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Solution and structural binding studies of phosphate with thiophene-based azamacrocycles

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Abstract

Two thiophene-based monocyclic receptors **L1** and **L2** have been studied for phosphate binding in solutions (D₂O and DMSO-*d*₆) by ¹H NMR and ³¹P NMR titrations, and in the solid state by single crystal X-ray analysis. Results from ¹H NMR titrations suggest that the ligands bind phosphate anions in a 1:2 binding mode in DMSO-*d*₆, with the binding constants of 5.25 and 4.20 (in log *K*), respectively. The binding of phosphate to **L1** and **L2** was further supported by ³¹P NMR in D₂O at pH = 5.2. The crystal structure of the phosphate complex of **L1** reveals unambiguous proof for the formation of a ditopic complex *via* multiple hydrogen bonds from NH...O and CH...O interactions.

Keywords

Azamacrocycle; Ditopic complex; Anion binding; Phosphate complex

Phosphate is a key building block of nucleic acids, playing critical roles in many biochemical processes [1]. The translocation of phosphate between DNA and proteins in living cells is an essential step in the regulation of metabolic processes [2]. Energy production and storage processes within the body are regulated by phosphorylated compounds, such as adenosine triphosphate [3]. It is known that the phosphate level in serum and saliva is linked to several diseases including hyperparathyroidism, vitamin D deficiency and *Fanconi* syndrome [4]. It is also widely used in fertilizer and drug-related industries [5]. Because of its significant roles in environment, healthcare and biochemical applications, molecular recognition of phosphate by synthetic molecules is a growing area of current research in supramolecular chemistry [6]. However, phosphate binding in water is unfavorable due to its high free energy of hydration ($G^{\circ} = -465$ J/mol) [7]. Polyamine-based receptors that are water soluble, are often used to bind phosphates in water over a wide range of pH [8]. For examples, Martell and coworkers synthesized hexaazamacrocyclic ligands and used them to encapsulate a pyrophosphate *via* four hydrogen bonds [9]. A hexaprotonated 26-membered polyammonium macrocycle reported by Bowman-James and

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Appendix A. Supplementary material CCDC 1035333 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif. Supplementary data associated with this article can be found, in the online version, at doi: ???.

coworkers was shown to form a complex with six di-hydrogen phosphate anions and two neutral phosphoric acid molecules, illustrating diprotic behavior of the receptor [10]. Bianchi, García-España, Paoletti and coworkers reported a tetraprotonated macrocycle [18]aneN₆ that binds two pyrophosphate anions *via* NH...O and CH...O bonds [11]. Lu and coworkers showed that an octa-protonated *p*-xylyl-based cryptand encapsulated a phosphate anion *via* multiple hydrogen bonds [12]. Other reported receptors that are neutral molecules including amides [13], thioamides [14], ureas [15], thioureas [16], pyrroles [17] and indoles [18] were shown to bind phosphate in organic solvents. Our recent studies on various macrocycle-based receptors indicated that hydrogen bonding and electrostatic interactions are major binding forces in stabilizing complexes; thus providing insight into the binding modes of anions and conformations of host molecules [19-21]. Herein, we report the binding aspects of two thiophene-based macrocycles **L1** and **L2** *via* ¹H NMR and ³¹P NMR in solutions. We also report the structural characterization of a phosphate complex with **L1**, forming a ditopic complex with one dihydrogen phosphate and one monohydrogen phosphate.

The hexamine ligands **L1** and **L2** were synthesized through Schiff-base condensation reaction (Scheme 1) of the corresponding dipodal amine with two equivalents of 2,5-thiophendicarboxaldehyde followed by the reduction with NaBH₄, as reported earlier [20-22]. The tosylate salts were prepared by reacting the free ligands with *p*-toluenesulfonic acid in methanol. The analysis of ¹H NMR data suggested the formation of an adduct with six tosylate groups providing six positive charges on the macrocyclic moiety. The phosphate complex was synthesized by the addition of a few drops of aqueous phosphoric acid to **L1** dissolved in methanol. Crystals suitable for X-ray analysis were grown from the slow evaporation of the solution at room temperature.

