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Acute Administration of Acyl, but not Desacyl Ghrelin, Decreases Blood Pressure in Healthy Humans

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Abstract

Objective—To compare the effects of acyl ghrelin (AG) and desacyl ghrelin (DAG) on blood pressure (BP), heart rate (HR), and other autonomic parameters in healthy humans and to elucidate hormonal mechanisms through which AG could exert cardiovascular (CV) effects.

Design—17 healthy participants underwent frequent monitoring of systolic (sBP) and diastolic blood pressure (dBP), HR, respiratory rate (RR), and body surface temperature (Temp) during continuous infusion of AG, DAG, combined AG+DAG, or saline control before and during an IV-glucose tolerance test on 4 separate days. Plasma catecholamines, renin, and aldosterone levels were also measured. Differences in outcome measures between treatment groups were assessed using mixed model analysis.

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Declaration of interest

The authors have nothing to disclose.

Author contributions

C.Z. wrote the manuscript and contributed to the discussion. M.B contributed to the assay performance, the discussion and editing of the manuscript. M.A. analyzed the data. L.P. contributed to the discussion and reviewed/edited the manuscript. D.D. contributed to the discussion and reviewed/edited the manuscript. M.T. contributed to the discussion and reviewed the manuscript. J.T. designed the study, collected the data, and reviewed/edited the manuscript.

Results—Compared to the saline control, AG and combined AG+DAG infusions decreased sBP, dBP, mean arterial blood pressure (MAP), HR, and Temp. In contrast, DAG infusion did not alter BP, RR, or Temp, but did decrease HR. The AG and AG+DAG infusions also raised plasma aldosterone levels compared to saline ($p < 0.001$) without affecting renin or catecholamine levels.

Conclusions—The decrease in BP, HR, RR, and Temp with AG infusion suggests mediation through the autonomic nervous system. The lack of response to DAG suggests that these autonomic effects require activation of the ghrelin receptor.

Keywords

ghrelin; blood pressure; cardiovascular system; circulation

Introduction

Ghrelin is an orexigenic peptide, synthesized primarily in the stomach (1), which is involved in multiple physiologic functions including growth hormone secretion, energy balance, glucose homeostasis, and cardiovascular (CV) regulation (2). Following translation but prior to secretion, intracellular ghrelin is acylated at the serine-3 residue by ghrelin *O*-acyltransferase (GOAT). Both acyl (AG) and desacyl ghrelin (DAG) are released into the circulation and under most physiologic settings DAG is the more abundant of the two isoforms (3). Acylation is required for binding and activation of the growth hormone secretagogue receptor type 1a (GHSR1a), the only known receptor for ghrelin. Therefore, most of the biologic actions attributed to ghrelin are considered to be mediated by AG.

AG rises prior to meals and falls after eating with levels varying 2–3 fold between the fasting and fed states (4). As it promotes hunger and feeding behavior, AG has been used in clinical trials as therapy for disease states associated with cachexia (5,6). When given as a continuous infusion, AG also worsens insulin secretion, glucose tolerance, and insulin sensitivity (7,8), which is perhaps a protective mechanism against fasting hypoglycemia. Therefore, AG antagonists have been proposed as potential treatments for obesity and type 2 diabetes, although they have only been studied in preclinical trials to date (9). DAG is proposed to have potential beneficial effects on glucose tolerance in rodents (10), which has not been consistently duplicated in humans (11,12). Despite this variability, clinical trials with DAG analogues are ongoing (13).

Recent studies also implicate a role for AG in CV function and blood pressure (BP) control, which has been highlighted in a recent review (14). The GHSR1a is expressed in the myocardium, aorta, coronary arteries, peripheral vasculature, and regions of the central nervous system (CNS) involved in CV regulation including spinal cord, brainstem, and midbrain autonomic preganglionic neurons (15,16). GOAT is similarly widely expressed in human tissues including the myocardium (17). In healthy humans and those with chronic heart failure, acute ghrelin administration decreases systemic vascular resistance while increasing cardiac output and stroke volume index (18,19). *In vitro*, AG has direct vasodilatory effects in humans and animals through nitric oxide dependent and independent mechanism (15,20,21). AG administered centrally also decreases blood pressure in rodents, possibly through suppression of sympathetic activity (22).

As noted above, emerging evidence suggests that DAG may have physiologic effects that oppose AG and are independent of GHSR1a signaling (23). Similar to AG, DAG can exert endothelial dependent and independent vasodilatory effects on human and rodent vasculature *in vitro* (15,24). In humans, lower levels of DAG are correlated with hypertension (25) and increased cardiovascular risk (26), but the direct effect of DAG on the human CV system has not been investigated prior to this study. Here, we compare the effects of AG and DAG infusions on BP and other autonomic parameters in healthy humans. We include a coadministration (AG+DAG) treatment arm to determine if DAG potentiates or antagonizes the effects of AG on the CV system. We also evaluate the effect of AG on various BP modulating hormones to determine mechanisms by which AG may potentiate its CV actions.

Subjects and methods

Subjects

Healthy participants between the ages of 18 and 45 years with a BMI between 18 and 29 kg/m² were recruited from the greater Cincinnati area to participate in a study of the effect of ghrelin on glucose tolerance (11). The data presented here were obtained contemporaneously during this study. Subjects with a history of impaired fasting glucose or diabetes mellitus, recent myocardial infarction, congestive heart failure, active liver or kidney disease, growth hormone deficiency or excess, neuroendocrine tumor, anemia, or who were on medications known to alter insulin sensitivity were excluded. Women were studied during their follicular phase.

Study procedures were conducted at the Clinical and Translational Research Center (CTRC) at Cincinnati Children's Hospital Medical Center. Study participants provided written informed consent for the study by signing a form approved by University of Cincinnati and Cincinnati Children's Hospital Medical Center Institutional Review Boards and all data collected and reported here are in compliance with the Declaration of Helsinki.

Experimental protocol

Subjects arrived at the CTRC between 0730 and 0800 after a 10–12 hour fast on four occasions separated by at least five days. IV catheters were placed in veins of both forearms for blood sampling and infusion of test substances. The arm with the sampling catheter was placed in a 55°C chamber to arterialize venous blood. BP, HR, RR, and Temp were monitored every 15 min using GE Dash 4000 vital sign monitors during the study procedure. Mean arterial blood pressure (MAP) was calculated using the equation $MAP = \frac{2}{3} \text{ diastolic BP} - \frac{1}{3} \text{ systolic BP}$.

