

HHS Public Access

J Thromb Haemost. Author manuscript; available in PMC 2018 January 01.

Published in final edited form as:

J Thromb Haemost. 2017 January ; 15(1): 150–154. doi:10.1111/jth.13541.

Plasminogen-receptor $_{\kappa\tau}$: Plasminogen activation and beyond

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Author manuscript

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Abstract

The cell surface orchestrates plasminogen activation through the concomitant binding of plasminogen and plasminogen activators to specific receptors. In this issue, Miles and colleagues describe their detailed phenotypic characterization of mice deficient in Plg-R_{KT}, a key plasminogen receptor expressed in numerous tissues, but highly expressed by proinflammatory macrophages. The analysis provides critical and surprising new insights into the biology of this receptor.

The plasminogen activation system is a versatile proteolytic system with essential functions in thrombolysis, extravascular fibrin surveillance, suppression of fibrin-associated inflammation, tissue remodeling, tissue regeneration, and more. In addition to these physiological functions, deregulation of the plasminogen activation system is linked to the genesis, progression, or morbidity of a wide variety of important human diseases, including bacterial infection, cancer, neurodegenerative disorders, fibrosis, muscular dystrophy, and rheumatoid arthritis [1-31].

Plasmin, the key effector of most functions of the plasminogen activation system, is a multidomain trypsin-like serine protease consisting of a pan-apple domain, five kringle domains, and a serine protease domain. It is formed by proteolytic conversion of the catalyticallyinactive protease zymogen, plasminogen, by an endoproteolytic cleavage within the activation site of the serine protease domain. Plasminogen is predominantly synthesized by the liver and is present in remarkably high concentrations $(1-2 \ \mu M)$ in plasma and in other extravascular fluids [32, 33]. Plasminogen is converted to plasmin either by tissue plasminogen activator (tPA) or by urokinase plasminogen activator (uPA), which are two closely related trypsin-like serine proteases that typically are synthesized, activated, and/or released after disruption of tissue homeostasis, leading to spatially and temporally restricted

Addendum

Disclosure of Conflict of Interests

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M. Flick and T. Bugge participated in the writing, revision, and final approval of this work.

The authors state that they have no conflict of interest.

plasmin generation [3]. Once generated, plasmin is inhibited primarily by the fast-acting and abundant serpin-type protease inhibitor, α2-antiplasmin, [34, 35], while tPA- and uPAmediated activation of plasminogen mainly is inhibited by the serpin-type protease inhibitor, plasminogen activator inhibitor-1 (PAI-1) [36, 37]. Following inhibition by their cognate inhibitors, plasmin and plasminogen activators are internalized by members of the low-density lipoprotein receptor family for lysosomal degradation [38, 39].

Although tPA and uPA both can activate plasminogen in solution, the activation is inefficient, and the newly generated plasmin is susceptible to rapid inhibition by α 2-antiplasmin. Rather, the molecular pathways that mediate the conversion of plasminogen to plasmin under physiological conditions involves the formation of ternary complexes between plasminogen and plasminogen activator, with the fibrin polymer or the surface of cells serving as the two principal sites for plasminogen activation. Fibrin strongly promotes activation of plasminogen by tPA by serving as a scaffold for the binding of tPA and plasminogen in a manner that brings the two molecules in close apposition and simultaneously protects the newly generated plasmin from inactivation by α 2-antiplasmin [40-43]. Both tPA and uPA mediate cell surface plasminogen activation through the binding to specific cellular receptors that may be constitutively expressed, or induced in response to disruption of tissue homeostasis.

Although cell surface binding has long been recognized to be critical to both the conversion of plasminogen to plasmin and for the subsequent physiological functions of plasmin, the identification and validation of specific cell surface receptors for plasminogen has proved to be a remarkably complex task and the subject of extensive and long-standing investigation (reviewed in [44]). Two factors seem to have contributed to this: The first is the peculiar ability of plasminogen to bind to proteins containing a C-terminal lysine residue via its kringle domains [45]. That this mode of binding is indeed employed by plasminogen during its activation on the cell surface was revealed in early studies, showing that treatment of cells with carboxypeptidase B, which removes C-terminal lysines from proteins, largely abolished the potentiation of plasminogen activation by cells [46]. However, it follows that a large number of cell surface-exposed proteins with C-terminal lysine residues will be amenable to plasminogen binding, although this binding may not be productive in terms of stimulating plasminogen activation or affording protection from α 2-antiplasmin. The second factor is the unusually high concentration of plasminogen $(1-2 \mu M)$ in plasma and interstitial fluids, which means that even cell surface proteins with relatively low affinity for plasminogen must be considered candidate receptors for productive plasminogen activation.

