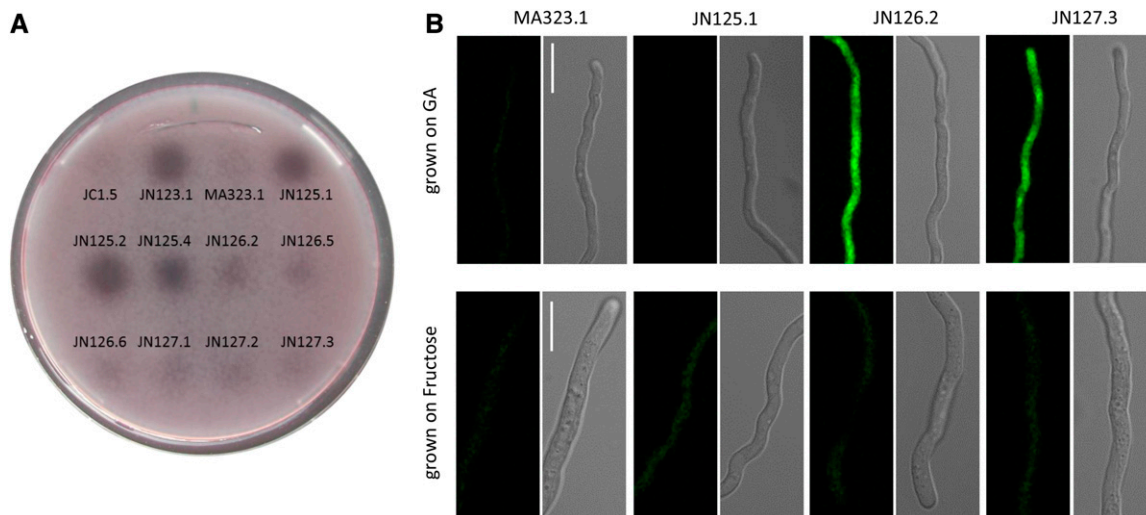
Like *gaaR*, *gaaX* is also missing in *A. glaucus*. BLASTP and synteny analysis between *A. niger* and *A. glaucus* revealed that the GaaR/GaaX encoding genes have been excised, as surrounding genes are conserved. Despite the loss of GaaX and GaaR, *A. glaucus* still possesses the GA-specific catabolic genes *gaaA* (Aspg11\_0124049), *gaaB* (Aspg11\_0091535), and *gaaC* (Aspg11\_0065497).

The GaaR transcriptional activator has previously been reported to be conserved in other Ascomycetes belonging to the Pezizomycotina subdivision, including members of the Eurotiomycetes (*Penicillium*, *Talaromyces* spp), Leotiomycetes (*Botrytis*, *Oidiodendron*), Sordariomycetes (*Neurospora*, *Myceliophthora*, *Magnaporthe*, *Trichoderma*, and *Fusarium* spp.), and Dothideomycetes [*Zyoseptoria* (*Mycosphaerella*), *Aureobasidium*, and *Cochliobolus* spp.] (Zhang et al. 2016). Synteny analysis of 17 species belonging to four classes of Pezizomycetes (Eurotiomycetes, Leotiomycetes, Sordariomycetes, and Dothideomycetes) revealed a strong conservation of the genomic clustering of *gaaR* and *gaaX* orthologs (Figure 5 and Table S5). For most fungal species analyzed, *gaaR* and *gaaX* are next to each other on the chromosome or close to each other and separated by one to five genes (Figure 5). The head to tail orientation of *gaaR*-*gaaX* driving expression of *gaaR* and *gaaX* from different promoters is conserved in all species except in *Oidiodendron maius*. Like GaaR, GaaX was found only in the Pezizomycotina and not in ascomycete yeasts, zygomycetes, or basidiomycetes.

The strategy to identify the responsible mutation by sequencing five independently obtained mutants has been

successful and efficient. Clearly, sequencing only a limited number of mutants leads only to successful identification when the mutants isolated in the screen all belong to a single complementation group. If more complementation groups are involved, more mutants would need to be sequenced. It is interesting to note that in addition to mutations in *gaaX* which were present in all five mutants, we noticed that two mutants (S1 and UV1) also contained allelic mutations in NRRL3\_06175 (Table S2). The protein encoded by this gene is predicted to encode a cocaine esterase and belongs to a protein subfamily of hydrolases that included cocaine esterase (CocE), several glutaryl-7-ACA acylases, and the putative diester hydrolase NonD of *Streptomyces griseus*. This family shows extensive, low-level similarity to a family of Xaa-Pro dipeptidyl-peptidases. Whether this gene also contributes to the constitutive expression of GA-dependent genes remains to be determined, but this is unlikely as mutants without mutations in this gene display essentially the same constitutive phenotype.

Previous studies have identified genes specifically induced by GA (Martens-Uzunova and Schaap 2008) and pectinolytic genes that were dependent on the GaaR transcriptional activator for induction by GA (Alazi et al. 2016). Eleven of the 15 GA-induced genes identified by Martens-Uzunova and Schaap were upregulated in the *gaaX* mutant (Figure 3 and Table 3). The three genes that are considered GA-inducible but not detected as differentially expressed in the *gaaX* mutant are predicted to encode a transporter (NRRL3\_04281), an exo-polygalacturonase (NRRL3\_09810, *pgxA*), and a pectin lyase (NRRL3\_00965, *pelA*). These three genes were not classified as differentially expressed according to the stringent statistical settings in our current study. The fourth gene



**Figure 4** (A) Complementation analysis of GaaX-GFP fusions. Polygalacturonase activities of *gaaX* deletion strains, *gaaX-GFP* and *GFP-gaaX* complementation strains, and their parental strains were detected by spotting 50  $\mu$ l supernatant from fructose-grown cultures on polygalacturonic acid agarose. (B) Subcellular localization of GaaX-GFP and GFP-GaaX in *A. niger* germlings. Strains were grown on coverslips in Petri dishes with minimal medium (pH 5.8) supplemented with either galacturonic acid or fructose as carbon source. Bar, 10  $\mu$ m.

