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Stereoselective Synthesis of Homoneryl and Homogeranyl Triazole Bisphosphonates

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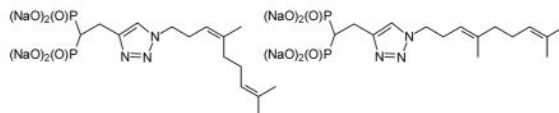
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Abstract

Isoprenoid-substituted bisphosphonates are known to serve as inhibitors of the enzyme geranylgeranyl diphosphate synthase, and their activity can be highly sensitive to olefin stereochemistry. A mixture of homogeranyl and homoneryl triazole bisphosphonates has previously demonstrated potent activity, and thus stereocontrolled syntheses of the individual isomers have been developed.

Graphical Abstract



Keywords

Homogeraniol; Homonerol; Bisphosphonate; Geranylgeranyl diphosphate synthase

Inhibitors of the enzyme farnesyl diphosphate synthase (FDPS), including pamidronate (**1**) and zoledronate (**2**), are in clinical use for treatment of osteoporosis and diseases of the bone such as Paget's disease,¹ and have become the standard of care for patients with multiple myeloma bone disease (Figure 1).² Inhibition of FDPS leads to depletion of the isoprenoid farnesyl diphosphate (FDP) as well as the downstream product geranylgeranyl diphosphate (GGDP), and there is evidence that these agents exert their pharmacological effects by virtue of the depletion of GGDP. The C₂₀ isoprenoid GGDP is formed from the C₁₅ FDP and the C₅ isopentenyl diphosphate (IPP) in a reaction mediated by the enzyme geranylgeranyl

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Notes.

The Authors declare the following competing financial interest(s): D. F. W. is a named inventor of intellectual property related to digeranyl bisphosphonate that is owned by the University of Iowa Research Foundation. He is a founder of Terpenoid Therapeutics, Inc., which has licensed this property.

Supporting Information Available: The ¹H and ¹³C NMR spectra of compounds **13**, **19**, **20**, **23**, **28**, **29**, and the ¹H NMR spectra of compounds **15** and **25**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

diphosphate synthase (GGDPS).³ If depletion of GGDP is the proximate cause of the biological activity of the nitrogenous bisphosphonates, then inhibition of GGDPS may represent a more direct way to achieve the desired biological effect.

Our earliest studies of GGDPS inhibition identified several isoprenoid bisphosphonates that affect this enzyme selectively, perhaps most notably digeranyl bisphosphonate (**3**, DGBP, Figure 2) that has an IC₅₀ of approximately 260 nM against the isolated enzyme.⁴ Several other dialkyl bisphosphonates have been reported to show comparable activity,^{5,6} and similar activity has been found more recently in the closely related ether **4**.⁷ While further improvements on the V-shaped motif of these compounds may yet be possible,⁸ in our recent efforts^{9,10} to secure more potent inhibitors we have examined bisphosphonates assembled through click chemistry.^{11,12} In this vein, the most potent inhibitor we have identified to date is the bisphosphonate **5**.¹³ This material is readily available via cycloaddition of the acetylene **6** and the azide **7**, with the azide prepared from the alcohol **8** which in turn is readily available through a ring opening rearrangement of the cyclopropyl carbinol **9**.¹⁴ Unfortunately, this rearrangement is known to give a mixture of olefin isomers in a ratio of ~3:1 (*E* to *Z*).¹⁴ Variations in the reaction conditions did improve the ratio somewhat in favor of the *E*-isomer,¹⁵ but the homonerol isomer was of special interest because the neryl analogue already was known to be more active than the corresponding geranyl isomer.⁷ As the homoallylic bromides, the olefin isomers were not readily separable and so the mixture was carried forward to provide material for initial bioassays. Those efforts were rewarded when the olefin mixture **5** was found to have an IC₅₀ of 46 nM in enzyme assays,¹³ approximately 4-fold more potent than DGBP, and attractive activity in cellular assays as well. The increased potency of this mixture relative to earlier compounds encouraged studies of the individual olefin isomers. In this report we describe the syntheses of both the *E*- and *Z*-olefin isomers of compound **5**.