Blood samples for hormone measurements were placed on ice, and plasma and serum were separated by centrifugation within 1 h and stored at –80°C until assay. Blood for AG and DAG measurements was collected directly into 4 mM 4-(2-aminoethyl) benzenesulfonyl fluoride hydrochloride (AEBSF), a protease and esterase inhibitor, and 200 µL 1 N HCl was added to every milliliter of plasma.

After withdrawal of fasting blood samples, a bolus dose of synthetic human AG (CS Bio, Menlo Park, CA; 0.28 µg/kg), synthetic human DAG (CS Bio, Menlo Park, CA; 1.1 µg/kg),

combined AG+DAG, or saline was given at time 0, followed by a continuous IV infusion of AG at 1.0 µg/kg/h, DAG at 4.0 µg/kg/h, combined AG (1.0 µg/kg/h) and DAG (4.0 µg/kg/h) or saline, respectively, for a total of 210 minutes. A higher concentration of DAG was used in the study to mimic the ratio of AG:DAG found in healthy humans in the fed state (3). Based on the pharmacokinetic data from previous studies (27), ghrelin levels in the circulation were expected to reach steady state within 30 minutes of ghrelin infusion.

After 30 min of peptide/saline infusion, an IV bolus of 50% dextrose solution (11.4 g/m² body surface area) was given as the commencement of an insulin-modified frequently sampled intravenous glucose tolerance test (IVGTT) (28). Subsequently, regular insulin (0.025 units/kg body wt) was infused intravenously over 5 min, starting 20 min after the glucose injection. AG and DAG levels were measured at -15, 0, 5, 15, 25, 30, 60, 90, 150, and 210 minutes; norepinephrine (NE) and epinephrine (Epi) levels were measured at -1, 29, 30 minutes; and renin and aldosterone levels were measured at -15, 29, 46, 60, 90, and 210 minutes.

Assays

Plasma NE and Epi were measured by high performance liquid chromatography with electrochemical detection. Plasma was absorbed onto alumina at a pH of 8.6, catecholamines eluted with dilute perchloric acid, and auto-injected onto a c18 reversed-phase column. An internal standard (dehydroxylbenzylamine; DHBA) was included with each extraction to monitor recovery and standard curves for NE and Epi. Results were quantified through a chromatography data station (29,30). Plasma aldosterone and plasma active renin were measured in 10 of the 17 subjects by automated chemiluminescence immunoassays (LIAISON, DiaSorin, Saluggia, Italy) as previously described (31,32).

Statistical analysis

Differences in baseline values of the outcome variables between treatment groups were assessed with Kruskal-Wallis test using GraphPad Prism v.6. Data was analyzed using Mixed Model regression where each of the main outcomes of interest (sBP, dBP, MAP, HR, RR, Temp) was modeled as a function of treatment group, time, the interaction between time and group, and the baseline outcome measure (at time 0). Both the difference between treatment groups and the difference across time were tested using this model. Comparisons between treatment groups were assessed using differences of least squares means (LSM) analysis with p-values adjusted for multiple comparisons for using the Tukey-Kramer method as implemented in the SAS software. To assess the effect of ghrelin infusions on levels of catecholamines, aldosterone, and renin, areas under the curve (AUC) were calculated using the trapezoid method for each of these hormones and compared using the repeated measures one-way ANOVA with adjustment of multiple comparisons. All data analysis except for the baseline comparison was performed using SAS (Cary, NC). Results are expressed as mean ± SEM unless otherwise noted.

Results

Subject Characteristics

17 healthy subjects (9 men and 8 women) aged 26.6 years (SD 8.3) with BMI of 24 kg/m² (SD 3.8) completed the study; 3 other subjects enrolled but did not complete all the infusions and were not included in the analysis. Physiologic parameters (sBP, dBP, MAP, HR, Temp, RR), and hormone levels measured at baseline (before the start of infusions) did not differ within subjects on the 4 study days.

Hemodynamic & Autonomic Effects of AG and DAG

Compared to saline control, continuous AG as well as the combined AG+ DAG infusion decreased sBP (LSM -3.84 , $p < 0.0001$ and LSM -3.73 , $p < 0.0001$, respectively), dBP (LSM -3.10 , $p < 0.0001$ and LSM -2.44 , $p < 0.0001$, respectively), and MAP (LSM -3.35 , $p < 0.0001$ and LSM -2.86 , $p < 0.0001$, respectively) for the duration of the study (Table 1; Figure 1 A–C). In contrast, DAG alone had no effect on sBP (LSM 0.45 , $p = 0.437$), dBP (LSM 0.66 saline vs. DAG, $p = 0.129$), or MAP (LSM 0.59 , $p = 0.169$) compared to saline (Table 1). AG and combined AG+DAG infusion also decreased HR (LSM -3.0 , $p < 0.0001$ and LSM -2.33 , $p = 0.0001$, respectively) when compared to saline control (Table 1; Figure 1D). DAG infusion led to smaller but significant decrease in HR (LSM -1.44 saline vs. DAG, $p = 0.029$ Table 1; Figure 1D).

AG and combined AG+DAG induced a decrease in body surface temperature compared to the saline control (LSM -0.11 , $p < 0.0001$ and LSM -0.08 , $p = 0.013$, respectively), while DAG alone had no significant effect on temperature (LSM -0.01 , $p = 0.709$) (Table 1, Figure 1E). AG led to a modest decrease in RR compared to saline (LSM -0.94 , $p = 0.022$) (Table 1, Figure 1F). DAG alone and combination AG + DAG had no effect on RR compared to saline (Table 1, Figure 1F).

Significant differences for BP and Temp, but not HR or RR, were observed between the combined treatment and DAG treatment (Table 1).

Effects of AG and DAG on BP Modulating Hormone Levels

Although renin levels were not different between all drug treatment groups compared to saline, plasma aldosterone level was significantly higher during AG infusion compared to saline and DAG treatment (AG vs. Saline LSM 30.2 , $p < 0.0001$, AG vs. DAG LSM 25.4 , $p = 0.0002$) (Table 2; Figure 2). AG, DAG, and saline infusions did not affect plasma NE and Epi levels (Table 2, Figure 2). AUC values for NE, Epi, renin, and aldosterone following ghrelin and saline infusions are shown in Figure 2 (E–H). Although neither were significant, NE and Epi AUC values show a trend to be increased by AG compared to DAG and saline.