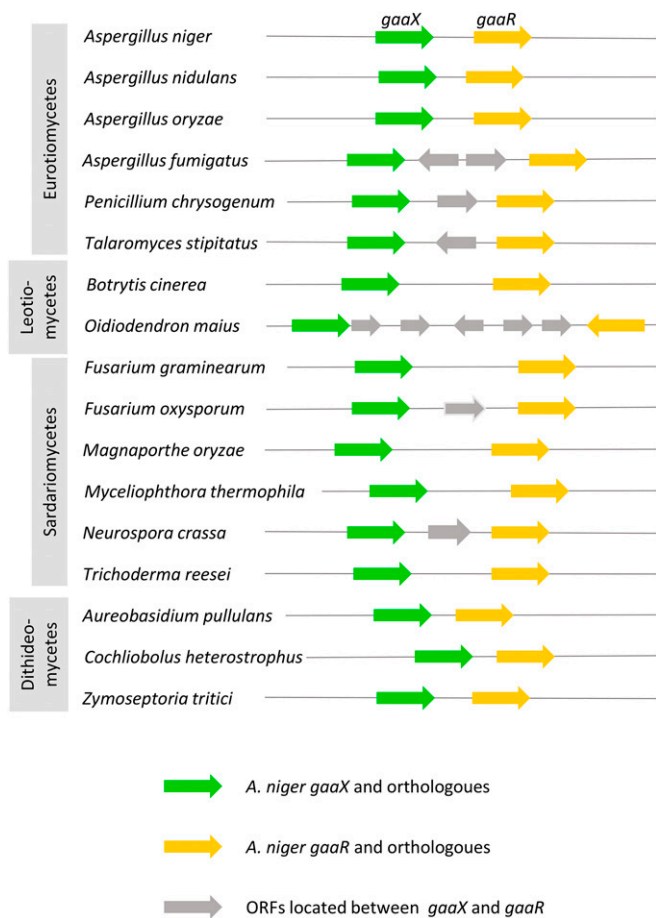
induced on GA in the study of Martens-Uzunova and Schaap (2008), but missing in our study, is *gaaX* itself.

In our recent study on the GaaR transcriptional activator, we identified 32 pectinolytic genes whose expression on GA was dependent on GaaR (Alazi *et al.* 2016). These genes overlap largely with the previously identified GA-responsive genes (Martens-Uzunova and Schaap 2008) (Figure 3 and Table 3), but also include 18 new potential GaaR target genes. Six of these genes [including NRRL3\_02479 (*lacB*), NRRL3\_05252 (*pmeC*), NRRL3\_08325 (*pmeA*), NRRL3\_07470 (*pmeB*), NRRL3\_10559 (*rgxC*), and NRRL3\_01237 (*pelD*)] were also found to be significantly upregulated in  $\Delta$ *gaaX* (Figure 3 and Table 3) and are therefore considered to be part of the core GA regulon. The remaining 12 genes identified as being GaaR dependent for induction on GA (Alazi *et al.* 2016) were not identified as differentially expressed based on the stringent settings in this study. Whether these genes are indeed directly controlled by GaaR and GaaX, and therefore part of the core GA regulon, awaits further study.

The GaaX protein is predicted to be 697 aa long and displays significant similarity to the last three domains in the C-terminal half of the AROM protein. AROM is a large (1586 aa in *A. niger*) pentafunctional protein composed of five domains and the individual domains are involved in five different enzymatic steps representing the prechorismate shikimate pathway, which is required for aromatic amino acid biosynthesis (Duncan *et al.* 1987; Hawkins and Smith 1991). The last three domains of the AROM protein encode the shikimate kinase, 3-dehydroquinase, and shikimate dehydrogenase and are homologous to the respective bacterial enzymes (*aroL*, *aroD*, and *aroE*) (Lamb *et al.* 1996). The AROM protein is present in fungi, including yeasts, and Euglena. The evolutionary origin of AROM is likely to be bacterial and it has been suggested that the AROM protein is the

result of gene fusion events (Richards *et al.* 2006). Sequence alignment and BLASTP searches showed that the GaaX protein has significant sequence homology with the last three domains of the AROM protein. The observation of a transcriptional activator (GaaR) located next to a possible repressor protein (GaaX) that displays significant homology to AROM is analogous to the clustered transcriptional activator/repressor module regulating quinic acid utilization (Geever *et al.* 1989; Lamb *et al.* 1990). Like GaaX, the quinate repressor protein shows significant sequence similarities with the last three C-terminal domains of AROM (Lamb *et al.* 1996).

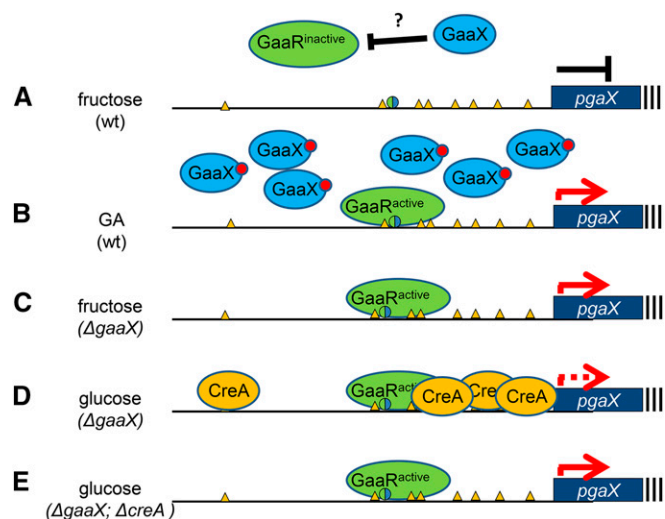
The regulation of metabolic enzymes required for quinic acid utilization has been a classical example of gene regulation both in *N. crassa* and *A. nidulans* (Geever *et al.* 1989; Leversley *et al.* 1996). In *A. nidulans* and *N. crassa*, the transcriptional activator and repressor are located in a gene cluster which consists of the activator and repressor and other genes involved in quinic acid catabolism and transport (Geever *et al.* 1989; Lamb *et al.* 1990). *A. niger* also has a quinic acid gene cluster that includes, besides the *qutA* gene (NRRL3\_11038) and *qutR* gene (NRRL3\_11039), a catabolic 3-dehydroquinase (NRRL3\_11037) and an Major facilitator superfamily (MFS) transporter possibly involved in quinate uptake (NRRL3\_11036). In contrast to the quinic acid gene cluster in which the regulatory genes (activator and repressor) are clustered with structural genes, no structural genes involved in GA utilization were clustered with GaaR and GaaX. Deletion of the *qutA* transcription factor (NRRL3\_11038) in *A. niger* results in a quinate nonutilizing mutant (M. Arentshorst and A. F. J. Ram, unpublished results). Both in *A. nidulans* and *N. crassa*, the regulation of genes involved in quinic acid metabolism has been studied in detail and is characterized by the presence of a transcriptional activator (named QutA in *A. nidulans*, and *qa-1F* in



