

Molecular Epidemiology of Human Immunodeficiency Virus Type 1 Sub-Subtype A3 in Senegal from 1988 to 2001

Seema Thakore Meloni,^{1*} Jean-Louis Sankalé,¹ Donald J. Hamel,¹ Geoffrey Eisen,¹
Aissatou Guéye-Ndiaye,² Souleymane Mboup,² and Phyllis J. Kanki¹

*Department of Immunology and Infectious Diseases, Harvard School of Public Health, Boston, Massachusetts,¹
and Laboratoire de Bactériologie-Virologie, Université Cheikh Anta Diop, Dakar, Senegal²*

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The global human immunodeficiency virus (HIV) epidemic is characterized by significant genetic diversity in circulating viruses. We have recently characterized a group of viruses that form a distinct sub-subtype within the subtype A radiation, which we have designated HIV type 1 (HIV-1) sub-subtype A, circulating in West Africa. A prospective study of a cohort of female sex workers (FSW) in Dakar, Senegal over an 18-year period indicated that an A3-specific sequence in the C2-V3 region of the env gene was found in 46 HIV-1-infected women. HIV-1 sub-subtype A3 appeared in the FSW population as early as 1988 and continued to be transmitted as of 2001. We also found that HIV-1 A3 is not confined to the FSW cohort in Senegal but is also circulating in the general population in Dakar. Furthermore, analyses of viral sequences from a few other West and Central African countries also demonstrated evidence of HIV-1 A3 sequence in isolates from HIV-1-infected people in Ivory Coast, Nigeria, Niger, Guinea Bissau, Benin, and Equatorial Guinea. Overall, because of the evidence of sub-subtype A3 in the general population in Senegal, as well as in a few neighboring West and Central African countries, along with the increasing incidence of infection with A3-containing viruses in the Dakar high-risk FSW population, we feel that HIV-1 sub-subtype A3 viruses are important to distinguish and monitor.

Molecular surveillance has revealed significant heterogeneity in the prevalence and geographic distribution of various subtypes worldwide (5, 35, 36). For example, subtype B viruses account for an estimated 12.3% of cases globally, but infections with this subtype are primarily seen in the Americas, Western Europe, and Australia (35). Conversely, subtype C viruses were estimated to have caused >47% of the worldwide infections, with the highest incidence in southern African countries, Ethiopia, and India (35). As with the nonrecombinant subtypes, circulating recombinant forms (CRFs) also show founder effects and uneven geographic distributions. For example, CRF01_AE appears to be the cause of a high proportion of infections in Southeast Asia and seems to be largely confined to that area of the world with relatively little spread into other populations (35). In contrast, CRF02_AG contributes to a major number of new infections in West Africa and appears to be associated with rapidly spreading epidemics in West African countries (3, 6, 7, 28, 31, 35, 37). The underlying causes of the varied geographic distributions are most likely founder effects with other contributing factors, such as human population movements, unregulated commercial sex work, and intravenous drug use.

Over the course of an 18-year prospective study monitoring a cohort of female sex workers (FSWs) in Dakar, Senegal, we found that the most prevalent circulating strain in the cohort is CRF02_AG, as seen in other parts of West Africa (2, 6, 7, 28, 31, 35, 37). However, recent work conducted in our lab re-

vealed that a unique group of viruses within the human immunodeficiency virus type 1 (HIV-1) subtype A radiation is also present in the FSW study population (40, 42). Through phylogenetic analyses, we have shown that this subcluster of viruses can be classified as HIV-1 sub-subtype A3 (30). The goal of the current investigation was to understand the distribution and dynamics of HIV-1 A3 in the Dakar FSW cohort. We examined the molecular epidemiology of sub-subtype A3 in Senegal, as well as other West and Central African countries. This investigation revealed that there were 46 women in the cohort with the A3 sequence in the C2-V3 region of the env gene. Furthermore, analyses of GenBank-submitted sequences from other West African countries revealed evidence of A3 sequence in isolates from Ivory Coast, Nigeria, Niger, Guinea Bissau, Benin, and Equatorial Guinea. Our findings also indicate that sub-subtype A3 viruses entered the high-risk population of Dakar FSWs as early as 1988, with continued transmission.

