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### Global change biology

## Spatio-temporal environmental variation mediates geographical differences in phenotypic responses to ocean acidification

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Phenotypic plasticity is expected to play a major adaptive role in the response of species to ocean acidification (OA), by providing broader tolerances to changes in  $pCO<sub>2</sub>$  conditions. However, tolerances and sensitivities to future OA may differ among populations within a species because of their particular environmental context and genetic backgrounds. Here, using the climatic variability hypothesis (CVH), we explored this conceptual framework in populations of the sea urchin Loxechinus albus across natural fluctuating  $pCO_2/pH$  environments. Although elevated  $pCO_2$  affected the morphology, physiology, development and survival of sea urchin larvae, the magnitude of these effects differed among populations. These differences were consistent with the predictions of the CVH showing greater tolerance to OA in populations experiencing greater local variation in seawater  $pCO<sub>2</sub>/pH$ . Considering geographical differences in plasticity, tolerances and sensitivities to increased  $pCO<sub>2</sub>$  will provide more accurate predictions for species responses to future OA.

## 1. Introduction

In an era of rapid environmental changes such as ocean acidification (OA), there is a pressing need to understand how organisms will respond to greater and less predictable variations in environmental conditions [\[1\]](#page-3-0). Intuitively, phenotypic plasticity seems a suitable strategy to cope with these changes by means of behavioural, physiological, life-history and morphological adjustments [\[2\]](#page-3-0). These plastic responses have long been recognized as important mechanisms by which organisms maximize fitness in heterogeneous environments [\[3\]](#page-3-0), facilitating the persistence of natural populations by providing broader tolerances to environmental conditions [\[4\]](#page-3-0). Despite this central role of phenotypic plasticity, standard models aimed to predict the effect of climatic change on species persistence and distribution (i.e. the climate envelope models) do not incorporate differences in plastic responses among populations [[5](#page-3-0)]. Geographical differences in plasticity may reflect contrasting selective pressures resulting from habitats with different environmental heterogeneity [[6](#page-3-0)]. In this context, the climatic variability hypothesis (CVH) offers a powerful conceptual framework with which to view the impact of future climate change (e.g. OA) on species persistence, by linking physiology, climate and biogeographic distributions [[5](#page-3-0)]. The CVH states that in more variable environments, organisms should have broader ranges of

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environmental tolerance and/or greater physiological flexibilities that enable them to cope with fluctuating environmental conditions [[7](#page-3-0)].

Environmental variability is an intrinsic characteristic of coastal ecosystems along the southern Pacific coast of South America, where spatio-temporal changes in  $CO<sub>2</sub>$ , pH, temperature, nutrients and other factors are the result of dynamic processes such as upwellings, riverine discharges and biological activity [\[8\]](#page-3-0). In this region, populations of marine organisms are exposed to natural variability in pH/  $pCO<sub>2</sub>$  with fluctuating  $CO<sub>2</sub>$ -supersaturated surface waters almost year-round in the northern section  $(18^{\circ} S - 30^{\circ} S)$ , seasonal CO<sub>2</sub>-supersaturated waters southward (30° S–39° S) and CO<sub>2</sub>-undersaturated surface waters in the southernmost section (more than  $42^{\circ}$  S) [[9](#page-3-0)]. Like in other coastal regions, this natural variability can be far greater in magnitude than the predicted change due to OA [[10](#page-3-0)], and may have prompted the evolution of a broad range of mechanisms by which coastal organisms can maintain the homeostasis for biological processes (e.g. calcification) [[11\]](#page-3-0). Homeostatic capacity and tolerances to  $CO<sub>2</sub>/pH$  changes are known to differ across geographical regions [\[12](#page-3-0)], highlighting the importance of this component in the understanding of species susceptibilities to future OA. In order to test the CVH, we explored morphophysiological and developmental responses to elevated  $pCO<sub>2</sub>$ , in natural populations of a keystone species of the Pacific coast of Chile, the sea urchin Loxechinus albus. We predict that populations experiencing greater local variation in  $CO<sub>2</sub>/pH$  (i.e. continuous and seasonal upwelling subregions) will be less susceptible to OA than populations from less variable  $CO<sub>2</sub>/pH$  environments.