**Figure 5** Schematic overview of the conservation of the *gaaX-gaaR* gene pair in 17 *Pezizomycotina* species. GaaX orthologs (green), GaaR orthologs (yellow), and ORFs between *gaaX* and *gaaR* (gray) are indicated. Arrow heads indicate the direction of transcription.

*N. crassa*) located next to a repressor protein (QutR in *A. nidulans*, and *qa-1S* in *N. crassa*). Loss of function of quinic acid repressor *qutR* or *qa-1S* in *A. nidulans* and *N. crassa*, respectively, leads to constitutive expression of quinic acid utilization genes (Giles *et al.* 1985; Lamb *et al.* 1996), very similar to the effect observed for the loss of function of GaaX, resulting in constitutive expression of GA utilization genes. Based on the phenotype of the *gaaX* mutant and the analogy to the organization of the quinic acid utilization gene cluster, our current working hypothesis is that *gaaX* encodes a repressor protein which is required to keep the transcriptional activator GaaR in an inactive form in the absence of the inducer molecule.

As noted earlier, *gaaX* (NRRL3\_08194) was identified as an upregulated gene when an *A. niger* culture pregrown for 18 hr with 2% fructose was transferred to a medium containing 1% GA as the sole carbon source (Martens-Uzunova and Schaap 2008). The expression of a functional GFP-tagged version of GaaX confirmed the induced expression and showed cytosolic localization of GaaX in the presence of GA (Figure 4). In the promoter region of *gaaX*, a GA-responsive element (GARE) was found, suggesting that activation of the



**Figure 6** Model for the regulation of GA-induced gene expression in *A. niger*. (A) GA-induced gene expression, with *pgaX* as an example, is controlled via interaction of the transcriptional activator (GaaR) and transcriptional repressor (GaaX) in combination with CreA-mediated carbon catabolite repression. (A) In the presence of fructose (a nonrepressible, noninducing carbon source) *pgaX* expression is prevented because GaaX inhibits GaaR activation. The question mark indicates that the mechanism by which GaaX controls GaaR activity is unknown. (B) In the presence of GA, GA itself or a derivative of GA is predicted to bind to GaaX. The binding of the inducer to GaaX is expected to activate GaaR. GaaX is induced and remains cytosolic but the presence of the inducer keeps GaaX inactive. (C) In the  $\Delta gaaX$  strain, GaaR is no longer kept inactive by GaaX and therefore is constitutively active, resulting in constitutive expression of *pgaX*. (D) In the  $\Delta gaaX$  strain, the presence of glucose leads to CreA-mediated repression leading to reduced expression of *pgaX* and possibly other pectinolytic genes. (E) Deletion of both *gaaX* and *creA* results in constitutive expression of *pgaX* even in the presence of glucose. The yellow triangles represent putative CreA binding sites. The green/blue circle represents a putative GaaR-binding site. The red circle represents the postulated inducing sugar.

transcription factor results in increased levels of repressor protein. Although this might seem contradictory at the first glance, it could be an elegant mechanism to ensure that the expression of GA-induced genes is tightly controlled and quickly responds to the presence or absence of GA. The induction of the expression of the repressor is partially analogous the activation/repression system of the *qa* cluster in *N. crassa*. In *N. crassa* it has been shown that both the activator (*qa-1F*) and the repressor (*qa-1S*) are transcriptionally induced in the presence of quinic acid (Patel *et al.* 1981; Giles *et al.* 1991). In the GA regulation system of *A. niger*, only the repressor protein is induced and not the activator. It should be noted that in almost all of 17 species analyzed, the *gaaX* and *gaaR* genes do not share the same promoter region (head to tail orientation; Figure 5), while the *qa-1S* and *qa-1F* genes of *N. crassa* share the same promoter region, which might function as a bidirectional promoter.

As a working model (Figure 6), we postulate that in the presence of GA, the inducer molecule, which could be GA or a derivative of GA, binds in the cytosol to repressor protein GaaX. Binding of the inducer to the GaaX repressor is posited

to result in the activation of the transcription factor GaaR. Active GaaR is expected to induce the expression of GA-responsive genes involved in GA release, uptake and metabolism, but also induces the expression of repressor protein. As long as the inducer is present in sufficient amounts, the GaaX repressor is predicted to be inactive as a repressor and thereby the GaaR transcription factor remains active. When the concentration of inducer decreases, it is reasonable to suggest that repressor proteins lacking bound inducer could inactivate the GaaR transcriptional activator, thereby restraining the expression of GA-responsive genes. Thus, high expression of the repressor could serve as a sensitive system to ensure that, when intracellular GA levels decrease, the cell can tightly turn off expression of GA-responsive genes. Moreover, this mechanism also ensures the rapid response to the presence of GA as it does not require *de novo* synthesis of GaaR. Induction simply requires the binding of inducer to the repressor and subsequent activation of GaaR via post-translational mechanisms, as the expression of GaaR is not dramatically induced by GA (Alazi *et al.* 2016) or in the *gaaX* mutant (this study). The expression of GA-induced genes is also controlled via CreA mediated CCR (de Vries *et al.* 2002; Niu *et al.* 2015). The analysis of the *PpgaX-amdS* reporter strain (Figure 2) suggests that the expression of *pgaX* is carbon catabolite repressed even in the  $\Delta gaaX$  strain. This suggests that CreA directly represses *pgaX* expression via CreA binding sites in the *pgaX* promoter, independent of GaaX repression (Figure 6).