¹H NMR titrations of [H₆**L1**]⁶⁺ and [H₆**L2**]⁶⁺ were performed to study the binding interactions with phosphate using *n*-Bu₄N⁺H₂PO₄⁻ (TBAH₂PO₄) in D₂O as well as in DMSO-*d*₆. The incremental addition of the anion (20 mM) to [H₆**L1**]⁶⁺ (2 mM) in D₂O (pH = 5.2) resulted in a downfield shift of aliphatic protons. The non-linear regression analysis of the shift change of several independent protons of [H₆**L1**]⁶⁺ showed moderate binding for phosphate (log *K* = 2.0 M⁻¹), providing a good fit of a 1:1 binding model [23]. However, under identical conditions, there was no significant change in any protons of [H₆**L2**]⁶⁺ upon the addition of TBAH₂PO₄ to the host solution, indicating weak host-guest interactions in D₂O. We, therefore, proceeded to investigate the binding properties of [H₆**L1**]⁶⁺ and [H₆**L2**]⁶⁺ in DMSO-*d*₆. Figure 1 displays the stacking of ¹H NMR titration spectra of [H₆**L1**]⁶⁺ obtained after the increasing amount of phosphate anion (ranging 0 to 10 equivalents), showing a gradual upfield shift of both the aromatic and the aliphatic protons of the macrocyclic moiety at room temperature. The changes in the chemical shift of the aromatic proton (*ArH*) as a function of the anion concentration are displayed in Figure 1. The non-linear regression analysis of the changes in the chemical shift of the ligand as recorded with an increasing amount of anionic solution provided the best fit for a 1:2 binding model [24]. The ligand [H₆**L2**]⁶⁺ also shows a similar binding trend with phosphate in DMSO-*d*₆. Association constants presented in Table 1 suggest that the binding process of each ligand involves the formation of both a 1:1 (ligand:anion) complex and a 1:2 (ligand:anion) complex. However, a 1:2 complex is stronger than a 1:1 complex in DMSO-

d_6 supporting the X-ray structure (discussed later). A similar binding mode was previously reported for phosphate with [26]aneN₆C₆ [10]. As shown in Table 1, the overall binding constant of **L1** for phosphate in DMSO- d_6 is slightly higher than that of **L2** which could be due to the reduced hydrogen bonding ability of the methylated compound.

The interaction of [H₆**L1**]⁶⁺ and [H₆**L2**]⁶⁺ with TBAH₂PO₄ in D₂O was also investigated by phosphorus ³¹P NMR at room temperature. Because of the lower sensitivity of ³¹P NMR compared to ¹H NMR, a higher concentration of TBAH₂PO₄ (10 mM) was loaded to an NMR tube, and a host solution (25 mM) was prepared as a titrant. The ³¹P resonance was calibrated against an aqueous phosphoric acid used in a sealed capillary tube. Figure 2 shows the ³¹P NMR spectra of TBAH₂PO₄ (10 mM) after the addition of two equivalents of [H₆**L1**]⁶⁺ and [H₆**L2**]⁶⁺ in D₂O. As clearly shown in Figure 2a, the signal at $\delta_P = 0.218$ ppm for the free TBAH₂PO₄ shifts upfield to $\delta_P = -1.635$ ppm ($\delta_P = 1.853$ ppm) after the addition of [H₆**L1**]⁶⁺ (2 equivalents). The other ligand [H₆**L2**]⁶⁺ also shows similar shifting to $\delta_P = -1.404$ ppm but to a lesser extent ($\delta_P = 1.622$ ppm), indicating a lower affinity for the phosphate anion than that with [H₆**L1**]⁶⁺. This upfield shift suggest the formation of the phosphate complex due to the strong electrostatic interactions, where the bound phosphate is shielded by the interacting macrocycle. Similar upfield shifts of phosphorus resonance were reported due to the complexation by polyamine-based hosts [12,25].

The phosphate complex of **L1** was obtained by reacting the free macrocycle with phosphoric acid. Crystals suitable for X-ray analysis were obtained by recrystallization of the salt in methanol/water. Crystallographic data are presented in Table 2. The structural analysis of the phosphate complex reveals that the salt crystallizes in the triclinic space group (*P1*) and the asymmetric unit consists of two hexaprotonated macrocycles, three monohydrogen phosphate (HPO₄²⁻), six di-hydrogen phosphate (H₂PO₄⁻) and eight water molecules to yield a molecular formula of 2(C₂₀H₄₀N₆S₂)⁶⁺·3(HPO₄)²⁻·6(H₂PO₄)⁻·8(H₂O). One hydrogen phosphate (P9) is disordered about 90:10 into two orientations. One water molecule (O8W) is also found to be disordered about 80:20. Each macrocycle is fully protonated and the total positive charges (12+) from the two hexaprotonated receptors are balanced by the negative charges provided by three doubly-charged monohydrogen phosphates and six singly-charged dihydrogen phosphate anions.