MATERIALS AND METHODS

Study population and sample collection. Since 1985, we have conducted a prospective study of registered FSWs in Dakar, Senegal; blood samples and questionnaire data were collected after obtaining informed consent. The specific details of the study recruitment procedures and methods have been described elsewhere (21, 22). Serostatus was determined by immunoblot on whole virus lysates and recombinant envelope peptides and by diagnostic HIV-1 and/or HIV-2 PCR. The time of infection for women who converted to HIV-positive serostatus while in the study was estimated as the midpoint between their last seronegative and first seropositive samples. HIV-1-positive person-years of observation was calculated as described previously (22).

DNA amplification and sequencing. Proviral DNA was extracted from peripheral blood mononuclear cells by using a kit from Qiagen, Inc. (Chatsworth, Calif.). The C2-V3 env region (350 bp) for all of our samples was amplified by nested PCR with primers and reaction conditions that have been described

* Corresponding author. Mailing address: Department of Immunology and Infectious Diseases, Harvard School of Public Health, 651 Huntington Ave., Boston, MA 02115. Phone: (617) 432-1267. Fax: (617) 432-3575. E-mail: pkanki@hsph.harvard.edu.

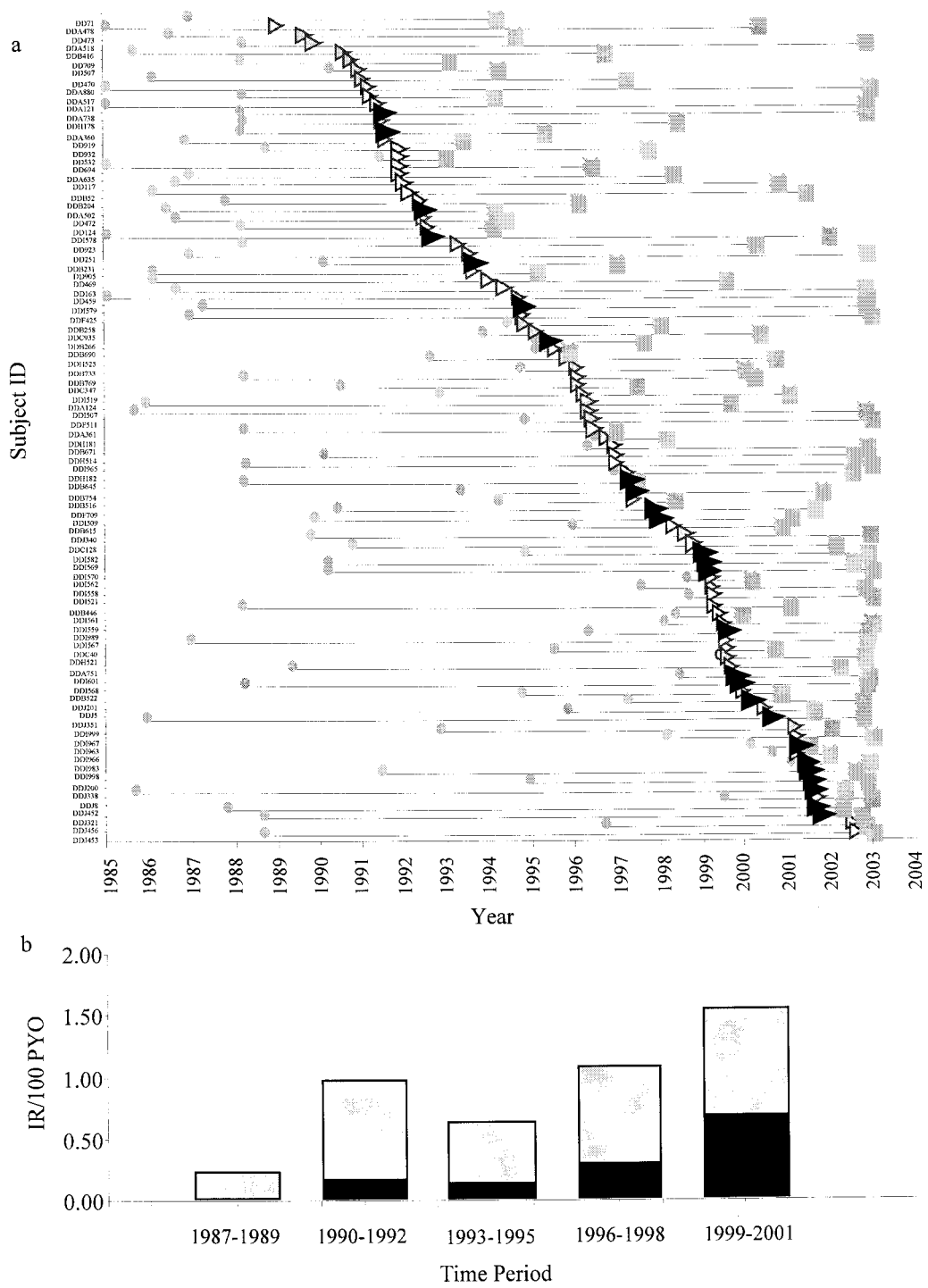


FIG. 1. (a) Distribution of HIV-1 CRF02_AG and A3 infections by year of seroconversion among Senegalese FSWs that were seroincident in the cohort. For each subject, the first bleed date, the time of seroconversion to HIV-1-positive status, and the last bleed date are plotted. The position of the triangles for each subject corresponds to the time of seroconversion. The shading and sizes of the triangles correspond to the HIV-1A sub-subtype or CRF with which the individual was infected: ○, first bleed date; ►, seroconversion date for HIV-1 A3-infected subjects; ▽, seroconversion date for HIV-1 CRF02_AG-infected subjects; □, last bleed date. (b) Annual