



# Whole-Genome Sequence and Annotation of *Salmonella enterica* subsp. *enterica* Serovar Enteritidis Phage Type 8 Strain EN1660

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**ABSTRACT** The genome of *Salmonella enterica* subspecies *enterica* serovar Enteritidis phage type 8 strain EN1660, isolated from an outbreak in Thunder Bay, Canada, was sequenced to 46-fold coverage using an Illumina MiSeq with 300-bp paired-end sequencing chemistry to produce 28 contigs with an  $N_{50}$  value of 490,721 bp.

The bacterial pathogen *Salmonella enterica* presents a significant health burden globally and remains a persistent challenge in North America, particularly for the agriculture and food production industries that work diligently to prevent *Salmonella* contamination. Nontyphoidal *Salmonella* is estimated to cost the U.S. economy \$3.7 billion per year due to hospitalizations and premature deaths (1). *Salmonella enterica* subsp. *enterica* serovar Enteritidis is the most common clinical isolate in the United States, yet *Salmonella* Enteritidis genomes are under-represented in DNA sequence databases. Here, we report the whole-genome sequencing of *Salmonella* Enteritidis strain EN1660, a clinical isolate from a 1992 community outbreak in Thunder Bay, Ontario, Canada, associated with consumption of contaminated poultry products. EN1660 was classified as phage type 8 (PT8) by the Ontario Laboratory Centre for Disease Control and was determined to be motile, capable of forming biofilm, and highly invasive in Caco-2 cell culture (2). EN1660 is classified as *Salmonella* sequence type 11 (ST11) by MLST version 1.8 (3).

Genomic DNA isolation was performed using a BioBasic Canada DNA isolation kit. DNA sequencing libraries were prepared using the NEBNext Ultra DNA library prep kit, with a final library size selection of 700 bp. Template sequencing was performed using an Illumina MiSeq and V3 300-bp paired-end sequencing chemistry. Demultiplexed sequencing data were filtered for phiX sequences using Bowtie2 (4) and the *Enterobacteria* phage phiX174 *sensu lato* reference sequence (RefSeq: NC\_001422.1) for comparison. Filtered sequencing reads were then quality trimmed using Trimmomatic (5), removing GC% skews from the beginnings and ends of the reads and trimming at an average Q score below 20 in a sliding window of 5 bp. This resulted in 985,195 high-quality paired-end reads for use in genome assembly.

The genome was assembled using “careful” mode with a  $k$ -mer size of 127 in SPAdes version 1.35 (6). The resulting assembly had a consensus length of 4,701,324 bp spanning 28 contigs, with an  $N_{50}$  value of 490,721 bp and an  $L_{75}$  value of five contigs. The average genome coverage was 46 $\times$  with a GC % of 52.1%. Annotation of the draft genome sequence was conducted using the NCBI Prokaryotic Genome Annotation Pipeline (7). Genome annotation predicted 4,478 coding sequences, eight rRNA operons, 78 tRNAs, 17 ncRNAs, and two CRISPR arrays.

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The EN1660 draft genome was screened for antibiotic resistance genes using the Antibiotic Resistance Gene Database (8), which predicted the presence of four resistance gene homologs: *mdtM*, *pbp2*, *acrB*, and *mdtL*. The draft genome sequence was also analyzed for prophage content using the PHAge Search Tool server (9). PHAST analysis revealed six prophage regions, of which two are intact (similar to Gifsy-2 and RE-2010); one more is potentially fully intact (similar to *Pseudomonas* phage B3); and three are incomplete. We used BLAST to confirm the presence of the broadly distributed *Salmonella* pathogenicity islands SPI-1, SPI-2, SPI-3, SPI-4, SPI-5, SPI-6, SPI-9, SPI-11, SPI-12, SPI-13, SPI-14, and SPI-16 in strain EN1660.

**Accession number(s).** The draft whole-genome sequence for *Salmonella enterica* subsp. *enterica* serovar Enteritidis EN1660 was deposited into the NCBI GenBank database under the accession number [LUUA00000000](https://www.ncbi.nlm.nih.gov/nuccore/LUUA00000000).

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## REFERENCES

1. Economic Research Service (ERS), U.S. Department of Agriculture (USDA). 7 October 2014. Cost estimates of foodborne illnesses. <http://ers.usda.gov/data-products/cost-estimates-of-foodborne-illnesses.aspx>.
2. Shah DH, Zhou X, Addwebi T, Davis MA, Call DR. 2011. *In vitro* and *in vivo* pathogenicity of *Salmonella enteritidis* clinical strains isolated from North America. *Arch Microbiol* 193:811–821. <https://doi.org/10.1007/s00203-011-0719-4>.
3. Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total genome sequenced bacteria. *J Clin Microbiol* 50:1355–1361. <https://doi.org/10.1128/JCM.06094-11>.
4. Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. *Nat Methods* 9:357–359. <https://doi.org/10.1038/nmeth.1923>.
5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Pribelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
7. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
8. Liu B, Pop M. 2009. ARDB—Antibiotic Resistance Genes Database. *Nucleic Acids Res* 37:D443–D447. <https://doi.org/10.1093/nar/gkn656>.
9. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <https://doi.org/10.1093/nar/gkr485>.