As shown in Figure 3, two identical macrocycles in the asymmetric unit possess a non-crystallographic inversion center, forming a sandwich type complex with four bound phosphate groups and two water molecules. Two water molecules serve as linkers to macrocycles as well as to bound dihydrogen phosphates locating between the macrocycles (Figure 3a). Each macrocycle adopts the shape of a flat ellipsoid and binds two phosphate groups; one is monohydrogen phosphate and the other is dihydrogen phosphate, located above and below the elliptical plane. The complex is stabilized through electrostatic interactions and multiple H-bonds. A list of H-bond distances with bond angles are shown in Table 3. Figure 3c shows one macrocycle of the asymmetric unit, with two bound phosphates held by hydrogen bonding interactions with the macrocyclic cation. The monohydrogen phosphate (P76) is held by two NH...O (2.684 and 2.747 Å) and two CH...O (3.266 and 3.388 Å) bonds, while the dihydrogen phosphate interacts through two strong NH...O bonds (N15H...O70 = 2.932 and N15H...O73 = 2.677 Å) and one weak NH...O

bonds ($N39H\cdots O80 = 3.266 \text{ \AA}$). As a result, the monohydrogen phosphate is closer to the cavity than the dihydrogen phosphate. The involvement of $CH\cdots O$ interactions is well documented in the literature [21]. Extensive hydrogen bonds between the macrocycle and water molecules are also present.

In conclusion, we report the phosphate binding properties of two simple thiophene-based monocyclic receptors **L1** and **L2** by 1H NMR and ^{31}P NMR studies in two different polar solvents (D_2O and $DMSO-d_6$). Both ligands show lessened interactions with the anion in competitive D_2O due to their weak hydrogen bonding ability in the highly polar solvent. However, after switching the solvent from D_2O to $DMSO-d_6$, the binding affinity is significantly increased, exhibiting the formation of 1:2 complexes. The ^{31}P NMR has been successfully used as a probe to identify the chemical environment of bound and free phosphates in D_2O . Structural characterization of the phosphate complex with **L1** suggests that both $NH\cdots O$ and $CH\cdots O$ interactions play roles in stabilizing the host-guest complex.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

1. (a) Mason, CF. *Biology of Freshwater Pollution*. Longman; New York: 1991. (b) Liebschner D, Elias M, Moniot S, Fournier B, Scott K, Jelsch C, Guillot B, Lecomte C, Chabrière E. *J Am Chem Soc.* 2009; 131:7879–7886. [PubMed: 19445459]
2. Smith E, Morowitz H. *Proc Natl Acad Sci USA.* 2004; 101:13168–73. [PubMed: 15340153]
3. Kivlehan F, Mace WJ, Moynihan HA, Arrigan DWM. *Analytica Chimica Acta.* 2007; 585:154–160. [PubMed: 17386660]
4. Berchmans S, Issa TB, Singh P. *Analytica Chimica Acta.* 2012; 729:7–20. [PubMed: 22595428]
5. Tiessen, H. *Phosphorus in the Global Environment: Transfers, Cycles, and Management*. Vol. 1995. Wiley; New York:
6. (a) Hargrove AE, Nieto S, Zhang T, Sessler JL, Anslyn EV. *Chem Rev.* 2011; 111:6603–6782. [PubMed: 21910402] (b) Tobey SL, Anslyn EV. *J Am Chem Soc.* 2003; 125:14807–14815. [PubMed: 14640656] (c) Barker JE, Liu Y, Martin ND, Ren T. *J Am Chem Soc.* 2003; 125:13332–13333. [PubMed: 14583009] (d) Iverson BL, Shreder K, Kral V, Sessler JL. *J Am Chem Soc.* 1993; 115:11022–11023. (e) Arranz P, Bianchi A, Cuesta R, Giorgi C, Godino ML, Gutiérrez MD, López R, Santiago A. *Inorg Chem.* 2012; 51:4883–4883. (f) Murugavel R, Kuppaswamy S, Randoll S. *Inorg Chem.* 2008; 47:6028–6039. [PubMed: 18500795] (g) Barker JE, Liu Y, Yee GT, Chen WZ, Wang G, Rivera VM, Tong R. *Inorg Chem.* 2006; 45:7973–7980. [PubMed: 16961393] (h) Warden AC, Warren M, Hearn MTW, Spiccia L. *Inorg Chem.* 2004; 43:6936–6943. [PubMed: 15500331] (i) Harris WR, Wang Z, Hamada YZ. *Inorg Chem.* 2003; 42:3262–3273. [PubMed: 12739968] (j) Jur ek O, Cametti M, Pontini M, Kolehmainen E, Rissanen K. *Org Biomol Chem.* 2013; 11:4585–4590. [PubMed: 23774968] (k) Arranz-Mascarós P, Bazzicalupi C, Bianchi A, Giorgi C, Godino-Salido ML, Gutiérrez-Valero MD, Lopez-Garzón R, Valtancoli B. *New J Chem.* 2011; 35:1883–1891. (l) Rajbanshi A, Wan S, Custelcean R. *Cryst Growth Des.* 2013; 13:2233–2237. (k) Gavette JV,