The proposed model for the mechanism by which GaaR and GaaX regulate gene expression resembles in some aspects the Gal3/Gal4/Gal80 module of *Saccharomyces cerevisiae*, but shows at least two important differences. Whereas the Gal4 regulatory system consists of three proteins (Gal4 as the transcriptional activator, Gal80 as the repressor, and Gal3 as possible galactose sensor), we have identified two genes/proteins involved in GA regulation and no evidence for a third member. Also in the regulation of quinate metabolism, no third regulatory gene has been identified even though saturating mutant screens have been performed. These observations do not exclude the possibility that a third factor is involved in the GA or quinic acid regulation, but it is unlikely with the available evidence. Whereas the sensor (Gal3)/repressor (Gal80) function is mediated via two different proteins in the Gal regulatory system in *S. cerevisiae*, in the GA and quinic acid regulatory systems, the sensor/repressor function might well be performed by a single protein, GaaX and QutR, respectively. Another important difference is that GaaX and QutR do not show homology to Gal80 or Gal3, nor do Gal80 or Gal3 display homology to AROM. Based on these observations, we suggest that the GAL repressor module has evolved independently from that of GaaX/QutR.

In addition to GaaX (NRRL3\_08194) and QutR (NRRL3\_11039), we identified two additional paralogues in the *A. niger* genome (NRRL3\_08276 and NRRL3\_07605). All four paralogues showed significant homology to the *A. niger* AROM protein, as well as limited homology towards

each other. Both NRRL3\_08276 and NRRL3\_07605 are also located next to predicted Zn(II)<sub>2</sub>Cys<sub>6</sub> domain transcription factors, NRRL3\_08275 and NRRL3\_07604, respectively. Whereas the function of the GaaR/GaaX and QutA/QutR modules are related to GA and quinic acid metabolism, respectively, the function of the two other pairs that are present in *A. niger* remains to be elucidated. The sequence similarity of NRRL3\_08276 and NRRL3\_07605 to QutR and GaaX and their genome clustering with predicted transcription factors suggest that the proposed activator/repressor modules observed for GaaR–GaaX and QutA–QutR is an evolutionarily conserved mechanism to control gene expression in filamentous ascomycete fungi. The number of similar activator/repressor modules varies among Pezizomycotina species (Figure S4 and Figure S5). Most Pezizomycotina species contain the galacturonic acid and quinic acid related transcriptional activator/repressor modules. It is interesting to note that some fungi, *e.g.*, *Talaromyces stipitatus* and *B. cinerea*, seem to have lost the quinic acid specific repressor, which suggests they might have lost the capacity to utilize quinic acid. The GaaR/GaaX and QutA/QutR activator/repressor modules and their variants are specific for Pezizomycotina and missing in ascomycete yeasts, zycomycetes, and basidiomycetes.