incidence of HIV-1A *env* sub-subtypes in the Senegalese FSW cohort. Incidence rate per 100 person-years of observation (PYO) of each of the subtype A *env* viruses is plotted in 3-year intervals. The y axis indicates the incidence rate per 100 person-years of observation, and the x axis indicates the time period to which the values correspond. Shading of rectangles corresponds to HIV-1 sub-subtype or CRF with which the individual was infected: ■, A3; □, CRF02_AG.

TABLE 1. Seroincident subjects in Senegalese FSW cohort with A3 sequence in the C2-V3 region (*env*)

Subject	Serohistory	Date (yr):	
		Enrolled in cohort	Diagnosed as HIV-1 ⁺
DDA360	HIV-1 only	1988	1991
DDA738	HIV-1 only	1988	1991
DDI578	HIV-1 only	1985	1992
DDA502	HIV-1 only	1986	1992
DDB231	HIV-1 only	1990	1993
DDI579	HIV-1 only	1987	1994
DDB266	HIV-1 only	1995	1995
DDB645	HIV-1 only	1988	1997
DDI509	HIV-1 only	1989	1997
DDF709	HIV-1 only	1990	1997
DDB754	HIV-1 only	1993	1997
DDI569	HIV-1 only	1990	1998
DDI582	HIV-1 only	1994	1998
DDI568	HIV-1 only	1988	1999
DDI570	HIV-1 only	1990	1999
DDI989	HIV-1 only	1996	1999
DDI601	HIV-1 only	1998	1999
DDJ351	HIV-1 only	1986	2000
DDJ321	HIV-1 only	1988	2000
DDJ201	HIV-1 only	1997	2000
DDJ338	HIV-1 only	1985	2001
DDJ452	HIV-1 only	1987	2001
DDI998	HIV-1 only	1991	2001
DDJ200	HIV-1 only	1995	2001
DDI963	HIV-1 only	2000	2001
DDI983	HIV-1 only	2001	2001
DDA482	HIV-1 and HIV-2	1985	1992
DDB252	HIV-1 and HIV-2	1993	1994

previously (22, 41, 48). The PCR products were purified and directly sequenced by using the second-round primers. In cases in which poor data or highly heterogeneous sequences were obtained from direct sequencing, purified products were cloned into the pCR2.1 vector (T/A Cloning; Invitrogen, San Diego, Calif.). Sequences for both strands of DNA were determined by dye terminator cycle sequencing with *Taq* polymerase (Perkin-Elmer Applied Biosystems Division, Foster City, Calif.) and an automatic sequencer (ABI 373; Perkin-Elmer Applied Biosystems Division).