Our initial efforts¹⁵ to prepare homonerol were based on use of the Wittig reagent prepared from the THP derivative of 3-bromopropanol,¹⁶ which has been shown to be *Z*-selective in its reactions with aldehydes.¹⁷ However, while condensation of this reagent with commercial 6-methyl-5-hepten-2-one gave the THP derivative of compound **8** as a 1.4:1 mixture of *Z*- and *E*-olefin isomers, those isomers were not readily separated either as the THP acetals or as the free alcohols.¹⁵ There are several reports on preparation of homonerol and homogeraniol¹⁷⁻¹⁹ or the corresponding carboxylic acids,²⁰ but to secure pure samples of the individual isomers a new sequence derived from research reported by Wessjohann and co-workers²¹ appeared to be particularly attractive (Scheme 1). Their strategy relied upon classical acetoacetate chemistry and recently was employed in a clever synthesis of epothilone D.²² For that effort, the *Z* stereochemistry of a trisubstituted olefin was derived from neryl bromide and cleanly preserved throughout their reaction sequence.

For our purposes, the β -keto acetate derivative **10** was prepared from neryl bromide and *t*-butyl 2-acetoxyacetoacetate according to the known procedure,²² and subsequent decarboxylation upon treatment with TsOH gave racemic acetate **11** (Scheme 1).²¹ Hydrolysis of this acetate proceeded smoothly to the acyloin **12**,²³ but then our efforts to bring about oxidative cleavage to the carboxylic acid went unrewarded. In contrast, reduction of compound **11** proceeded smoothly upon treatment with LiAlH₄ to give the diol

13 as a mixture of stereoisomers, and oxidative cleavage by treatment with sodium periodate on silica gel gave the expected aldehyde **14** in nearly quantitative yield.

Once homoneral (**14**) was in hand, the remaining steps in the synthetic sequence proceeded as expected. Reduction of aldehyde **14** proceeded smoothly to give the homoallylic alcohol **15** as a single olefin isomer. After conversion of this alcohol to the corresponding mesylate (**16**) and a subsequent reaction with sodium azide to obtain the alkyl azide **17**, the click reaction with acetylene **18** gave the desired triazole **19**. Hydrolysis of the tetraester **19** under standard McKenna conditions²⁴ gave the final product **20**, with no apparent formation of the isomeric olefin. The final product as well as the late stage intermediates were identified as single olefin isomers on the basis of their ¹³C NMR spectra. The C-4 methylene group of nerol and geraniol have significantly different resonances in their ¹³C NMR spectra (32.2 and 39.7 ppm, respectively),²⁵ and the corresponding C-5 methylene resonances for homoneral (32.2 ppm) and homogeraniol (40.0 ppm) are virtually identical. In compound **19**, which was soluble in CDCl₃, this resonance was observed at 32.2 ppm; in compound **20**, which was soluble in D₂O, it was observed at 31.5 ppm. While the resonance of the alpha carbon in the bisphosphonate also is found in this range, it can be unambiguously identified by the large coupling constant with the two phosphorus atoms (~120 Hz).

A parallel reaction sequence was employed to obtain the corresponding homogeranyl isomer (Scheme 2). In this series, the early reactions with the geranyl derivatives proceeded under parallel conditions and in nearly the same yields as those with the neryl isomer. For experimental convenience, the mesylate **26** again was converted directly to the azide **27** and then the azide was carried immediately into the click reaction with alkyne **18** to give the triazole **28** as the tetra ethyl ester. Hydrolysis then proceeded under parallel conditions to give the desired salt **29**. This sequence also gave a single olefin isomer throughout. For the ester **28**, the resonance for the key C-5 methylene group was observed at 39.8 ppm (CDCl₃) while the corresponding resonance in the salt **29** was found at 39.3 ppm (D₂O).

In conclusion, isomerically pure samples of both the *Z*- and *E*-olefin isomers **20** and **29** have been prepared through short synthetic sequences based on classic acetoacetate chemistry. These sequences can be conducted on a gram scale, and have provided materials appropriate for further biological investigations. Based on the initial Western blot analyses, the homoneryl isomer does appear to be more potent as an inhibitor of GGDPS than the homogeranyl isomer, which is consistent with the relative activity of the neryl/geranyl pair,⁷ but more quantitative comparisons will require further studies.

