



Review

The TGF- β /Smad4 Signaling Pathway in Pancreatic Carcinogenesis and Its Clinical Significance

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Academic Editor: Andrei Turtoi

Received: 25 October 2016; Accepted: 27 December 2016; Published: 5 January 2017

Abstract: Pancreatic ductal adenocarcinoma (PDAC) is one of the most fatal human cancers due to its complicated genomic instability. PDAC frequently presents at an advanced stage with extensive metastasis, which portends a poor prognosis. The known risk factors associated with PDAC include advanced age, smoking, long-standing chronic pancreatitis, obesity, and diabetes. Its association with genomic and somatic mutations is the most important factor for its aggressiveness. The most common gene mutations associated with PDAC include KRas2, p16, TP53, and Smad4. Among these, Smad4 mutation is relatively specific and its inactivation is found in more than 50% of invasive pancreatic adenocarcinomas. Smad4 is a member of the Smad family of signal transducers and acts as a central mediator of transforming growth factor beta (TGF- β) signaling pathways. The TGF- β signaling pathway promotes many physiological processes, including cell growth, differentiation, proliferation, fibrosis, and scar formation. It also plays a major role in the development of tumors through induction of angiogenesis and immune suppression. In this review, we will discuss the molecular mechanism of TGF- β /Smad4 signaling in the pathogenesis of pancreatic adenocarcinoma and its clinical implication, particularly potential as a prognostic factor and a therapeutic target.

Keywords: TGF- β ; Smad4; pancreatic ductal adenocarcinoma; prognosis; therapy

1. Introduction

In humans, transforming growth factor β (TGF- β) plays an important role in both physiological and pathological processes. TGF- β is a cytokine that resides in the extracellular matrix and is synthesized by macrophages, lymphocytes, fibroblasts, epithelial cells, and platelets [1,2]. Physiologically, it is involved in prenatal and postnatal development, reconstruction, maintenance of normal organ structure, and wound healing [3]. TGF- β suppresses tumor formation by blocking cell cycle progression and maintaining tissue hemostasis. However, the tumor suppressive function is often lost in pancreatic adenocarcinoma by inactivation of the TGF- β signaling mediator, Smad4 [4–6].

Pancreatic ductal adenocarcinoma (PDAC) is one of the most aggressive cancers with a five-year survival of less than 5% due to its high recurrence rate. Surgical resection provides the patient with a chance of a cure, but only 20% of patients present early enough to be surgical candidates [7]. Moreover, in the setting of metastatic metastasis, only 5% of tumors are resectable [7]. Despite multimodality treatment with surgery and adjuvant therapy [8], the five-year survival is only increased to 20%–30% [9], thus early detection is vital in improving survival. Because the TGF- β pathway has an important role in the pancreatic carcinogenesis, many studies have focused not only on the impact of this pathway on the development of pancreatic cancers, but also on its potential clinical use in aiding in clinical management, including early detection, prognostication, and as a therapeutic option.

2. TGF- β Signaling

2.1. Smad-Dependent Pathway

TGF- β , a multifunctional cytokine exists in three different isoforms in mammals (TGF- β 1, TGF- β 2, and TGF- β 3). TGF- β 1 is the most abundant and well-studied isoform. After being stimulated, active TGF- β dimers mediate signaling through TGF- β Type-I and Type-II receptors (T β RI and T β RII), which are serine/threonine kinases. The functional receptor is composed of a heterotetramer of two T β RI and two T β RII molecules [10,11]. Once this complex forms, the T β RII kinase phosphorylates a specific serine residue of T β RI, and this in turn activates the T β RI serine-threonine kinase [12]. Activation of kinase then propagates the signal transduction through phosphorylation of Smad proteins [13]. The Smad proteins include receptor-regulated Smads (R-Smad), the common mediator Smad (co-Smad), and the inhibitory Smad (I-Smad). R-Smads include Smad1, Smad2, Smad3, Smad5, and Smad8. R-Smads act as direct substrates of specific Type-I receptors, while Smad1, Smad5, and Smad8 are targets of bone morphogenic protein (BMP) receptors. Smad2 and Smad3 are substrates of TGF- β receptors and the activin receptor, another member of the TGF- β superfamily [14–16]. Once phosphorylated, R-Smads associate with the common Smad, Smad4, also known as DPC4 (deleted in pancreatic cancer 4), and mediate nuclear translocation of the heterotetrameric complex (Figure 1) [17]. The Smad complex in the nucleus then regulates the expression of different genes, such as integrin, E-cadherin, collagen, and others through interaction with DNA and DNA-binding proteins [18,19]. Immediate activation of inhibitory Smads (Smad6/7) negatively regulate the signaling pathway at several levels such as T β RI degradation, Smad2/Smad3 phosphorylation, and at the level of Smad complex binding with chromatin [20]. This Smad-dependent TGF- β signaling pathway is recognized as tumor-suppressive due to the activation of cell cycle arrest, apoptosis of epithelial cells, and the maintenance of genomic integrity [21]. In pancreatic adenocarcinoma, this tumor suppressive action is often lost by the inactivation of Smad4-dependent TGF- β signaling.

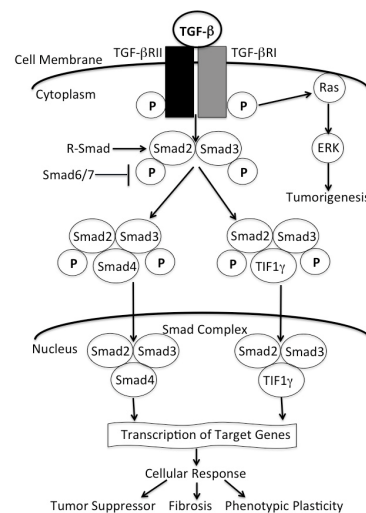


Figure 1. The TGF- β /Smad4 signaling pathway. The Ligand TGF- β binds a complex of transmembrane receptor serine/threonine kinases (Types I and II) in the cell surface and induces transphosphorylation of the receptors. The consequently activated receptors phosphorylate selected Smads at C-terminal serines, and these receptor-activated Smads (R-Smads) then form a complex with a common Smad4. Activated Smad complexes translocate into the nucleus, where they regulate transcription of target genes, through physical interaction and functional cooperation with DNA-binding transcription factors. Besides the Smad4-mediated signaling, Smad2/3 form a complex with Tifly and Smad complexes then translocate into nucleus, thus regulating the transcription of target genes. Activation of R-Smads by Type-I receptor kinases is inhibited by Smad6 or Smad7. Phosphorylated TGF- β receptors also activate Ras and ERK in a Smad-independent manner and induced tumorigenesis.