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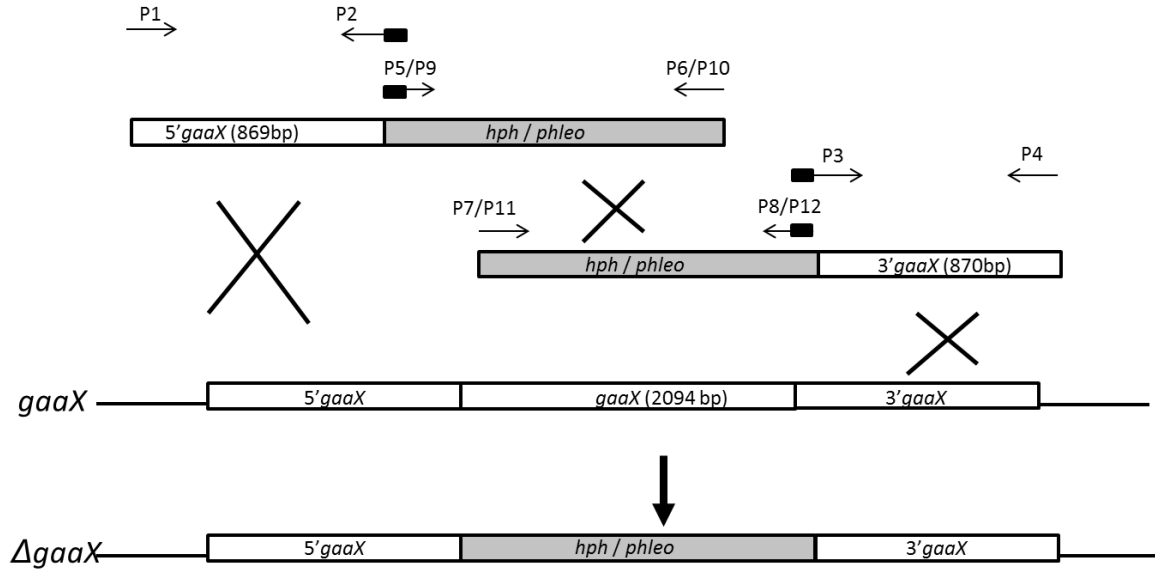
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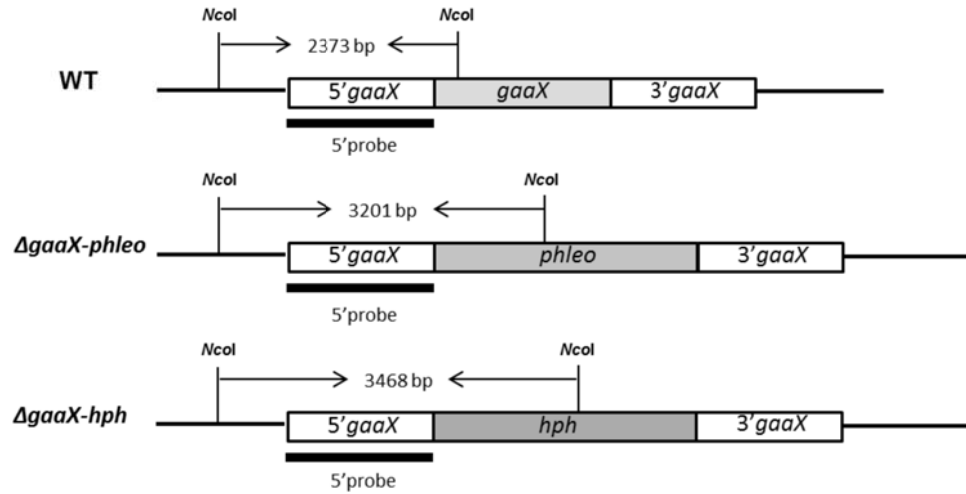
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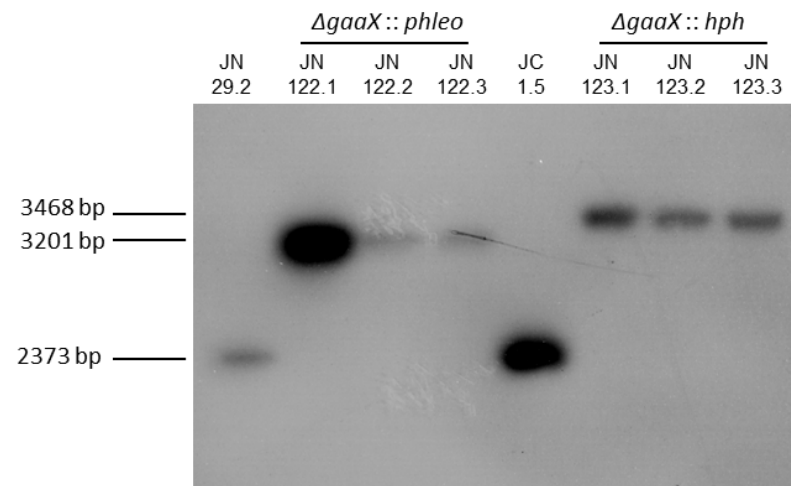
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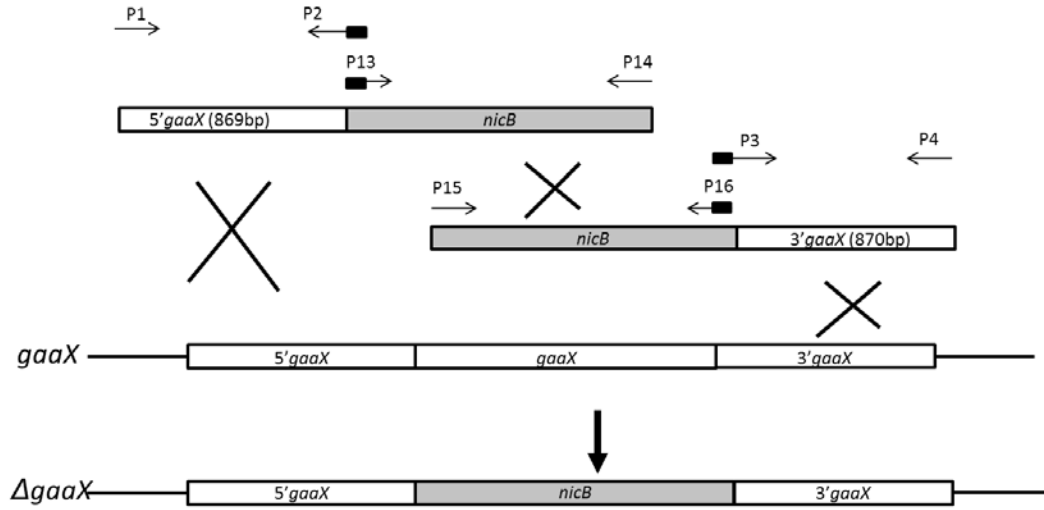


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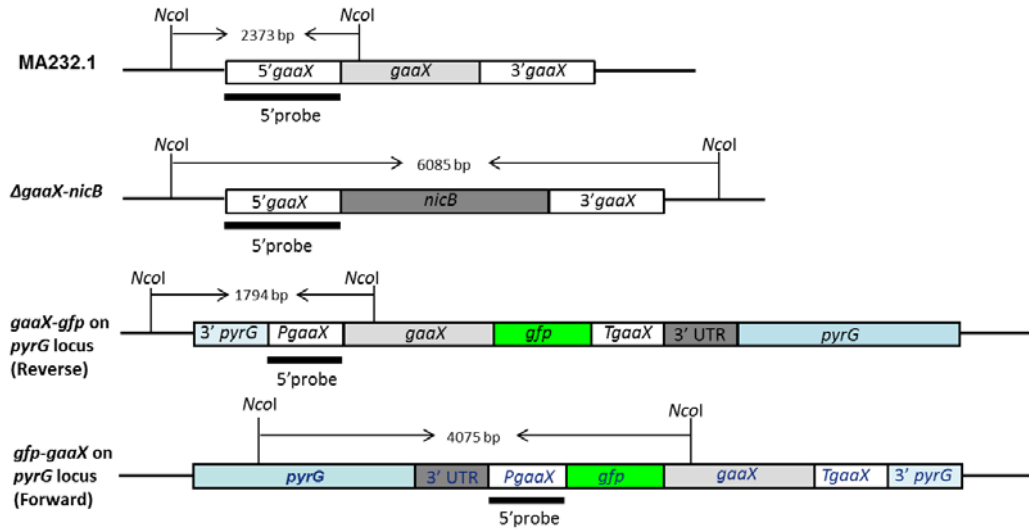