- Mills NS, Zakharov LN, Johnson CA, Johnson DW, Haley MM. *Angew Chem.* 2013; 125:10460–10464. (l) Law al AT, Adeloju SB. *Talanta.* 2013; 114:191–203. [PubMed: 23953460] (l) Liu J, Wu K, Li X, Han Y, Xia M. *RSC Adv.* 2013; 3:8924–8928.
7. Blades AT, Ho Y, Kebarle P. *J Phys Chem.* 1996; 100:2443–2446.
 8. García-España E, Díaz P, Llinares JM, Bianchi A. *Coord Chem Rev.* 2006; 250:2952–2988.
 9. Nation DA, Reibenspies J, Martell AE. *Inorg Chem.* 1996; 35:4597–4603.
 10. Gerasimchuk OA, Mason S, Llinares JM, Song M, Alcock NW, Bowman-James K. *Inorg Chem.* 2000; 39:1371–1375. [PubMed: 12526438]
 11. Bazzicalupi C, Bencini A, Bianchi A, Cecchi M, Escuder B, Fusi V, Garcia-España E, Giorgi C, Luis SV, Maccagni G, Marcelino V, Paoletti P, Valtancoli B. *J Am Chem Soc.* 1999; 121:6807–6815.
 12. Yang LZ, Li Y, Jiang L, Feng XL, Lu TB. *CrystEngComm.* 2009; 11:2375–2380.
 13. Bondy CR, Loeb SJ. *Coord Chem Rev.* 2003; 240:77–99.
 14. (a) Hossain MA, Kang SK, Llinares LM, Powell D, Bowman-James K. *Inorg Chem.* 2003; 42:5043–5045. [PubMed: 12924873] (b) Inoue Y, Kanbara T, Yamamoto T. *Tetrahedron Lett.* 2003; 44:5167–5169.
 15. (a) Custelcean R, Moyer BA, Hay BP. *Chem Commun.* 2005:5971–5973. (b) Ravikumar I, Lakshminarayanan PS, Arunachalam M, Suresh E, Ghosh P. *Dalton Trans.* 2009:4160–4168. [PubMed: 19452065] (c) Bell KJ, Westra AN, Warr RJ, Chartres J, Ellis R, Tong CC, Blake AJ, Tasker PA, Schroder M. *Angew Chem Int Ed.* 2008; 47:1745–1748. (d) Busschaert N, Wenzel M, Light ME, Iglesias-Hernandez P, Perez-Tomas R, Gale PA. *Am J Chem Soc.* 2011; 133:14136–14148. (e) Bose P, Dutta R, Ghosh P. *Org Biomol Chem.* 2013; 11:4581–4584. [PubMed: 23764916] (f) Basu A, Das G. *J Org Chem.* 2014; 79:2647–2656. [PubMed: 24558963] (g) Pramanik A, Powell DR, Wong BM, Hossain MA. *Inorg Chem.* 2012; 51:4274–4284. [PubMed: 22409384]
