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# Loss of MPZL3 Function Causes Seborrheic Dermatitis-like Phenotype in Mice

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### **Keywords**

sebaceous gland; barrier; inflammation; alopecia

# Background

Seborrheic Dermatitis (SD) is a common inflammatory skin disorder. It is chronic and relapsing, affecting 1–3% of the general adult population [1]. SD presents as pink to red greasy-looking skin with yellowish scales in seborrheic areas such as the scalp, face (nasolabial folds, upper lip, eyelids and eyebrows), retro-auricular area, and the upper chest. It can be pruritic and socially embarrassing. Current treatment includes antifungals, antiinflammatory, and immune modulators, but treatment in some patients may cause adverse effects such as skin atrophy and telangiectasia if used long term. Sebaceous secretion, yeast Malassezia infection, and individual susceptibilities such as host immunity and epidermal barrier integrity have all been identified as predisposing factors, and may work together to worsen disease. In 2006, Birnbaum et al demonstrated that a frame-shift mutation in ZNF750, a zinc finger transcription factor and master regulator of epidermal differentiation, causes early onset (<10 years of age) autosomal dominant seborrhea-like psoriasiform dermatitis with 100% penetrance (OMIM #610227) [2]. Yet, how ZNF750 dysfunction causes the SD phenotype remains unknown.

We have previously generated knockout mice to determine MPZL3 (Myelin Protein Zerolike 3) function in the skin. Mpzl3 encodes an immunoglobulin protein and is expressed in the suprabasal layers of mouse epidermis, the sebaceous gland, and anagen hair follicles [3,4]. Not surprisingly, Mpzl3 knockout mice showed various skin abnormalities, including

#### Conflict of interest

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V.L. Perez and T.C. Wikramanavake designed the study and experiments, T.C. Wikramanavake, L.J. Borda, Y. Wang, S. Duffort, A. Reyes-Capo, M. Urbieta and A. Barsam performed the experiments and data analysis, T.C. Wikramanayake and R.S. Kirsner wrote the paper and other authors reviewed/revised the paper.

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epidermal and sebaceous hyperplasia and hair loss [3–5]. Interestingly, these mice also developed early onset skin inflammation with dandruff-like flakes [4]. Importantly, ZNF750 was recently shown to directly bind to *MPZL3* promoter and activate its transcription in cultured human keratinocytes [6]. Therefore, *Mpzl3* knockout mice can serve as a useful model to understand SD pathogenesis caused by ZNF750 dysfunction.

#### Questions addressed

In this study, we further investigated the inflammatory skin phenotype of *Mpz13* knockout mice to better understand the pathobiology of SD.

### **Experimental design**

See Supporting Information.

#### Results

To investigate the effects of loss of MPZL3 function in skin inflammation, we examined homozygous *Mpzl3* knockout (–/–) mice and their wild type (+/+) or heterozygous (+/–) littermates. Skin inflammation became apparent in *Mpzl3*<sup>-/–</sup> mice within 3 weeks after birth, with 100% penetrance, when areas of alopecia on the animals' face, eyelid, and upper chest appeared red and swollen (Fig. 1a–k). Histology of affected skin in *Mpzl3*<sup>-/–</sup> mice showed epidermal hyperplasia, hyperkeratosis, infiltrates including neutrophils, and with occasional parakeratosis at the hair canal opening ("shoulder parakeratosis") (Fig. 11, m). Spongiosis typical of SD, especially during the acute phase, was seen only occasionally. Interestingly, spongiosis was not detected in the family with SD and the ZNF750 mutation [2].

Prior to apparent skin inflammation, significant levels of aberrant keratin 6 expression were already detected in the hyperplastic epidermis of  $Mpzl3^{-/-}$  mice 12 days after birth (P12) (Fig. S1). Expression of filaggrin was also increased in  $Mpzl3^{-/-}$  epidermis (Fig. 1p, q). PCNA immunohistochemistry demonstrated increased basal cell proliferation in  $Mpzl3^{-/-}$  epidermis (Fig. 1r, s). We also detected MPZL3 protein expression in the granular layers of human epidermis (Fig. 1u), consistent with previous reports in mouse (Fig. 1t) [4] and human skin [6].

Immunofluorescent staining demonstrated increased T-helper cells (CD4<sup>+</sup>) and macrophages (CD11-b<sup>+</sup>) (Fig. 2a–d), but not cytotoxic T cells (CD8<sup>+</sup>) or B cells (CD19<sup>+</sup>) (Fig. S1g–j), in the dermal infiltrates of  $Mpzl3^{-/-}$  eyelids. To determine the role of adaptive immunity in skin inflammation,  $Mpzl3^{-/-}$  mice were bred with  $Rag2^{-/-}$  mice, which lack mature B and T lymphocytes [7].  $Mpzl3^{-/-}|Rag2^{-/-}$  double knockout mice developed skin inflammation and alopecia similar to that observed in  $Mpzl3^{-/-}$  mice (Fig. 2e–j). These observations suggest that while adaptive immunity participated in skin inflammation (increased CD4<sup>+</sup> cell infiltrates) in the  $Mpzl3^{-/-}$  mice, it was not required for initiation (inflammation developed in the absence of adaptive immunity in the  $Rag2^{-/-}$  background).