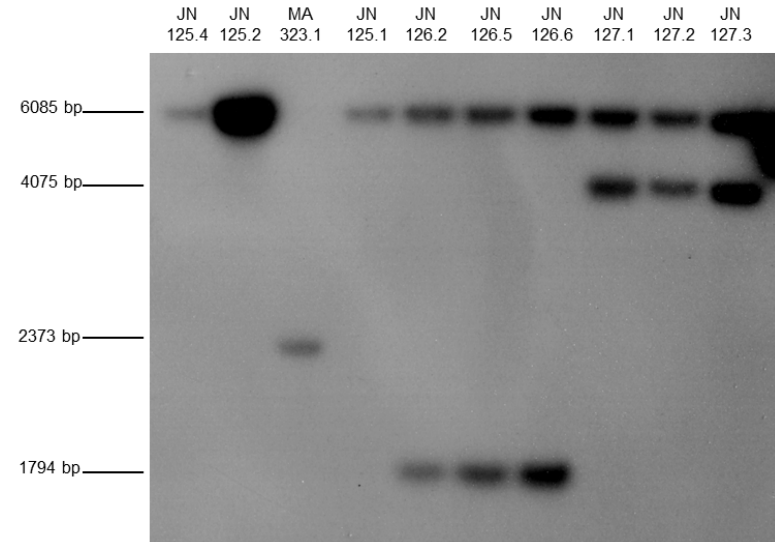
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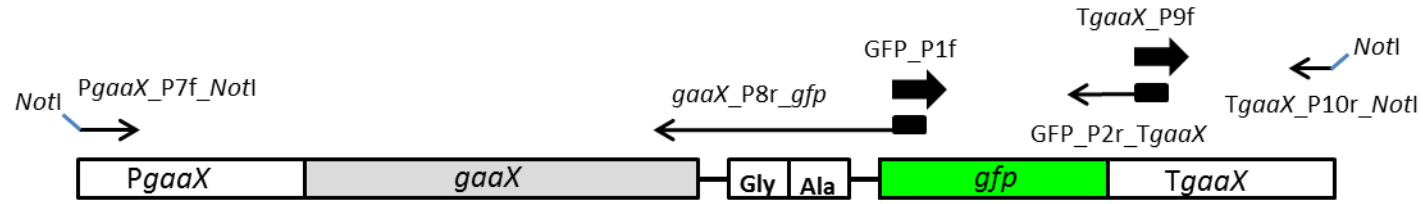


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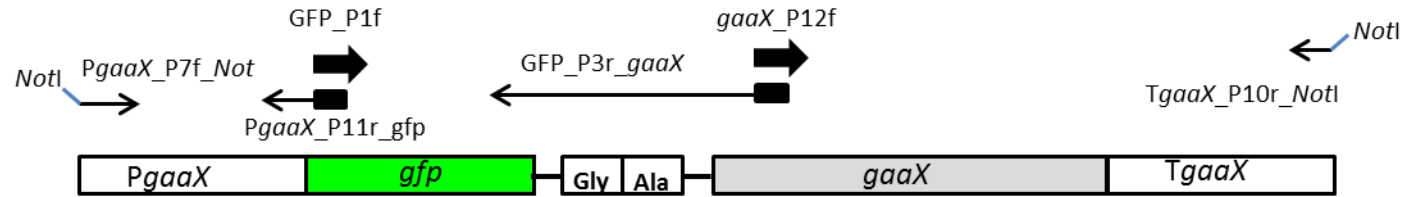