Sequence analysis and statistical methods. Multiple alignments of all of the consensus sequences to reference sequences from all of the major subtypes and sub-subtypes (A1, A2, A3, B, C, D, F1, F2, G, H, J, and K), as well as CRF02_AG, were performed by using the ClustalX package (45). Manual adjustments, when necessary, were made by using the MacClade version 4.05 program (Sinauer Associates, Inc., Sunderland, Mass.). For reference A3 sequences, we used AY521629 (DDI579), AY521630 (DDJ360), and AY521631 (DDJ369) (30). Phylogenetic analyses were performed, using the SEQBOOT, DNADIST, NEIGHBOR, and CONSENSE programs in the Phylip version 3.6 package (11). Neighbor-joining trees were generated by using the F84 model of substitution and a transition/transversion ratio of 1.42 (24). Alignments were gap stripped prior to analyses. Trees were visualized by using NJPlot (38).

In order to determine whether there was evidence of A3 sequence in any other regions of the world, a BLAST (<http://www.ncbi.nlm.gov/BLAST>) search was conducted with reference A3 sequences. Once a subset of sequences was obtained, more detailed phylogenetic analyses were performed. Neighbor-joining trees, using an F84 model of evolution and a transition/transversion ratio of 1.42 were generated. For these trees, all major subtype, sub-subtype, and CRF02_AG reference sequences were included in the analyses.

Sequence information. The sequences have been deposited into the GenBank database under accession numbers AY646116 to AY646151.

RESULTS

From 1987 through 2001, 3,681 women were enrolled in the Dakar FSW cohort, 491 (13.3%) of whom were or became

TABLE 2. Seroprevalent subjects in Senegalese FSW cohort with A3 sequence in the C2-V3 region (*env*)

Subject	Serohistory	Yr enrolled in cohort
DD170	HIV-1 only	1988
DD737	HIV-1 only	1993
DDB107	HIV-1 only	1993
DDA399	HIV-1 only	1993
DDA467	HIV-1 only	1993
DDC571	HIV-1 only	1995
DDH188	HIV-1 only	1995
DDH189	HIV-1 only	1996
DDH394	HIV-1 only	1998
DDI944	HIV-1 only	1998
DDJ371	HIV-1 only	1998
DDJ426	HIV-1 only	2000
DDJ330	HIV-1 only	2000
DDI979	HIV-1 only	2001
DDB85	HIV-1 and HIV-2	1989
DD935	HIV-1 and HIV-2	1989
DDB501	HIV-1 and HIV-2	1992
DDE303	HIV-1 and HIV-2	1997

infected with HIV-1. Of the HIV-1-infected in the cohort, 175 (35.6%) were seroincident with HIV-1 only and 253 (51.5%) were seroprevalent with HIV-1 only; the remaining subjects were dually infected with HIV-1 and HIV-2.

Epidemiology of HIV-1 A3 in Senegalese FSW cohort. Of the HIV-1-infected women, including those that were dually infected, we identified 301 (61%) women whose HIV-1 sequences were subtype A in the *env*. Of the 301 women, 46 (21%) had sequences that clustered distinctly from CRF02_AG as well as HIV-1 sub-subtypes A1 and A2, as HIV-1 sub-subtype A3 viruses. Overall, 9.4% of the HIV-1 infections in the Dakar FSW were attributed to HIV-1 A3. Of the 46 women whose viral sequences clustered as sub-subtype A3 (30), 26 (56.5%) were seroincident with HIV-1 only, 2 (4.3%) were seroincident and dually infected (Table 1), 14 (30.4%) were seroprevalent and infected with HIV-1 only, and the remaining 4 (8.7%) were seroprevalent and dually infected (Table 2).

