

miR-195 enhances the radiosensitivity of colorectal cancer cells by suppressing CARM1

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Background: microRNAs (miRNAs) can regulate the sensitivity of cancer cells to chemotherapy and radiotherapy. Aberrant expression of miR-195 has been found to be involved in colorectal cancer (CRC); however, its function and underlying mechanism in the radioresistance of CRC remains unclear.

Methods: The levels of miR-195 and CARM1 were detected by quantitative reverse transcription-polymerase chain reaction and Western blot analysis in HCT-116 and HT-29 cells, respectively. Colony survival and apoptosis were determined by clonogenic assay and flow cytometry analysis, respectively. The apoptosis-related proteins Bax, Bcl-2, and γ -H2AX were detected using Western blot. The targets of miR-195 were identified by bioinformatic prediction and luciferase reporter assays. CRC cells in vitro and in vivo were exposed to different doses of X-ray radiations.

Results: miR-195 was downregulated, and CARM1 was upregulated in HCT-116 and HT-29 cells. miR-195 overexpression or CARM1 knockdown suppressed colony survival, induced apoptosis, promoted expression of Bax and γ -H2AX, and inhibited Bcl-2 expression in CRC cells. CARM1 was identified and validated to be a functional target of miR-195. Moreover, restored expression of CARM1 reversed the enhanced radiosensitivity of CRC cells induced by miR-195. Furthermore, miR-195 increased the sensitivity of CRC cells to radiation in vivo.

Conclusion: miR-195 enhances radiosensitivity of CRC cells through suppressing CARM1. Therefore, miR-195 acts as a potential regulator of radioresistance for CRC cells and as a promising therapeutic target for CRC patients.

Keywords: miR-195, CARM1, colorectal cancer, radiosensitivity

Introduction

Colorectal cancer (CRC) is the most common malignant tumor, and it is one of the major causes of cancer deaths.¹ The total rate of CRC risk is generally about 5%, and the 5-year survival rate remains at around 50%.² To date, surgery is a mainstay of treatment for CRC; however, surgery alone is insufficient, due to the fact that CRC is often diagnosed at an advanced stage.³ Although preoperative treatment with chemoradiotherapy in combination with conventional surgery was developed, it failed to universally improve outcomes as a result of chemoresistance and radioresistance. Therefore, there is urgent need to improve radiosensitivity in patients with CRC and understand the mechanisms involved in radiation resistance of CRC.

Coactivator-associated arginine methyltransferase 1 (CARM1), a protein with arginine-specific histone methyltransferase activity, was initially identified as an SRC-2 binding protein in a yeast two-hybrid screening, and it functioned as a coactivator for many nuclear receptors and transcription factors.^{4,5} A previous study reported that CARM1 could enhance breast cancer cell migration and metastasis by promoting the methylation of BAF155.⁶ Moreover, another report found that CARM1 could increase

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the chemosensitivity of breast cancer cells by methylation of MED 12.⁷ However, the function of CARM1 on radioresistance was unclear.

microRNAs (miRNAs), a class of small noncoding RNA containing about 18–24 nucleotides, function as posttranscriptional regulators and usually lead to translational suppression or RNA degradation via specific binding to 3'-untranslated region (3'-UTR) of their target genes.^{8,9} miRNAs could serve as oncogenes or tumor suppressor genes involved in nearly all cancers.¹⁰ Recent studies have reported that some miRNAs were involved in regulating radiation response of different cancers to increase susceptibility or protect cells from radiation effect.^{11–15} For instance, Lin et al¹⁵ found that the miR-200c expression was downregulated in breast cancer cells, and miR-200c overexpression could increase the radiosensitivity of breast cancer cells by inhibiting cell proliferation, and increasing apoptosis and DNA double-strand breaks through directly targeting TANK-binding kinase 1.

miR-195 expression varies from one cancer to another, and its functional effect is also diverse in different malignancies.^{16,17} In addition, aberrant expression of miR-195 was found in CRC cells. For example, miR-195 was downregulated in CRC cells, and restoration of miR-195 reduced cell viability, promoted cell apoptosis, and suppressed tumorigenicity by directly targeting Bcl-2.¹⁸ Moreover, Zhu et al¹⁹ illustrated that overexpression of miR-195 increased the radiosensitivity of breast cancer cells through inhibiting Bcl-2. However, the role and molecular mechanism of miR-195 in CRC radioresistance is still unclear.

In the present study, we aimed to investigate the effect of miR-195 on the radiosensitivity of CRC cells and its underlying molecular mechanisms.

Materials and methods

Cell culture and transfection

Normal human colonic epithelial cell line (HCEpiC) and human CRC cell lines (HCT-116 and HT-29) were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and cultured at 37°C in humidified 5% CO₂. All cells were grown in RPMI 1640 (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (FBS; Gibco), 100 IU/mL penicillin (Invitrogen, Carlsbad, CA, USA), and 100 µg/mL streptomycin (Invitrogen). miR-control, miR-195 mimics, and miR-195 inhibitor (anti-miR-195) were purchased from Dharmacon (Lafayette, CO, USA). Empty pcDNA vector (pcDNA-control), small interfering RNA against CARM1 (si-CARM1), and its negative control (si-control) were obtained from Dharmacon.

Cells (1×10⁴/well) were inoculated in a 96-well plate and cultured for 24 h so that they could attach to the bottom of the well, and then transfection was performed using Lipofectamine 2000 (Invitrogen) in accordance with the protocol provided by the manufacturer. Cells were used for further analysis 48 h post transfections.

Quantitative reverse transcription-polymerase chain reaction (qRT-PCR)

Total RNA was extracted from HCEpiC, HCT-116, and HT-29 cells with TRIzol Reagent (Invitrogen). M-MLV reverse transcriptase (Invitrogen) was used to synthesize specific cDNA according to the instruction provided by the manufacturer. The expression of miR-195 and CARM1 mRNA was quantified using the SYBR Green qRT-PCR master mix on ABI PRISM 7900HT Sequence Detection System (Applied Biosystems, Foster City, CA, USA). U6 snRNA and β-actin served as endogenous controls. The primer sequences were as follows: for miR-195 5'-CGTAGCAGCACAGAAAT-3' (forward) and 5'-GTGCAGGGTCCGAGGT-3' (reverse); for CARM1 5'-TTGATGTTGGCTGTGGCTCTGG-3' (forward) and 5'-ATGGGCTCCGAGATGATGATGTCC-3' (reverse); for U6 5'-CTCGCTTCGGCAGCACACA-3' (forward) and 5'-AACGCTTCACGAATTTGCGT-3' (reverse); for β-actin 5'-GCGAGAAGATGACCCAGATC-3' (forward) and 5'-CGGTTGGCCTTAGGGTTCAGGGGGG-3' (reverse). The relative expression of mRNAs was calculated and normalized to the control gene using the 2^{-ΔΔCt} method.