A

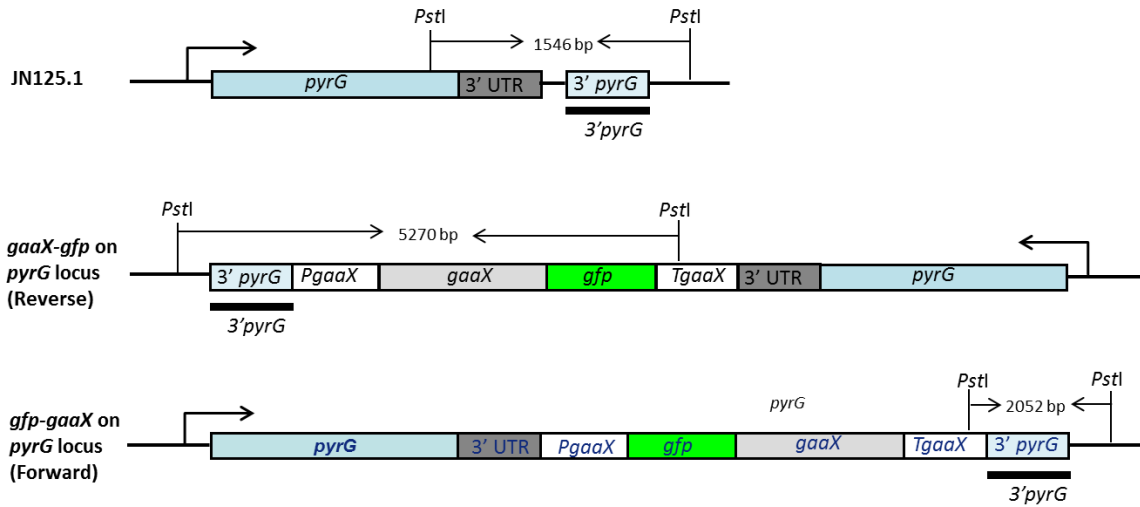
PgaaX-gaaX::gfp-TgaaX



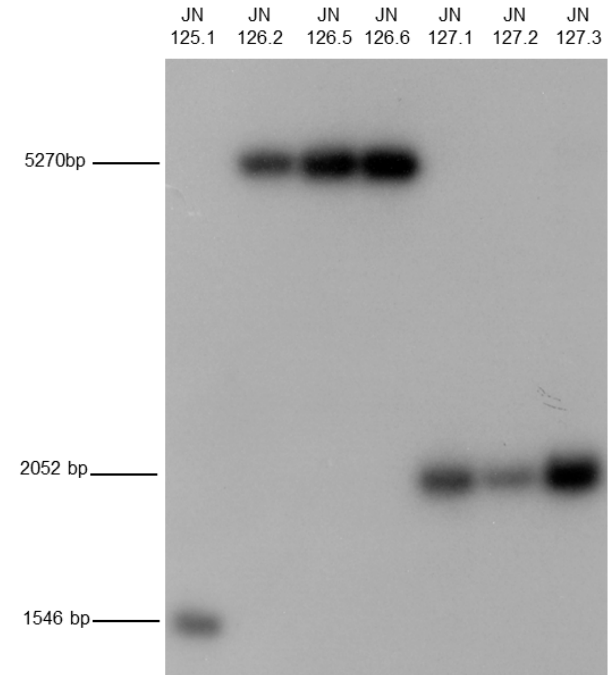
PgaaX-gfp::gaaX-TgaaX



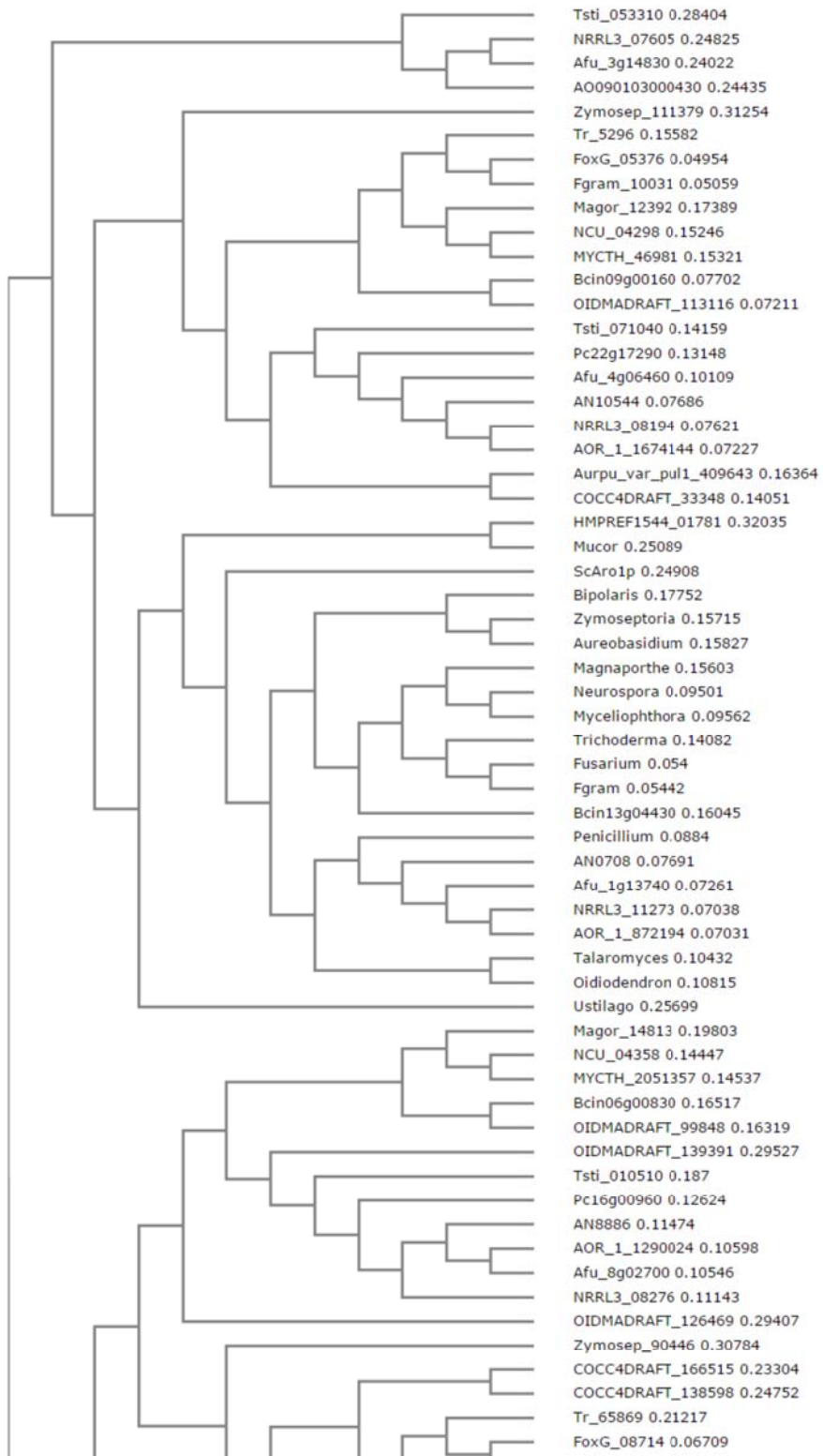
B



C







NRRL3\_07605

NRRL3\_08194  
(GaaX)

NRRL3\_11273  
(Arom)

NRRL3\_08276

Tsti\_053310 0.28404  
 NRRL3\_07605 0.24825  
 Afu\_3g14830 0.24022  
 AO090103000430 0.24435  
 Zymosep\_111379 0.31254  
 Tr\_5296 0.15582  
 FoxG\_05376 0.04954  
 Fgram\_10031 0.05059  
 Magor\_12392 0.17389  
 NCU\_04298 0.15246  
 MYCTH\_46981 0.15321  
 Bcin09g00160 0.07702  
 OIADMADRAFT\_113116 0.07211  
 Tsti\_071040 0.14159  
 Pc22g17290 0.13148  
 Afu\_4g06460 0.10109  
 AN10544 0.07686  
 NRRL3\_08194 0.07621  
 AOR\_1\_1674144 0.07227  
 Aurpu\_var\_pul1\_409643 0.16364  
 COCC4DRAFT\_33348 0.14051  
 HMPREF1544\_01781 0.32035  
 Mucor 0.25089  
 ScAro1p 0.24908  
 Bipolaris 0.17752  
 Zymoseptoria 0.15715  
 Aureobasidium 0.15827  
 Magnaporthe 0.15603  
 Neurospora 0.09501  
 Myceliophthora 0.09562  
 Trichoderma 0.14082  
 Fusarium 0.054  
 Fgram 0.05442  
 Bcin13g04430 0.16045  
 Penicillium 0.0884  
 AN0708 0.07691  
 Afu\_1g13740 0.07261  
 NRRL3\_11273 0.07038  
 AOR\_1\_872194 0.07031  
 Talaromyces 0.10432  
 Oidiodendron 0.10815  
 Ustilago 0.25699  
 Magor\_14813 0.19803  
 NCU\_04358 0.14447  
 MYCTH\_2051357 0.14537  
 Bcin06g00830 0.16517  
 OIADMADRAFT\_99848 0.16319  
 OIADMADRAFT\_139391 0.29527  
 Tsti\_010510 0.187  
 Pc16g00960 0.12624  
 AN8886 0.11474  
 AOR\_1\_1290024 0.10598  
 Afu\_8g02700 0.10546  
 NRRL3\_08276 0.11143  
 OIADMADRAFT\_126469 0.29407  
 Zymosep\_90446 0.30784  
 COCC4DRAFT\_166515 0.23304  
 COCC4DRAFT\_138598 0.24752  
 Tr\_65869 0.21217  
 FoxG\_08714 0.06709