2.2. Non-Smad-Dependent Pathway

Accumulation of TGF- β and activation of receptors activate the non-Smad pathways either by phosphorylation or by direct activation (i.e., ligand occupied receptors modulate downstream cellular response). It can be activated in parallel to Smads either downstream or upstream of Smad signaling. In Erk MAP kinase pathway, TGF- β activates Erk through activation of Ras [22,23]. Erk activation by TGF- β /ALK5 has also been shown to be mediated by the tyrosine kinase function of ALK5 [24]. Erk activation plays an important role in epithelial mesenchymal transition (EMT), which is one of the major biological functions of TGF- β [25,26]. TGF- β also induces the JNK/p38 MAPK signaling pathway. Active JNK/p38 acts in combination with the Smad pathway to regulate EMT. JNK also regulates the Smad pathway through the regulation of R-Smad, and influences TGF- β induced apoptosis [27]. Rho-like GTPases play an important role in TGF- β mediated cytoskeletal organization, cell motility, and gene expression [28]. RhoA and Rac can be activated by TGF- β via Smad-dependent or Smad-independent pathways to induce EMT. In addition, the P13/Akt pathway activated by increased TGF- β plays a role in fibroblast proliferation and morphological transformation [29].

3. TGF- β /Smad4 Signaling in Pancreatic Cell as a Tumor Suppressor

The TGF- β pathway is crucial in maintaining gastrointestinal homeostasis and contributes to the regulation of gastrointestinal carcinogenesis [30–32]. In the GI tract, TGF- β maintains homeostasis by immune modulation and suppresses tumor formation by keeping a balance between cell renewal, cell differentiation, and loss. Loss of the balance results in tumor promotion [33]. In normal pancreatic cells, TGF- β /Smad4 signaling induces a tumor suppressive effect mediated through Smad4-regulated genes [34]. Alternatively, in tumor cells from some PDAC patients, TGF- β loses its tumor suppressive effect and acts as a tumor promoter [35]. The transformation of function is due to mutations of TGF- β transduction and the loss of Smad4 signaling. TGF- β /Smad4-dependent cell cycle arrest and apoptosis in pancreatic cells is reduced as a result of the down-regulation of Smad4 in the early stage of pancreatic carcinogenesis [36]. TGF- β blocks mitogenic growth signals through Smad4, inhibiting cell growth and proliferation [37,38]. It also induces programmed cell death or apoptosis in pancreatic cell lines by regulation of TIEG, a zinc-finger gene induced by TGF- β /Smad4 signaling and acts as a tumor suppressor [39,40]. Conversely, the loss of Smad4 with a resultant increase in TGF- β aids in tumor progression through the activation of Smad4-independent signaling pathways. In advanced cancer, overexpression of TGF- β activates Ras/Erk, P13K/Akt, p38 MAPK, and Rho-GTPase pathways, which all play a role in tumorigenesis [34,41].

Over 50% of PDAC patients present with a mutation in the TGF- β pathway, with the most common mutation found in Smad4. Smad4 is located in chromosome 18q2, deletion or inactivation of which occurs in the late adenoma-to-carcinoma sequence. Homozygous deletion of Smad4 is found in about 30% of pancreatic cancer patients, inactivation of Smad4 in 20% of patients, and allelic loss of its chromosome is found in 90% [42]. Overall inactivation or loss of Smad4 is found in about 60%–90% of pancreatic adenocarcinomas [43]. Allelic loss of the Smad4 gene results in its mutation and degradation, thus triggering TGF- β independent pathways, which leads to a decrease in TGF- β cell cycle arrest and apoptosis and promotes the epithelial to mesenchymal transition [44]. However, its role in tumor progression and metastasis is complex and still under study. Studies on genetically engineered mouse models of pancreatic cancer with Smad4 mutation as well as Kras allowed a successful analysis on PDAC development and prognosis [45]. Smad4 loss alone did not initiate human pancreatic cancer formation since conditional deletion of Smad4 was not sufficient to induce either pancreatic intraepithelial lesions or invasive cancer, emphasizing the importance of multiple genetic hits [46–48]. Further studies have shown that Smad4 loss markedly promotes tumor development initiated by Kras G12D activation and Kras G12D/Smad4 $-/-$ tumors exhibited both increased proliferation and tumor stromal formation [46,49]. In a pancreatic cancer mouse model, Smad4 $-/-$ tumors metastasized more frequently than Smad4 $+/+$ tumors [48].

The TGF- β signaling pathway plays an important role in cancer progression. It drives progression by immune suppression, by angiogenesis, and by mesenchymal transition, which is an important factor for cancer metastasis [50,51]. EMT is a normal physiological process necessary for embryonic development and transition from epithelial cells to mesenchymal cells with expression of several mesenchymal markers such as vimentin, snail, and *N*-cadherin [24,25,52,53]. In EMT, cells lose polarity and cell-to-cell contact, and acquire enhanced motility and invasiveness. TGF- β is a regulator of this transition process [54]. In pancreatic ductal adenocarcinoma, EMT is an important transition that leads to the progression and metastasis of cancer cells [50]. In the Smad4-dependent pathway, Smad3/mad4 complex induces transcription of snail protein and decreases expression of epithelial junction protein E-cadherin [55]. TGF- β also increases the expression of ZEB transcription factors through Smad4-dependent pathways, thus further increasing the EMT response [56]. In the late stage of tumorigenesis, TGF- β promotes tumor growth by a combined effect of Smad4-dependent and -independent effects on EMT [57,58]. These abovementioned activities emphasize that TGF- β induces pancreatic cancer progression not only by immunosuppression but also, and more importantly, by EMT [51].

Another way of promoting metastasis in cancer is through angiogenesis, and TGF- β acts on endothelial cell proliferation and migration, as well as capillary formation and thereby angiogenesis, promoting vascular metastasis [59]. A major factor for this vascularization is vascular endothelial growth factor (VEGF), which is induced by TGF- β . The major stimulation for this expression is hypoxia, which is a common microenvironment in a growing tumor [59]. TGF- β promotes the secretion of proangiogenic factors, such as matrix-metalloproteinases2 (MMP2) and MMP-9, and downregulates the expression of anti-angiogenic factors such as protease inhibitor TIMP [10,21,60,61] through the recruitment of inflammatory cells in the tumor environment [59]. TGF- β further promotes angiogenesis by inducing connective tissue growth factor (CTGF) in addition to VEGF [10,60,62].