Western blot analysis

The primary antibody, anti-CARM1 (1:1,000; sc5418), was purchased from Upstate Biotechnology (Lake Placid, NY, USA). Anti-Bax (1:1,000; clone 6A7), anti-Bcl-2 (1:1,000; Cat sc-509), and anti-γ-H2AX (1:1,000; Ser139) were acquired from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Anti-β-actin (1:10,000; A1978) was obtained from Sigma-Aldrich (Louis, MO, USA). All these antibodies are monoclonal antibodies. Cells were harvested and total proteins were isolated with RIPA reagents (Thermo Scientific, Rockford, IL, USA), according to the manufacturer's instruction. Subsequently, protein concentrations were detected using the BCA kit (Pierce, Rockford, IL, USA). Western blot analysis was performed following previous protocols.²⁰ Briefly, protein specimens were isolated by a 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to polyvinylidene fluoride membranes (Millipore, Billerica, MA, USA). After blocking in Tris Buffered saline with Tween (TBST) buffer with 5% BSA for 2 h at 37°C, the

membranes were incubated with primary antibodies overnight at 4°C. Following washing in TBST, the membranes were further incubated with secondary antibody labeled with horseradish peroxidase for 1 h at room temperature. The proteins were visualized using the Image Quant software (Molecular Dynamics, Sunnyvale, CA, USA).

Clonogenic assay

Cells (2×10^5 /well) were inoculated in 6-well plates, and transfection was carried out at 24 h. Forty-eight hours post transfection, cells were seeded into 100-cm dishes and treated with variable doses (0, 2, 4, 6, and 8 Gy) of X-rays. After incubation for 14 days, colonies formed were stained with 1% crystal violet, and colonies consisting of more than 50 cells were counted. Plate efficiency was calculated using the following formula: (number of colonies/number of cells inoculated) \times 100%. Survival fraction was normalized to the plating efficiency of control groups. The data were fitted to linear-quadratic model using Sigmaplot software (Systat, San Jose, CA, USA).

Flow cytometry

Transfected cells were irradiated with a dose of 4 Gy and cultured for 24 h. Then the cells were harvested by trypsinization, washed twice with cold phosphate-buffered saline, and suspended in 500 mL of binding buffer and stained using 5 μ L Annexin V/FITC and 5 μ L PI (Sigma, CA, USA) for 15 min at room temperature in the dark. Stained cells were identified and analyzed using a FACScan flow cytometer (Becton Dickinson, CA, USA). The flow cytometry data were analyzed using BD FACS Diva software V6.1.3 (BD Biosciences, San Jose, CA, USA).

Luciferase reporter assays

The wild CARM1 3'-UTR and mutated CARM1 3'-UTR (a site-directed mutagenesis of miR-195 binding site introduced in CARM1 3'-UTR) were amplified and cloned into downstream of the luciferase gene in a pGL3 control plasmid. The constructed vectors were named as WT-CARM1-PGL3 and MUT-CARM1-PGL3, respectively. HCT-116 and HT-29 cells co-transfected with WT(MUT)-CARM-PGL3 and miR-control or miR-195 mimics and cultured for 48 h. Then, luciferase activities were measured using the Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA), according to the protocol provided by the manufacturer.

Tumor formation in nude mice

A total of 5×10^6 HT-29 cells transfected with miR-195 or miR-control were subcutaneously injected into 4-

6-week-old female BALB/c nude mice (n=6 per group). The mice were exposed to a dose of 8 Gy X-rays when the tumor volume was about 100 mm³. The tumor volumes were measured with calipers every 5 days after radiation according to the formula: volume = $0.5 \times \text{length} \times \text{width}^2$. At Day 20 after radiation, mice were sacrificed and imaged, and tumor weight was detected. All animal procedures were performed with the approval of the Local Medical Experimental Animal Care Commission. All animal-handling procedures were performed according to the *Guide for the Care and Use of Laboratory Animals* of the National Institutes of Health and followed the guidelines of the *Animal Welfare Act*.

Statistical analysis

All analyses were performed using SPSS 19.0 software (IBM, Armonk, NY, USA). The data were expressed as mean \pm SD. The Student's *t*-test or one-way ANOVA was used to evaluate the statistical significance. A *P*-value of less than 0.05 was considered statistically significant.

Results

miR-195 is downregulated and CARM1 is upregulated in CRC cell lines

The level of expression of miR-195 and CARM1 in human corneal epithelial cells (HCEpiC) and CRC cell lines (HCT-116 and HT-29) was measured using qRT-PCR and Western blot. miR-195 expression was significantly lower in HCT-116 and HT-29 cells compared with HCEpiC cells (Figure 1A). CARM1 protein levels were significantly higher in HCT-116 and HT-29 cells compared with HCEpiC cells (Figure 1B). These results suggested that miR-195 and CARM1 may be involved in the pathogenesis of CRC.

miR-195 enhances radiosensitivity of CRC cells

The colony survival assay is regarded as a typical standard to determine radiosensitivity.²¹ Moreover, the anti-apoptosis ability of cancer cells is closely related to radioresistance. To examine whether miR-195 affected radiosensitivity of CRC cells, HCT-116 and HT-29 cells transfected with miR-195 mimics or miR-control were exposed to different doses of ionizing radiation (0, 2, 4, 6, and 8 Gy). The results of clonogenic assay indicated that overexpression of miR-195 significantly reduced the survival fractions of cellular colony at each dose (2, 4, 6, and 8 Gy) in HCT-116 (Figure 2A) and HT-29 cells (Figure 2B) compared with controls. Furthermore, cells transfected with miR-195 mimics or miR-control were treated with a dose of 4 Gy

