



Prevalence and Molecular Characterization of Intestinal Trichomonads in Pet Dogs in East China

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Abstract: The trichomonad species *Tritrichomonas foetus* and *Pentatrichomonas hominis* were recently detected in the feces of dogs with diarrhea. However, little information is available on the prevalence and pathogenicity of these parasites in the canine population. Therefore, the aim of this study was to determine the prevalence and molecular characterization of trichomonads infecting pet dogs in Anhui and Zhejiang provinces, east China. In total, 315 pet dogs, with or without diarrhea, from 7 pet hospitals were included in this epidemiological survey. Microscopy and PCR detected *P. hominis* in 19.7% (62/315) and 31.4% (99/315) of fecal samples, respectively. *T. foetus* infection was detected in 0% (0/315) of samples with microscopy and in 0.6% (2/315) with PCR. The prevalence of *P. hominis* was significantly higher in young dogs (≤ 12 months) than in adult dogs (> 12 months), and was significantly higher in diarrheic dogs (50.6%) than in non-diarrheic dogs (24.3%; $P < 0.05$). Infection with *T. foetus* did not correlate with any risk factors evaluated in this study. A sequence analysis of the *P. hominis* PCR products showed minor allelic variations between our sequences and those of *P. hominis* strains from other hosts in different parts of the world. Type CC1 was the most common strain in dogs in east China. The internal transcribed spacer 1 (ITS1)-5.8S rRNA gene sequences from the 2 *T. foetus* isolates detected in this study displayed 100% identity and were homologous to the sequences of other strains isolated from domestic cats in other countries.

Key words: *Pentatrichomonas hominis*, *Tritrichomonas foetus*, pet dog, Anhui, Zhejiang province, China

INTRODUCTION

Trichomonads consist of both pathogenic and presumably non-pathogenic species and are frequently encountered in veterinary medicine [1,2]. These single-celled obligate organisms are characterized morphologically by multiple anterior flagella and a single recurrent flagellum that functions as an undulating membrane [3]. They commonly inhabit warm, moist, anaerobic locations within the gastrointestinal and genitourinary tracts of a variety of vertebrate and invertebrate hosts [2,4,5]. Two trichomonad species found in dogs have received scientific attention, *Tritrichomonas foetus* (family Tritrichomonadidae) and *Pentatrichomonas hominis* (family Trichomonadidae) [1,2,6-10].

T. foetus is mainly known as the causative agent of bovine

trichomoniasis, which can lead to infertility and occasionally abortion [3,11]. A recent study suggested that *T. foetus* is the primary cause of chronic large-bowel diarrhea in domestic cats [12,13]. *T. foetus* has been identified as a synonym of *T. suis*, which is recognized as a facultative pathogen in the large intestine of pigs [14,15].

In contrast, *P. hominis* has frequently been reported in dogs [1,2,6-10] and is presumed to be a commensal organism that may overgrow opportunistically in dogs with diarrhea from other causes [1,2]. However, several authors have described *P. hominis* as the probable causative agent of gastrointestinal disturbances in children [16,17], and this protozoan has also been found occasionally outside its natural habitat, in patients with liver abscesses [18] or empyema thoracis [9,19]. This suggests the zoonotic potential of *P. hominis* and the existence of host-specific genotypes, as found among *T. foetus* isolates from cats and cattle [20,21].

The total number of dogs in China at present is vast, and the number of pet dogs alone approaches 200 million [22]. Pet dogs have played a vital role in human life in most cities and rural regions of China. Unfortunately, few data are available

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on the epidemiology of trichomonads in pets in China. Recently, our laboratory identified *P. hominis* infections in dogs [2] and investigated their prevalence in dogs in northeast China [10]. Given China's vast territory, the occurrence of the parasite may vary greatly in different regions. Until now, no study has investigated *T. foetus* infections in dogs, and it is unknown whether *P. hominis* or *T. foetus* is more common in dogs in China. Therefore, the aims of the present study were (1) to determine the prevalence of trichomonads infecting pet dogs in east China; (2) to evaluate the risk factors for trichomonad infections in dogs; and (3) to investigate the genetic diversity of trichomonad isolates identified in the Chinese pet dog population.