The list of candidate receptors for plasminogen reported thus far is exhaustive, and includes the membrane-associated proteins S100A10 (in complex with annexin A2 within the annexin A2 heterotetramer) [47] and Plg-R_{KT} (see below), as well as, surprisingly, proteins with a normally intracellular location and function including cytoplasmic proteins (α enolase [46], cytokeratin 8 [48], actin [49]) and nuclear proteins (TIP49a [50] and histone H2B [51]). A subset of integrins have also been identified as plasminogen receptors, including $\alpha_V\beta_3$, $\alpha_M\beta_2$, and $\alpha_{IIb}\beta_3$. The role of these integrins as plasminogen receptors is notable in that these receptors do not engage plasminogen through a C-terminal lysine and do not significantly enhance plasminogen activation (reviewed in [44]).

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Originally identified from a membrane proteome screen of differentiated mouse macrophages, the 147- amino acid Plg- R_{KT} is unique compared to previously described plasminogen receptors, as it is present exclusively on the cell surface, being synthesized as an integral membrane protein that supports plasminogen binding through a C-terminal lysine. Plg- R_{KT} is highly conserved across mammalian species with homologs also in *Xenopus, Drosophila*, and zebrafish, and, importantly, all of the mammalian orthologs of Plg- R_{KT} contain a C-terminal lysine residue [52]. Like many of the proposed plasminogen receptors, Plg- R_{KT} is broadly expressed in mammalian tissues and notably in many hematopoietic-derived cells. Plg- R_{KT} also significantly enhances plasminogen activation by supporting binding of plasminogen activators. tPA binds Plg- R_{KT} through the same Cterminal lysine domain and, thus, can enhance plasmin generation through plasminogen bound to an adjacent Plg- R_{KT} molecule. In addition, Plg- R_{KT} is clustered on the cell surface with uPA when bound to its receptor uPAR. This colocalization of plasminogen activators and plasminogen has been proposed as a key mechanism by which Plg- R_{KT} regulates cell surface associated plasmin generation [44, 52, 53].

The broad expression pattern of plasminogen receptors in general, and Plg-R_{KT} in particular, coincides with the numerous physiologic and pathophysiologic processes in which cell-surface associated plasmin generation has been proposed to participate (*i.e.*, thrombus resolution, inflammation, bacterial infection, wound healing, neuronal function, tumor progression, metastasis, muscle injury and repair, and bone homeostasis). However, the numerous identified candidates, as well as the overlapping cellular expression pattern of these proteins, complicate defining the precise roles of specific plasminogen receptors in various processes. In order to address this concept and provide a valuable new tool for *in vivo* analysis of the PA system, Miles and colleagues recently generated Plg-R_{KT} knockout (Plg-R_{KT}^{-/-}) mice through a standard homologous recombination strategy in mouse embryonic stem cells (see pages XXX of this issue of JTH). The baseline characterization of these animals supports key roles for plasmin(ogen) that are linked to Plg-R_{KT} binding and plasmin functions that are independent of Plg-R_{KT}. Furthermore, unexpected phenotypes suggest novel functions for Plg-R_{KT} independent of plasminogen itself.

Like plasminogen knockout (Plg^{-/-}) mice, Plg-R_{KT}^{-/-} mice are viable and fertile. Although Plg-R_{KT}^{-/-} female mice can carry a litter to term, they are incapable of supporting even an initial litter of neonates to weaning. This failure of pup survival was linked to a severe lactation defect where milk production in Plg-R_{KT}^{-/-} females was severely decreased within 2 days postpartum. Plg^{-/-} mice also have a documented impairment in lactational competence, however it is markedly less severe. Indeed, many Plg^{-/-} females can support a first litter to weaning, but routinely fail to support a second litter indicating a significant deleterious event following the first mammary gland involution episode. The mammary gland defect observed in Plg^{-/-} mice was linked to persistent fibrin accumulation [31, 54]. The fact that the phenotype in Plg-R_{KT}^{-/-} mice was more severe than that observed for Plg^{-/-} mice suggests a plasminogen-independent mechanism. It is possible that Plg-R_{KT} serves as a receptor for a second, as yet unidentified, protease that functions to modify the ECM during mammary gland development. Alternatively, it is possible that Plg-R_{KT} works in concert with other cell surface or integral membrane proteins to support mammary gland development. To this end, it is notable that mammary epithelial cell deficiencies in either β 1

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integrin or the gap junction protein connexin 43 have similar mammary gland defects with diminished milk production as that described for Plg- $R_{KT}^{-/-}$ mice [55, 56]. The precise cellular and molecular basis of lactation incompetence in Plg- $R_{KT}^{-/-}$ remains to be established.