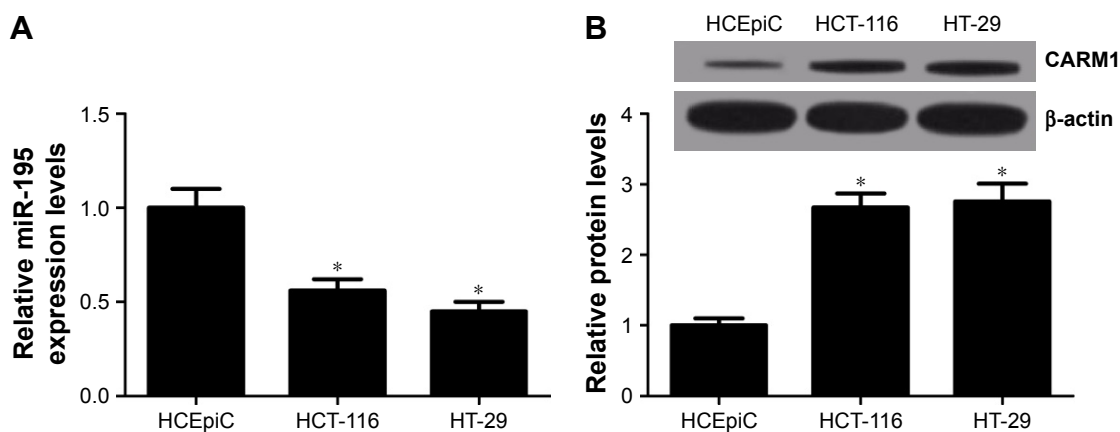


Figure 1 The expression of miR-195 and CARM1 in CRC cell lines HCT-116 and HT-29.

Notes: (A) qRT-PCR analysis of the expression levels of miR-195 in HCT-116, HT-29 cells, and HCEpiC cells. (B) The protein level of CARM1 was detected and quantified in HCT-116, HT-29, and HCEpiC cells by Western blot. * $P < 0.05$ vs control group.

Abbreviations: qRT-PCR, quantitative reverse transcription-polymerase chain reaction; CRC, colorectal cancer.

ionizing radiation. Flow cytometry analysis showed that miR-195 overexpression strikingly increased the apoptosis rates of HCT-116 (Figure 2C and D) and HT-29 cells (Figure 2E and F), and the effect was more pronounced in the presence of ionizing radiation. Western blot was performed to detect the expression of apoptosis-related proteins Bax, Bcl-2, and γ -H2AX. The results demonstrated that miR-195 overexpression increased the levels of Bax and γ -H2AX, and reduced Bcl-2 expression in HCT-116 (Figure 2G) and HT-29 cells (Figure 2H), and the effect was much stronger when combined with ionizing radiation. Taken together, these results revealed that miR-195 enhanced sensitivity of CRC cells to radiation treatment.

CARM1 knockdown enhances radiosensitivity of CRC cells

To confirm the effect of CARM1 on radiosensitivity of CRC cells, HCT-116 and HT-29 cells transfected with si-control or si-CARM1 were treated with various doses of ionizing radiation (0, 2, 4, 6, and 8 Gy), respectively. The clonogenic assay indicated that the CARM1 knockdown by si-CARM1 significantly reduced the survival fractions of cellular colony at each dose (2, 4, 6, and 8 Gy) in HCT-116 (Figure 3A) and HT-29 cells (Figure 3B) compared with controls. Western blot analysis showed that radiation treatment could lower the level of CARM1 in HCT-116 and HT-29 cells (Figure 3C and D). Moreover, flow cytometry analysis showed that a combination of si-CARM1 and radiation treatment resulted in a higher apoptosis rate than a single treatment (si-CARM1 or radiation) in HCT-116 (Figure 3E) and HT-29 cells (Figure 3F). Furthermore, CARM1 knockdown by si-CARM1 upregulated the level of Bax and γ -H2AX expression, and downregulated Bcl-2 expression in HCT-116

(Figure 3G) and HT-29 cells (Figure 3H), and the effect was much stronger when co-treated with ionizing radiation. These results suggested that CARM1 knockdown enhanced sensitivity of CRC cells to radiation treatment.

CARM1 is a direct target of miR-195 in CRC cells

miRNAs exert functional effects mainly by base-pairing with the complementary sequence of their targets. An miRNA target analysis tool, TargetScan, was used to predict the possible targets of miR-195. The results showed that the binding site of miR-195 was in the 3'-UTR of CARM1 (Figure 4A). Then dual-luciferase reporter assay was used to identify the relationship between CARM1 and miR-195. Results revealed that luciferase activity in HCT-116 and HT-29 cells co-transfected with WT-CARM1-PGL3 and miR-195 was strikingly decreased compared with controls, while cells co-transfected with MUT-CARM1-PGL3 and miR-195 showed no obvious changes in luciferase activity (Figure 4B and C). Moreover, miR-195 inhibition reduced the luciferase activity of WT-CARM1-PGL3, but had no effect on the luciferase activity of MUT-CARM1-PGL3 (Figure 4D and E). To further confirm whether miR-195 can affect the expression of CARM1, mi-control, miR-195, or anti-miR-195 were transfected into cells to investigate the mRNA and protein levels of CARM1 using qRT-PCR and Western blot. Results showed that enforced expression of miR-195 significantly reduced the protein level of CARM1, whereas miR-195 downregulation increased the CARM1 expression in HCT-116 and HT-29 cells (Figure 4G and H). However, it was found that there were no obvious differences in CARM1 mRNA level among miR-195, anti-miR-195, and miR-control groups (Figure 4F). Taken together, these results validated that miR-195 targetedly