In addition to its effect on tumor epithelial cells, TGF- β modulates tumor development and progression by changing the tumor microenvironment [63]. TGF- β suppresses immune and inflammatory processes through the inhibition of CD⁺ cytotoxic T cells, macrophages, dendritic cells, and NK cells [63]. These changes lead to an elevated TGF- β in 50% of TGF- β mutated PDAC [64]. In PDAC, calcium binding inflammatory proteins, S100A8/A9, are overexpressed by tumor-infiltrating inflammatory cells as well as in PDAC cells [65–67]. Smad4 depletion is associated with reduced S100A8 positive infiltrating inflammatory cells [65,66]; conversely, these proteins are expressed by PDAC cell lines only in areas of Smad4 inactivation [67]. It has recently been found that inflammatory transcription factor NF- κ B activation in PDAC is closely involved in driving tumor progression, especially when its activation is sustained. Thus, Smad4 plays a key role in linking inflammation and cancer [68]. Intact Smad4, S100A8, S100A9, and S100A8/A9 share an overall inhibitory effect on NF- κ B, while these molecules do not affect NF- κ B in the presence of Smad4 homozygous deletion.

Deposition of extracellular matrix (ECM) or desmoplastic reaction is a hallmark of PDAC. ECM is composed of structural proteins such as fibronectin collagen [69,70]. In addition to ECM, endothelial cells, immune cells, and the fibroblast of tumor microenvironments contribute to tumor growth invasion and chemoresistance [71,72]. TGF- β expression enhances the release of multiple ECM molecules including fibronectin, collagen fibulins, and elastin [73,74]. Overexpression of TGF- β is a major factor of fibrosis in many tumors [2,75,76]. In cell culture conditions, TGF- β stimulates proliferation of fibroblast in skin and lung cells and increases collagen synthesis in pancreatic and liver cells [1,70,74]. TGF- β also inhibits degradation of newly synthesized ECM by inhibiting the synthesis of MMP and by inhibiting the expression of genes responsible for the production of MMP [77].

4. Transcriptional Intermediary Factor 1 Gamma (Tif1 γ) in the Regulation of the TGF- β Pathway

Tif1 γ (or Ectodermin/PTC/RFG7/TRIM33) is a transcriptional cofactor competing with Smad2/Smad3 for binding to Smad4, or targeting Smad4 for degradation, although its role in carcinogenesis is unclear. It has a critical role in regulation of the TGF- β signaling pathway and

can have a positive or negative role in this pathway as shown in the literature. It can act as a negative regulator of the pathway by controlling Smad4 function. Smad4 regulation is the biological target of Tif1 γ . It is shown in the mouse pancreatic model that tumor suppressive effects of Tif1 γ could be independent of Smad4 [78]. Overexpression of Tif1 γ can lead to inhibition of the TGF- β signaling pathway [79,80]. Tif1 γ antagonizes transcriptional response of TGF- β by forming a complex with Smad4, also shown in pancreatic cell line [79,81]. Tif1 γ affects the stability of Smad4 by causing its degradation by ubiquitin-proteasome pathway in human and *Xenopus* cells [79]. Tif1 γ also regulates localization of Smad4 (nuclear versus cytoplasmic) and its depletion leads to nuclear localization of Smad4 [79]. Ligr et al. showed inverse relationship between levels of Tif1 γ and Smad4 in the pancreatic cells [82].

A study has demonstrated that Tif1 γ could cause up-regulation of the TGF- β signaling pathway by acting through Smad4-independent pathway and competing with Smad4 to bind with phosphorylated Smad2/3 to transduce signals [83]. Tif1 γ has a significant role in pancreatic carcinogenesis. Recent studies have shown its role as a tumor suppressor in pancreatic cancer. Tif1 γ expression has shown to be decreased in pancreatic ductal adenocarcinomas with the use of RT-PCR and immunohistochemistry [84]. Tif1 γ inactivation in the presence of activated Kras mutation can result in cystic pancreatic tumors in the mouse model [84]. Ligr et al showed imbalanced expression of Tif1 γ has anti-proliferative effect on pancreatic ductal epithelial cells. Tif1 γ overexpression as well as under expression can result in inhibition of pancreatic cell growth by arresting cell cycle.

5. TGF- β /Smad4 as a Prognostic Marker of PDAC

TGF- β level is elevated in the tissue and plasma of PDAC. Plasma levels of TGF- β are correlated with the presence of metastases in CRC, PDAC, and some non-gastrointestinal tumors such as prostate and breast cancers [85]. The association of Smad4 inactivation with poor prognosis may relate to an increased propensity of PDAC to metastasize widely [86]. A recent study conducted on 76 rapidly autopsied patients with PDCA found that Smad4 negative patients had widespread metastasis [87]. Similarly, colorectal cancer with Smad loss was associated with progression to metastasis [88].

A study was done to assess the prognostic significance of the TGF- β level in PDAC patients. TGF- β level was determined in the serum of 146 PDAC patients and 58 patients with benign pancreatic conditions. In healthy controls, serum levels of TGF- β were 57.6 ± 23.2 ng/mL (mean \pm SD); in benign pancreatic conditions, it was 64.5 ± 27.4 ng/mL; in PDAC, it was 237 ± 45.3 ng/mL. In this study, they found that serum levels of TGF- β were significantly higher in PDAC patients than patients with benign pancreatic conditions. They also concluded that high levels of TGF- β were associated with increased tumor size, metastasis (lymphatic and distant), and higher tumor stage [89]. The five-year median overall survival was 21.7% in the low TGF- β group and 15.5% in the high TGF- β group ($p < 0.01$) [89]. The data demonstrate that an elevated level of TGF- β decreases the patient survival rate. Investigators also proved that patients with elevated TGF- β correlated with the risk of death. The correlation between TGF- β and increased invasion of pancreatic cancer was also observed in several studies. Accumulated data support that PDAC patients with high TGF- β levels have an increased risk of metastasis and poor prognosis, and the level of TGF- β can be used as a prognostic marker [89].

Many studies have shown that loss or inactivation of SMAD4 is associated with poor prognosis. In a study of more than 200 patients with PDAC, intact expression of SMAD4 detected by immunohistochemistry was associated with a significantly improved median survival and five-year survival (19.2 months and 20.5 months for intact expression compared to 14.7 months and 13.7 months for loss of expression, respectively). In the multivariate Cox model, SMAD4 status was an independent prognostic factor [90]. A meta-analysis of 4247 patients in 20 published articles concluded that the immunohistochemical loss of SMAD4 predicted a poor overall survival in both Asian and Caucasian patients with pancreatic cancer, but did not correlate with tumor size, differentiation, or lymph node metastasis [91]. Another meta-analysis of 1762 patients from 14 studies found that loss of SMAD4 correlated significantly with poor overall survival. The multivariate analysis showed that the loss

of SMAD4 predicted poor prognosis in patients with less advanced disease (likely Stage I to Stage II pancreatic cancer) [92].