Perhaps the most striking spontaneous phenotype for Plg^{-/-} mice is a severe wasting disease leading to early mortality experienced by both male and female mice. Survival analysis of Plg- $R_{KT}^{-/-}$ mice indicated survival patterns similar to wildtype mice and body wasting was not observed, at least for male mice. Female Plg- $R_{KT}^{-/-}$ did show a progressive reduction in weight gain over time starting at 4.5 weeks of age, however the mechanistic basis for this gender-specific diminution in growth rate remains undefined. The severe wasting disease and early mortality characteristic of Plg^{-/-} mice is mechanistically linked to multi-organ, persistent fibrin accumulation, as superimposing fibrinogen-deficiency on Plg^{-/-} mice rescued both the progressive weight loss and early mortality [15]. A multi-organ histological survey revealed no evidence of extravascular fibrin deposits in Plg-R_{KT}^{-/-} mice, and Plg- $R_{KT}^{-/-}$ mice did not display other related phenotypes typically observed in Plg^{-/-} mice (*e.g.*, rectal prolapse, ligneous conjunctivitis) [5, 14-16]. Interestingly, mice deficient in either annexin A2 or S100A10 have increased microvascular fibrin deposition in multiple organs suggesting that, unlike Plg-R_{KT}, the annexin A2-S100A10 plasminogen receptor plays a key role in baseline plasmin-mediated fibrin surveillance and clearance [57]. Annexin A2-S100A10-deficient mice also display a compromised ability to clear arterial thrombi following injury [58, 59]. Whether or not Plg-RKT plays any role in thrombus clearance, be it the clearance of arterial thrombi or venous thrombi, remains to be established. However, Plg- $R_{KT}^{-/-}$ mice provide an ideal tool for addressing these very questions in an *in vivo* model system.

The initial identification of Plg-RKT from macrophages implicated the receptor in macrophage function and inflammation. Indeed, $Plg-R_{KT}^{-/-}$ mice were shown to have an ~80% reduction in macrophage recruitment to the peritoneal cavity of mice using the thioglycollate model. This finding was consistent with previous studies showing that systemic administration of an antibody against Plg-R_{KT} reduced macrophage trafficking to the peritoneal cavity following thioglycollate injection by 49% [53]. Modifying inflammation and macrophage activity appears to be a point of commonality for many of the plasminogen receptors, as multiple plasminogen receptors, including annexin A2-S100A10, enolase-1, histone 2B, TATA-binding protein interacting protein, $\alpha_M\beta_2$, and Plg-R_{KT}, are expressed on the cell surface of monocytoid cells. Accordingly, targeting other individual plasminogen receptors similarly reduced macrophage migration in the mouse thioglycollate model. Antibodies directed against histone 2B and enolase-1 significantly reduced macrophage accumulation by 48% and 24%, respectively [60]. Additionally, $S100A10^{-/-}$ mice display 53% less macrophage recruitment following thioglycollate stimulation [59]. Each of these findings aligned well with results of thioglycollate challenge in Plg^{-/-} mice. A 65% reduction in macrophage recruitment following thioglycollate challenge in Plg^{-/-} mice compared to wildtype mice was observed [61]. That elimination or blockade of any one plasminogen receptor on macrophages results in a significant diminution of migration following an identical chemotactic stimulus is intriguing and suggests one of two possibilities: (1) individual plasminogen receptors may be working coordinately as part of a

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complex, such that loss of any one member significantly impacts the functionality of all or (*ii*) individual receptors may support different critical aspects of the migratory process with each step requiring plasminogen. Indeed, a more in depth characterization of the migratory process with respect to the role of plasminogen and receptors, including the impact of combinatorial loss of receptors is warranted.