Discussion

The role of AG in the regulation of cardiovascular function has been extensively studied, but the precise mechanism for its BP lowering effect has not been elucidated. DAG has been reported to have GHSR independent effects on the CV system in rodents that are largely protective (33,34), but its effect in humans is not well understood. Our study is novel in its

use of a DAG infusion to explore the direct effect of this hormone on CV tone. We also investigated the role of AG and DAG on autonomic parameters beyond BP and HR, which have not been previously studied in humans. Finally, we attempted to determine hormonal mechanisms through which DAG and AG could exert cardiovascular effects. Interestingly, the DAG infusion had no effect on any autonomic parameter except HR. We found that AG infusion led to a decrease in sBP, dBP, and MAP, as expected, with new findings that it also lowered HR, RR, and Temp. The BP lowering effect of AG was unaffected by coadministration of DAG. Thus, our findings imply that the effect of AG on the cardiovascular system is likely dependent on GHSR receptor activation.

Several clinical studies have demonstrated decreased arterial pressure following AG given as IV bolus or infusion (19,35,36). Our study, novel in its continuous monitoring of systemic BP and HR over a 3-hour infusion period, supports these previous findings as AG infusion lowered both sBP and dBP. However, the effect was modest. The average decreases in sBP and dBP from baseline were 5.2 ± 1.7 and 5.9 ± 1.5 mmHg, respectively. We found a similarly modest, but significant decrease in HR that has been noted in some, but not all, human studies as well as rodent studies (35,37). The discrepancies in HR effects between studies may be due to differences in AG doses or mode of administration. However, in reviewing studies that found no changes in HR despite decreased BP, we do note a nonsignificant trend of lower heart rate with AG (18,19).

The precise mechanism(s) by which AG lowers blood pressure is not known but the possibilities include: (1) central effects on the sympathetic and parasympathetic nervous system (22,37), (2) peripheral effects through direct action on blood vessels (15,20), and/or (3) hormonally mediated effects on either of these systems. AG can cross the blood-brain barrier (38) and the ghrelin receptor (GHSR) is present in the autonomic preganglionic neurons (16) and the nucleus tractus solitarius (NTS) (22), a brain region that regulates autonomic outflow. Direct ghrelin injection into the NTS has been shown to decrease HR and MAP in rodents (22). Peripheral ghrelin administration can suppress cardiac sympathetic nervous activity, increase parasympathetic activity, and prevent early left ventricular remodeling in rats after myocardial infarction (39). Similarly in humans, peripheral AG bolus injection decreases parameters of sympathetic nervous activity while increasing measures of parasympathetic nervous activity during Holter monitoring (35). Our findings of simultaneous decreases in BP and HR suggest that AG decreases sympathetic and/or increases parasympathetic outflow to the CV system. The negative effect of AG on catecholamine secretion observed in the current study could be due to the insufficient sample size. With the decrease in BP, we would expect a temporary rise in catecholamine levels; however, this effect may be offset by decreased sympathetic tone, which should lower catecholamines. Although increased levels of Epi have been demonstrated by studies using AG intravenous boluses (19,36), this neutral effect of AG on Epi has been found previously in healthy humans given AG infusions at doses similar to those used in our study (7). As for the second hypothesis, although AG is also reported to act peripherally on the vasculature by decreasing tone and peripheral vascular resistance (15,20), we would expect this effect to result in increased HR along with decreased BP instead of the decreased HR that we observed.

Not much is known about the effect of ghrelin on body temperature or respiratory drive in humans. In rodents, intracerebroventricular (ICV) injection of AG (40) and DAG (41) decreases core body temperature. This effect could be mediated through decreased brown adipose tissue sympathetic nerve activity resulting in decreased thermogenesis (42) and/or activation of the hypothalamic thermoregulatory center (43). Furthermore, neuronal deletion of GHSR increases energy expenditure, locomotor activity, and thermogenesis (44). The physiological significance of the observed effect of AG on Temp needs further investigation. No studies have assessed the effects of ghrelin on RR. GHSR is expressed in brain centers that control respiration (i.e. medulla oblongata and pons) (45) and more relevant to our other findings, respiration is controlled by the autonomic nervous system. We speculate that the decreased RR with AG at least partly results from decreased sympathetic outflow or increased parasympathetic tone.

We also examined hormonal mechanisms through which AG or DAG could affect BP and HR. Changes in catecholamine levels and the renin-angiotensin-aldosterone system in response to sympathetic signaling are key mediators of short-term cardiovascular homeostasis. Previous studies have shown increased levels of Epi, ACTH, and cortisol with AG administration in humans (19,36). However, we did not observe significant changes in plasma Epi with AG infusion, nor did we see changes in NE or renin levels. DAG infusion did not affect any hormone levels in our study, which is consistent with previous findings that DAG did not alter ACTH, GH, or prolactin levels in humans (46).

However, we did observe an increase in aldosterone levels with AG infusion, a finding that has multiple potential explanations. AG is known to stimulate release of pituitary hormones including growth hormone, prolactin, and ACTH (46). We have also reported previously that AG, but not DAG, stimulates cortisol secretion (11). Although ACTH levels were not measured during this study, the elevated cortisol levels support an increase in ACTH which may have stimulated aldosterone release as well. Consistent with this hypothesis, although AG fails to stimulate aldosterone secretion from freshly isolated adrenocortical cells (47), GOAT inhibition has been shown to decrease plasma ACTH, aldosterone, and corticosterone levels *in vivo* in rats (48).

Alternatively, decreased preload resulting from the AG infusion may have decreased levels of the natriuretic peptides, atrial natriuretic peptide (ANP) and brain natriuretic peptide (BNP). The natriuretic peptides are important inhibitors of aldosterone secretion (49); thus, lower levels may have resulted in increased aldosterone. Because the natriuretic peptides can also inhibit catecholamine release as well as renin (49), we might have expected to see similar rises in these hormones. However, a study of salt-induced hypertension in rats also found no effect of chronic ghrelin administration on renin or urine catecholamine levels, despite improved BP in the ghrelin treated group (50). Although aldosterone levels were not measured in that study, ghrelin treatment did significantly decrease ANP mRNA expression in the myocardium of the high salt group (50). Additional work is needed to define the effects of AG related BP changes on natriuretic peptide and aldosterone levels.