6. TGF- β /Smad4 as a Therapeutic target for PDAC

All of these abovementioned studies have supported a strong association of TGF- β with the development of pancreatic adenocarcinoma (50% of PDAC is due to mutations of TGF- β), metastasis, and prognosis. This would make TGF- β a potential therapeutic target for PDAC [69]. PDAC mouse models have shown that T β RII neutralization could reduce the metastasis and proliferation of cancer cells significantly while increasing apoptosis in the primary tumor [69].

TGF- β signaling through T β RII is a prerequisite pathway for tumor cells. The neutralization of T β RII with a monoclonal antibody 2G8 resulted in a decrease in fibroblast maturation and collagen deposition. It also changes the tumor microenvironment by increasing the epithelial differentiation more than mesenchymal differentiation, thereby reducing metastasis [69]. This strongly suggests that the 2G8 monoclonal antibody has a therapeutic potential for PDAC.

Several studies have demonstrated that TGF- β can mediate responses through a Smad-independent pathway, and that some of these responses are found in conjunction with increased expression of T β R and TGF- β isoforms in pancreatic cancer [93,94]. In a different study on Smad4 deficient PDAC cell lines, PDAC cells displayed constitutive activation of the T β R system as a result of autocrine production and activation of TGF- β [4]. The study demonstrates that PDAC cell lines have escaped the tumor suppressive function and constitutively elevated the level of phosphorylated R-Smads (pSmad4), which is dependent on the rate of T β RI kinase [95,96]. In an in vitro study, the investigators observed that constitutive activation of endogenous TGF- β receptor signaling drives cell migration and invasion in a cell-autonomous manner. When the cell lines were treated with T β RI kinase inhibitor, SD-093, they found significant inhibition of cellular migration and invasiveness, whereas treatment of the same cell lines with exogenous TGF- β further stimulates their invasiveness in vitro [4]. All of these findings highlight a potential of targeting T β RI kinase to treat an aggressive subtype of PDAC.

The therapeutic significance of Smads is unclear. Although inactivation or loss of Smad4 occurs in the majority of pancreatic cancer, targeting Smad4 or other Smads as treatment of PDAC may not be successful due to presence of the Smad-independent TGF- β signaling pathway.

7. Conclusions

Current research has shed light on the biological pathways of TGF- β and its role in carcinogenesis. The TGF- β signaling pathway is involved in tumor suppression and promotion, through the activation of early and late genes, but the complicated mechanism of transition from precancerous cells to cancerous cells is still unknown, particularly in pancreatic carcinogenesis. It is known that TGF- β is a growth factor for gastrointestinal tract and pancreatic cancer, similar to EGFR in non-small cell lung cancer. Elevated levels of TGF- β are found in noncancerous pancreatic tissue and in tumor tissue, and studies have shown that it may serve well as a marker for tumor progression and poor survival in patients with PDAC [97]. There are a myriad of proteins including Smads and Tifl γ that TGF- β interacts with in the signal transduction pathways that promote or inhibit cancer growth. The findings imply that TGF- β and its signaling pathway may be a potential target of therapy and information related to this signaling pathway can be helpful in management of patients with PDAC.

Acknowledgments: The cost for publication is supported in part by the Department of Pathology, NYU School of Medicine/Langone Medical center.

Author Contributions: S.A. and R.U. formulated the original hypothesis, designed the study and wrote the manuscript. M.Z.D designed the figure and wrote the manuscript. A.-D.B. and S.G. wrote the parts of manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Kato, Y.; Inoue, H.; Yoshioka, U.; Fujiyama, Y.; Bamba, T. Effects of transforming growth factor β 1, interleukin-1 β , tumor necrosis factor α and platelet-derived growth factor on the collagen synthesis and the proliferation of periacinar fibroblastoid cells isolated and cultured from rat pancreatic acini. *Pathophysiology* **1999**, *3*, 175–179. [[CrossRef](#)]
2. Krzemiński, S.; Knapczyk, P. Current review on the role of transforming growth factor beta (TGF- β) in some pathological disorders. *Wiad. Lek.* **2005**, *58*, 536–539. [[PubMed](#)]
3. Kajdaniuk, D.; Marek, B.; Borgiel-Marek, H.; Kos-Kudła, B. Transforming growth factor β 1 (TGF β 1) in physiology and pathology. *Endokrynol. Polska* **2013**, *64*, 384–396. [[CrossRef](#)] [[PubMed](#)]
4. Subramanian, G.; Schwarz, R.E.; Higgins, L.; McEnroe, G.; Chakravarty, S.; Dugar, S.; Reiss, M. Targeting endogenous transforming growth factor beta receptor signaling in SMAD4-deficient human pancreatic carcinoma cells inhibits their invasive phenotype. *Cancer Res.* **2004**, *64*, 5200–5211. [[CrossRef](#)] [[PubMed](#)]
5. Kim, J.E.; Lee, K.T.; Lee, J.K.; Paik, S.W.; Rhee, J.C.; Choi, K.W. Clinical usefulness of carbohydrate antigen 19-9 as a screening test for pancreatic cancer in an asymptomatic population. *J. Gastroenterol. Hepatol.* **2004**, *19*, 182–186. [[CrossRef](#)] [[PubMed](#)]
6. Zhao, H.W.; Li, Y.W.; Feng, R.; Yu, J.B.; Li, J.; Zhang, Y.; Li, J.C.; Wang, Y.X. TGF- β /Smad2/3 signal pathway involves in U251 cell proliferation and apoptosis. *Gene* **2015**, *562*, 76–82. [[CrossRef](#)] [[PubMed](#)]
7. Hidalgo, M. Pancreatic Cancer. *N. Engl. J. Med.* **2010**, *362*, 1605–1617. [[CrossRef](#)] [[PubMed](#)]
8. Gong, Z.; Holly, E.A.; Bracci, P.M. Survival in population-based pancreatic cancer patients: San Francisco Bay area, 1995–1999. *Am. J. Epidemiol.* **2011**, *174*, 1373–1381. [[CrossRef](#)] [[PubMed](#)]
9. Sultana, A.; Cox, T.; Ghaneh, P.; Neoptolemos, J.P. Adjuvant therapy for pancreatic cancer. *Recent Results Cancer Res.* **2012**, *196*, 65–88. [[PubMed](#)]
10. Katz, L.H.; Li, Y.; Chen, J.S.; Muñoz, N.M.; Majumdar, A.; Chen, J.; Mishra, L. Targeting TGF- β signaling in cancer. *Expert Opin. Ther. Targets* **2013**, *17*, 743–760. [[CrossRef](#)] [[PubMed](#)]
11. Todorović-Raković, N.; Milovanović, J.; Nikolić-Vukosavljević, D. TGF- β and its coreceptors in cancerogenesis: An overview. *Biomark. Med.* **2011**, *5*, 855–863. [[CrossRef](#)] [[PubMed](#)]
12. López-Casillas, F.; Wrana, J.L.; Massagué, J. Betaglycan presents ligand to the TGF beta signaling receptor. *Cell* **1993**, *73*, 1435–1444. [[CrossRef](#)]
13. Shi, Y.; Massagué, J. Mechanisms of TGF- β signaling from cell membrane to the nucleus. *Cell* **2003**, *113*, 685–700. [[CrossRef](#)]
14. Kretzschmar, M.; Doody, J.; Massagué, J. Opposing BMP and EGF signalling pathways converge on the TGF-beta family mediator Smad1. *Nature* **1997**, *389*, 618–622. [[PubMed](#)]
15. Wrana, J.L.; Attisano, L.; Wieser, R.; Ventura, F.; Massagué, J. Mechanism of activation of the TGF-beta receptor. *Nature* **1994**, *370*, 341–347. [[CrossRef](#)] [[PubMed](#)]
16. Liu, F.; Pouppnot, C.; Massagué, J. Dual role of the Smad4/DPC4 tumor suppressor in TGF β -inducible transcriptional complexes. *Genes Dev.* **1997**, *11*, 3157–3167. [[CrossRef](#)] [[PubMed](#)]
17. Zhang, Y.; Musci, T.; Derynck, R. The tumor suppressor Smad4/DPC 4 as a central mediator of Smad function. *Curr. Biol.* **1997**, *7*, 270–276. [[CrossRef](#)]
18. Chen, Y.; Lebrun, J.J.; Vale, W. Regulation of transforming growth factor β - and activin-induced transcription by mammalian Mad proteins. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 12992–12997. [[CrossRef](#)] [[PubMed](#)]
19. Zhang, Y.; Feng, X.H.; Derynck, R. Smad3 and Smad4 cooperate with c-Jun/c-Fos to mediate TGF-beta-induced transcription. *Nature* **1998**, *394*, 909–913. [[PubMed](#)]
20. Akhurst, R.J.; Hata, A. Targeting the TGF β signalling pathway in disease. *Nat. Rev. Drug Discov.* **2012**, *11*, 790–811. [[CrossRef](#)] [[PubMed](#)]
21. Derynck, R.; Akhurst, R.J.; Balmain, A. TGF- β signaling in tumor suppression and cancer progression. *Nat. Genet.* **2001**, *29*, 117–129. [[CrossRef](#)] [[PubMed](#)]
22. Mulder, K.M.; Morris, S.L. Activation of p21ras by transforming growth factor beta in epithelial cells. *J. Biol. Chem.* **1992**, *267*, 5029–5031. [[PubMed](#)]
23. Yan, Z.; Winawer, S.; Friedman, E. Two different signal transduction pathways can be activated by transforming growth factor-beta-1 in epithelial cells. *J. Biol. Chem.* **1994**, *269*, 13231–13237. [[PubMed](#)]