Beyond simple studies of macrophage migration, the broader role of Plg-RKT and other plasminogen receptors in inflammation and inflammatory disease remains an open question. Inflammatory stimuli increase surface expression of several plasminogen receptors (annexin A2-S100A10, enclase-1, histone 2B, and Plg-RKT) and multiple plasminogen receptors (annexin A2, enolase-1, histone 2B) are targets of autoantibody production in the context of autoimmune diseases [62-65]. Plg- $R_{KT}^{-/-}$ mice do not appear to be particularly susceptible to spontaneous infectious or inflammatory events. Note that a modest increase in dermatitis was reported for Plg- $R_{KT}^{-/-}$ mice but strain C57Bl/6 animals are inherently susceptible to dermatitis [66]. However, perhaps a more interesting open question is to understand the contribution of cell surface-associated plasmin activity and Plg-RKT to specific inflammatory diseases, particularly those where macrophages play a preeminent role in pathogenesis (e.g., microglial cells and neuroinflammatory disease, Kupffer cells and hepatotoxic injury, M1-type adipose tissue macrophages and obesity). Indeed, given that extravascular fibrin deposits are a near universal feature of inflammatory foci, it will be interesting to determine whether plasmin-Plg-RKT plays a role in clearing inflammationassociated pathological fibrin deposits or whether Plg-RKT-restricted plasmin activity functions in inflammation through fibrin-independent mechanisms. Further, the PA system and macrophages each are implicated in tumor progression. Plg- R_{KT} is upregulated in many tumor types and may play a role in this disease. It is also possible that Plg-R_{KT} may function in cancer through a mechanism linked to M2-type tumor promoting macrophages. Such possibilities highlight both the complexity of the PA system in disease and the need for more in-depth analyses.

In conclusion, Plg- $R_{KT}^{-/-}$ mice provide a valuable new reagent for understanding the contribution of cell surface-associated plasminogen activation in physiological and pathological processes. These mice should help provide clarity to the expanding and complicated field of plasminogen receptor biology by better defining roles for the most unique member of this receptor family. Further, these mice have the capacity to expand our knowledge of basic cell biology, as the current data suggests Plg- R_{KT} may serve vital biological roles through mechanisms independent from plasminogen binding and activation. That Plg- $R_{KT}^{-/-}$ mice are viable and largely phenotype-free in the absence of a specific challenge suggests that pharmacological targeting of Plg- R_{KT} in a pathological context would be a viable and attractive novel therapeutic strategy.

Acknowledgments

This research was supported by NHLBI (R01 HL112603) P.I. Matthew J. Flick, and by the Intramural Research Program of the NIH, NIDCR.