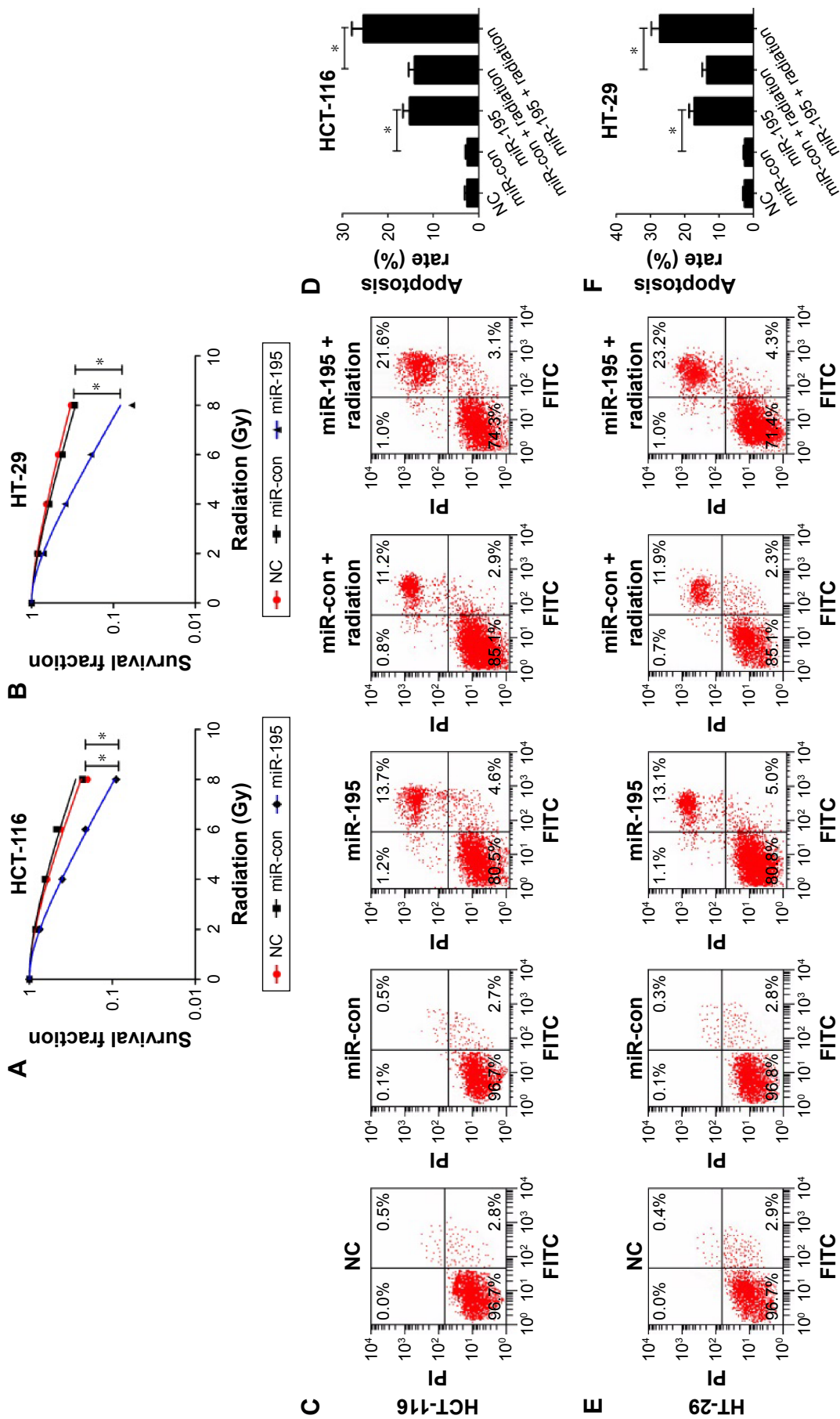


Figure 2 (Continued)

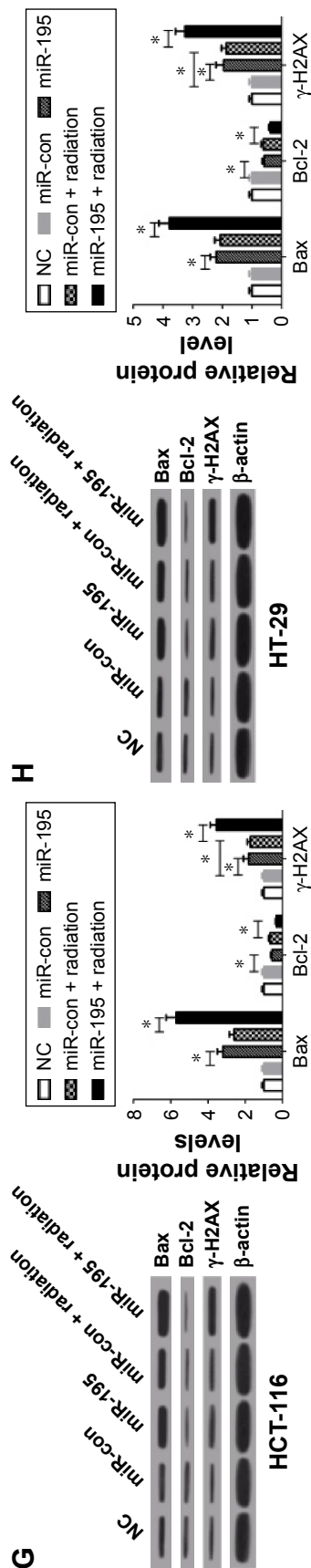


Figure 2 miR-195 mediates radiosensitivity of CRC cells. **Notes:** HCT-116 and HT-29 cells were transfected with miR-control (miR-con) or miR-195 mimics at 24 h of incubation. Survival fractions were calculated in transfected HCT-116 (A) and HT-29 cells (B) after treatment with 0, 2, 4, 6, and 8 Gy of ionizing radiation for 24 h. Cell apoptosis was detected by flow cytometry in transfected HCT-116 (C and D) and HT-29 (E and F) cells at 24 h after treatment with or without ionizing radiation (4 Gy). FITC, fluorescein isothiocyanate. The levels of apoptosis-related proteins Bax, Bcl-2, and γ -H2AX were measured by Western blot in transfected HCT-116 (G) and HT-29 cells (H) after treatment with or without ionizing radiation (4 Gy) for 24 h. * $p < 0.05$ vs miR-control. **Abbreviations:** NC, negative control; PI, propidium iodide; FITC, fluorescein isothiocyanate; CRC, colorectal cancer.

regulated CARM1 expression at the posttranscriptional level in HCT-116 and HT-29 cells.

Restoration of CARM1 expression relieves the enhanced radiosensitivity of CRC cells mediated by miR-195

To further elucidate whether miR-195 enhanced radiosensitivity of CRC cells through regulating CARM1, HCT-116 and HT-29 cells were transfected with miR-control or miR-195 mimics or co-transfected miR-195 mimics with pcDNA-control or pcDNA-CARM1. Western blot analysis showed that pcDNA-CARM1 could rescue the reduced level of CARM1 by miR-195 overexpression in HCT-116 and HT-29 cells (Figure 5A), which suggested that the rescue assay was successful. Transfected cells were irradiated with 0, 2, 4, 6, and 8 Gy X-rays for clonogenic assay or a single dose of 4-Gy X-ray for apoptosis assay. The clonogenic assay and flow cytometry demonstrated that ectopic expression of CARM1 by pcDNA-CARM1 significantly relieved the reduced survival fractions of cellular colony and the increased apoptosis triggered by miR-195 in HCT-116 and HT-29 cells (Figure 5B and C). In addition, Western blot manifested that restoration of CARM1 significantly reversed the effects of miR-195 on Bax, Bcl-2, and γ -H2AX expression in HCT-116 (Figure 5D) and HT-29 cells (Figure 5E). These results indicated that miR-195 enhanced radiosensitivity of CRC cells by targeting CARM1.