With a clinical trial reporting beneficial effects of DAG on glucose metabolism distinct from the actions of AG (13), we sought to explore whether DAG could also exert GHSR

independent effects on BP. To our knowledge, prior investigations assessing the CV effects of DAG have been limited to animal models and *in vitro* settings. DAG was comparable to AG in reversing endothelin-1 induced vasoconstriction in isolated human coronary arteries (15) and decreasing MAP and HR when administered centrally to the NTS in rats (51). More recently, lower levels of DAG have been associated with an increased risk of CV events (26) in older hypertensive humans as well as increased BP in men with hypertension (25). We show that in humans, peripheral DAG infusion did not alter systemic BP, RR, or Temp acutely or counteract the effects of AG. Therefore, the observed ghrelin effects on the CV system are likely to be GHSR dependent. Interestingly, DAG infusion alone did lead to a modest decrease in HR compared to saline control with no significant changes in BP. This isolated bradycardic effect of DAG has not previously been seen in humans and could be mediated by DAG-specific receptors on cardiomyocytes, which have been described in mice (52). Future studies are needed to confirm this finding.

There are limitations of our study. First, BP, HR, RR, and Temp measurements were obtained as part of a study that involved an insulin-modified frequently sampled IVGTT. The IV glucose bolus (11.4 g/m²) and/or insulin infusion (0.025 units/kg) could have affected our CV parameters. However, since the IVGTT occurred on all study days, any confounding effects should have been equivalent on all days. Secondly, the hormonal measures (i.e. Epi, NE, renin, and aldosterone) were only obtained in 10 of the 17 participants. Therefore, as mentioned previously, we cannot exclude a type II error in our negative findings on catecholamine or renin secretion.

Overall, our study confirms past findings that AG infusion decreases systemic BP in healthy humans and yields additional findings of decreases in HR, RR, and Temp. We believe the most likely explanation for these CV effects is through CNS actions of AG to depress sympathetic tone and/or stimulate parasympathetic activity. Furthermore, we report the novel finding that continuous DAG infusion in humans does not significantly change BP nor does it counteract the CV effects of AG. However, DAG did mildly lower HR, which suggests a GHSR independent mechanism and may have clinical implications, as GOAT inhibitors and DAG mimetics are being actively pursued as therapeutic options for type 2 diabetes and obesity. These ongoing trials highlight the importance of understanding the effect of ghrelin on the CV system.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

1. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature*. 1999; 402:656–660. [PubMed: 10604470]
2. Müller TD, Nogueiras R, Andermann ML, Andrews ZB, Anker SD, Argente J, Batterham RL, Benoit SC, Bowers CY, Broglio F, et al. Ghrelin. *Molecular Metabolism*. 2015; 4:437–460. [PubMed: 26042199]
3. Liu J, Prudom CE, Nass R, Pezzoli SS, Oliveri MC, Johnson ML, Veldhuis P, Gordon DA, Howard AD, Witcher DR, et al. Novel ghrelin assays provide evidence for independent regulation of ghrelin acylation and secretion in healthy young men. *The Journal of Clinical Endocrinology and Metabolism*. 2008; 93:1980–1987. [PubMed: 18349056]
4. Cummings DE, Frayo RS, Marmonier C, Aubert R, Chapelot D. Plasma ghrelin levels and hunger scores in humans initiating meals voluntarily without time- and food-related cues. *American Journal of Physiology Endocrinology and Metabolism*. 2004; 287:E297–304. [PubMed: 15039149]
5. Miki K, Maekura R, Nagaya N, Nakazato M, Kimura H, Murakami S, Ohnishi S, Hiraga T, Miki M, Kitada S, et al. Ghrelin treatment of cachectic patients with chronic obstructive pulmonary disease: a multicenter, randomized, double-blind, placebo-controlled trial. *PloS one*. 2012; 7:e35708. [PubMed: 22563468]
6. Strasser F, Lutz TA, Maeder MT, Thuerlimann B, Bueche D, Tschoep M, Kaufmann K, Holst B, Brandle M, von Moos R, et al. Safety, tolerability and pharmacokinetics of intravenous ghrelin for cancer-related anorexia/cachexia: a randomised, placebo-controlled, double-blind, double-crossover study. *British Journal of Cancer*. 2008; 98:300–308. [PubMed: 18182992]
7. Tong J, Prigeon RL, Davis HW, Bidlingmaier M, Kahn SE, Cummings DE, Tschoep MH, D'Alessio D. Ghrelin suppresses glucose-stimulated insulin secretion and deteriorates glucose tolerance in healthy humans. *Diabetes*. 2010; 59:2145–2151. [PubMed: 20584998]
8. Tong J, Davis HW, Gastaldelli A, D'Alessio D. Ghrelin Impairs Prandial Glucose Tolerance and Insulin Secretion in Healthy Humans Despite Increasing GLP-1. *The Journal of Clinical Endocrinology and Metabolism*. 2016; 101:2405–2414. [PubMed: 27055279]
9. Barnett BP, Hwang Y, Taylor MS, Kirchner H, Pfluger PT, Bernard V, Lin YY, Bowers EM, Mukherjee C, Song WJ, et al. Glucose and weight control in mice with a designed ghrelin O-acyltransferase inhibitor. *Science*. 2010; 330:1689–1692. [PubMed: 21097901]
10. Gauna C, Kiewiet RM, Janssen JA, van de Zande B, Delhanty PJ, Ghigo E, Hofland LJ, Themmen AP, van der Lely AJ. Unacylated ghrelin acts as a potent insulin secretagogue in glucose-stimulated conditions. *American Journal of Physiology Endocrinology and Metabolism*. 2007; 293:E697–704. [PubMed: 17578884]
11. Tong J, Davis HW, Summer S, Benoit SC, Haque A, Bidlingmaier M, Tschoep MH, D'Alessio D. Acute Administration of Unacylated Ghrelin Has No Effect on Basal or Stimulated Insulin Secretion in Healthy Humans. *Diabetes*. 2014; 63:2309–2319. [PubMed: 24550190]
12. Benso A, St-Pierre DH, Prodam F, Gramaglia E, Granata R, van der Lely AJ, Ghigo E, Broglio F. Metabolic effects of overnight continuous infusion of unacylated ghrelin in humans. *European Journal of Endocrinology*. 2012; 166:911–916. [PubMed: 22379116]
13. Allas S, Delale T, Ngo N, Julien M, Sahakian P, Ritter J, Aribat T, van der Lely AJ. Safety, Tolerability, Pharmacokinetics and Pharmacodynamics of AZP-531, a First-in-class Analogue of Unacylated Ghrelin (UAG), in Healthy, Overweight/Obese, and Type 2 Diabetes Subjects. *Diabetes, Obesity and Metabolism*. 2016
14. Virdis A, Lerman LO, Regoli F, Ghiadoni L, Lerman A, Taddei S. Human Ghrelin: A Gastric Hormone with Cardiovascular Properties. *Current Pharmaceutical Design*. 2016; 22:52–58. [PubMed: 26581223]
15. Kleinz MJ, Maguire JJ, Skepper JN, Davenport AP. Functional and immunocytochemical evidence for a role of ghrelin and des-octanoyl ghrelin in the regulation of vascular tone in man. *Cardiovascular Research*. 2006; 69:227–235. [PubMed: 16226234]
16. Ferens DM, Yin L, Bron R, Hunne B, Ohashi-Doi K, Kitchener PD, Sanger GJ, Witherington J, Shimizu Y, Furness JB. Functional and in situ hybridization evidence that preganglionic