References

- Sun H, Ringdahl U, Homeister JW, Fay WP, Engleberg NC, Yang AY, Rozek LS, Wang X, Sjobring U, Ginsburg D. Plasminogen is a critical host pathogenicity factor for group A streptococcal infection. Science. 2004; 305:1283–6. [PubMed: 15333838]
- Coleman JL, Gebbia JA, Piesman J, Degen JL, Bugge TH, Benach JL. Plasminogen is required for efficient dissemination of B. burgdorferi in ticks and for enhancement of spirochetemia in mice. Cell. 1997; 89:1111–9. [PubMed: 9215633]
- Dano K, Andreasen PA, Grondahl-Hansen J, Kristensen P, Nielsen LS, Skriver L. Plasminogen activators, tissue degradation, and cancer. Adv Cancer Res. 1985; 44:139–266. [PubMed: 2930999]
- Dano K, Romer J, Nielsen BS, Bjorn S, Pyke C, Rygaard J, Lund LR. Cancer invasion and tissue remodeling--cooperation of protease systems and cell types. APMIS : acta pathologica, microbiologica, et immunologica Scandinavica. 1999; 107:120–7.
- Drew AF, Kaufman AH, Kombrinck KW, Danton MJ, Daugherty CC, Degen JL, Bugge TH. Ligneous conjunctivitis in plasminogen-deficient mice. Blood. 1998; 91:1616–24. [PubMed: 9473227]
- 6. Schott D, Dempfle CE, Beck P, Liermann A, Mohr-Pennert A, Goldner M, Mehlem P, Azuma H, Schuster V, Mingers AM, Schwarz HP, Kramer MD. Therapy with a purified plasminogen concentrate in an infant with ligneous conjunctivitis and homozygous plasminogen deficiency. The New England journal of medicine. 1998; 339:1679–86. [PubMed: 9834305]
- 7. Hidayat AA, Riddle PJ. Ligneous conjunctivitis. A clinicopathologic study of 17 cases. Ophthalmology. 1987; 94:949–59. [PubMed: 3658371]
- Tefs K, Gueorguieva M, Klammt J, Allen CM, Aktas D, Anlar FY, Aydogdu SD, Brown D, Ciftci E, Contarini P, Dempfle CE, Dostalek M, Eisert S, Gokbuget A, Gunhan O, Hidayat AA, Hugle B, Isikoglu M, Irkec M, Joss SK, et al. Molecular and clinical spectrum of type I plasminogen deficiency: A series of 50 patients. Blood. 2006; 108:3021–6. [PubMed: 16849641]
- Schuster V, Mingers AM, Seidenspinner S, Nussgens Z, Pukrop T, Kreth HW. Homozygous mutations in the plasminogen gene of two unrelated girls with ligneous conjunctivitis. Blood. 1997; 90:958–66. [PubMed: 9242524]
- Pantanowitz L, Bauer K, Tefs K, Schuster V, Balogh K, Pilch BZ, Adcock D, Cirovic C, Kocher O. Ligneous (pseudomembranous) inflammation involving the female genital tract associated with type-1 plasminogen deficiency. Int J Gynecol Pathol. 2004; 23:292–5. [PubMed: 15213608]
- Ciftci E, Ince E, Akar N, Dogru U, Tefs K, Schuster V. Ligneous conjunctivitis, hydrocephalus, hydrocele, and pulmonary involvement in a child with homozygous type I plasminogen deficiency. European journal of pediatrics. 2003; 162:462–5. [PubMed: 12719968]
- 12. Baykul T, Bozkurt Y. Destructive membranous periodontal disease (ligneous periodontitis): a case report and 3 years follow-up. Br Dent J. 2004; 197:467–8. [PubMed: 15547600]
- Watts P, Suresh P, Mezer E, Ells A, Albisetti M, Bajzar L, Marzinotto V, Andrew M, Massicotle P, Rootman D. Effective treatment of ligneous conjunctivitis with topical plasminogen. Am J Ophthalmol. 2002; 133:451–5. [PubMed: 11931777]
- Bugge TH, Flick MJ, Daugherty CC, Degen JL. Plasminogen deficiency causes severe thrombosis but is compatible with development and reproduction. Genes & development. 1995; 9:794–807. [PubMed: 7705657]
- Bugge TH, Kombrinck KW, Flick MJ, Daugherty CC, Danton MJ, Degen JL. Loss of fibrinogen rescues mice from the pleiotropic effects of plasminogen deficiency. Cell. 1996; 87:709–19. [PubMed: 8929539]
- Ploplis VA, Carmeliet P, Vazirzadeh S, Van Vlaenderen I, Moons L, Plow EF, Collen D. Effects of disruption of the plasminogen gene on thrombosis, growth, and health in mice. Circulation. 1995; 92:2585–93. [PubMed: 7586361]
- Cole HA, Ohba T, Nyman JS, Hirotaka H, Cates JM, Flick MJ, Degen JL, Schoenecker JG. Fibrin accumulation secondary to loss of plasmin-mediated fibrinolysis drives inflammatory osteoporosis in mice. Arthritis & rheumatology. 2014; 66:2222–33. 10.1002/art.38639. [PubMed: 24664548]