Effect of miR-195 on radiosensitivity of xenografted tumor in vivo

To investigate the role of miR-195 on radiosensitivity in a mouse xenotransplant model of colon tumor, HT-29 cells overexpressing miR-195 or miR-control were injected subcutaneously into nude mice. The mice were treated with a single dose of 8-Gy X-rays when the tumor volume was about 100 mm³. The body weight and tumor volume were measured every 5 days after radiation. As shown in Figure 6A, no significant difference was observed in the body weight among different groups (Figure 6A). Exposed to the same dose of radiation (8 Gy), the weight (Figure 6C) and size (Figure 6B and D) of xenografts derived from miR-195-overexpressed cells were dramatically decreased compared with those derived from miR-control treated cells. These results suggested that miR-195 increased the radiosensitivity of CRC cells in vivo.

Discussion

Radiotherapy is an important clinical treatment for patients with CRC. However, radioresistance is a challenging

obstacle to this therapy. Therefore, more and more research studies should be conducted to deal with this issue. Radioresistance is involved in many complicated processes, including autophagy,²² angiogenesis,²³ cancer stem cells,²⁴ aberrant activation of signaling pathways,²⁵ and abnormal expressions of DNA repair proteins.²⁶ Several previous studies have reported that miRNAs could increase

or decrease radiosensitivity of various tumors,²⁷⁻²⁹ which suggests that miRNAs may play vital roles in radiation resistance. Moreover, miRNAs could regulate radiation sensitivity and resistance of CRC.³⁰⁻³² Zhang et al³² found that miR-630 mediated by CREB could enhance radiosensitivity in CRC cell lines by inducing cell apoptosis and death through targeting BCL2L2 and TP53RK. miR-124 could

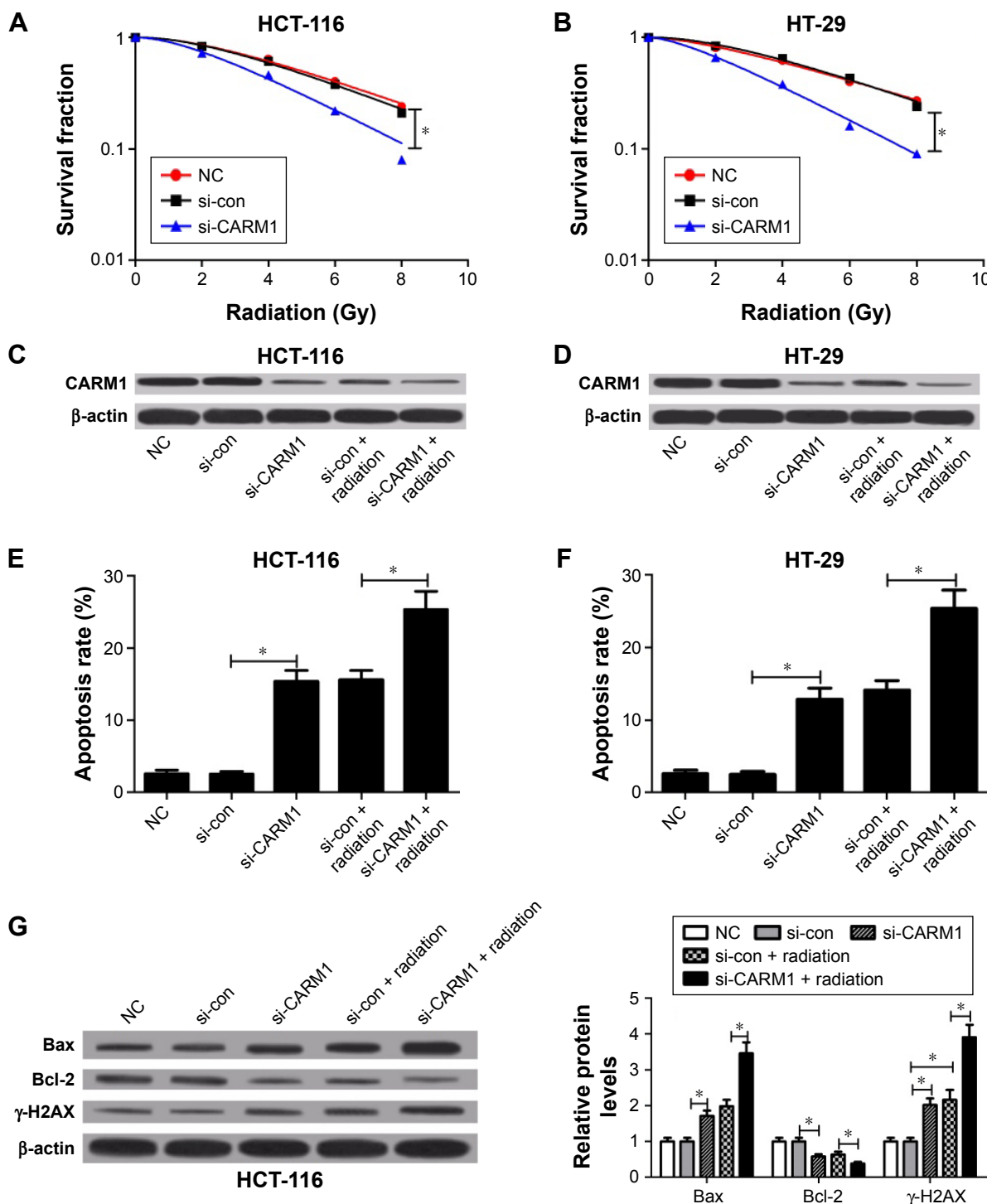


Figure 3 (Continued)