### 2. Material and methods

Adult sea urchins were collected from nine localities along the upwelling system and fjords of the Pacific coast of Chile, spanning approximately 4500 km (electronic supplementary material, figure S1). Animals were maintained in flowing seawater aquaria (13–14 $^{\circ}$ C) and fed *ad libitum* with kelp until experiments. Gametes were obtained by standard methods [\[13\]](#page-3-0) within 2-3 days after collection. From each locality, 20 independent crosses of a single male with pooled eggs of three females were developed to avoid male– female incompatibility. Eggs were fertilized using filtered  $(0.1 \mu m)$  seawater at ambient conditions (13 $\degree$ C and 390  $\mu$ atm  $pCO_2$ ) and distributed in the experimental  $pCO<sub>2</sub>$  treatments (current global: 390  $\mu$ atm and projected OA: 1200  $\mu$ atm [\[10,14](#page-3-0)]) with controlled temperature (13–14 $^{\circ}$ C) at a concentration of approximately 0.7 embryos  $ml^{-1}$ . Each cross was cultured separately in two replicate buckets for each treatment, in which the seawater carbonate chemistry was maintained using a semi-automatic flow-through  $CO<sub>2</sub>$ mixing system [\[15\]](#page-3-0), modified following [\[16\]](#page-3-0) (electronic supplementary material, table S1). Embryos were sampled daily to record developmental progression (DP) and survival until early pluteus (4 arms, 80–84 h) when physiological (metabolic rate (MR) by oxygen uptake- $VO<sub>2</sub>$ ) and morphometric (total larval length and postoral arm length) analyses were performed (see the electronic supplementary material). Traits were analysed using linear mixed models in the 'lme4' package of R v.3.3 (R Core Team, 2016), with  $CO<sub>2</sub>$  treatment and subregion as fixed factors and locality as random factor. Significance tests were performed with the 'lmerTest' package. Post hoc comparisons for mixed effects models were done with the 'multcomp' and 'lsmeans' packages. DP curves were fitted and analysed with GraphPad Prism software (GraphPad, San Diego, CA, USA).

### 3. Results

There were significant effects of high  $pCO<sub>2</sub>$  on total larval length (TL:  $F_{1,348} = 32.4$ ,  $p < 0.05$ ), postoral arm length (POL:  $F_{1,348} = 221.7$ ,  $p < 0.05$ ), size-corrected MR ( $F_{1,348} =$ 35.8,  $p < 0.05$ ), DP ( $F_{1,1792} = 11.35$ ,  $p < 0.05$ ) and survival  $(F<sub>1,348</sub> = 494.9, p < 0.05)$  of L. albus. These effects varied among subregions (significant interaction) for all of the morpho-physiological ( $X_{\text{LRT}}^2 > 15$ ,  $p < 0.05$ ) and developmental traits (electronic supplementary material, table S2). Although POL, survival and DP were significantly affected  $(F_{2,348} = 9.03, F_{2,348} = 33.1 \text{ and } F_{20,1776} = 66.2; p < 0.05) \text{ by}$ elevated  $pCO<sub>2</sub>$  in populations from the three subregions (figures [1](#page-2-0) and [2](#page-3-0)), the major negative effects were detected in those within the  $CO<sub>2</sub>$ -undersaturated subregion (post hoc  $t_{\rm ratio} < -12$ ,  $p < 0.05$ ). These three populations were the only ones with negative effects of high  $pCO<sub>2</sub>$  on TL and MR (post hoc  $t_{\text{ratio}} = -6.5$  and  $-4.4$ , respectively,  $p < 0.05$ ; [figure 1](#page-2-0)a). Under low  $pCO<sub>2</sub>$  conditions, most of the phenotypic traits did not differ among populations and subregions (TL:  $F_{2,177} = 1.08$ ,  $p = 0.34$ ; POL:  $F_{2,177} = 1.12$ ,  $p = 0.32$ ; MR:  $F_{2,177} = 1.92$  $F_{2,177} = 1.92$  $F_{2,177} = 1.92$ ,  $p = 0.15$ ; survival:  $F_{2,177} = 2.23$ ,  $p = 0.11$ ; figures 1 and [2](#page-3-0)). However, DP differed geographically  $(F_{8,864} = 6.634,$  $p < 0.05$ ; [figure 2](#page-3-0)), showing a faster rate in the southernmost population (figures [1](#page-2-0) and [2](#page-3-0)a).