Experimental

General Experimental Procedures

Diethyl ether was freshly distilled from sodium and benzophenone, whereas methylene chloride was distilled from calcium hydride prior to use. All other reagents and solvents were purchased from commercial sources and used without further purification. All reactions in nonaqueous solvents were conducted in flame-dried glassware under a positive pressure of argon and with a magnetic stir bar. NMR spectra were obtained at 300 MHz for ¹H, 75 MHz for ¹³C NMR, and 121 MHz for ³¹P NMR, in CDCl₃ with (CH₃)₄Si (¹H,

0.00 ppm) or CDCl₃ (¹H, 7.26 ppm; ¹³C NMR; 77.0 ppm) for non-aqueous samples or D₂O (¹H, 4.80 ppm) and 1,4-dioxane (¹³C, 67.19) for aqueous samples, as the internal standards. High resolution mass spectra were obtained by GC-TOF.

(5Z)-3-Acetoxy-6,10-dimethyl-5,9-undecadien-2-one (11)

According to the published procedure,²¹ *p*-TsOH·H₂O (260 mg, 1.37 mmol) was added to a stirred solution of β-keto ester **10** (4.69 g, 13.3 mmol) in benzene (40 mL) at room temperature. The resulting solution was heated at 78 °C for 90 minutes and then was allowed to stir for two days at room temperature. The reaction mixture then was filtered through a bed of silica, the silica was rinsed with EtOAc, and the combined filtrate was concentrated *in vacuo* to afford the desired keto acetate **11** (3.34 g, 99%) as an orange oil which was used without further purification. The ¹H and ¹³C NMR data were consistent with the literature data.²¹

(5Z)-2,3-Dihydroxy-6,10-dimethyl-5,9-undecadiene (13)

LiAlH₄ (247 mg, 6.51 mmol) was added to a stirred solution of keto acetate **11** (912 mg, 3.61 mmol) in Et₂O (18 mL) at 0 °C. After it was stirred for 2 hours and allowed to warm slowly, the reaction then was cooled to 0 °C, quenched by slow addition of 1N HCl followed by H₂O, and extracted with Et₂O (3x). The combined organic extracts were dried (Na₂SO₄), and filtered, and the filtrate was concentrated *in vacuo* to afford the desired diol **13** (685 mg, 89%) as a yellow oil. This mixture of diastereomers was used in the next step without further purification: ¹H NMR (300 MHz, CDCl₃) δ 5.23–5.05 (m, 2H), 3.89–3.31 (m, 2H), 2.28–2.16 (m, 2H), 2.12–2.04 (m, 4H), 2.02–1.98 (m, 1H), 1.94–1.88 (m, 1H), 1.74 (d, *J* = 1.2 Hz, 3H), 1.68 (s, 3H), 1.61 (d, *J* = 1.2 Hz, 3H), 1.22–1.16 (m, 3H); ¹³C NMR (75 MHz, CDCl₃) for the major isomer δ 137.8, 131.6, 124.1, 120.7, 76.1, 70.3, 32.1, 26.5, 25.7, 23.5, 19.4, 17.6, 16.6; HRMS (ES⁺) *m/z* calcd for C₁₃H₂₄O₂Na (M + Na)⁺ 235.1674, found 235.1679.

(Z)-4,8-Dimethylnona-3,7-dienal (14)

In a manner similar to the published procedure,²⁶ sodium periodate (2.57 g, 12.03 mmol) was dissolved in hot water (5 mL, 70 °C), and silica gel (10.04 g) was added, followed by vigorous shaking to yield sodium periodate coated silica gel. The prepared periodate reagent (510 mg, 0.35 mmol) was suspended in CH₂Cl₂ (2.7 mL) and allowed to stir for 10 minutes. To the suspension was added diol **13** (51 mg, 0.24 mmol), and the mixture was allowed to stir for 30 minutes, followed by filtration of the reaction mixture through diatomaceous earth, and removal of solvent using a rotary evaporator to yield the desired aldehyde **14** (39 mg, 100%) as a yellow oil that was used without further purification. The ¹H and ¹³C NMR data were consistent with the literature data.²⁷

(Z)-4,8-Dimethylnona-3,7-dien-1-ol (15)

To a stirred suspension of LiAlH₄ (95%, 93 mg, 2.44 mmol) in Et₂O (27 mL) at 0 °C, aldehyde **14** (677 mg, 4.07 mmol) was added as a solution in Et₂O (1 mL) over four minutes. The reaction was allowed to warm to room temperature and stirred overnight. The reaction was cooled to 0 °C, quenched by slow addition of 1N HCl followed by H₂O, and extracted with Et₂O. The combined organic extracts were dried (Na₂SO₄) and filtered, and

the filtrate was concentrated *in vacuo*. Final purification by column chromatography (5% EtOAc in hexanes) afforded the desired alcohol **15** (654 mg, 95%) as a clear oil with a ^1H NMR spectrum in agreement with known data;²⁸ ^{13}C NMR (75 MHz, CDCl_3) δ 139.1, 132.1, 124.3, 121.0, 62.8, 32.2, 31.6, 26.8, 25.9, 23.7, 17.9.