A time-lapse graph was generated to examine the appearance and prevalence of A3 in the cohort (Fig. 1a). This graph included all of the seroincident subjects in the Senegalese FSW cohort who had been identified as infected with either A3 or CRF02_AG. For each subject, we plotted first bleed date, seroconversion date, and last bleed date. The data indicated that the first evidence of HIV-1 seroconversion with A3 occurred in 1991. However, when examining study entry dates for seroprevalent subjects (Table 2), we found evidence of A3 infection as early as 1988. Overall, the annual incidence of A3 has steadily increased from 0% in the time period of 1987 through 1989 to an average of 0.67% in the time period of 1999 through 2001 in this FSW population (Fig. 1b).

Kaplan-Meier analyses were conducted to determine whether there was any difference between women in our cohort infected with A3 and those infected with CRF02_AG in terms of progression to AIDS. We did not find a statistically significant difference between the two groups (data not shown). Interestingly, nonparametric univariate analyses indicated that women infected with A3 had a longer time to HIV-1 infection from the date at which they self-reported starting commercial sex work

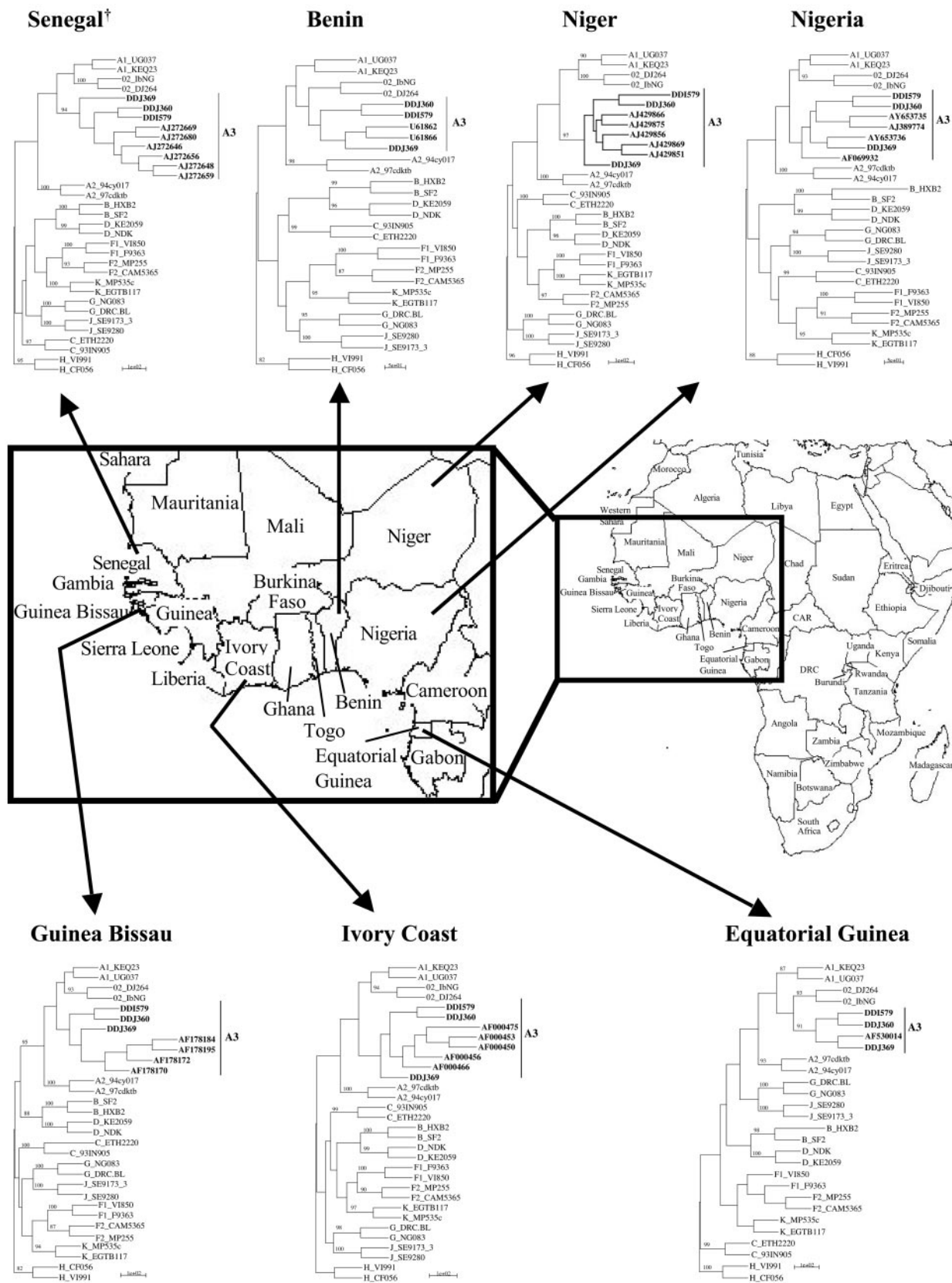


FIG. 2. Evaluation of sequences from studies conducted on other cohorts in Senegal, as well as those in other countries in West Africa. Evidence of sequences clustering with A3 viruses identified in our Senegalese FSW cohort was found in another Senegalese cohort (46), as well as in the Ivory Coast (9), Nigeria (29, 37), Niger (26), Guinea-Bissau (3), Benin (14), and Equatorial Guinea (34). Phylogenetic trees including GenBank-submitted sequences from each of these respective locations are shown. All trees were generated by using an F84 model of evolution, a transition/transversion ratio of 1.42 and references from all published subtypes and sub-subtypes (A1, A2, B-D, F1, F2, G, H, J, and K), as well CRF02_AG. For reference A3 sequences, AY521629 (DDI579), AY521630 (DDJ360), and AY521631 (DDJ369) were used. Only bootstrap values of ≥ 80 are indicated at the major branch points. The A3 sequences are in boldface. Arrows link phylogenetic trees with the appropriate sequences to their corresponding geographic sources.