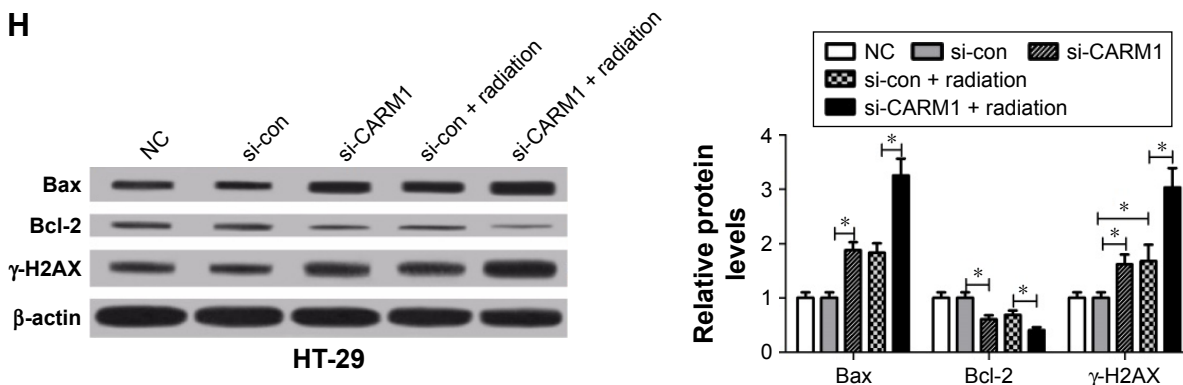


Figure 3 Effect of CARM1 knockdown on the radiosensitivity of CRC cells.
Notes: HCT-116 and HT-29 cells were transfected with si-control (si-con) and si-CARM1 at 24 h of incubation. Survival fractions were calculated in transfected HCT-116 (A) and HT-29 cells (B) after treatment with 0, 2, 4, 6, or 8 Gy of ionizing radiation for 24 h. The CARM1 level was detected using Western blot in HCT-116 (C) and HT-29 cells (D) after treatment with or without ionizing radiation (4 Gy) for 24 h. Cell apoptosis was detected by flow cytometry in transfected HCT-116 (E) and HT-29 (F) cells after treatment with or without ionizing radiation (4 Gy) for 24 h. The levels of apoptosis-related proteins Bax, Bcl-2, and γ -H2AX were determined by Western blot in transfected HCT-116 (G) and HT-29 cells (H) after treatment with or without ionizing radiation (4 Gy) for 24 h. * $P < 0.05$ vs si-control.
Abbreviations: NC, negative control; CRC, colorectal cancer.

increase the radiosensitivity of CRC cells by suppressing the expression of PRRX1, and further reverse epithelial mesenchymal transition (EMT) and simultaneously weaken the self-renewal properties of cells.³⁰ miR-100 upregulation

could radiosensitize CCL-244 cells in CRC by leading to apoptosis and DNA double-strand breaks.³³ The improvements of miRNAs in CRC radiosensitivity were consistent with our findings. In the present study, we revealed that

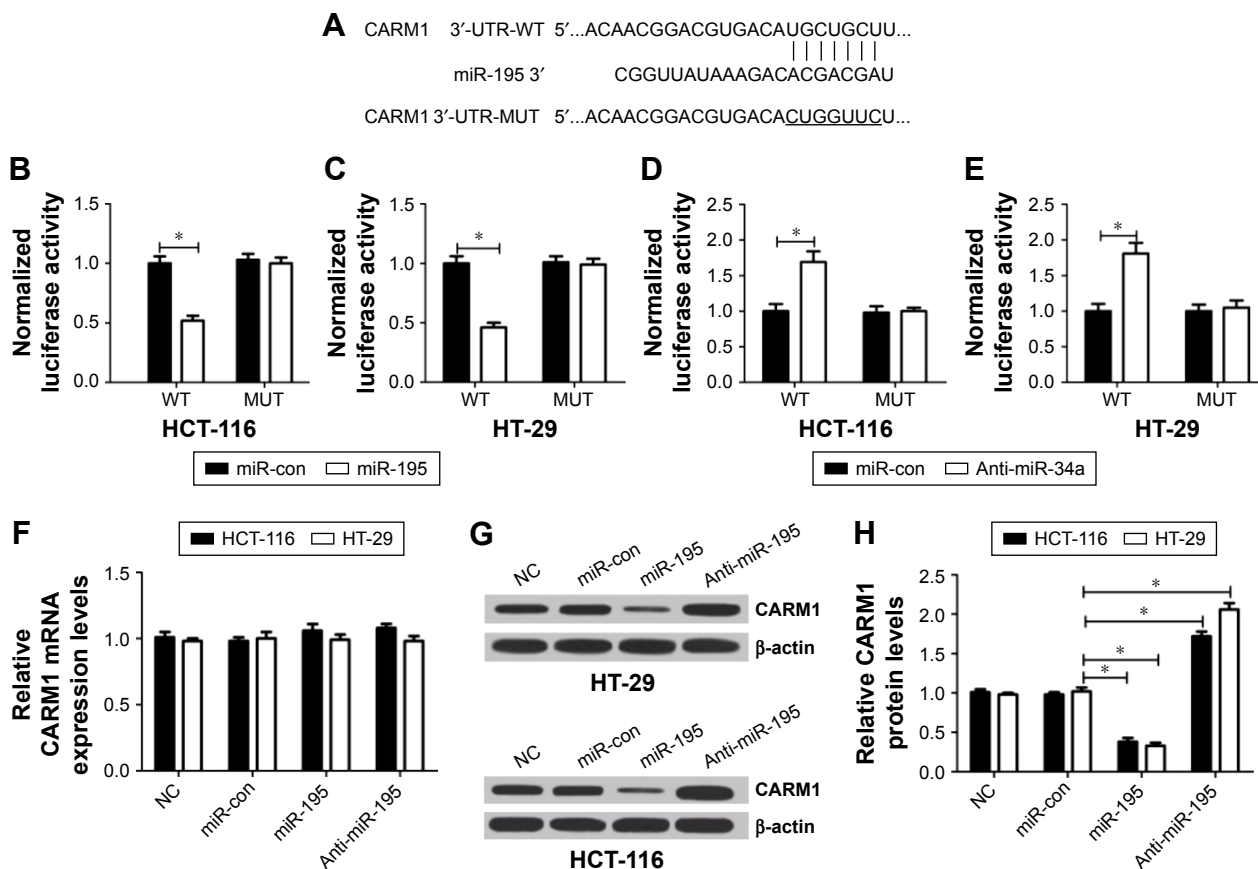


Figure 4 miR-195 directly targets 3'-UTR of CARM1 and regulates its expression in CRC cells.
Notes: (A) Wild type (WT) and mutant (MUT) 3'-UTR binding sites for miR-195 are shown. The mutated bases are labeled with a horizontal line. (B and C) The relative luciferase activity was measured in HCT-116 and HT-29 cells co-transfected WT or MUT CARM1 3'-UTR with miR-195 mimic or miR-control. (D and E) The relative luciferase activity was measured in HCT-116 and HT-29 cells co-transfected WT or MUT CARM1 3'-UTR with anti-miR-195 or anti-miR-control. The relative mRNA (F) and protein (G and H) level of CARM1 was detected in HCT-116 and HT-29 cells transfected with miR-195 mimics or anti-miR-195. * $P < 0.05$ vs miR-control.
Abbreviations: NC, negative control; CRC, colorectal cancer.