(Z)-4,8-Dimethylnona-3,7-dien-1-yl methanesulfonate (16)

In a manner similar to a published procedure,²⁹ NEt_3 (0.25 mL, 1.78 mmol) was added to a stirred solution of homonerol (**15**, 178 mg, 1.06 mmol) in CH_2Cl_2 (10 mL) at 0 °C. After the reaction was allowed to stir at 0 °C for twenty minutes, methanesulfonyl chloride (0.13 mL, 1.68 mmol) was added dropwise to the reaction mixture. The reaction was allowed to stir for three hours at 0 °C, followed by addition of H_2O to quench the reaction mixture. The resulting mixture was washed with 1N HCl (2x), brine (1x), and NaHCO_3 (2x). The combined organic extracts were dried (Na_2SO_4) and filtered, and the filtrate was concentrated *in vacuo* to afford desired mesylate **16** (247 mg, 95%) as a yellow oil that was carried immediately to the next step.

Tetraethyl (3Z)-(2-(1-(4,8-dimethylnon-3,7-dien-1-yl)-1H-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (19)

In a manner similar to published procedures,^{11,30} NaN_3 (473 mg, 7.28 mmol) was added to a stirred solution of homoneryl mesylate (**16**, 944 mg, 3.83 mmol) in DMF (20 mL) under an argon atmosphere. The mixture was heated to 40 °C and allowed to stir overnight. The reaction then was diluted with Et_2O , washed with water (5x) and brine, dried (Na_2SO_4) and filtered, and the filtrate was concentrated *in vacuo*, to yield homoneryl azide (**17**, 626 mg, 84%) as a yellow oil. The material was used immediately in the following reaction without further purification. To a stirred solution of tetraethyl but-3-yne-1,1-diylidiphosphonate³¹ (**18**, 770 mg, 2.36 mmol) and homoneryl azide (**17**, 616 mg, 3.19 mmol) in *t*-BuOH/ H_2O (4:1, 25 mL total), saturated CuSO_4 (0.01 mL) and sodium ascorbate (140 mg, 0.71 mmol) were added in sequence. The resulting reaction mixture was allowed to stir overnight at room temperature, and then the solvent was removed *in vacuo*. The resulting residue was dissolved in brine and extracted with EtOAc (5x). The combined organic extracts were washed with 5% NH_4OH , dried (Na_2SO_4) and filtered, and the filtrate was concentrated *in vacuo*. Final purification by column chromatography (10% EtOH in hexanes) afforded triazole **19** (982 mg, 80%) as a yellow oil: ^1H NMR (300 MHz, CDCl_3) δ 7.46 (s, 1H), 5.12–5.03 (m, 2H), 4.26 (t, $J = 7.5$ Hz, 2H), 4.19–4.06 (m, 8H), 3.39–3.24 (td, $J_{\text{HP}} = 16.1$ Hz, $J = 6.4$ Hz, 2H), 3.06–2.85 (m, 1H), 2.60–2.51 (m, 2H), 2.04–1.96 (m, 4H), 1.69 (s, 3H), 1.67 (s, 3H), 1.59 (s, 3H), 1.32–1.36 (m, 12H); ^{13}C NMR (75 MHz, CDCl_3) δ 145.2 (t, $J_{\text{CP}} = 8.8$ Hz), 139.6, 132.2, 124.0, 122.4, 119.6, 63.0 (d, $J_{\text{CP}} = 6.6$ Hz, 2C), 62.7 (d, $J_{\text{CP}} = 6.4$ Hz, 2C), 50.5, 36.9 (t, $J_{\text{CP}} = 133.0$ Hz), 32.2, 29.2, 26.5, 25.9, 23.6, 22.3 (t, $J_{\text{CP}} = 4.9$ Hz), 17.9, 16.6 (d, $J_{\text{CP}} = 3.7$ Hz, 2C), 16.6 (d, $J_{\text{CP}} = 3.4$ Hz, 2C); ^{31}P NMR (121 MHz, CDCl_3) 22.5 ppm; HRMS (ES^+) m/z calcd for $\text{C}_{23}\text{H}_{44}\text{N}_3\text{O}_6\text{P}_2$ ($\text{M} + \text{H}$)⁺ 520.2705, found 520.2698.