Toluidine blue permeability assay detected increased dye permeability of *Mpzl3<sup>-/-</sup>* skin, suggestive of a defective epidermal barrier (Fig. 2k, 1). As expected, gene expression

profiling detected correlated changes in genes related to barrier function and inflammation (Table S1), including small proline-rich protein (Sprr) genes, keratins 6 and 16, inflammatory mediators and antimicrobial peptides (defensins and lipocalin), chemokines (ccls), and inflammatory cytokines and receptors (interleukins and receptors) (Table S1) [S1–S3]. These observations suggest a two-fold consequence of loss of MPZL3 function in the skin: defective barrier and recruitment of immune cells.

#### Conclusions

In this study, we demonstrated seborrheic dermatitis-like clinical features with associated histology in the *Mpzl3* knockout mice. *Mpzl3<sup>-/-</sup>* skin showed increased macrophage and CD4<sup>+</sup> lymphocyte infiltration, however adaptive immunity was not required for the onset of skin inflammation. Furthermore, we detected epidermal barrier defects as suggested by increased dye permeability and altered gene expression in *Mpzl3<sup>-/-</sup>* embryos and pup skin. Taken together, these findings suggest MPZL3 plays an essential role in epidermal barrier and immunity in SD.

Previous studies of SD have focused on *Malassezia* yeast colonization and the immunologic abnormalities [Note, 1, S4–S8]. Our data suggest that epidermal barrier defects and the ensuing innate immune response likely initiated SD-like phenotype in *Mpz13<sup>-/-</sup>* mice. *In toto*, our results suggest a more primary role of barrier maintenance in the pathogenesis of SD.

The SD-like clinical and histologic features in the  $Mpzl3^{-/-}$  mice also resemble those in patients carrying a frame-shift mutation in ZNF750, a key regulator of epidermal differentiation and a transcriptional activator of the MPZL3 gene [2, 6]. Therefore, we conclude that the ZNF750/MPZL3 pathway plays a critical role in the pathogenesis of SD, and a better understanding of skin inflammation and barrier restoration in the  $Mpzl3^{-/-}$  mice will provide insight into the pathogenesis and treatment/prevention of recurrent SD.

# **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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#### Figure 1.

Early onset skin inflammation with features of seborrheic dermatitis in  $Mpz13^{-/-}$  mice (a–s) and MPZL3 protein expression in human epidermis (t). (a–k)  $Mpz13^{-/-}$  mice start to show greasy hair 2 weeks after birth, and develop skin inflammation by 3 weeks of age that persists. They show red, swollen skin on the vibrissae area (block arrows) and margin of the eyelid (arrows), accompanied by hair loss, and dilated blood vessels in the ears (arrowhead). (l, m) H&E staining of  $Mpz13^{-/-}$  skin shows epidermal hyperplasia, hyperkeratosis, parakeratosis at the hair canal opening ("shoulder parakeratosis") (white arrows), infiltrates, and occasional spongiosis. Image in (m) is enlarged from inset in (l). (n–q) Aberrant Keratin 6 expression (n, o) and increased Filaggrin expression (p, q) in hyperplastic  $Mpz13^{-/-}$  epidermis. (r, s) Immunohistochemistry of PCNA expression (brown nuclei, arrows) in Mpz13 +/- and -/- epidermis. (t,u) MPZL3 protein expression in the granular layers

(brackets) of mouse (t) and human (u) epidermis. Signal in mouse stratum corneum (t) is staining artifact (Leiva *et al*, 2014).



#### Figure 2.

Skin inflammation and epidermal barrier defects in the  $Mpzl3^{-/-}$  mice. (a–d) Increased Thelper cell (CD4 positive) (a, b) and macrophage (CD11-b positive) (c, d) infiltrates in the dermis of  $Mpzl3^{-/-}$  eyelids. (e–j) Gross phenotype (e–g) and hematoxylin and eosin staining of skin sections (h–j) showing skin inflammation and alopecia (arrows) in  $Mpzl3^{-/-}|Rag2^{-/-}$ mice (g, j) similar to that of  $Mpzl3^{-/-}$  mice (f, i), compared with wild type mice (e, h). (k, l) Increased toluidine blue dye permeability +/- in E17.0  $Mpzl3^{-/-}$  embryos.