24. Lee, M.K.; Pardoux, C.; Hall, M.C.; Lee, P.S.; Warburton, D.; Qing, J.; Smith, S.M.; Derynck, R. TGF-beta activates Erk MAP kinase signalling through direct phosphorylation of ShcA. *EMBO J.* **2007**, *26*, 3957–3967. [[CrossRef](#)] [[PubMed](#)]
25. Thiery, J.P. Epithelial-mesenchymal transitions in development and pathologies. *Curr. Opin. Cell Biol.* **2003**, *15*, 740–746. [[CrossRef](#)] [[PubMed](#)]
26. Lee, J.M.; Dedhar, S.; Kalluri, R.; Thompson, E.W. The epithelial-mesenchymal transition: New insights in signaling, development, and disease. *J. Cell Biol.* **2006**, *172*, 973–981. [[CrossRef](#)] [[PubMed](#)]
27. Zhang, Y.E. Non-Smad pathways in TGF-beta signaling. *Cell Res.* **2009**, *19*, 128–139. [[PubMed](#)]
28. Jaffe, A.B.; Hall, A. Rho GTPases: Biochemistry and biology. *Annu. Rev. Cell Dev. Biol.* **2005**, *21*, 247–269. [[CrossRef](#)] [[PubMed](#)]
29. Wilkes, M.C.; Mitchell, H.; Penheiter, S.G.; Doré, J.J.; Suzuki, K.; Edens, M.; Sharma, D.K.; Pagano, R.E.; Leof, E.B. Transforming growth factor-beta activation of phosphatidylinositol 3-kinase is independent of Smad2 and Smad3 and regulates fibroblast responses via p21-activated kinase-2. *Cancer Res.* **2005**, *65*, 10431–10440. [[CrossRef](#)] [[PubMed](#)]
30. Tang, Y.; Katuri, V.; Dillner, A.; Mishra, B.; Deng, C.X.; Mishra, L. Disruption of transforming growth factor-β signaling in ELF β-spectrin-deficient mice. *Science* **2003**, *299*, 574–577. [[CrossRef](#)] [[PubMed](#)]
31. Mishra, L.; Derynck, R.; Mishra, B. Transforming growth factor-β signaling in stem cells and cancer. *Science* **2005**, *310*, 68–71. [[CrossRef](#)] [[PubMed](#)]
32. Tang, Y.; Katuri, V.; Srinivasan, R.; Fogt, F.; Redman, R.; Anand, G.; Said, A.; Fishbein, T.; Zasloff, M.; Reddy, E.P.; et al. Transforming growth factor-beta suppresses nonmetastatic colon cancer through Smad4 and adaptor protein ELF at an early stage of tumorigenesis. *Cancer Res.* **2005**, *65*, 4228–4237. [[CrossRef](#)] [[PubMed](#)]
33. Moses, H.L.; Roberts, A.B.; Derynck, R. The discovery and early days of TGF-β: A historical perspective. *Cold Spring Harb. Perspect. Biol.* **2016**. [[CrossRef](#)] [[PubMed](#)]
34. Pickup, M.; Novitskiy, S.; Moses, H.L. The roles of TGFβ in the tumour microenvironment. *Nat. Rev. Cancer* **2013**, *13*, 788–799. [[CrossRef](#)] [[PubMed](#)]
35. Derynck, R.; Zhang, Y.E. Smad-dependent and Smad-independent pathways in TGF-β family signalling. *Nature* **2003**, *425*, 577–584. [[CrossRef](#)] [[PubMed](#)]
36. Furukawa, T.; Sunamura, M.; Horii, A. Molecular mechanisms of pancreatic carcinogenesis. *Cancer Sci.* **2006**, *97*, 1–7. [[CrossRef](#)] [[PubMed](#)]
37. Duda, D.G.; Sunamura, M.; Lefter, L.P.; Furukawa, T.; Yokoyama, T.; Yatsuoka, T.; Abe, T.; Inoue, H.; Motoi, F.; Egawa, S.; et al. Restoration of SMAD4 by gene therapy reverses the invasive phenotype in pancreatic adenocarcinoma cells. *Oncogene* **2003**, *22*, 6857–6864. [[CrossRef](#)] [[PubMed](#)]
38. Evan, G.I.; Vousden, K.H. Proliferation, cell cycle and apoptosis in cancer. *Nature* **2001**, *411*, 342–348. [[CrossRef](#)] [[PubMed](#)]
39. Lecanda, J.; Ganapathy, V.; D’Aquino-Ardalan, C.; Evans, B.; Cadacio, C.; Ayala, A.; Gold, L.I. TGFβ prevents proteasomal degradation of the cyclin-dependent kinase inhibitor p27kip1 for cell cycle arrest. *Cell Cycle* **2009**, *8*, 742–756. [[CrossRef](#)] [[PubMed](#)]
40. Alvarez, C.; Bass, B.L. Role of transforming growth factor-beta in growth and injury response of the pancreatic duct epithelium in vitro. *J. Gastrointest. Surg.* **1999**, *3*, 178–184. [[CrossRef](#)]
41. Tachibana, I.; Imoto, M.; Adjei, P.N.; Gores, G.J.; Subramaniam, M.; Spelsberg, T.C.; Urrutia, R. Overexpression of the TGFbeta-regulated zinc finger encoding gene, TIEG, induces apoptosis in pancreatic epithelial cells. *J. Clin. Invest.* **1997**, *99*, 2365–2374. [[CrossRef](#)] [[PubMed](#)]
42. Wagner, M.; Kleeff, J.; Lopez, M.E.; Bockman, I.; Massaqué, J.; Korc, M. Transfection of the type I TGF-beta receptor restores TGF-β responsiveness in pancreatic cancer. *Int. J. Cancer* **1998**, *78*, 255–260. [[CrossRef](#)]
43. Moustakas, A.; Heldin, C.H. Non-Smad TGF-beta signals. *J. Cell Sci.* **2005**, *118*, 3573–3584. [[CrossRef](#)] [[PubMed](#)]
44. Lin, X.; Feng, X.H. Abrogation of transforming growth factor-β signaling in pancreatic cancer. *World J. Surg.* **2005**, *29*, 312–316. [[CrossRef](#)] [[PubMed](#)]
45. Xia, X.; Wu, W.; Huang, C.; Cen, G.; Jiang, T.; Cao, J.; Huang, K.; Qiu, Z. SMAD4 and its role in pancreatic cancer. *Tumour Biol.* **2015**, *36*, 111–119. [[CrossRef](#)] [[PubMed](#)]
46. Pérez-Mancera, P.A.; Guerra, C.; Barbacid, M.; Tuveson, D.A. What we have learned about pancreatic cancer from mouse models. *Gastroenterology* **2012**, *142*, 1079–1092. [[CrossRef](#)] [[PubMed](#)]