Sodium (3Z)-2-(1-(4,8-dimethylnon-3,7-dien-1-yl)-1H-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (20)

In a manner similar to published procedures,^{9,24} to a stirred solution of bisphosphonate ester **19** (938 mg, 1.81 mmol) in CH₂Cl₂ (30 mL) at 0 °C, collidine (1.67 mL, 12.6 mmol) and TMSBr (97%, 1.97 mL, 15.2 mmol) were added dropwise in succession. The reaction was allowed to stir overnight while it warmed to room temperature, and the solvent then was removed *in vacuo*. The resulting residue was diluted with toluene (40 mL) and concentrated *in vacuo* to remove any excess TMSBr (3x). It then was treated with 2N NaOH (6.05 mL, 12.1 mmol) and allowed to stir overnight at room temperature. Anhydrous acetone was added and the mixture was placed in the freezer for 20 minutes. The resulting solid was collected by filtration, dissolved in water, reprecipitated by addition of anhydrous acetone and the mixture was placed in the freezer for 20 minutes. The resulting solid was collected by filtration, dissolved in water, and lyophilized to provide the desired salt **20** (501 mg, 56%) as a white powder: ¹H NMR (300 MHz, D₂O) δ 7.84 (s, 1H), 5.21–5.10 (m, 2H), 4.39 (t, *J* = 6.6 Hz, 2H), 3.21 (td, *J*_{HP} = 15.2 Hz, *J* = 6.6 Hz, 2H), 2.63–2.55 (m, 2H), 2.18–1.86 (m, 5H), 1.68 (m, 6H), 1.61 (s, 3H); ¹³C NMR (125 MHz, D₂O) δ 147.5, 140.8, 134.3, 124.8, 124.7, 120.6, 50.9, 40.1 (t, *J*_{CP} = 116.7 Hz), 31.5, 28.9, 26.3, 25.5, 23.1, 22.3 (t, *J*_{CP} = 4.0 Hz), 17.5; ³¹P NMR (121 MHz, D₂O) 18.7 ppm; HRMS (ES⁻) *m/z* calcd for C₁₅H₂₆N₃O₆P₂ (M - H)⁻ 406.1297, found 406.1289.

(5E)-2,3-Dihydroxy-6,10-dimethyl-5,9-undecadiene (23)

According to the procedure for preparation of diol **13**, keto ester **22** (1.26 g, 5.00 mmol) was treated with LiAlH₄ (595 mg, 14.9 mmol) in Et₂O (34 mL) at 0 °C. A parallel work up afforded the desired diol **23** (1.04 g, 98%) as a yellow oil that was used without further purification: ¹H NMR (300 MHz, CDCl₃) δ 5.20–5.12 (m, 1H), 5.10–5.01 (m, 1H), 3.90–3.76 (m, 1H), 3.69–3.54 (m, 1H), 2.10–1.96 (m, 6H), 1.68 (s, 3H), 1.64 (s, 3H), 1.60 (s, 3H), 1.25 (br s 3H); ¹³C NMR (75 MHz, CDCl₃) δ 139.7, 132.1, 124.3, 113.2, 75.9, 70.6, 40.0, 29.9, 26.7, 25.9, 19.5, 17.9, 17.4; HRMS (ES⁺) *m/z* calcd for C₁₃H₂₄O₂Na (M + Na)⁺ 235.1674, found 235.1658.