- Raghu H, Jone A, Cruz C, Rewerts CL, Frederick MD, Thornton S, Degen JL, Flick MJ. Plasminogen is a joint-specific positive or negative determinant of arthritis pathogenesis in mice. Arthritis & rheumatology. 2014; 66:1504–16. 10.1002/art.38402. [PubMed: 24574269]
- de Giorgio-Miller A, Bottoms S, Laurent G, Carmeliet P, Herrick S. Fibrin-induced skin fibrosis in mice deficient in tissue plasminogen activator. The American journal of pathology. 2005; 167:721– 32. [PubMed: 16127152]
- Drew AF, Tucker HL, Liu H, Witte DP, Degen JL, Tipping PG. Crescentic glomerulonephritis is diminished in fibrinogen-deficient mice. American journal of physiology Renal physiology. 2001; 281:F1157–63. [PubMed: 11704568]
- 21. Sachs BD, Baillie GS, McCall JR, Passino MA, Schachtrup C, Wallace DA, Dunlop AJ, MacKenzie KF, Klussmann E, Lynch MJ, Sikorski SL, Nuriel T, Tsigelny I, Zhang J, Houslay MD, Chao MV, Akassoglou K. p75 neurotrophin receptor regulates tissue fibrosis through inhibition of plasminogen activation via a PDE4/cAMP/PKA pathway. The Journal of cell biology. 2007; 177:1119–32. [PubMed: 17576803]
- 22. Schachtrup C, Lu P, Jones LL, Lee JK, Lu J, Sachs BD, Zheng B, Akassoglou K. Fibrinogen inhibits neurite outgrowth via beta 3 integrin-mediated phosphorylation of the EGF receptor. Proc Natl Acad Sci U S A. 2007; 104:11814–9. [PubMed: 17606926]
- 23. Akassoglou K, Adams RA, Bauer J, Mercado P, Tseveleki V, Lassmann H, Probert L, Strickland S. Fibrin depletion decreases inflammation and delays the onset of demyelination in a tumor necrosis factor transgenic mouse model for multiple sclerosis. Proc Natl Acad Sci U S A. 2004; 101:6698–703. [PubMed: 15096619]
- 24. Vidal B, Serrano AL, Tjwa M, Suelves M, Ardite E, De Mori R, Baeza-Raja B, Martinez de Lagran M, Lafuste P, Ruiz-Bonilla V, Jardi M, Gherardi R, Christov C, Dierssen M, Carmeliet P, Degen JL, Dewerchin M, Munoz-Canoves P. Fibrinogen drives dystrophic muscle fibrosis via a TGFbeta/alternative macrophage activation pathway. Genes & development. 2008; 22:1747–52. [PubMed: 18593877]
- 25. Flick MJ, LaJeunesse CM, Talmage KE, Witte DP, Palumbo JS, Pinkerton MD, Thornton S, Degen JL. Fibrin(ogen) exacerbates inflammatory joint disease through a mechanism linked to the integrin alphaMbeta2 binding motif. The Journal of clinical investigation. 2007; 117:3224–35. [PubMed: 17932565]
- 26. Kao WW, Kao CW, Kaufman AH, Kombrinck KW, Converse RL, Good WV, Bugge TH, Degen JL. Healing of corneal epithelial defects in plasminogen- and fibrinogen-deficient mice. Investigative ophthalmology & visual science. 1998; 39:502–8. [PubMed: 9501859]
- Busso N, Peclat V, Van Ness K, Kolodziesczyk E, Degen J, Bugge T, So A. Exacerbation of antigen-induced arthritis in urokinase-deficient mice. The Journal of clinical investigation. 1998; 102:41–50. [PubMed: 9649555]
- Suelves M, Lopez-Alemany R, Lluis F, Aniorte G, Serrano E, Parra M, Carmeliet P, Munoz-Canoves P. Plasmin activity is required for myogenesis in vitro and skeletal muscle regeneration in vivo. Blood. 2002; 99:2835–44. [PubMed: 11929773]
- 29. Akassoglou K, Yu WM, Akpinar P, Strickland S. Fibrin inhibits peripheral nerve remyelination by regulating Schwann cell differentiation. Neuron. 2002; 33:861–75. [PubMed: 11906694]
- Akassoglou K, Kombrinck KW, Degen JL, Strickland S. Tissue plasminogen activator-mediated fibrinolysis protects against axonal degeneration and demyelination after sciatic nerve injury. The Journal of cell biology. 2000; 149:1157–66. [PubMed: 10831618]
- Green KA, Nielsen BS, Castellino FJ, Romer J, Lund LR. Lack of plasminogen leads to milk stasis and premature mammary gland involution during lactation. Developmental biology. 2006; 299:164–75. [PubMed: 16949567]
- Zhang L, Seiffert D, Fowler BJ, Jenkins GR, Thinnes TC, Loskutoff DJ, Parmer RJ, Miles LA. Plasminogen has a broad extrahepatic distribution. Thromb Haemost. 2002; 87:493–501. [PubMed: 11916082]
- Collen D, Lijnen HR. The fibrinolytic system in man. Crit Rev Oncol Hematol. 1986; 4:249–301. [PubMed: 2420482]
- Lijnen HR, Collen D. Protease inhibitors of human plasma. Alpha-2-antiplasmin. J Med. 1985; 16:225–84. [PubMed: 2430039]