MATERIALS AND METHODS

Sample collection and processing

A total of 315 fecal samples were collected between April and December 2013. The study animals were from 7 pet hospitals distributed in the cities Hefei (1), Xuancheng (1), Chuzhou (2), Bengbu (1), and Suzhou (1) of Anhui province, and in the city Hangzhou (1) of Zhejiang province (Fig. 1). The

data used to evaluate the possible risk factors associated with infection were recorded with a written survey at the time of sample submission, and included the age, sex, source, fecal form classified as consistent (normal) or not consistent (pasty, poorly formed, or liquid), and medications used. Permission was obtained from the dog owners before the fecal samples were collected, and the experimental protocol was approved by the Animal Care and Welfare Committee of Anhui Science and Technology University. All fecal samples were submitted to the laboratory, stored at room temperature, and preliminarily screened within 3 hr with light microscopy. The remaining portions of all the samples were stored at 4°C until DNA extraction.

DNA extraction

Genomic DNA was extracted directly from a 200-mg sample of each preserved fecal specimen using the Stool DNA Kit (Tiangen, Beijing, China), according to the manufacturer's instructions. The DNA samples were stored at -20°C until analysis.

PCR analysis of *P. hominis* and *T. foetus*

All the extracted fecal DNAs were subjected to single-tube

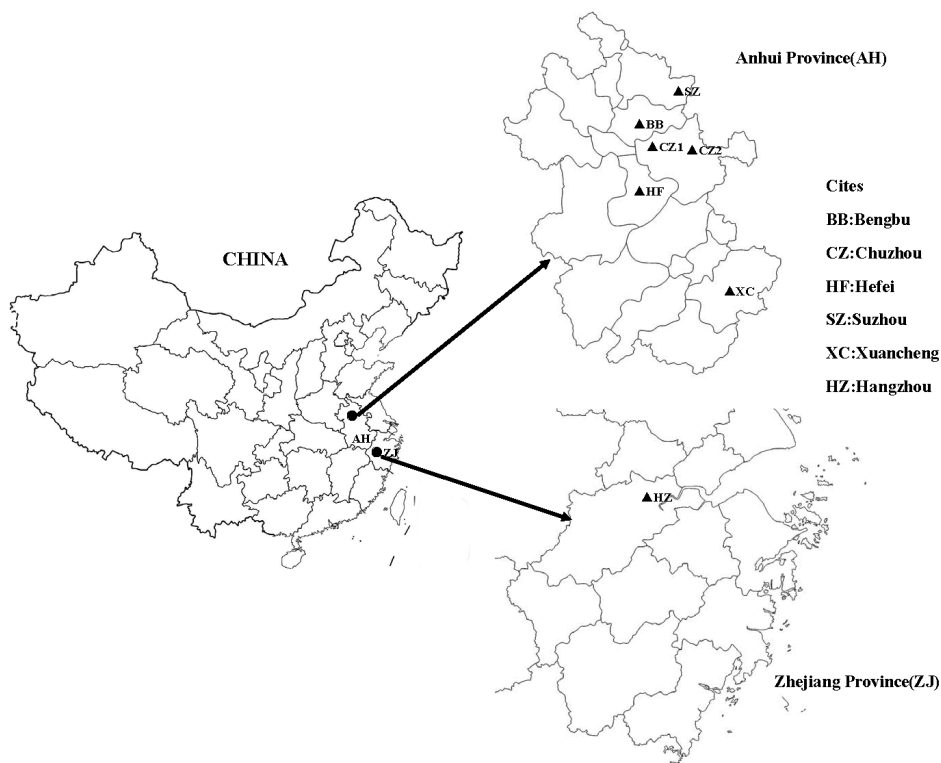


Fig. 1. Specific locations at which specimens were collected in this study. ▲ Study locations.

nested PCR amplification of a 339-bp sequence of the partial 18S rRNA gene of *P. hominis*, using previously published reaction conditions and primer sequences [10]. Each fecal DNA was then subjected to another single-tube nested PCR targeting the ITS1-5.8S rRNA gene of *T. foetus*, as previously described [3,23]. To analyze the genetic diversity of *T. foetus*, the *T. foetus*-positive isolates identified in this study were amplified again using the trichomonad-specific sense primer TRICHO-F and antisense primer TRICHO-R targeting the ITS1-5.8S rRNA-ITS2 region, as previously described [19,24].

DNA sequence analysis

The secondary PCR products were analyzed by electrophoresis on a 1.5% agarose gel and visualized with ethidium bromide staining. The target PCR products were purified with a Biospin Gel Extraction Kit (Bioer, Hangzhou, China). The purified DNA fragments were directly sequenced in both directions on an ABI 377 automated DNA sequencer (Applied Biosystems, Foster City, California, USA) using the primer sets Th3/Th5 [10] or TRICHO-FBIS/TRICHO-RBIS [24]. The sequences were analyzed and aligned with trichomonad reference sequences available in databases, using the BioEdit v 7.1.3.0 soft-

ware (Ibis Biosciences, Carlsbad, California, USA). Because many of the nucleotide sequences determined in this study were identical, only representative sequences have been deposited in GenBank under accession nos. KX136876-KX136894 for *P. hominis* and KX136895-KX136896 for *T. foetus*.