 16. (a) Khansari ME, Wallace KD, Hossain MA. *Tetrahedron Lett.* 2014:438–440. (b) Bose P, Dutta R, Santra S, Chowdhury B, Ghosh P. *Eur Inorg J Chem.* 2012:5791–5801.
 17. (a) Sessler JL, Cho DG, Stepien M, Lynch V, Waluk J, Yoon ZS, Kim D. *J Am Chem Soc.* 2006; 128:12640–12641. [PubMed: 17002345] (b) Katayev EA, Sessler JL, Khrustalev VN, Ustyniuk YA. *J Org Chem.* 2007; 72:7244–7252. [PubMed: 17705541]
 18. (a) Jiang QQ, Darhkijav B, Liu H, Wang F, Li Z, Jiang YB. *Chem An Asian J.* 2010; 5:543–549. (b) Liu WX, Yang R, Li AF, Li Z, Gao YF, Luo XX, Ruan YB, Jiang YB. *Org Biomol Chem.* 2009; 7:4021–4028. [PubMed: 19763306]
 19. (a) Gibson D, Dey KR, Fronczek FR, Hossain MA. *Tetrahedron Lett.* 2009; 50:6537–6539. [PubMed: 20526453] (b) Saeed MA, Wong BM, Fronczek FR, Venkatraman R, Hossain MA. *Cryst Growth Des.* 2010; 10:1486–1488. (c) Mendy JS, Saeed MA, Fronczek FR, Powell DR, Hossain MA. *Inorg Chem.* 2010; 49:7223–7225. [PubMed: 20690729]
 20. Saeed MA, Thompson JJ, Fronczek FR, Hossain MA. *CrystEngComm.* 2010; 12:674–676. [PubMed: 20228881]
 21. (a) Mendy JS, Pilate M, Horne T, Day VW, Hossain MA. *Chem Commun.* 2010; 46:6084–6086. (b) Saeed MA, Fronczek FR, Powell DR, Hossain MA. *Tetrahedron Lett.* 2010; 51:4233–4236. [PubMed: 20640221] (c) Rhaman MM, Ahmed L, Wang J, Powell DR, Leszczynski J, Hossain MA. *Org Biomol Chem.* 2014; 12:2045–2048. [PubMed: 24554233]
 22. Saeed MA, Pramanik A, Hossain MA. *Inorg Chem Commun.* 2012; 21:32–24. [PubMed: 23874145]
 23. Schneider HJ, Kramer R, Simova S, Schneider U. *J Am Chem Soc.* 1988; 110:6442–6448.
 24. Hynes MJ. *J Chem Soc Dalton Trans.* 1993:311–312.
 25. Hossain MA, I klan M, Pramanik A, Saeed MA, Fronczek FR. *Cryst Growth Des.* 2012; 12:567–571.