(E)-4,8-Dimethylnona-3,7-dienal (24)

According to the procedure for the preparation of aldehyde **14**, diol **23** (1.78 g, 8.38 mmol) was added to a stirred suspension of periodate coated silica gel (17.67 g, 12.1 mmol) in CH₂Cl₂ (56 mL). A parallel work up afforded the desired aldehyde **24** (1.39 g, 100%) as a yellow oil that was used without further purification. The ¹H NMR data was consistent with the literature data.²⁷

(E)-4,8-Dimethylnona-3,7-dien-1-ol (25)

According to the procedure for the preparation of homoallylic alcohol **15**, aldehyde **24** (1.39 g, 8.38 mmol) was added to a suspension of LiAlH₄ (180 mg, 5.03 mmol) in Et₂O (55 mL) at 0 °C. A parallel work up afforded homogeraniol (**25**, 1.35 g, 96%) as a yellow oil that was used without further purification. The ¹H NMR data was consistent with the literature data for material prepared by a different method.^{32,32}

(E)-4,8-Dimethylnon-3,7-dien-1-yl methanesulfonate (26)

According to the procedure for the preparation of mesylate **16**, homogeraol (**25**, 419 mg, 2.49 mmol) was treated with NEt_3 (0.59 mL, 4.20 mmol) followed by MsCl (0.31 mL, 4.01 mmol) in CH_2Cl_2 (25 mL) at 0 °C. A parallel work up afforded the desired mesylate **26** (542 mg, 88%) as a yellow oil that was used immediately without further purification.

Tetraethyl (3E)-(2-(1-(4,8-dimethylnon-3,7-dien-1-yl)-1H-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (28)

According to the procedure for the preparation of triazole **19**, mesylate **26** (161 mg, 0.65 mmol) was treated with NaN_3 (80.1 mg, 1.23 mmol), and the resulting azide (**27**, 111 mg, 0.57 mmol) was isolated and immediately treated with acetylene bisphosphonate **18** (109 mg, 0.33 mmol), saturated CuSO_4 (0.01 mL), and sodium ascorbate (25 mg, 0.13 mmol) in sequence. A parallel workup and purification afforded the desired triazole **28** (112 mg, 65%) as a yellow oil: ^1H NMR (300 MHz, CDCl_3) δ 7.40 (s, 1H), 5.06–4.95 (m, 2H), 4.24–4.18 (t, $J = 7.3$ Hz, 2H), 4.12–4.04 (m, 8H), 3.32–3.19 (td, $J_{\text{HP}} = 16.2$ Hz, $J = 6.5$ Hz, 2H), 3.00–2.80 (tt, $J_{\text{HP}} = 23.5$ Hz, $J = 6.3$ Hz, 1H), 2.54–2.46 (dt, $J = 7.9$ Hz, 7.3 Hz, 2H), 2.00–1.88 (m, 4H), 1.61 (s, 3H), 1.53 (s, 3H), 1.48 (s, 3H), 1.26–1.19 (m, 12H); ^{13}C NMR (75 MHz, CDCl_3) δ 145.0, 139.7, 131.9, 124.1, 122.5, 118.8, 63.1 (d, $J_{\text{CP}} = 6.5$ Hz, 2C), 62.8 (d, $J_{\text{CP}} = 6.5$ Hz, 2C), 50.3, 39.8, 36.8, 29.4, 26.7, 25.9, 22.3 (t, $J_{\text{CP}} = 4.9$ Hz), 17.9, 16.5 (d, $J_{\text{CP}} = 3.4$ Hz, 2C), 16.4 (d, $J_{\text{CP}} = 3.7$ Hz, 2C), 16.3; ^{31}P NMR (121 MHz, CDCl_3) 22.5 ppm; HRMS (ES^+) m/z calcd for $\text{C}_{23}\text{H}_{44}\text{N}_3\text{O}_6\text{P}_2$ ($\text{M} + \text{H}$) $^+$ 520.2705, found 520.2703.

Sodium (3E)-(2-(1-(4,8-dimethylnon-3,7-dien-1-yl)-1H-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (29)