Statistical analysis

To explore the relationships between trichomonad infection and specific risk factors, statistical analyses were performed with a χ^2 test for age and sex and with Fisher's test for clinical symptoms, using SPSS for Windows (release 13.0 standard version, SPSS Inc., Chicago, Illinois, USA). Differences were considered statistically significant at $P < 0.05$.

RESULTS

Prevalence of trichomonad infection

A total of 315 fecal samples from dogs in 7 pet hospitals were screened, and 62 (19.7%) and 99 (31.4%) were positive for *P. hominis* on microscopy and PCR, respectively (Table 1). The highest rates of *P. hominis* infection were 25.8% and 48.4%, on microscopy and PCR, respectively, in the Bengbu

Table 1. Prevalence of *Pentatrichomonas hominis* infection in pet dogs in east China

Location	No. samples	Microscopy		PCR	
		No. positive	Prevalence (%)	No. positive	Prevalence (%)
Hefei	29	5	17.2	10	34.5
Xuancheng	25	6	24.0	9	36.0
Chuzhou 1	17	3	17.6	7	41.2
Chuzhou 2	72	15	20.8	21	29.2
Bengbu	31	8	25.8	15	48.4
Suzhou	41	10	24.4	16	39.0
Hangzhou	100	15	15.0	21	21.0
Total	315	62	19.7	99	31.4

Table 2. Prevalence of *Pentatrichomonas hominis* and *Tritrichomonas foetus* infection in pet dogs by age, gender and clinical symptoms using PCR

Variable		Age (months) ^a		Gender ^b		Clinical symptoms ^c	
		≤ 12	> 12	Male	Female	Diarrheic	Non-diarrheic
<i>P. hominis</i>	No. samples	45	270	150	165	85	230
	No. positive	24	75	46	53	43	56
	Prevalence (%)	53.3 ^d	27.8	30.7	32.1	50.6 ^d	24.3
<i>T. foetus</i>	No. samples	45	270	150	165	85	230
	No. positive	0	2	1	1	1	1
	Prevalence (%)	0	0.7	0.7	0.6	1.8	0.4

^aCompared with pet dogs over > 12 months.

^bCompared with female dogs.

^cCompared with non-diarrheic dogs.

^d $P < 0.05$.