- sympathetic vasoconstrictor neurons express ghrelin receptors. *Neuroscience*. 2010; 166:671–679. [PubMed: 20060438]
17. Lim CT, Kola B, Grossman A, Korbonits M. The expression of ghrelin O-acyltransferase (GOAT) in human tissues. *Endocrine Journal*. 2011; 58:707–710. [PubMed: 21646729]
 18. Nagaya N, Miyatake K, Uematsu M, Oya H, Shimizu W, Hosoda H, Kojima M, Nakanishi N, Mori H, Kangawa K. Hemodynamic, renal, and hormonal effects of ghrelin infusion in patients with chronic heart failure. *The Journal of Clinical Endocrinology and Metabolism*. 2001; 86:5854–5859. [PubMed: 11739451]
 19. Nagaya N, Kojima M, Uematsu M, Yamagishi M, Hosoda H, Oya H, Hayashi Y, Kangawa K. Hemodynamic and hormonal effects of human ghrelin in healthy volunteers. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*. 2001; 280:R1483–1487.
 20. Wiley KE, Davenport AP. Comparison of vasodilators in human internal mammary artery: ghrelin is a potent physiological antagonist of endothelin-1. *British Journal of Pharmacology*. 2002; 136:1146–1152. [PubMed: 12163347]
 21. Iantorno M, Chen H, Kim JA, Tesauro M, Lauro D, Cardillo C, Quon MJ. Ghrelin has novel vascular actions that mimic PI 3-kinase-dependent actions of insulin to stimulate production of NO from endothelial cells. *American Journal of Physiology Endocrinology and Metabolism*. 2007; 292:E756–764. [PubMed: 17106060]
 22. Lin Y, Matsumura K, Fukuhara M, Kagiya S, Fujii K, Iida M. Ghrelin acts at the nucleus of the solitary tract to decrease arterial pressure in rats. *Hypertension*. 2004; 43:977–982. [PubMed: 14993197]
 23. Heppner KM, Tong J. Mechanisms in endocrinology: regulation of glucose metabolism by the ghrelin system: multiple players and multiple actions. *European Journal of Endocrinology*. 2014; 171:R21–32. [PubMed: 24714083]
 24. Moazed B, Quest D, Gopalakrishnan V. Des-acyl ghrelin fragments evoke endothelium-dependent vasodilatation of rat mesenteric vascular bed via activation of potassium channels. *European Journal of Pharmacology*. 2009; 604:79–86. [PubMed: 18957289]
 25. Gibas-Dorna M, Nowak D, Piatek J, Pupek-Musialik D, Krauss H, Kopczynski P. Plasma ghrelin and interleukin-6 levels correlate with body mass index and arterial blood pressure in males with essential hypertension. *Journal of Physiology and Pharmacology*. 2015; 66:367–372. [PubMed: 26084218]
 26. Yano Y, Nakazato M, Toshinai K, Inokuchi T, Matsuda S, Hidaka T, Hayakawa M, Kangawa K, Shimada K, Kario K. Circulating des-acyl ghrelin improves cardiovascular risk prediction in older hypertensive patients. *American Journal of Hypertension*. 2014; 27:727–733. [PubMed: 24363280]
 27. Tong J, Dave N, Mugundu GM, Davis HW, Gaylinn BD, Thorner MO, Tschop MH, D'Alessio D, Desai PB. The pharmacokinetics of acyl, des-acyl, and total ghrelin in healthy human subjects. *European Journal of Endocrinology*. 2013; 168:821–828. [PubMed: 23482590]
 28. Utzschneider KM, Tong J, Montgomery B, Udayasankar J, Gerchman F, Marcovina SM, Watson CE, Ligueros-Saylan MA, Foley JE, Holst JJ, et al. The dipeptidyl peptidase-4 inhibitor vildagliptin improves beta-cell function and insulin sensitivity in subjects with impaired fasting glucose. *Diabetes Care*. 2008; 31:108–113. [PubMed: 17909087]
 29. Goldstein DS, Feuerstein G, Izzo JL Jr, Kopin IJ, Keiser HR. Validity and reliability of liquid chromatography with electrochemical detection for measuring plasma levels of norepinephrine and epinephrine in man. *Life Sciences*. 1981; 28:467–475. [PubMed: 7207028]
 30. Anton AH, Sayre DF. A study of the factors affecting the aluminum oxide-trihydroxyindole procedure for the analysis of catecholamines. *Journal of Pharmacology and Experimental Therapeutics*. 1962; 138:360–375. [PubMed: 14013351]
 31. Manolopoulou J, Fischer E, Dietz A, Diederich S, Holmes D, Junnila R, Grimminger P, Reincke M, Morganti A, Bidlingmaier M. Clinical validation for the aldosterone-to-renin ratio and aldosterone suppression testing using simultaneous fully automated chemiluminescence immunoassays. *Journal of Hypertension*. 2015; 33:2500–2511. [PubMed: 26372319]
 32. Morganti A. European study group for the validation of DiaSorin LDRA. A comparative study on inter and intralaboratory reproducibility of renin measurement with a conventional enzymatic