47. Bardeesy, N.; Cheng, K.H.; Berger, J.H.; Chu, G.C.; Pahler, J.; Olson, P.; Hezel, A.F.; Horner, J.; Lauwers, G.Y.; Hanahan, D.; et al. Smad4 is dispensable for normal pancreas development yet critical in progression and tumor biology of pancreas cancer. *Genes Dev.* **2006**, *20*, 3130–3146. [[CrossRef](#)] [[PubMed](#)]
48. Ijichi, H.; Chytil, A.; Gorska, A.E.; Aakre, M.E.; Fujitani, Y.; Fujitani, S.; Wright, C.V.; Moses, H.L. Aggressive pancreatic ductal adenocarcinoma in mice caused by pancreas-specific blockade of transforming growth factor-beta signaling in cooperation with active Kras expression. *Genes Dev.* **2006**, *20*, 3147–3160. [[CrossRef](#)] [[PubMed](#)]
49. Izeradjene, K.; Combs, C.; Best, M.; Gopinathan, A.; Wagner, A.; Grady, W.M.; Deng, C.X.; Hruban, R.H.; Adsay, N.V.; Tuveson, D.A.; et al. Kras^{G12D} and Smad4/Dpc4 haploinsufficiency cooperate to induce mucinous cystic neoplasms and invasive adenocarcinoma of the pancreas. *Cancer Cell.* **2007**, *11*, 229–243. [[CrossRef](#)] [[PubMed](#)]
50. Leung, L.; Radulovich, N.; Zhu, C.Q.; Wang, D.; To, C.; Ibrahimov, E.; Tsao, M.S. Loss of canonical Smad4 signaling promotes KRAS driven malignant transformation of human pancreatic duct epithelial cells and metastasis. *PLoS ONE* **2013**, *8*, e84366. [[CrossRef](#)] [[PubMed](#)]
51. Krantz, S.B.; Shields, M.A.; Dangi-Garimella, S.; Munshi, H.G.; Bentrem, D.J. Contribution of epithelial-to-mesenchymal transition and cancer stem cells to pancreatic cancer progression. *J. Surg. Res.* **2012**, *173*, 105–112. [[CrossRef](#)] [[PubMed](#)]
52. Ikushima, H.; Miyazono, K. TGFbeta signalling: A complex web in cancer progression. *Nat. Rev. Cancer* **2010**, *10*, 415–424. [[CrossRef](#)] [[PubMed](#)]
53. Thiery, J.P.; Acloque, H.; Huang, R.Y.; Nieto, M.A. Epithelial-mesenchymal transitions in development and disease. *Cell* **2009**, *139*, 871–890. [[CrossRef](#)] [[PubMed](#)]
54. Polyak, K.; Weinberg, R.A. Transitions between epithelial and mesenchymal states: Acquisition of malignant and stem cell traits. *Nat. Rev. Cancer* **2009**, *9*, 265–273. [[CrossRef](#)] [[PubMed](#)]
55. Meulmeester, E.; Ten Dijke, P. The dynamic roles of TGF- β in cancer. *J. Pathol.* **2011**, *223*, 205–218. [[CrossRef](#)] [[PubMed](#)]
56. Vincent, T.; Neve, E.P.; Johnson, J.R.; Kukalev, A.; Rojo, F.; Albanell, J.; Pietras, K.; Virtanen, I.; Philipson, L.; Leopold, P.L.; et al. A SNAIL1-SMAD3/4 transcriptional repressor complex promotes TGF- β mediated epithelial-mesenchymal transition. *Nat. Cell Biol.* **2009**, *11*, 943–950. [[CrossRef](#)] [[PubMed](#)]
57. Bracken, C.P.; Gregory, P.A.; Kolesnikoff, N.; Bert, A.G.; Wang, J.; Shannon, M.F.; Goodall, G.J. A double-negative feedback loop between ZEB1-SIP1 and the microRNA-200 family regulates epithelial-mesenchymal transition. *Cancer Res.* **2008**, *68*, 7846–7854. [[CrossRef](#)] [[PubMed](#)]
58. Derynck, R.; Akhurst, R.J. Differentiation plasticity regulated by TGF- β family proteins in development and disease. *Nat. Cell Biol.* **2007**, *9*, 1000–1004. [[CrossRef](#)] [[PubMed](#)]
59. Davies, M.; Robinson, M.; Smith, E.; Huntley, S.; Prime, S.; Paterson, I. Induction of an epithelial to mesenchymal transition in human immortal and malignant keratinocytes by TGF- β 1 involves MAPK, Smad and AP-1 signalling pathways. *J. Cell. Biochem.* **2005**, *95*, 918–931. [[CrossRef](#)] [[PubMed](#)]
60. Pardali, E.; ten Dijke, P. Transforming growth factor-beta signaling and tumor angiogenesis. *Front. Biosci.* **2009**, *14*, 4848–4861. [[CrossRef](#)]
61. Padua, D.; Massagué, J. Roles of TGF β in metastasis. *Cell Res.* **2009**, *19*, 89–102. [[CrossRef](#)] [[PubMed](#)]
62. Hagedorn, H.G.; Bachmeier, B.E.; Nerlich, A.G. Synthesis and degradation of basement membranes and extracellular matrix and their regulation by TGF- β in invasive carcinomas (Review). *Int. J. Oncol.* **2001**, *18*, 669–681. [[CrossRef](#)] [[PubMed](#)]
63. Sánchez-Elsner, T.; Botella, L.M.; Velasco, B.; Corbí, A.; Attisano, L.; Bernabéu, C. Synergistic cooperation between hypoxia and transforming growth factor-beta pathways on human vascular endothelial growth factor gene expression. *J. Biol. Chem.* **2001**, *276*, 38527–38535. [[CrossRef](#)] [[PubMed](#)]
64. Achyut, B.R.; Yang, L. Transforming growth factor- β in the gastrointestinal and hepatic tumor microenvironment. *Gastroenterology* **2011**, *141*, 1167–1178. [[CrossRef](#)] [[PubMed](#)]
65. Sheikh, A.A.; Vimalachandran, D.; Thompson, C.C.; Jenkins, R.E.; Nedjadi, T.; Shekouh, A.; Campbell, F.; Dodson, A.; Prime, W.; Crnogorac-Jurcevic, T.; et al. The expression of S100A8 in pancreatic cancer-associated monocytes is associated with the Smad4 status of pancreatic cancer cells. *Proteomics* **2007**, *11*, 1929–1940. [[CrossRef](#)] [[PubMed](#)]