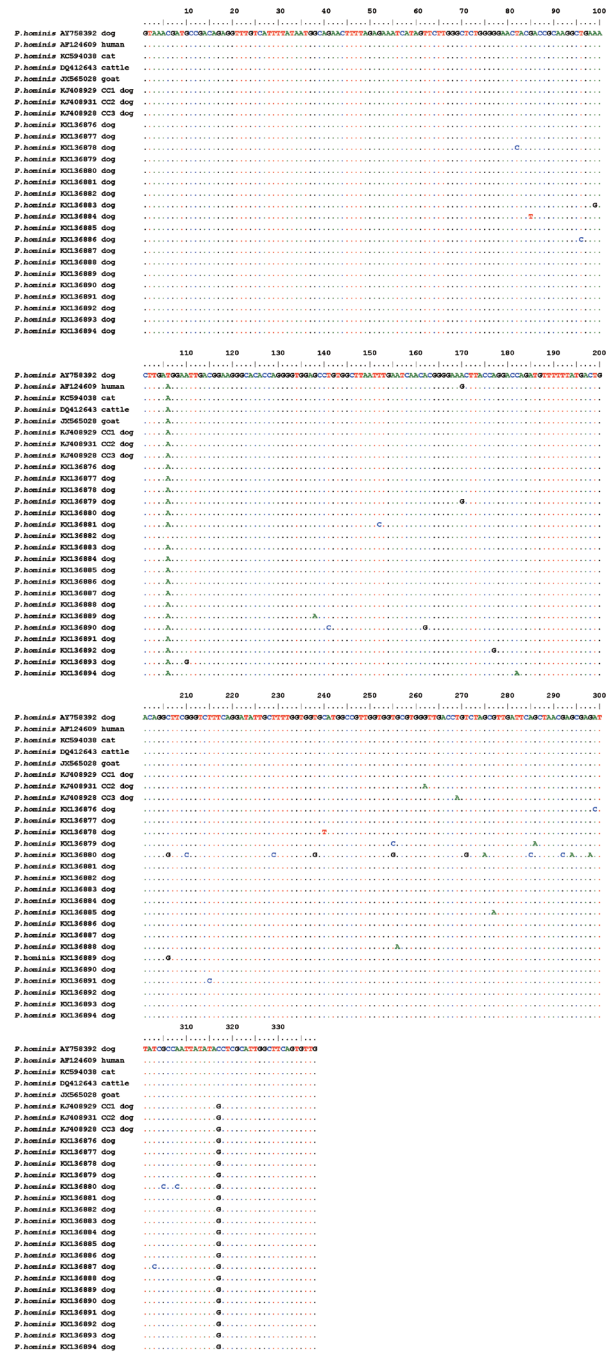


Fig. 2. Alignment of the 18S rRNA sequences of the *P. hominis* isolates analyzed in this study and *P. hominis* strains of interest. Only the nucleotides that differ from those in the reference sequence (no. AY758392) are indicated. Dots represent the consensus sequence of all the *P. hominis* strains used in this figure.

City. Among the samples that were *P. hominis*-positive on microscopy, 10 were not amplified with PCR, probably because either PCR inhibitors were present in the fecal samples, or the

numbers of parasites were low in the fecal samples randomly selected for DNA extraction.

Two samples from 2 pet hospitals (located in the Bengbu City and the Hangzhou City) were positive for *T. foetus* on PCR, with an average infection rate of 0.6% (data not shown). Thus, the prevalence of *T. foetus* in the Bengbu City and the Hangzhou City was 3.2% and 1.00%, respectively. Two samples that were *T. foetus*-positive on PCR were negative on microscopy, and the 2 *T. foetus*-positive samples were both *P. hominis*-negative on microscopy and PCR.

Risk factors associated with infection and clinical signs

The mean age of the screened dogs was 24.0 months, whereas the mean ages of the dogs infected with *P. hominis* and *T. foetus* were 20.0 and 14.0 months, respectively. From our statistical analysis, *P. hominis* infection in pet dogs was significantly associated with 2 factors: clinical symptoms (diarrhea or no diarrhea) and age (≤ 12 or > 12 months). However, the *P. hominis* infection rates in males and females did not differ significantly. Infection with *T. foetus* did not correlate with any risk factors evaluated in this study (Table 2).

Molecular characterization of trichomonad isolates

Nested PCR resulted in specific bands of approximately 339 bp (*P. hominis*) and 363 bp (*T. foetus*). All the trichomonad-positive samples were successfully sequenced. With the exception of isolate KX136880 (isolated from the Bengbu City), an alignment of the amplification products of the 18S rRNA gene demonstrated that the dog *P. hominis* isolates identified in this study and obtained from other hosts, including humans, cats, cattle, goats, and dogs had almost identical sequences (98.8-99.7%), and only differed from the reference sequence (AY 758392) at fewer than 5 variable positions. However, KX136880 showed only 95.6% identity and differed at 15 positions from no. AY758392 (Fig. 2). Interestingly, nucleotide 317 in all the 18S rRNA sequences from dogs in China was G, whereas it was C in the sequences from other hosts in other countries (Fig. 2). Among the 99 *P. hominis*-positive isolates identified in this study, the sequences of 62 (62.6%) from the 7 pet hospitals displayed 100% identity to type CC1 (KJ4 08929). The remaining 18 sequences isolated in this work were all different from the type CC1.

For *T. foetus*, the 2 isolates of *T. foetus* identified in this study showed 100% homology. In the common part of our alignment, these sequences also showed 100% identity to the corre-