- method and a new chemiluminescent assay of immunoreactive renin. *Journal of Hypertension*. 2010; 28:1307–1312. [PubMed: 20224432]
33. Pei XM, Yung BY, Yip SP, Chan LW, Wong CS, Ying M, Siu PM. Protective effects of desacyl ghrelin on diabetic cardiomyopathy. *Acta Diabetologica*. 2015; 52:293–306. [PubMed: 25192951]
34. Pei XM, Yung BY, Yip SP, Ying M, Benzie IF, Siu PM. Desacyl ghrelin prevents doxorubicin-induced myocardial fibrosis and apoptosis via the GHSR-independent pathway. *American Journal of Physiology Endocrinology and Metabolism*. 2014; 306:E311–323. [PubMed: 24326424]
35. Soeki T, Koshiba K, Niki T, Kusunose K, Yamaguchi K, Yamada H, Wakatsuki T, Shimabukuro M, Minakuchi K, Kishimoto I, et al. Effect of ghrelin on autonomic activity in healthy volunteers. *Peptides*. 2014; 62:1–5. [PubMed: 25265271]
36. Krapalis AF, Reiter J, Machleidt F, Iwen KA, Dodt C, Lehnert H, Sayk F. Ghrelin modulates baroreflex-regulation of sympathetic vasomotor tone in healthy humans. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*. 2012; 302:R1305–1312.
37. Matsumura K, Tsuchihashi T, Fujii K, Abe I, Iida M. Central ghrelin modulates sympathetic activity in conscious rabbits. *Hypertension*. 2002; 40:694–699. [PubMed: 12411464]
38. Banks WA, Tschoop M, Robinson SM, Heiman ML. Extent and direction of ghrelin transport across the blood-brain barrier is determined by its unique primary structure. *Journal of Pharmacology and Experimental Therapeutics*. 2002; 302:822–827. [PubMed: 12130749]
39. Soeki T, Kishimoto I, Schwenke DO, Tokudome T, Horio T, Yoshida M, Hosoda H, Kangawa K. Ghrelin suppresses cardiac sympathetic activity and prevents early left ventricular remodeling in rats with myocardial infarction. *American Journal of Physiology Heart and Circulatory Physiology*. 2008; 294:H426–432. [PubMed: 18024547]
40. Lawrence CB, Snape AC, Baudoin FM, Luckman SM. Acute central ghrelin and GH secretagogues induce feeding and activate brain appetite centers. *Endocrinology*. 2002; 143:155–162. [PubMed: 11751604]
41. Inoue Y, Nakahara K, Maruyama K, Suzuki Y, Hayashi Y, Kangawa K, Murakami N. Central and peripheral des-acyl ghrelin regulates body temperature in rats. *Biochemical and Biophysical Research Communications*. 2013; 430:278–283. [PubMed: 23159626]
42. Yasuda T, Masaki T, Kakuma T, Yoshimatsu H. Centrally administered ghrelin suppresses sympathetic nerve activity in brown adipose tissue of rats. *Neuroscience Letters*. 2003; 349:75–78. [PubMed: 12946556]
43. Wiedmer P, Strasser F, Horvath TL, Blum D, Dimarchi R, Lutz T, Schurmann A, Joost HG, Tschoop MH, Tong J. Ghrelin-induced hypothermia: a physiological basis but no clinical risk. *Physiology & Behavior*. 2011; 105:43–51. [PubMed: 21513721]
44. Bahn RS, Burch HB, Cooper DS, Garber JR, Greenlee MC, Klein I, Laurberg P, McDougall IR, Montori VM, Rivkees SA, et al. Hyperthyroidism and other causes of thyrotoxicosis: management guidelines of the American Thyroid Association and American Association of Clinical Endocrinologists. *Endocrine Practice*. 2011; 17:456–520. [PubMed: 21700562]
45. Zigman JM, Jones JE, Lee CE, Saper CB, Elmquist JK. Expression of ghrelin receptor mRNA in the rat and the mouse brain. *Journal of Comparative Neurology*. 2006; 494:528–548. [PubMed: 16320257]
46. Broglio F, Gottero C, Prodam F, Gauna C, Muccioli G, Papotti M, Aribat T, Van Der Lely AJ, Ghigo E. Non-acylated ghrelin counteracts the metabolic but not the neuroendocrine response to acylated ghrelin in humans. *The Journal of Clinical Endocrinology and Metabolism*. 2004; 89:3062–3065. [PubMed: 15181099]
47. Rucinski M, Ziolkowska A, Tyczewska M, Malendowicz LK. Expression of prepro-ghrelin and related receptor genes in the rat adrenal gland and evidences that ghrelin exerts a potent stimulating effect on corticosterone secretion by cultured rat adrenocortical cells. *Peptides*. 2009; 30:1448–1455. [PubMed: 19416745]
48. Rucinski M, Ziolkowska A, Szyszka M, Hochol A, Malendowicz LK. Evidence suggesting that ghrelin O-acyl transferase inhibitor acts at the hypothalamus to inhibit hypothalamo-pituitary-adrenocortical axis function in the rat. *Peptides*. 2012; 35:149–159. [PubMed: 22543218]

49. Porzionato A, Macchi V, Rucinski M, Malendowicz LK, De Caro R. Natriuretic peptides in the regulation of the hypothalamic-pituitary-adrenal axis. *International Review of Cell and Molecular Biology*. 2010; 280:1–39. [PubMed: 20797680]
50. Aoki H, Nakata M, Dezaki K, Lu M, Gantulga D, Yamamoto K, Shimada K, Kario K, Yada T. Ghrelin counteracts salt-induced hypertension via promoting diuresis and renal nitric oxide production in Dahl rats. *Endocrine Journal*. 2013; 60:571–581. [PubMed: 23328675]
51. Tsubota Y, Owada-Makabe K, Yukawa K, Maeda M. Hypotensive effect of des-acyl ghrelin at nucleus tractus solitarii of rat. *Neuroreport*. 2005; 16:163–166. [PubMed: 15671869]
52. Lear PV, Iglesias MJ, Feijoo-Bandin S, Rodriguez-Penas D, Mosquera-Leal A, Garcia-Rua V, Gualillo O, Ghe C, Arnoletti E, Muccioli G, et al. Des-acyl ghrelin has specific binding sites and different metabolic effects from ghrelin in cardiomyocytes. *Endocrinology*. 2010; 151:3286–3298. [PubMed: 20410201]