66. Ang, C.W.; Nedjadi, T.; Sheikh, A.A.; Tweedle, E.M.; Tonack, S.; Honap, S.; Jenkins, R.E.; Park, B.K.; Schwarte-Waldhoff, I.; Khattak, I.; et al. Smad4 loss is associated with fewer S100A8-positive monocytes in colorectal tumors and attenuated response to S100A8 in colorectal and pancreatic cancer cells. *Carcinogenesis* **2010**, *9*, 1541–1551. [[CrossRef](#)] [[PubMed](#)]
67. Basso, D.; Greco, E.; Padoan, A.; Fogar, P.; Scorzeto, M.; Fadi, E.; Bozzato, D.; Moz, S.; Navaglia, F.; Zambon, C.F.; et al. Altered intracellular calcium fluxes in pancreatic cancer induced diabetes mellitus: Relevance of the S100A8 N-terminal peptide (NT-S100A8). *J. Cell. Physiol.* **2011**, *226*, 456–468. [[CrossRef](#)] [[PubMed](#)]
68. Gurumurthy, S.; Bardeesy, N. Uncapping NF- κ B activity in pancreatic cancer. *EMBO J.* **2011**, *30*, 1–2. [[CrossRef](#)] [[PubMed](#)]
69. Hagopian, M.M.; Brekken, R.A. Stromal TGF β R2 signaling: A gateway to progression for pancreatic cancer. *Mol. Cell. Oncol.* **2014**, *2*, e975606. [[CrossRef](#)] [[PubMed](#)]
70. Danen, E.H.; Yamada, K.M. Fibronectin, integrins, and growth control. *J. Cell. Physiol.* **2001**, *189*, 1–13. [[CrossRef](#)] [[PubMed](#)]
71. Eliceiri, B.P. Integrin and growth factor receptor crosstalk. *Circ. Res.* **2001**, *89*, 1104–1110. [[CrossRef](#)] [[PubMed](#)]
72. Weis, S.M.; Cheresh, D.A. Tumor angiogenesis: Molecular pathways and therapeutic targets. *Nat. Med.* **2011**, *17*, 1359–1370. [[CrossRef](#)] [[PubMed](#)]
73. Mao, Y.; Keller, E.T.; Garfield, D.H.; Shen, K.; Wang, J. Stromal cells in tumor microenvironment and breast cancer. *Cancer Metastasis Rev.* **2013**, *32*, 303–315. [[CrossRef](#)] [[PubMed](#)]
74. Kuang, P.P.; Joyce-Brady, M.; Zhang, X.H.; Jean, J.C.; Goldstein, R.H. Fibulin-5 gene expression in human lung fibroblasts is regulated by TGF- β and phosphatidylinositol 3-kinase activity. *Am. J. Physiol. Cell Physiol.* **2006**, *291*, C1412–C1421. [[CrossRef](#)] [[PubMed](#)]
75. Igotz, R.A.; Massagué, J. Transforming growth factor-beta stimulates the expression of fibronectin and collagen and their incorporation into the extracellular matrix. *J. Biol. Chem.* **1986**, *261*, 4337–4345. [[PubMed](#)]
76. Kajdaniuk, D.; Marek, B.; Borgiel-Marek, H.; Kos-Kudła, B. Vascular endothelial growth factor (VEGF)—Part 1: In physiology and pathophysiology. *Endokrynol. Polska* **2011**, *62*, 444–455.
77. Kajdaniuk, D.; Marek, B.; Foltyn, W.; Kos-Kudła, B. Vascular endothelial growth factor (VEGF)—Part 2: In endocrinology and oncology. *Endokrynol. Polska* **2011**, *62*, 456–464.
78. Kuwahara, F.; Kai, H.; Tokuda, K.; Kai, M.; Takeshita, A.; Egashira, K.; Imaizumi, T. Transforming growth factor-beta function blocking prevents myocardial fibrosis and diastolic dysfunction in pressure-overloaded rats. *Circulation* **2002**, *106*, 130–135. [[CrossRef](#)] [[PubMed](#)]
79. Vincent, D.F.; Gout, J.; Chuvin, N.; Arfi, V.; Pommier, R.M.; Bertolino, P.; Jonckheere, N.; Ripoche, D.; Kaniewski, B.; Martel, S.; et al. Tif1 suppresses murine pancreatic tumoral transformation by a Smad4-independent pathway. *Am. J. Pathol.* **2012**, *180*, 2214–2221. [[CrossRef](#)] [[PubMed](#)]
80. Dupont, S.; Zacchigna, L.; Cordenonsi, M.; Soligo, S.; Adorno, M.; Rugge, M.; Piccolo, S. Germ-layer specification and control of cell growth by Ectodermin, a Smad4 ubiquitin ligase. *Cell* **2005**, *121*, 87–99. [[CrossRef](#)] [[PubMed](#)]
81. Dupont, S.; Mamidi, A.; Cordenonsi, M.; Montagner, M.; Zacchigna, L.; Adorno, M.; Martello, G.; Stinchfield, M.J.; Soligo, S.; Morsut, L.; et al. FAM/USP9x, a deubiquitinating enzyme essential for TGF β signaling, controls Smad4 monoubiquitination. *Cell* **2009**, *136*, 123–135. [[CrossRef](#)] [[PubMed](#)]
82. Ligr, M.; Wu, X.; Daniels, G.; Zhang, D.; Wang, H.; Hajdu, C.; Wang, J.; Pan, R.; Pei, Z.; Zhang, L.; et al. Imbalanced expression of Tif1 γ inhibits pancreatic ductal epithelial cell growth. *Am. J. Cancer Res.* **2014**, *4*, 196–210. [[PubMed](#)]
83. He, W.; Dorn, D.C.; Erdjument-Bromage, H.; Tempst, P.; Moore, M.A.; Massague, J. Hematopoiesis controlled by distinct TIF1 γ and Smad4 branches of the TGF β pathway. *Cell* **2006**, *125*, 929–941. [[CrossRef](#)] [[PubMed](#)]
84. Vincent, D.F.; Yan, K.-P.; Treilleux, I.; Gay, F.; Arfi, V.; Kaniewski, B.; Marie, J.C.; Lepinasse, F.; Martel, S.; Goddard-Leon, S.; et al. Inactivation of TIF1 γ cooperates with Kras to induce cystic tumors of the pancreas. *PLoS Genet.* **2009**, *5*, e1000575. [[CrossRef](#)]
85. Massagué, J. TGF β in Cancer. *Cell* **2008**, *134*, 215–230. [[CrossRef](#)] [[PubMed](#)]