than did those women infected with CRF02_AG (12.9 ± 9.4 versus 7.4 ± 6.2 , respectively; $P = 0.02$). We also found that, on average, women infected with A3 were older at time of seroconversion than those infected with CRF02_AG (39.7 versus 35.4 , respectively; $P = 0.04$). There was no statistically significant difference between those infected with A3 and those infected with CRF02_AG with respect to age at which sex work began (27.4 versus 27.9 , respectively; $P = 0.67$). There were no statistically significant differences between the two groups with regard to nationality, religion, ethnicity, or level of education.

Evidence of HIV-1 A3 in other West and Central African populations. In order to determine whether A3 was specific to the high-risk population in Dakar, we obtained GenBank-submitted sequences from a study conducted by Toure-Kane et al. in Senegal (46) that looked at sequences from HIV-1-infected people attending a hospital in Dakar, Senegal. The sequences, which had previously been designated "A" (A prime) by Toure-Kane et al. (AJ272646, AJ272648, AJ272656, AJ272659, AJ272669, and AJ272680) (46), clustered with our reference A3 sequences. In addition, we wanted to examine whether there was evidence of spread of A3 to other neighboring African countries (3, 4, 9, 14, 26, 27, 33, 34, 43, 44). We first conducted a BLAST search to identify a subset of GenBank submitted sequences, which might have originally been classified as subtype A, that would potentially cluster with sub-subtype A3 reference sequences. Phylogenetic analyses indicated that a few of the previously characterized sequences from Ivory Coast (AF000450, AF000453, AF000456, AF000466, and AF000475) (9), Equatorial Guinea (AF530014) (34), Niger (AJ429851, AJ429856, AJ429866, AJ429869, and AJ429875) (26), Guinea-Bissau (AF178170, AF178172, AF178184, and AF178195) (3), Nigeria (AF069932, AJ389774, AY653735, and AY653736) (29, 37), and Benin (U61862 and U61866) (14) clustered with sub-subtype A3 viruses identified in the Dakar FSW cohort (Fig. 2). Finally, we also found evidence of A3 in West Brittany, France (AF461904) (47). Although this sequence had previously been characterized as sub-subtype A1, we found that it clustered with our A3 reference sequences, but with a relatively low bootstrap value (data not shown).