- 35. Coughlin PB. Antiplasmin: the forgotten serpin? The FEBS journal. 2005; 272:4852–7. [PubMed: 16176259]
- Loskutoff DJ. A slice of PAI. The Journal of clinical investigation. 1993; 92:2563. [PubMed: 8254010]
- Schneiderman J, Loskutoff DJ. Plasminogen activator inhibitors. Trends in cardiovascular medicine. 1991; 1:99–102. 10.1016/1050-1738(91)90001-U. [PubMed: 21239322]
- Strickland DK, Kounnas MZ, Argraves WS. LDL receptor-related protein: a multiligand receptor for lipoprotein and proteinase catabolism. FASEB journal : official publication of the Federation of American Societies for Experimental Biology. 1995; 9:890–8. [PubMed: 7615159]
- Herz J, Strickland DK. LRP: a multifunctional scavenger and signaling receptor. The Journal of clinical investigation. 2001; 108:779–84. [PubMed: 11560943]
- Thorsen S. The mechanism of plasminogen activation and the variability of the fibrin effector during tissue-type plasminogen activator-mediated fibrinolysis. Annals of the New York Academy of Sciences. 1992; 667:52–63. [PubMed: 1309072]
- Hoylaerts M, Rijken DC, Lijnen HR, Collen D. Kinetics of the activation of plasminogen by human tissue plasminogen activator. Role of fibrin. The Journal of biological chemistry. 1982; 257:2912–9. [PubMed: 7199524]
- Collen D. On the regulation and control of fibrinolysis. Edward Kowalski Memorial Lecture. Thromb Haemost. 1980; 43:77–89. [PubMed: 6450468]
- 43. Collen D, Lijnen HR. Thrombolytic agents. Thromb Haemost. 2005; 93:627–30. [PubMed: 15841305]
- 44. Miles LA, Parmer RJ. Plasminogen receptors: the first quarter century. Seminars in thrombosis and hemostasis. 2013; 39:329–37. 10.1055/s-0033-1334483. [PubMed: 23532575]
- 45. Miles LA, Plow EF. Binding and activation of plasminogen on the platelet surface. The Journal of biological chemistry. 1985; 260:4303–11. [PubMed: 3920216]
- Miles LA, Dahlberg CM, Plescia J, Felez J, Kato K, Plow EF. Role of cell-surface lysines in plasminogen binding to cells: identification of alpha-enolase as a candidate plasminogen receptor. Biochemistry. 1991; 30:1682–91. [PubMed: 1847072]
- 47. Kassam G, Le BH, Choi KS, Kang HM, Fitzpatrick SL, Louie P, Waisman DM. The p11 subunit of the annexin II tetramer plays a key role in the stimulation of t-PA-dependent plasminogen activation. Biochemistry. 1998; 37:16958–66. 10.1021/bi9817131. [PubMed: 9836589]
- Hembrough TA, Kralovich KR, Li L, Gonias SL. Cytokeratin 8 released by breast carcinoma cells in vitro binds plasminogen and tissue-type plasminogen activator and promotes plasminogen activation. The Biochemical journal. 1996; 317(Pt 3):763–9. [PubMed: 8760360]
- Dudani AK, Ganz PR. Endothelial cell surface actin serves as a binding site for plasminogen, tissue plasminogen activator and lipoprotein(a). British journal of haematology. 1996; 95:168–78. [PubMed: 8857956]
- Hawley SB, Tamura T, Miles LA. Purification, cloning, and characterization of a profibrinolytic plasminogen-binding protein, TIP49a. The Journal of biological chemistry. 2001; 276:179–86. 10.1074/jbc.M004919200. [PubMed: 11027681]
- 51. Herren T, Burke TA, Das R, Plow EF. Identification of histone H2B as a regulated plasminogen receptor. Biochemistry. 2006; 45:9463–74. 10.1021/bi060756w. [PubMed: 16878981]
- 52. Andronicos NM, Chen EI, Baik N, Bai H, Parmer CM, Kiosses WB, Kamps MP, Yates JR 3rd, Parmer RJ, Miles LA. Proteomics-based discovery of a novel, structurally unique, and developmentally regulated plasminogen receptor, Plg-RKT, a major regulator of cell surface plasminogen activation. Blood. 2010; 115:1319–30. 10.1182/blood-2008-11-188938. [PubMed: 19897580]
- Lighvani S, Baik N, Diggs JE, Khaldoyanidi S, Parmer RJ, Miles LA. Regulation of macrophage migration by a novel plasminogen receptor Plg-R KT. Blood. 2011; 118:5622–30. 10.1182/ blood-2011-03-344242. [PubMed: 21940822]
- Lund LR, Bjorn SF, Sternlicht MD, Nielsen BS, Solberg H, Usher PA, Osterby R, Christensen IJ, Stephens RW, Bugge TH, Dano K, Werb Z. Lactational competence and involution of the mouse mammary gland require plasminogen. Development. 2000; 127:4481–92. [PubMed: 11003846]