**Supplemental Table 1** Primers used in this study. Overlapping sequences for fusion PCR are written in bold.

Primer name	Sequence (5' to 3')	Used for
gaaRP7f	TCCTCTTCGGCTTCTGCTTC	amplification and sequencing of <i>gaaR</i>
gaaRP8r	CATGTGATCATTTCGTGGCCT	amplification and sequencing of <i>gaaR</i>
gaaRP9r	GCATACTCCAGGCTCCCTTG	sequencing of <i>gaaR</i>
gaaRP10f	CCTACATGGGAAGCCCTATTGG	sequencing of <i>gaaR</i>
gaaRP11r	CGATAGAGCAGACAGTGCACGA	sequencing of <i>gaaR</i>
gaaRP12f	CCCTGTTGGTCGACATTACGC	sequencing of <i>gaaR</i>
gaaRP13r	AAATCGGGTTCGGTCCAGTG	sequencing of <i>gaaR</i>
gaaRP14f	AGGACTTCCTGGACCGGTAT	sequencing of <i>gaaR</i>
gaaXP1f (P1)	TGAGAAATGAGGATGGAAGAGG	amplification of <i>gaaX</i> 5' flank
gaaXP2r (P2)	<b>CAATTCCAGCAGCGGCTT</b> CTCATACTTGAATGCGACCGTT	amplification of <i>gaaX</i> 5' flank
gaaXP3f (P3)	<b>ACACGGCACAATTATCCAT</b> CGAAAAGCAATCACGGCAACTGG	amplification of <i>gaaX</i> 3' flank
gaaXP4r (P4)	ACAGCTGGGACGTAACCGAT	amplification of <i>gaaX</i> 3' flank
gaaXP5f	CGTTGGTTCGATGTAAATGGG	diagnostic PCR to confirm deletion of <i>gaaX</i>
gaaXP6r	GTAAGTGTTTCGGGGTTGGAG	diagnostic PCR to confirm deletion of <i>gaaX</i>
hygP6f (P5)	<b>AAGCCGCTGCTGGAATT</b> GGGCTCTGAGGTGCAGTGGAT	amplification of <i>hygB</i> marker
hygP9r (P6)	GGCGTCGGTTTCCACTATC	amplification of <i>hygB</i> marker
hygP8f (P7)	AAAGTTCGACAGCGTCTCC	amplification of <i>hygB</i> marker
hygP7r (P8)	<b>CGATGGATAATTGTG</b> CCGTGTTGGGTGTTACGGAGCATTC	amplification of <i>hygB</i> marker
phleoP4f (P9)	<b>AAGCCGCTGCTGGAATT</b> GCTCTTTCTGGCATGCGGAG	amplification of phleomycin marker
phleoP7r (P10)	CACGAAGTGCACGCAGTTG	amplification of phleomycin marker
phleoP6f (P11)	AAGTTGACCAGTGCCGTTCC	amplification of phleomycin marker
phleoP5r (P12)	<b>CGATGGATAATTGTG</b> CCGTGTGGAGCATTCACTAGGCAACCA	amplification of phleomycin marker
PgaaX-P7f-NotI	ATAAGAATGCGGCCCGCGTTGGTTCGATGTAAATGGG	amplification of the promoter of <i>gaaX</i>
PgaaX-P11r	<b>TCCTCGCCCTTGCT</b> ACCATCTCATTGCATACGCAGTATA	amplification of the promoter of <i>gaaX</i>
GFP-P1f	ATGGTGAGCAAGGGCGAGGA	amplification of GFP
GFP-P3r	<b>TGAAGGGCTGTCTGC</b> ATCATGGCGCCCTTGACAGCTCGTCCATGC	amplification of GFP
gaaXP12f	ATGATGCAGACAGCCCTTCA	amplification of <i>gaaX</i> and the terminator of <i>gaaX</i>
TgaaX-P10r-NotI	ATAGTTTACGCGCCGCGTAAGTGTTTCGGGGTTGGAG	amplification of <i>gaaX</i> and the terminator of <i>gaaX</i>
gaaX-P8r	<b>TCCTCGCCCTTGCT</b> CACCATGGCGCCAAACACCCGTCGATGCCA	amplification of <i>gaaX</i> and the promoter of <i>gaaX</i>
GFP-P2r	<b>TATACTATATAAGAAAA</b> ATCCTACTTGTACAGCTCGTCCATGC	amplification of GFP
ABpyrGP11f	AAGGAAAAAAGCGGCCCGCGTCGCGTGATAAGGGTTG	amplification of <i>pyrG</i> 3' flank for probe
ABpyrGP13r	CGAGCTCGGCGCGCCTCGGGTCAATTTCTCTGTTG	amplification of <i>pyrG</i> 3' flank for probe
pgaX_SP1f	GCTTATCAGCGGCCAGTTCT	amplification of the promoter of <i>pgaX-amdS</i>
AmdS_SP1r	TGGGAATGGAACGATGGAAGG	amplification of the promoter of <i>pgaX-amdS</i>

Table S2. SNPs and indels detected in genomes of mutants. (.xlsx, 28 KB)

[www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS2.xlsx](http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS2.xlsx)

Table S3.A) TPM values of NRRL3 predicted ORFs (NRRL3\_00001-NRRL3\_11846). B)

Differentially expressed genes between WT and  $\Delta gaaX$ . (.xlsx, 956 KB)

[www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS3.xlsx](http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS3.xlsx)

Table S4. Expression data of genes proposed to be part of the GaaR-GaaX panregulon. (.xlsx, 21 KB)

[www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS4.xlsx](http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS4.xlsx)



Table S5. Synteny of the GaaR-GaaX orthologs among 17 *Pezizomycotina*. (.xlsx, 13 KB)

[www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS5.xlsx](http://www.genetics.org/lookup/suppl/doi:10.1534/genetics.116.194050/-/DC1/TableS5.xlsx)