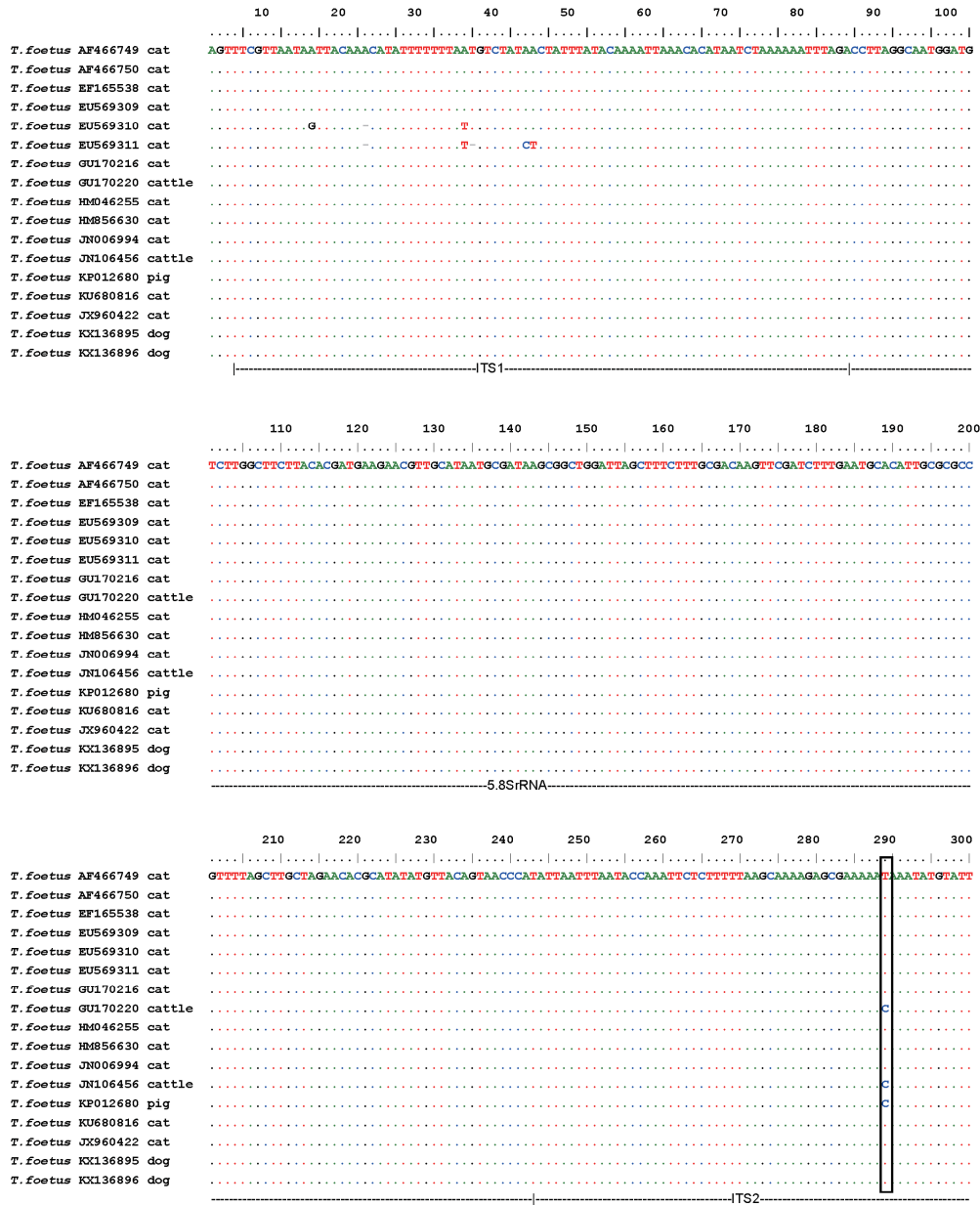


Fig. 3. Alignment of the ITS1-5.8S rRNA-ITS2 sequences of the *T. foetus* isolates analyzed in this study and *T. foetus* strains of interest. Only the nucleotides that differ from those in the consensus sequence used are indicated. Dots represent the consensus sequence of all the *T. foetus* strains used in this figure, and dashes are gaps inserted to optimize the alignment. The black-boxed residues indicate differences in the ITS2 region.

sponding sequences in *T. foetus* strains isolated from domestic cats in the USA (AF466749, AF466750, and EU569309), Switzerland (JN006994), Norway (HM856630 and EF165538), Australia (GU170216 and HM046255), Brazil (KU680816), and France (JX960422). The sequences from the domestic cats and dogs differed only by a single-nucleotide polymorphism (SNP) from those isolated from cattle (JN106456 in China,

and GU170220 in Australia) and pigs (KP012680 in Australia) in the ITS2 region (Fig. 3).

DISCUSSION

Because it is difficult to distinguish different trichomonad species that share similar morphological features using light

microscopy [25,26], molecular tools have been widely used for the identification of trichomonad species and strains. The 18S rRNA [26], ITS1-5.8S rRNA-ITS2 [27,28], and some protein-coding genes, such as glyceraldehyde 3-phosphate dehydrogenase (GAPDH), malate dehydrogenase (MDH), and enolase [29], are commonly described genetic markers. To our knowledge, the present study is the first to report the occurrence of *T. foetus* in pet dogs and confirms that *P. hominis* infection is more frequent than *T. foetus* infection in pet dogs in China. This finding provides objective data to support the previous assumption that trichomoniasis in dogs is most commonly attributable to *P. hominis* infection [1,7,9].