expression of miR-195 was significantly downregulated in CRC cell lines HCT-116 and HT-29 compared with HCEpiC cells. Moreover, overexpression of miR-195 could enhance the radiosensitivity of CRC, evidenced by reduced survival fractions and induced apoptosis after miR-195

overexpression. Similarly, a previous study revealed that miR-195 enhanced the radiosensitivity of breast cancer cells by inhibiting the anti-apoptosis protein BCL-2.¹⁹ This study and our study both suggested that miR-195 could modify the outcome of radiation by the induction of apoptosis. Contrary

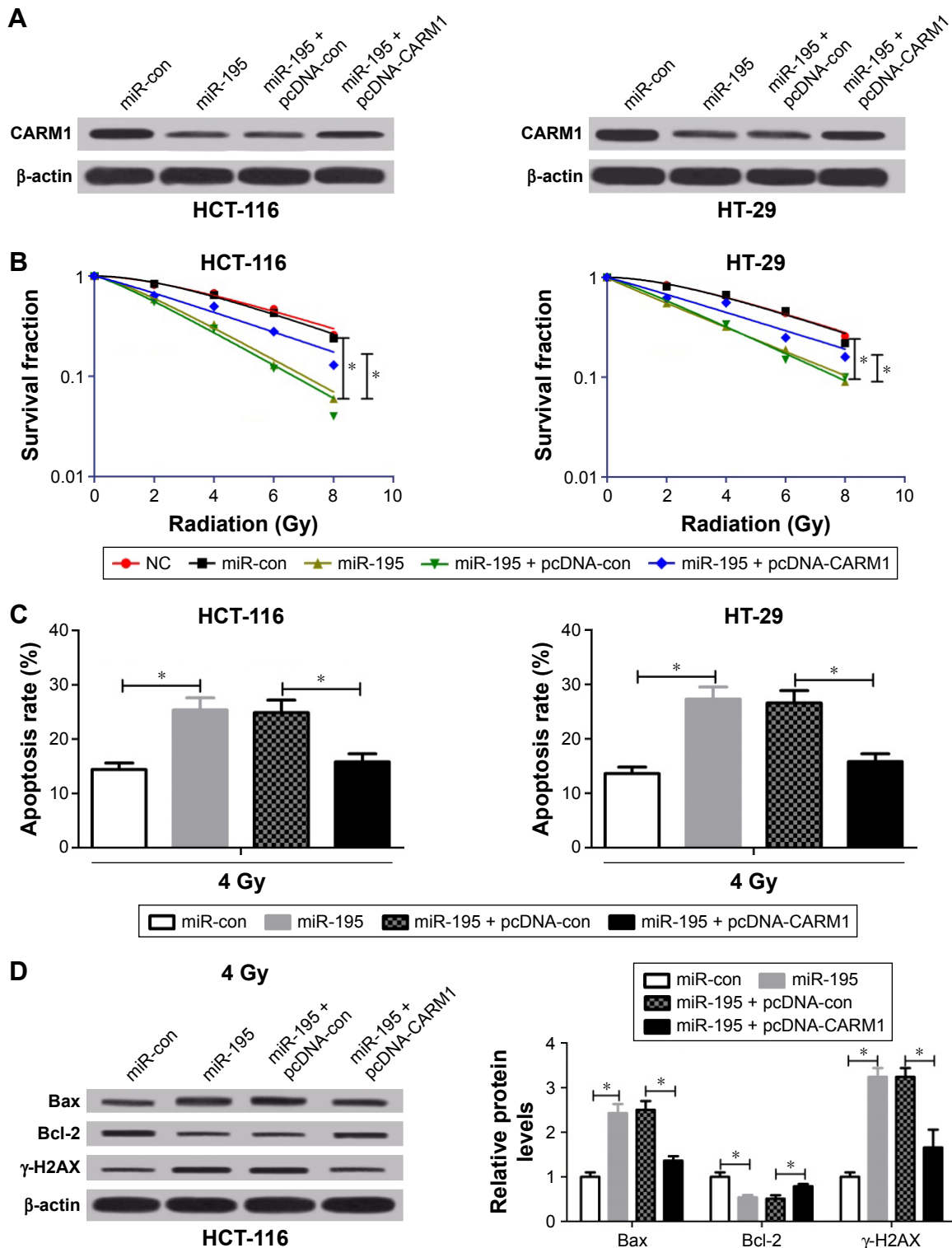


Figure 5 (Continued)