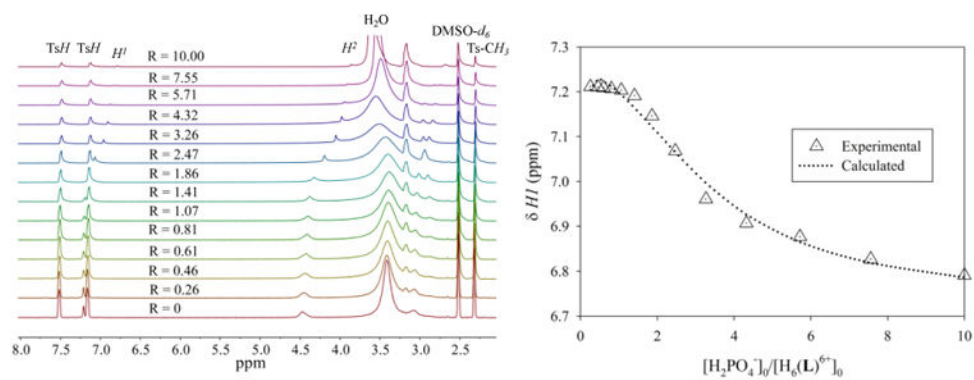


Figure 1.

(a) ^1H NMR spectra of $[\text{H}_6\text{L1}] (\text{Ts})_6$ (2 mM) with an increasing amount of TBAH_2PO_4 (20 mM) in $\text{DMSO-}d_6$. (b) Titration curve of $[\text{H}_6\text{L1}] (\text{Ts})_6$ with TBAH_2PO_4 showing the change in the chemical shift of Ar-H(H1) against an increasing concentration of phosphate in $\text{DMSO-}d_6$.

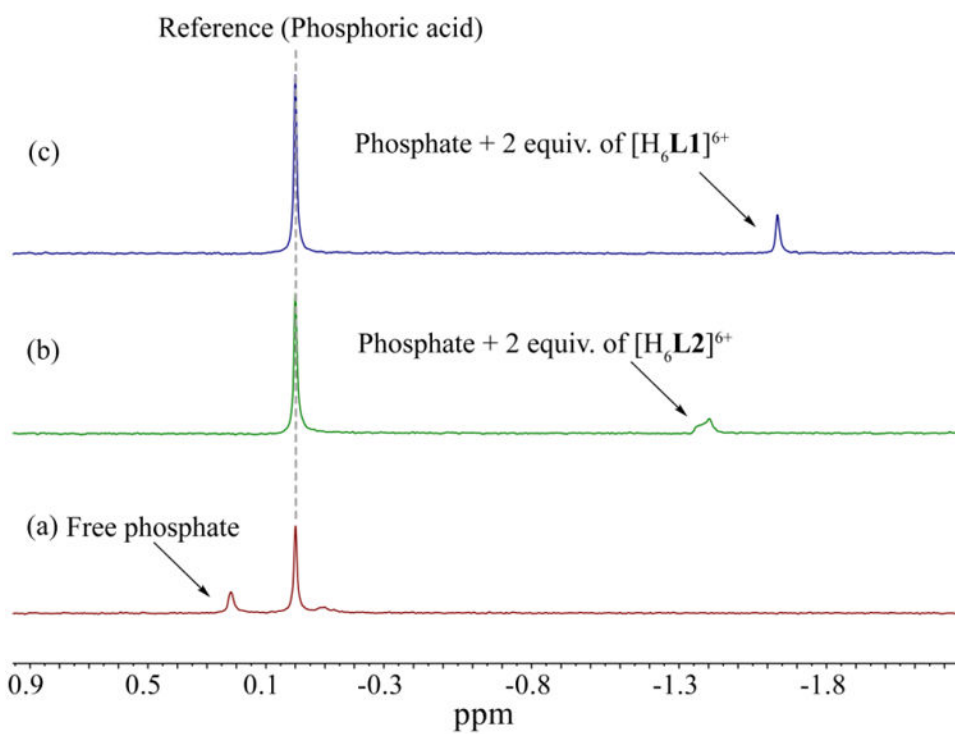


Figure 2. ^{31}P NMR spectra of $n\text{-Bu}_4N^+H_2PO_4^-$ in D_2O recorded at room temperature (a) free $TBAH_2PO_4$ ($\delta_P = 0.218$ ppm) (10 mM) (b) $TBAH_2PO_4 + 2$ equivalents of $[H_6L2] (Ts)_6$ ($\delta_P = -1.404$ ppm) and (c) $TBAH_2PO_4 + 2$ equivalents of $[H_6L1] (Ts)_6$ ($\delta_P = -1.6357$ ppm)