In this study, 99 samples were positive for *P. hominis* and 2 were positive for *T. foetus* when PCR was used, whereas only 62 were positive for *P. hominis* and none for *T. foetus* when microscopy was used because the trophozoites were rapidly destroyed in the feces or impurities in the feces interfered with the PCR reaction. These results clearly show that PCR is more sensitive than microscopy in detecting these parasites.

A study of the prevalence of intestinal trichomonads in breeding kennels in France found that 15.8% (34/215) of the puppies and 20.0% (5/25) of the breeding kennels tested were positive for trichomonads and that *P. hominis* was the only trichomonad infecting the canine population [9]. Our laboratory previously found that the prevalence of *P. hominis* infections in dogs in northeast China was 27.4% (69/252) [10]. In contrast, in this study, we detected a higher rate of *P. hominis* infections (31.4%) in pet dogs in east China than in northeast China.

In the present survey, the prevalence of *P. hominis* was significantly higher in dogs younger than 12 months (53.3%) than in dogs older than 12 months (27.8%; $P < 0.05$). This result is consistent with previous studies by [6,7,9,10], which suggested that puppies were more susceptible to *P. hominis* infection than adult dogs, probably because their immune systems are immature [9]. Previous studies have reported that dogs presenting with *P. hominis* infections are generally younger than 9 months [6,7]. However, it is noteworthy that the mean age of the dogs infected with *P. hominis* was 20.0 months in the present study. The 2 dogs identified with *T. foetus* infections were also older than 12 months (mean age, 14.0 months). A similar observation was made previously [1], which reported that *T. foetus*-infected dogs ranged in age from 10 weeks to 10 years.

Only a handful of case studies or surveys, which have included only a limited number of animals, have reported that *P.*

hominis is more frequent in diarrheic dogs than in non-diarrheic dogs [1,2,6,7]. In our study, *P. hominis* infection was significantly associated with abnormal feces, supporting the assertion that the liquid or semiliquid anaerobic environment created by diarrhea may favor the opportunistic overgrowth of *P. hominis*. This result is consistent with previous reports of *T. foetus* infections in cats [30]. Before the univariate risk factor analysis performed in this study, it was unclear whether the observed diarrhea was directly attributable to *P. hominis* infection or to the presence of other common enteropathogens can also cause clinical diarrhea. The pathogenic potential of *P. hominis* warrants investigation, as does the enteritic coinfection status of dogs infected with *P. hominis* [1,9].

In the present survey, *T. foetus* infection was identified infrequently. This result is quite similar to those found by Tolbert et al. [1] and Grellet et al. [9]. In contrast to *P. hominis* infection, *T. foetus* infection in dogs did not correlate with any of the risk factors evaluated in this study. This may be because the number of cases identified was low or because the number of animals included in the study was limited [31].

Of the 99 18S rRNA gene sequences determined for *P. hominis* isolates from dogs, 62.6% belonged to type CC1, as described previously by our laboratory [10], and no type CC2 (KJ408931) or type CC3 (KJ408928) was detected in the present study. These data suggest that type CC1 is most common in dogs in east China, which is quite similar to the distribution found in northeast China [10]. The remaining 18 sequences identified in this work differed from types CC1, CC2, and CC3, and may be new types of *P. hominis*. The few differences detected between the PCR products indicate that they were all derived from the same strain, and can be attributed to the expected variation within the multiple copies of the 18S rRNA genes in any given genome [9]. The high degree of similarity between the sequences isolated in this study and those of *P. hominis* strains from different hosts implies that they all belong to the same species [9]. Therefore, our data support the conclusion that the same *P. hominis* species colonizes the digestive tracts of several mammal hosts [17] and imply the potential zoonotic transmission of *P. hominis* between its human and animal hosts.

The ITS1-5.8S rRNA gene sequences of *T. foetus* obtained from 315 samples showed 100% identity, strongly suggesting that they were all derived from the same *T. foetus* strain. A comparative analysis with homologous sequences of *T. foetus* isolated from domestic cats in different countries also showed

100% identity. As previously described by [31], the sequences of the dog isolates determined in our study and all the sequences from domestic cat isolates currently available in GenBank show the same SNP in the ITS2 region relative to the *T. foetus* sequences from cattle (T for felids and C for bovids), which could distinguish the “cat genotype” from the “cattle genotype” [20,21]. This result suggests that domestic cats represent an important source of infection for dogs, given the high frequency of contact between dogs and cats in China.