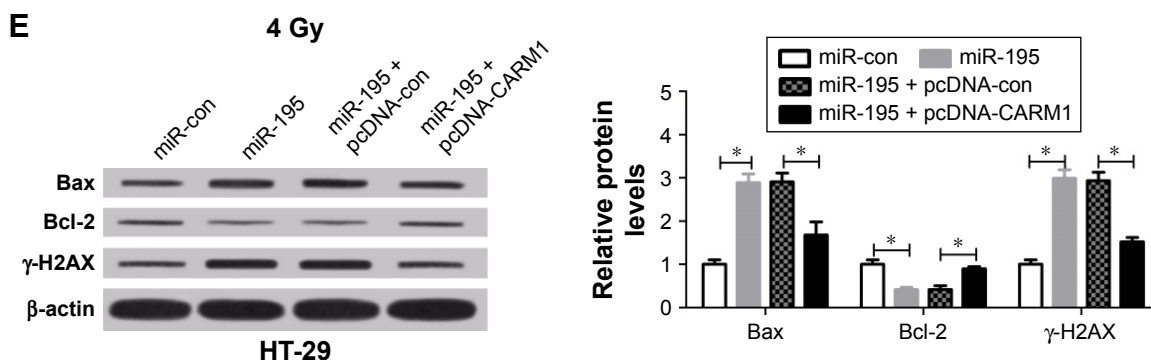


Figure 5 miR-195 enhances radiosensitivity of CRC cells by regulating CARM1. **Notes:** HCT-116 and HT-29 cells were transfected with miR-control or miR-195 mimics or co-transfected miR-195 mimics with pcDNA-control (pcDNA con) or pcDNA-CARM1. **(A)** The level of CARM1 was detected by Western blot. **(B)** Survival fractions were estimated in transfected HCT-116 and HT-29 cells after treatment with 0, 2, 4, 6, or 8 Gy of ionizing radiation. **(C)** Cell apoptosis assay of flow cytometry in transfected HCT-116 and HT-29 cells after treatment with ionizing radiation (4 Gy). Apoptosis-related proteins Bax, Bcl-2, and γ-H2AX were detected by Western blot in transfected HCT-116 **(D)** and HT-29 **(E)** cells after treatment with ionizing radiation (4 Gy). **P*<0.05 vs control groups. **Abbreviations:** NC, negative control; CRC, colorectal cancer.

to our results, Xue et al³⁴ found that miR-221 could reduce the radiosensitivity of CRC cells by regulating PTEN, negatively regulating the AKT/PKB signaling pathway. In addition, miR-106b could induce cell radioresistance in CRC by directly targeting PTEN and p21, accompanied by tumor-initiating cell capacity enhancement.³¹

A previous study indicated that CARM1 played pleiotropic roles in cell proliferation and survival in breast cancer.³⁵ A study also showed that CARM1 was highly overexpressed in CRC.³⁶ However, the role of CARM1 in radiosensitivity of cancers is unknown. In the present study, we found that knockdown of CARM1 inhibited colony

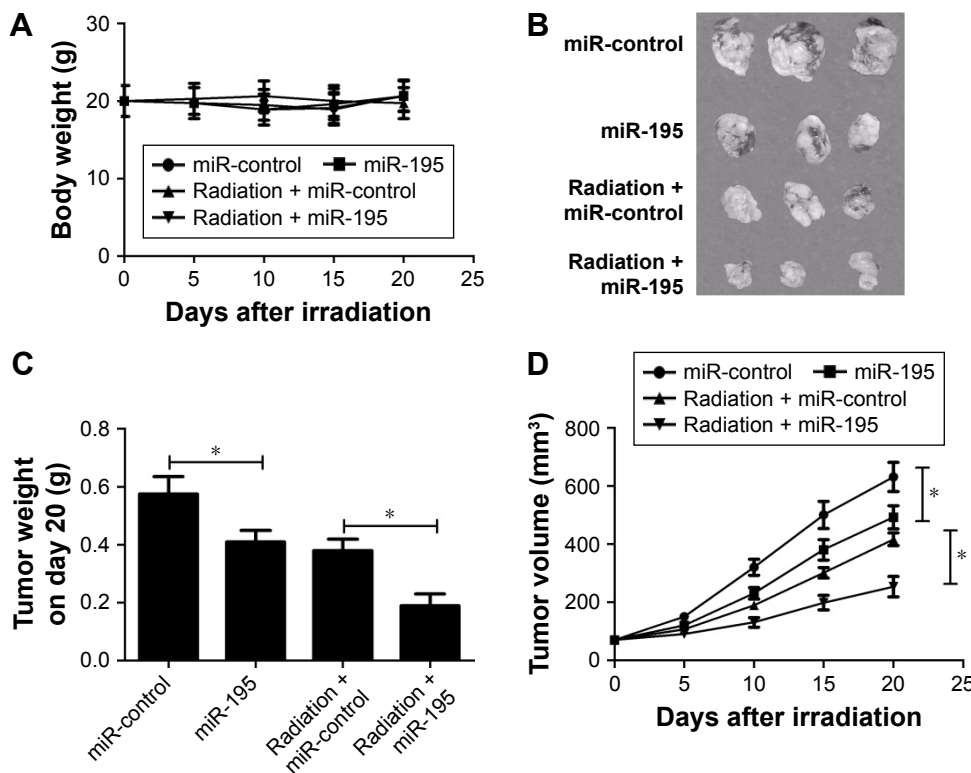


Figure 6 miR-195 upregulation increases CRC cell radiosensitivity in vivo. **Notes:** HT-29 cells overexpressing miR-195 or miR-control were subcutaneously injected into nude mice aged 4–6 weeks. Radiation was performed with 8 Gy X-rays when the tumor volume was about 100 mm³. **(A)** Body weight of the mouse was determined every 5 days after radiation in different groups. **(B)** Image of representative tumor in different groups at Day 20 after radiation. **(C)** Weight of tumor in different groups at Day 20 after radiation. **(D)** Tumor growth curve in different groups. **P*<0.05 vs control groups. **Abbreviation:** CRC, colorectal cancer.

survival and induced apoptosis. A previous study indicated that CARM1 exerted a biological effect in colon cancer cells by regulating p53 and NF- κ B target gene expression,³⁶ which were involved in the radiosensitivity of CRC cells.^{37,38} These studies suggested that CARM1 may regulate the radiosensitivity of CRC cells through p53 and NF- κ B; this needs further studies to confirm. Furthermore, a dual-luciferase assay was performed and CARM1 was verified to be a target of miR-195. The expression of CARM1 at protein levels was regulated by miR-195, and the restoration of CARM1 expression reversed the enhanced radiosensitivity of CRC cells induced by miR-195. Moreover, a previous study found a regulatory loop between CARM1 and miR-223 that controls the differentiation of CD³⁴⁺ toward the myeloid lineage.³⁹ Our findings suggest that miR-195 enhances the radiosensitivity of CRC cells by inhibiting the CARM1 expression, providing a theoretical foundation for the clinical application of miRNAs in therapy of radioresistance.

Conclusion

In summary, the expression of miR-195 was downregulated in CRC cell lines HCT-116 and HT-29. Enforced expression of miR-195 increased the radiosensitivity of CRC cells, at least partially, by inhibiting CARM1 expression. Our findings suggest that the restoration of miR-195 could be a promising therapeutic strategy for the radioresistance of CRC.

Disclosure

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