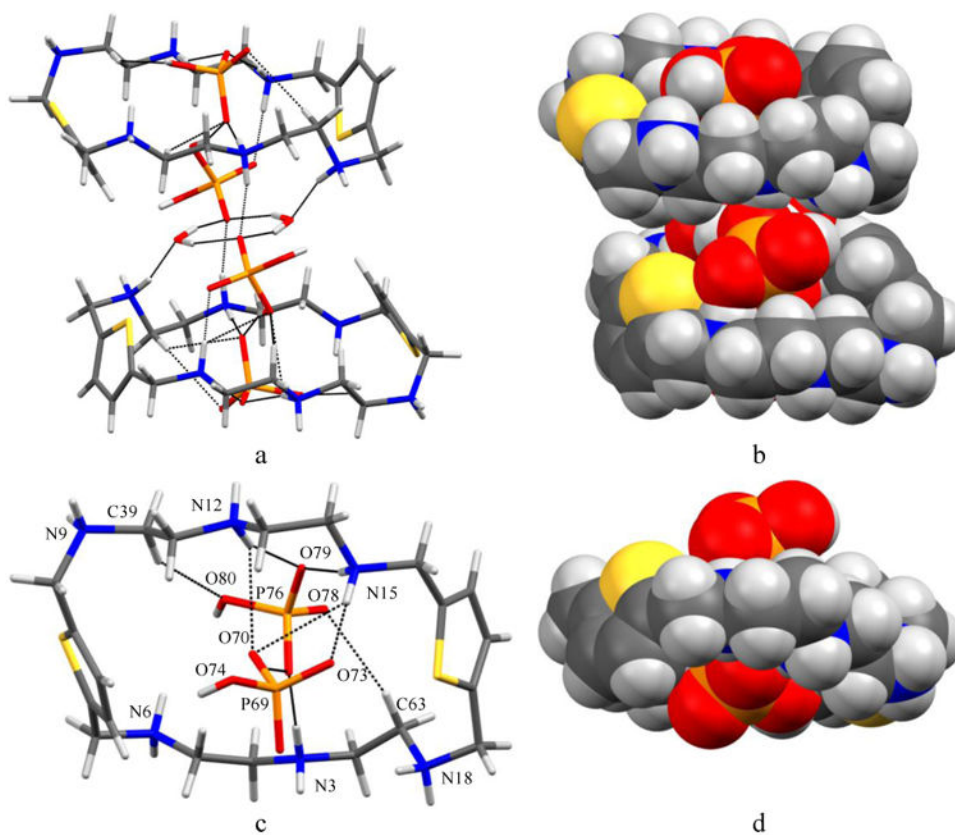
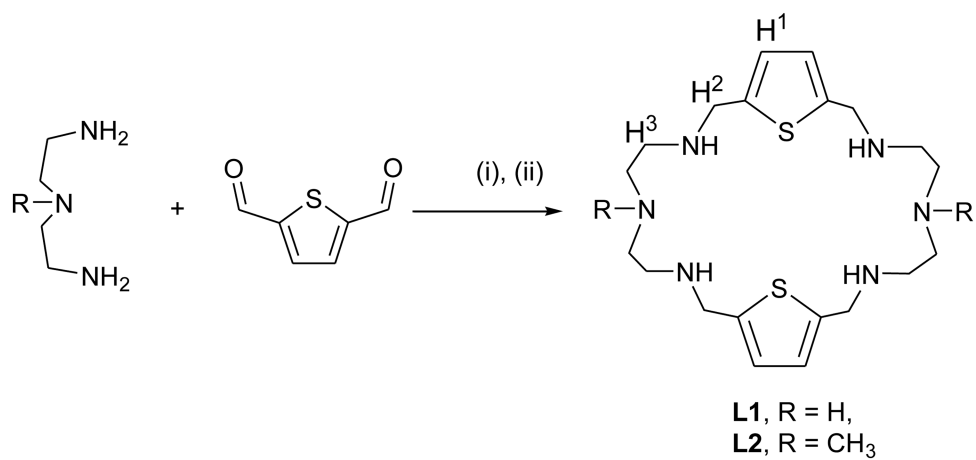


Figure 3. Top: crystal structure of the phosphate complex showing $[(H_6L1)_2 \cdot (HPO_4)_2 \cdot (H_2PO_4)_2 \cdot (H_2O)_2]^{6+}$ moiety in capped-stick view (a), and space filling view (b). Bottom: crystal structure of the phosphate complex showing $[(H_6L1) \cdot (HPO_4) \cdot (H_2PO_4)]^{3+}$ moiety in capped-stick view (c), and space filling view (d).

**Scheme 1.**

Synthetic scheme for the macrocycles **L1** and **L2**: Reaction conditions: (i) methanol, 0 °C, 24 hr stirring; (ii) NaBH₄, methanol, overnight stirring.

Table 1Association constants of the ligands for phosphate as determined by ^1H NMR titrations.^a

| Ligand | Log <i>K</i> | |
|--------|------------------|-----------------------------|
| | D ₂ O | DMSO- <i>d</i> ₆ |
| L1 | 2.0 (1:1) | 1.54 (1:1) |
| | | 5.25 (1:2) |
| L2 | <i>b</i> | 1.92 (1:1) |
| | | 4.20 (1:2) |

^aEstimated error was less than 15%^bNo significant NMR shift was observed.

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