In conclusion, we have shown the occurrence of *T. foetus* in pet dogs for the first time and confirmed that *P. hominis* is a widespread parasite in pet dogs in east China. A comparative analysis of the 18S rRNA sequences from numerous isolates suggested that *P. hominis* has high zoonotic potential. A sequence alignment of the ITS1-5.8S rRNA of *T. foetus* revealed the complete genetic identity between feline isolates and the canine isolates in our study, and confirmed that domestic cats may represent an important source of infection for dogs. In this study, the age and clinical symptoms of pet dogs were risk factors for *P. hominis* infection. However, because other enteropathogens that can cause diarrhea are frequently also present, further experimental studies are required to confirm the pathogenicity of trichomonads in dogs.

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CONFLICT OF INTEREST

The authors declare that they have no competing interests.

REFERENCES

1. Tolbert MK, Leutenegger CM, Lobetti R, Birrell J, Gookin JL. Species identification of trichomonads and associated coinfections in dogs with diarrhea and suspected trichomonosis. *Vet Parasitol* 2012; 187: 319-322.
2. Li WC, Gong PT, Ying M, Li JH, Yang J, Li H, Yang ZT, Zhang GC, Zhang XC. *Pentatrichomonas hominis*: first isolation from the feces of a dog with diarrhea in China. *Parasitol Res* 2014; 113: 1795-1801.
3. Gookin JL, Birkenheuer AJ, St John V, Spector M, Levy MG. Molecular characterization of trichomonads from feces of dogs with diarrhea. *J Parasitol* 2005; 91: 939-943.
4. Felleisen RS. Host-parasite interaction in bovine infection with *Tritrichomonas foetus*. *Microbes Infect* 1999; 1:807-816.
5. Ibanez-Escribano A, Nogal-Ruiz JJ, Delclaux M, Martinez-Nevaldo E, Ponce-Gordo F. Morphological and molecular identification of *Tetratrichomonas* flagellates from the giant anteater (*Myrmecophaga tridactyla*). *Res Vet Sci* 2013; 95: 176-181.
6. Gookin JL, Stauffer SH, Levy MG. Identification of *Pentatrichomonas hominis* in feline fecal samples by polymerase chain reaction assay. *Vet Parasitol* 2007; 145: 11-15.
7. Kim YA, Kim HY, Cho SH, Cheun HI, Yu JR, Lee SE. PCR detection and molecular characterization of *Pentatrichomonas hominis* from feces of dogs with diarrhea in the Republic of Korea. *Korean J Parasitol* 2010; 48: 9-13.
8. Dimasuy KG, Rivera WL. Molecular characterization of trichomonads isolated from animal hosts in the Philippines. *Vet Parasitol* 2013; 196: 289-295.
9. Grellet A, Brunopolack, Feugier A, Boucraut-Baralon C, Grandjean D, Vandewynckel L, Cian A, Meloni D, Viscogliosi E. Prevalence, risk factors of infection and molecular characterization of trichomonads in puppies from French breeding kennels. *Vet Parasitol* 2013; 197: 418-426.
10. Li WC, Ying M, Gong PT, Li JH, Yang J, Li H, Zhang XC. *Pentatrichomonas hominis*: prevalence and molecular characterization in humans, dogs, and monkeys in Northern China. *Parasitol Res* 2016; 115: 569-574.
11. Parsonson IM, Clark BL, Dufty JH. Early pathogenesis and pathology of *Tritrichomonas foetus* infection in virgin heifers. *J Comp Pathol* 1976; 86: 59-66.
12. Frey CF, Schild M, Hemphill A, Stünzi P, Müller N, Gottstein B, Burgener IA. Intestinal *Tritrichomonas foetus* infection in cats in Switzerland detected by in vitro cultivation and PCR. *Parasitol Res* 2009; 104: 783-788.
13. Yao C, Köster LS. *Tritrichomonas foetus* infection, a cause of chronic diarrhea in the domestic cat. *Vet Res* 2015; 46: 35.
14. Lun ZR, Chen XG, Zhu XQ, Li XR, Xie MQ. Are *Tritrichomonas foetus* and *Tritrichomonas suis* synonyms? *Trends Parasitol* 2005; 21: 122-125.
15. Mostegl MM, Richter B, Nedorost N, Maderner A, Dinhopf N, Weissenböck H. Investigations on the prevalence and potential pathogenicity of intestinal trichomonads in pigs using in situ