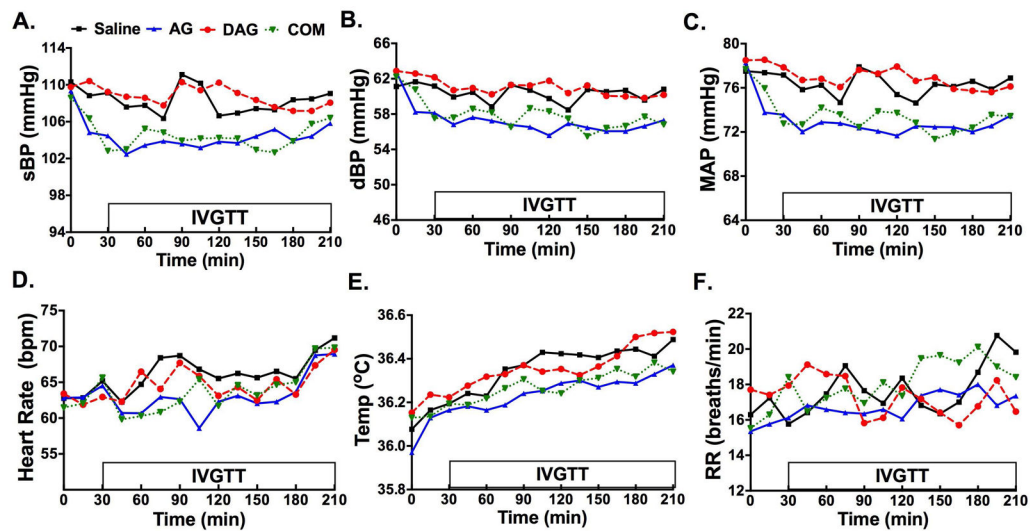


Figure 1. Blood Pressure, Heart Rate, Respiratory Rate Responses to Drug Infusion

Systolic blood pressure (sBP; Fig. 1A), diastolic blood pressure (dBP; Fig. 1B), mean arterial pressure (MAP; Fig. 1C), heart rate (HR; Fig. 1D), body surface temperature (Temp; Fig. 1E), and respiratory rate (RR; Fig. 1F) values during acyl ghrelin (AG; 1 $\mu\text{g}/\text{kg}/\text{h}$), desacyl ghrelin (DAG; 4 $\mu\text{g}/\text{kg}/\text{h}$), combined (COM) AG (1 $\mu\text{g}/\text{kg}/\text{h}$) and DAG (4 $\mu\text{g}/\text{kg}/\text{h}$), or saline infusions between 0 and 210 minutes in healthy men and women. An intravenous glucose tolerance test (IVGTT) was administered between 30 and 210 min after the peptide infusions began. Blood pressure is presented in mmHg, heart rate as beats per minute (bpm), Temp as $^{\circ}\text{C}$, and RR as breaths per minute (breaths/min).

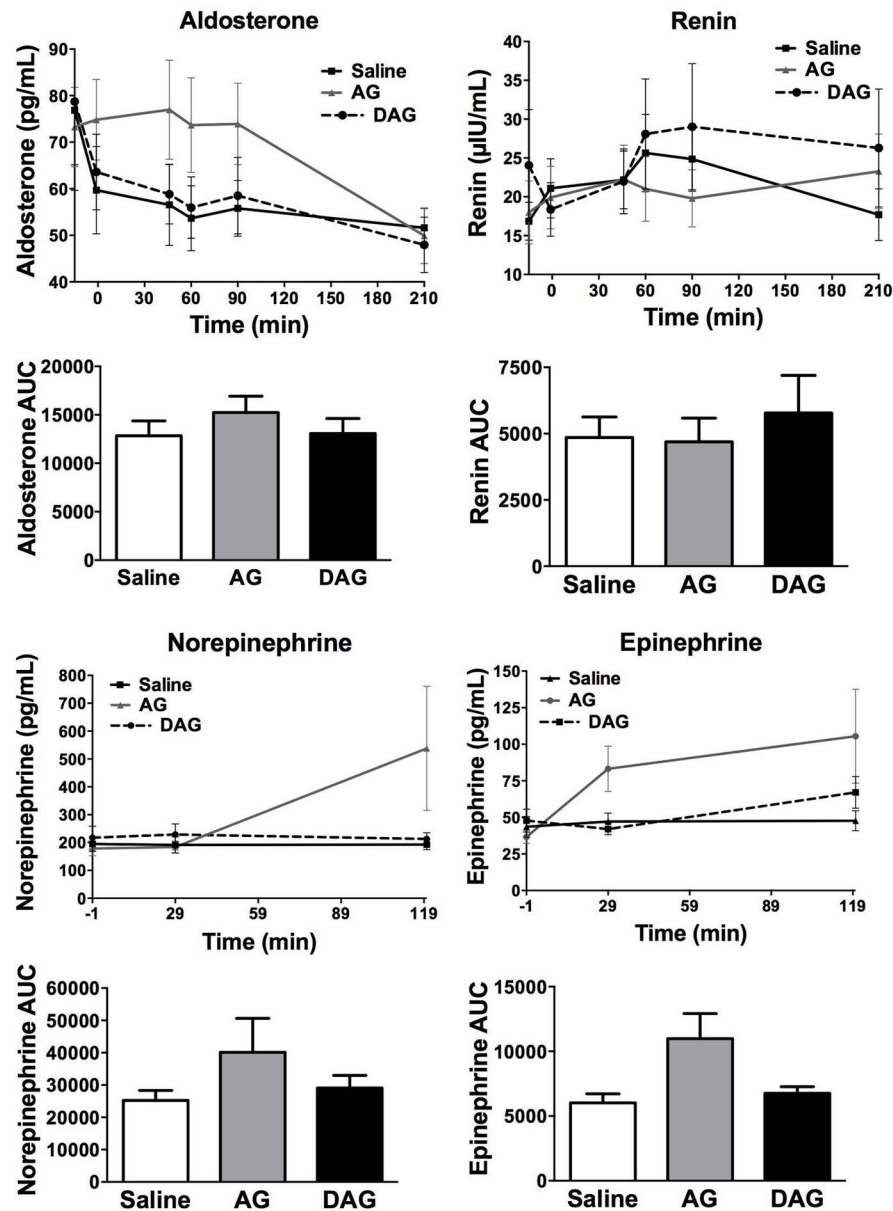


Figure 2. Study 1 Effects of AG and DAG on Catecholamines, Renin, and Aldosterone Plasma aldosterone (pg/mL; Fig. 2A), renin (μ IU/mL; Fig. 2B), norepinephrine (NE in pg/mL; Fig. 2C), and epinephrine (Epi in pg/mL; Fig. 2D) concentrations during a 210-min infusion of saline, acyl ghrelin (AG), desacyl ghrelin (DAG), or combined AG + DAG in 10 healthy men and women. The NE and Epi concentrations were only assayed until the 120 minute time point. An intravenous glucose tolerance test (IVGTT) was also performed between 30 and 210 min after the peptide infusions began. The area under the curve (AUC) is shown for NE, Epi, renin, and aldosterone.