86. Blackford, A.; Serrano, O.K.; Wolfgang, C.L.; Parmigiani, G.; Jones, S.; Zhang, X.; Parsons, D.W.; Lin, J.C.; Leary, R.J.; Eshleman, J.R.; et al. SMAD4 gene mutations are associated with poor prognosis in pancreatic cancer. *Clin. Cancer Res.* **2009**, *15*, 4674–4679. [[CrossRef](#)] [[PubMed](#)]
87. Iacobuzio-Donahue, C.A.; Fu, B.; Yachida, S.; Luo, M.; Abe, H.; Henderson, C.M.; Vilardell, F.; Wang, Z.; Keller, J.W.; Banerjee, P.; et al. DPC4 gene status of the primary carcinoma correlates with patterns of failure in patients with pancreatic cancer. *J. Clin. Oncol.* **2009**, *27*, 1806–1813. [[CrossRef](#)] [[PubMed](#)]
88. Maitra, A.; Molberg, K.; Albores-Saavedra, J.; Lindberg, G. Loss of Dpc4 expression in colonic adenocarcinomas correlates with the presence of metastatic disease. *Am. J. Pathol.* **2000**, *157*, 1105–1111. [[CrossRef](#)]
89. Zhao, J.; Liang, Y.; Yin, Q.; Liu, S.; Wang, Q.; Tang, Y.; Cao, C. Clinical and prognostic significance of serum transforming growth factor-beta1 levels in patients with pancreatic ductal adenocarcinoma. *Braz. J. Med. Biol. Res.* **2016**. [[CrossRef](#)] [[PubMed](#)]
90. Tascilar, M.; Skinner, H.G.; Rosty, C.; Sohn, T.; Wilentz, R.E.; Offerhaus, G.J.; Adsay, V.; Abrams, R.A.; Cameron, J.L.; Kern, S.E.; et al. The Smad4 protein and prognosis of pancreatic ductal adenocarcinoma. *Clin. Cancer Res.* **2001**, *12*, 4115–4121.
91. Du, Y.; Zhou, X.; Huang, Z.; Qiu, T.; Wang, J.; Zhu, W.; Wang, T.; Liu, P. Meta-Analysis of the Prognostic Value of Smad4 Immunohistochemistry in Various Cancers. *PLoS ONE* **2014**, *10*, e110182. [[CrossRef](#)] [[PubMed](#)]
92. Xing, S.; Yang, H.; Liu, J.; Zheng, X.; Feng, J.; Li, X.; Li, W. Prognostic Value of SMAD4 in Pancreatic Cancer: A Meta-Analysis. *Transl. Oncol.* **2016**, *1*, 1–7.
93. Friess, H.; Yamanaka, Y.; Büchler, M.; Berger, H.G.; Kobrin, M.S.; Baldwin, R.L.; Korc, M. Enhanced expression of the type II transforming growth factor β receptor in human pancreatic cancer cells without alteration of type III receptor expression. *Cancer Res.* **1993**, *53*, 2704–2707. [[PubMed](#)]
94. Friess, H.; Yamanaka, Y.; Büchler, M.; Ebert, M.; Beger, H.G.; Gold, L.I.; Korc, M. Enhanced expression of transforming growth factor beta isoforms in pancreatic cancer correlates with decreased survival. *Gastroenterology* **1993**, *105*, 1846–1856. [[CrossRef](#)]
95. Inman, G.J.; Nicolás, F.J.; Hill, C.S. Nucleocytoplasmic shuttling of Smads 2, 3, and 4 permits sensing of TGF- β receptor activity. *Mol. Cell* **2002**, *10*, 283–294. [[CrossRef](#)]
96. Nicolás, F.J.; Hill, C.S. Attenuation of the TGF-beta-Smad signaling pathway in pancreatic tumor cells confers resistance to TGF-beta-induced growth arrest. *Oncogene* **2003**, *22*, 3698–3711. [[CrossRef](#)] [[PubMed](#)]
97. Song, K.; Hu, W.; Yue, F.; Zou, J.; Li, W.; Chen, Q.; Yao, Q.; Sun, W.; Liu, L. Transforming Growth Factor TGF β Increases Levels of Microtubule-Associated Protein MAP1S and Autophagy Flux in Pancreatic Ductal Adenocarcinomas. *PLoS ONE* **2015**, *10*, e0143150. [[CrossRef](#)] [[PubMed](#)]



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