According to the procedure for the preparation of sodium salt **20**, the bisphosphonate ester **28** (1.10 g, 2.12 mmol) was treated with TMSBr (97%, 2.30 mL, 17.7 mmol), collidine (1.95 mL, 14.6 mmol) and then 2N NaOH (7.2 mL). A parallel work up and precipitation provided the desired sodium salt **29** (1.05 g, 70%) as a white powder: ^1H NMR (300 MHz, D_2O) δ 7.71 (s, 1H), 5.08–4.98 (m, 2H), 4.27–4.20 (t, $J = 6.8$ Hz, 2H), 3.10–2.95 (td, $J_{\text{HP}} = 15.0$ Hz, $J = 6.8$ Hz, 2H), 2.52–2.42 (m, 2H), 2.10–1.80 (m, 5H), 1.56 (s, 3H), 1.48 (s, 3H), 1.36 (s, 3H); ^{13}C NMR (75 MHz, D_2O) δ 150.5 (t, $J_{\text{CP}} = 7.3$ Hz), 140.4, 134.0, 124.8, 124.4, 119.6, 50.5, 41.8 (t, $J_{\text{CP}} = 118.1$ Hz), 39.3, 28.8, 26.2, 25.4, 24.3, 17.5, 15.6; ^{31}P NMR (121 MHz, D_2O) 18.7 ppm; HRMS (ES^-) m/z calcd for $\text{C}_{15}\text{H}_{26}\text{N}_3\text{O}_6\text{P}_2$ ($\text{M} - \text{H}$) $^-$ 406.1297, found 406.1304.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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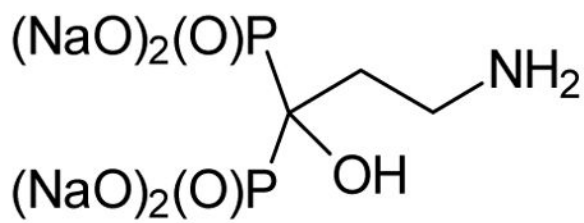
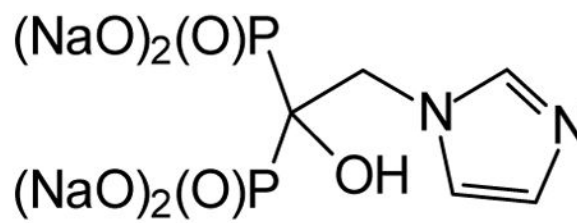
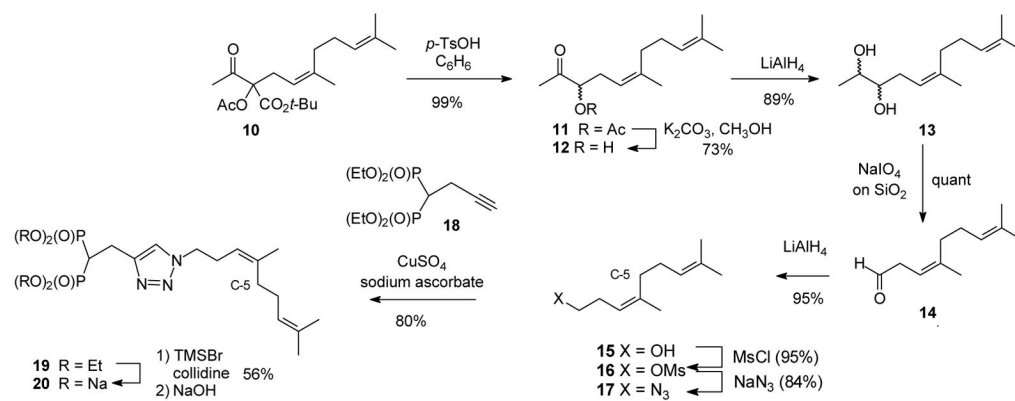
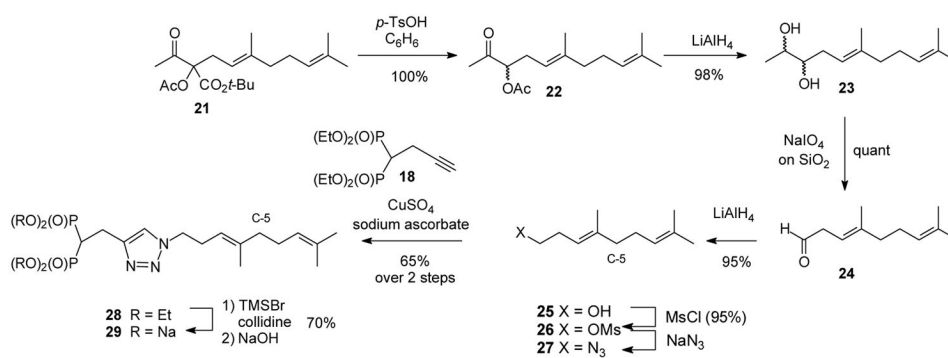
**1** (pamidronate)**2** (zoledronate)

Figure 1.
Inhibitors of FDPS in clinical use.



Scheme 1.
Synthesis of the homoneryl isomer **20**.



Scheme 2.
Synthesis of the homogeranyl isomer **29**.