- 55. Naylor MJ, Li N, Cheung J, Lowe ET, Lambert E, Marlow R, Wang P, Schatzmann F, Wintermantel T, Schuetz G, Clarke AR, Mueller U, Hynes NE, Streuli CH. Ablation of beta1 integrin in mammary epithelium reveals a key role for integrin in glandular morphogenesis and differentiation. The Journal of cell biology. 2005; 171:717–28. 10.1083/jcb.200503144. [PubMed: 16301336]
- 56. Stewart MK, Gong XQ, Barr KJ, Bai D, Fishman GI, Laird DW. The severity of mammary gland developmental defects is linked to the overall functional status of Cx43 as revealed by genetically modified mice. The Biochemical journal. 2013; 449:401–13. 10.1042/BJ20121070. [PubMed: 23075222]
- Ling Q, Jacovina AT, Deora A, Febbraio M, Simantov R, Silverstein RL, Hempstead B, Mark WH, Hajjar KA. Annexin II regulates fibrin homeostasis and neoangiogenesis in vivo. The Journal of clinical investigation. 2004; 113:38–48. [PubMed: 14702107]
- Surette AP, Madureira PA, Phipps KD, Miller VA, Svenningsson P, Waisman DM. Regulation of fibrinolysis by S100A10 in vivo. Blood. 2011; 118:3172–81. 10.1182/blood-2011-05-353482. [PubMed: 21768297]
- O'Connell PA, Surette AP, Liwski RS, Svenningsson P, Waisman DM. S100A10 regulates plasminogen-dependent macrophage invasion. Blood. 2010; 116:1136–46. 10.1182/ blood-2010-01-264754. [PubMed: 20424186]
- Das R, Burke T, Plow EF. Histone H2B as a functionally important plasminogen receptor on macrophages. Blood. 2007; 110:3763–72. 10.1182/blood-2007-03-079392. [PubMed: 17690254]
- Ploplis VA, French EL, Carmeliet P, Collen D, Plow EF. Plasminogen deficiency differentially affects recruitment of inflammatory cell populations in mice. Blood. 1998; 91:2005–9. [PubMed: 9490683]
- Cesarman-Maus G, Rios-Luna NP, Deora AB, Huang B, Villa R, Cravioto Mdel C, Alarcon-Segovia D, Sanchez-Guerrero J, Hajjar KA. Autoantibodies against the fibrinolytic receptor, annexin 2, in antiphospholipid syndrome. Blood. 2006; 107:4375–82. 10.1182/ blood-2005-07-2636. [PubMed: 16493010]
- Tomaino B, Cappello P, Capello M, Fredolini C, Sperduti I, Migliorini P, Salacone P, Novarino A, Giacobino A, Ciuffreda L, Alessio M, Nistico P, Scarpa A, Pederzoli P, Zhou W, Petricoin Iii EF, Liotta LA, Giovarelli M, Milella M, Novelli F. Circulating autoantibodies to phosphorylated alphaenolase are a hallmark of pancreatic cancer. Journal of proteome research. 2011; 10:105–12. 10.1021/pr100213b. [PubMed: 20455595]
- 64. Monestier M, Decker P, Briand JP, Gabriel JL, Muller S. Molecular and structural properties of three autoimmune IgG monoclonal antibodies to histone H2B. The Journal of biological chemistry. 2000; 275:13558–63. [PubMed: 10788471]
- Hasegawa M, Sato S, Kikuchi K, Takehara K. Antigen specificity of antihistone antibodies in systemic sclerosis. Annals of the rheumatic diseases. 1998; 57:470–5. [PubMed: 9797552]
- Andrews AG, Dysko RC, Spilman SC, Kunkel RG, Brammer DW, Johnson KJ. Immune complex vasculitis with secondary ulcerative dermatitis in aged C57BL/6NNia mice. Veterinary pathology. 1994; 31:293–300. [PubMed: 8053123]