Table 1
Comparison of Hemodynamic & Autonomic Parameters in Response to AG and DAG

Differences of Least Squares Means (LSM) comparison of systolic blood pressure (sBP), diastolic blood pressure (dBp), mean arterial pressure (MAP), heart rate (HR), temperature (Temp), and respiratory rate (RR) between acyl ghrelin (AG; 1 µg/kg/h), desacyl ghrelin (DAG; 4 µg/kg/h), combined AG (1 µg/kg/h) + DAG (4 µg/kg/h), and saline infusions in healthy men and women. Compared to saline control, AG and combined AG + DAG treatment significantly decreased sBP, dBp, and MAP ($p < 0.0001$ for sBP, dBp, and MAP). AG treatment alone decreased HR, Temp, and RR compared to saline ($p < 0.0001$ for HR and Temp, $p = 0.022$ for RR). DAG infusion had no effect on blood pressure parameters, Temp, or RR but decreased HR compared to saline control ($p = 0.029$).

Treatment	Comparison	LSM	SE	DF	T-Value	Pr > t
AG	Saline	-3.84	0.54	48	-7.05	<0001
	DAG	-4.29	0.54	48	-7.88	<0001
	Com	-0.11	0.54	48	-0.2	0.84
Systolic BP	Com	-3.73	0.54	48	-6.86	<0001
	DAG	-4.18	0.54	48	-7.7	<0001
	Saline	0.45	0.54	48	0.83	0.437
AG	Saline	-3.1	0.38	48	-8.14	<0001
	DAG	-3.76	0.38	48	-9.88	<0001
	Com	-0.66	0.38	48	-1.75	0.129
Diastolic BP	Com	-2.44	0.38	48	-6.41	<0001
	DAG	-3.1	0.38	48	-8.15	<0001
	Saline	0.66	0.38	48	1.73	0.129
AG	Saline	-3.35	0.38	48	-8.87	<0001
	DAG	-3.94	0.38	48	-10.43	<0001
	Com	-0.49	0.38	48	-1.3	0.247
MAP	Com	-2.86	0.38	48	-7.58	<0001
	DAG	-3.45	0.38	48	-9.15	<0001
	Saline	0.59	0.38	48	1.55	0.169

Treatment	Comparison	LSM	SE	DF	T-Value	Pr > t
HR	AG	-3.00	0.59	48	-5.11	<.0001
	DAG	-1.56	0.59	48	-2.66	0.019
	Com	-0.67	0.59	48	-1.14	0.301
Temp	AG	-2.33	0.59	48	-3.98	0.001
	DAG	-0.89	0.59	48	-1.52	0.173
	Com	-1.44	0.59	48	-2.45	0.029
RR	AG	-0.11	0.03	48	-4.07	<.0001
	DAG	-0.12	0.03	48	-4.47	<.0001
	Com	-0.03	0.03	48	-1.25	0.263
Temp	AG	-0.08	0.03	48	-2.83	0.013
	DAG	-0.09	0.03	48	-3.23	0.004
	Com	0.01	0.03	48	0.4	0.709
RR	AG	-0.94	0.36	48	-2.59	0.022
	DAG	-0.62	0.36	48	-1.7	0.132
	Com	-1.29	0.36	48	-3.58	0.002
Temp	AG	0.36	0.36	48	0.99	0.37
	DAG	0.68	0.36	48	1.88	0.105
	Com	-0.32	0.36	48	-0.89	0.413

* AG = acyl ghrelin, DAG = desacyl ghrelin, Com = Combined AG + DAG, BP = blood pressure, MAP = mean arterial pressure, HR = heart rate, RR = respiratory rate, Temp = body surface temperature, LSM = differences of Least Squares Means; SE = Standard Error

Table 2
Comparison of Hormone Concentrations in Response to AG and DAG

Differences of Least Squares Means (LSM) comparison of plasma norepinephrine (NE) and epinephrine between AG (1 µg/kg/h), DAG (4 µg/kg/h), and saline infusions in 10 healthy men and women. Differences of LSM comparison of renin and aldosterone levels with AG, combined AG + DAG, and saline infusion are also presented. No significant differences in NE, epinephrine, renin, and aldosterone were observed between saline control and peptide infusion treatments. AG and combined AG + DAG treatment significantly increased plasma aldosterone levels compared to control ($p < 0.0001$ for AG vs Saline; $p = 0.0002$ for AG vs Combined).

	Treatment	Comparison	LSM	SE	DF	T-Value	Pr > t
Aldosterone	AG	Saline	30.242	5.60	16	5.40	<0001
		DAG	25.440	5.20	16	4.89	0.0002
	DAG	Saline	4.802	5.59	16	0.86	0.4028
Renin	AG	Saline	2.165	2.39	16	0.91	0.3783
		DAG	3.960	2.20	16	1.80	0.0910
	DAG	Saline	-1.795	2.39	16	-0.75	0.4642
NE	AG	Saline	32.543	81.58	18	0.40	0.6946
		DAG	95.668	110.69	18	0.86	0.3988
	DAG	Saline	63.123	81.30	18	0.78	0.4476
Epi	AG	Saline	18.576	16.67	18	1.11	0.2799
		DAG	26.571	16.52	18	1.61	0.1252
	DAG	Saline	7.995	16.60	18	0.48	0.6360

* AG = acyl ghrelin, DAG = desacyl ghrelin, Com = Combined AG + DAG, NE = norepinephrine, Epi = epinephrine, DF = Degrees of freedom, LSM = differences of Least Squares Means; SE = Standard Error