- hybridization. *Vet Parasitol* 2011; 178: 58-63.
16. Yang CR, Meng ZD, Wang X, Li YL, Zhang YX, Zhao QP. Diarrhoea surveillance in children aged under 5 years in a rural area of Hebei Province, China. *J Diarrhoeal Dis Res* 1990; 8: 155-159.
 17. Meloni D, Mantini C, Goustille J, Desoubeaux G, Maakaroun-Vermesse Z, Chandener J, Gantois N, Duboucher C, Fiori PL, Dei-Cas E, Duong TH, Viscogliosi E. Molecular identification of *Pentatrichomonas hominis* in two patients with gastrointestinal symptoms. *J Clin Pathol* 2011; 64: 933-935.
 18. Jakobsen EB, Friis-Møller A, Friis J. *Trichomonas* species in a sub-hepatic abscess. *Eur J Microbiol* 1987; 6: 296-297.
 19. Jongwutiwes SU, Silachamroon U, Putapornpip C. *Pentatrichomonas hominis* in empyema thoracis. *Trans R Soc Trop Med Hyg* 2000; 94: 185-186.
 20. Slapeta J, Craig S, McDonnell D, Emery D. *Tritrichomonas foetus* from domestic cats and cattle are genetically distinct. *Exp Parasitol* 2010; 126: 209-213.
 21. Reinmann K, Müller N, Kuhnert P, Campero CM, Leitsch D, Hess M, Henning K, Fort M, Müller J, Gottstein B, Frey CF. *Tritrichomonas foetus* isolates from cats and cattle show minor genetic differences in unrelated loci ITS-2 and EF-1 α . *Vet Parasitol* 2012; 185: 138-144.
 22. Chen J, Xu MJ, Zhou DH, Song HQ, Wang CR, Zhu XQ. Canine and feline parasitic zoonoses in China. *Parasit Vectors* 2012; 5: 152.
 23. Gookin JL, Birkenheuer AJ, Breitschwerdt EB, Levy MG. Single-tube nested PCR for detection of *Tritrichomonas foetus* in feline feces. *J Clin Microbiol* 2002; 40: 4126-4130.
 24. Duboucher C, Caby S, Dufenez F, Chabé M, Gantois N, Delgado-Viscogliosi P, Billy C, Barré E, Torabi E, Capron M, Pierce RJ, Dei-Cas E, Viscogliosi E. Molecular identification of *Tritrichomonas foetus*-like organisms as coinfecting agents of human *Pneumocystis pneumonia*. *J Clin Microbiol* 2006; 44: 1165-1168.
 25. Cobo ER, Campero CM, Mariante RM, Benchimol M. Ultra-structural study of a tetratrichomonad species isolated from prepuccial smegma of virgin bulls. *Vet Parasitol* 2003; 117: 195-211.
 26. Walker RL, Hayes DC, Sawyer SJ, Nordhausen RW, Van Hoosear KA, Bon-Durant RH. Comparison of the 5.8 S rRNA gene and internal transcribed spacer regions of trichomonadid protozoa recovered from the bovine preputial cavity. *J Vet Diagn Invest* 2003; 15: 14-20.
 27. Kleina P, Bettim-Bandinelli J, Bonatto SL, Benchimol M, Bogo MR. Molecular phylogeny of Trichomonadidae family inferred from ITS-1, 5.8S rRNA and ITS-2 sequences. *Int J Parasitol* 2004; 34: 963-970.
 28. Grabensteiner E, Bilic I, Kolbe T, Hess M. Molecular analysis of clonal trichomonad isolates indicate the existence of heterogenic species present in different birds and within the same host. *Vet Parasitol* 2010; 172: 53-64.
 29. Malik SB, Brochu CD, Bilic I, Yuan J, Hess M, Logsdon JM Jr, Carlton JM. Phylogeny of parasitic parabasalids and free-living relatives inferred from conventional markers vs. Rpb1, a single-copy gene. *PLoS ONE* 2011; 6, e20774.
 30. Gookin JL, Copple CN, Papich MG, Poore MF, Stauffer SH, Birkenheuer AJ, Twedt DC, Levy MG. Efficacy of ronidazole for treatment of feline *Tritrichomonas foetus* infection. *J Vet Intern Med* 2006; 20: 536-543.
 31. Profizi C, Cian A, Meloni D, Hugonnard M, Lambert V, Groud K, Gagnon AC, Viscogliosi E, Zenner L. Prevalence of *Tritrichomonas foetus* infections in French catteries. *Vet Parasitol* 2013; 196: 50-55.