DISCUSSION

We have determined that 9.4% of the HIV-1-infected women in the Senegalese FSW cohort are infected sub-subtype A3. We have shown that HIV-1 A3 has been circulating in the Dakar FSW study population from as early as 1988 and that the incidence of HIV-1 infections due to A3 in this cohort is increasing. Our analyses also indicate that there is evidence of HIV-1 A3 in other populations, including those in some neighboring West and Central African countries.

Although we are uncertain as to when A3 was actually introduced into the cohort, the time-lapse graph indicates that the A3 sub-subtype has been newly infecting our population since 1991. Furthermore, our records indicate that a seroprevalent A3-infected subject first entered the cohort in 1988, which dates sub-subtype A3 to the early part of the HIV-1 epidemic in Senegal. Because this first case of A3 in our population was seroprevalent, her date of seroconversion could have been

much earlier, so it is possible that A3 has been present in the population for even longer. Regarding the origin of sub-subtype A3, it is also interesting that we have identified sequences from a few other West and Central African countries that cluster with A3 viruses found in the Senegalese FSW cohort. This finding was not surprising given that Senegal has important trade and travel links with other parts of West Africa (20, 46). It had been suggested that HIV-1 originally entered the Senegalese population through these routes via migrant workers (20). In addition, the evidence of A3 sequence in West Brittany, France, was also initially surprising and was the first indication of this virus in Europe. Vallet et al. (47) suggest that the presence of non-B subtype sequences in northwest France is likely due to either African immigrants in the region or returning fishermen who had traveled to west African countries. Therefore, as seen in West Africa, the transmission of HIV-1 from Africa to northwest France was not unlikely.

Understanding the genetic diversity and molecular epidemiology of the various circulating viral strains is important for a number of reasons. First of all, these studies can help us evaluate the patterns of disease spread and provide us with a tool for tracking the virus. As we have shown, there is evidence of A3 not only in various countries of West and Central Africa but also in northwest France. Second, characterizing the epidemiology of the different viruses assists in studies regarding biological characteristics of these viruses and the biologic consequences of these viral interactions. Although a number of studies have shown associations between viral genotype and disease phenotype (12, 13, 15–19, 22, 25, 32, 39), some cross-sectional studies have not been able to corroborate these findings (1, 10, 23). Several studies have indicated a slower disease progression for CRF02_AG or subtype A-infected individuals relative to other non-A subtypes (17, 18, 22). Furthermore, in a study that teased out the distinctions between subtype A viruses, Sarr et al. (42) uncovered different levels of interaction with HIV-2. These authors found that the *in vivo* interaction between HIV-1 and HIV-2 is influenced by HIV-1 subtype and that the prevalence of A3 viruses (previously referred to as HIV-1 subtype A subcluster 2 viruses) was significantly higher in dually infected individuals compared to women who were singly infected with HIV-1 (42). This suggested that the potential protective effect of HIV-2 would be less against A3 viruses relative to the predominant CRF02_AG. It is possible that future studies that similarly make more accurate subtype and sub-subtype distinctions will better clarify findings regarding the association between viral genotype and biological phenotype. Finally, the further characterization of the predominant HIV-1 subtypes, sub-subtypes, and circulating recombinant forms in a given population will enhance our understanding of viral diversity critical to the informed design of interventions, therapies, and vaccines (8). Although the importance of matching a vaccine candidate to regional circulating strains is yet unclear, incorporation of local strains might maximize the efficacy of a potential vaccine candidate (8).

We have characterized and described a new sub-subtype of HIV-1 termed A3. We have found evidence of sub-subtype A3 in Senegal, as well as in other neighboring West and Central African countries. In addition, we have identified the presence of A3 in northwest France but note that this case is likely traceable to West Africa. We have documented the presence

of A3 in the Senegalese high-risk populations in the late 1980s, with significant spread within that population to the present time. Continued monitoring and future molecular epidemiologic studies will elucidate the role of HIV-1 sub-subtype A3 in the global epidemic. It will also allow the study of viral variation and its impact on the distribution, dynamics, and consequences of virus infection in humans.

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