### 4. Discussion

This study highlights the role of naturally fluctuating  $pCO<sub>2</sub>/$ pH environments in determining geographical differences in phenotypic responses to projected OA. Although elevated  $pCO<sub>2</sub>$  affected larval morphology, physiology, development and survival, the magnitude of these effects differed among the three main subregions. The lack of clinal trends in phenotypic responses to simulated OA suggests that geographical differences in average  $pCO<sub>2</sub>$  are not driving differences in tolerances among sea urchin populations. Instead, phenotypic differences in L. albus were consistent with the predictions of the CVH, showing greater tolerance to OA in populations experiencing greater local variation in seawater  $CO<sub>2</sub>/pH$ [[7,17](#page-3-0)]. Total larval length, postoral arm length (a proxy of larval calcification [\[18](#page-4-0)]), DP and survival were less affected (approx.  $3\%$  change) in populations within the  $CO<sub>2</sub>$ -supersaturated subregions (i.e. continuous and seasonal; 18-39° S) than in populations from the CO<sub>2</sub>-undersaturated subregion (approx.  $9\%$  change; more than  $40^{\circ}$  S). This geographical pattern of phenotypic responses may result from spatial differences in OA-induced energetic costs for maintenance, growth and survival under elevated  $pCO<sub>2</sub>$  conditions [\[19](#page-4-0)]. In fact, larval MR was only affected by acidified seawater in populations within the  $CO<sub>2</sub>$ -undersaturated subregion, supporting the idea that changes in energy allocation can be the main drivers of the negative effects of OA in L. albus [[18](#page-4-0),[19](#page-4-0)]. Under present-day  $pCO<sub>2</sub>$  conditions, larvae of L. albus showed similar morpho-physiological characteristics among populations. Nonetheless, the faster growth rate observed in the southernmost population is likely due to the rearing temperature, which was slightly higher (approx.  $2^{\circ}C$ ) than the local conditions during its spawning/growing season (11 $^{\circ}$ C) [\[13\]](#page-3-0).

From a theoretical perspective, geographical differences in tolerances and sensitivities to high  $pCO<sub>2</sub>$  may reflect contrasting selective pressures along the spatial distribution of L. albus [\[6\]](#page-3-0). Within the  $CO<sub>2</sub>$ -supersaturated subregions,

<span id="page-2-0"></span>

Figure 1. Geographical responses in (a) total larval length, (b) postoral arm length and (c) size-corrected metabolic rate under  $pC_2$  treatments. Mean  $\pm$  s.d. S. Ant., St Antonio; P., Punta. (Online version in colour.)

local selection may have promoted highly plastic phenotypes with broad tolerance to pH variation [\[20\]](#page-4-0), and greater fitness (e.g. survival) under high  $pCO<sub>2</sub>$  conditions [[21](#page-4-0)] in comparison with phenotypes from the  $CO<sub>2</sub>$ -undersaturated subregion. Similar geographical differences in performance and fitness

have been documented in other marine species along upwelling systems (e.g. [[12](#page-3-0)[,22](#page-4-0)]), suggesting that regional differences in carbonate chemistry may have acted as selective pressures maintaining phenotypic and genetic variation necessary for adaptive responses to changing pH [[21,22](#page-4-0)]. Although gene 3

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<span id="page-3-0"></span>

Figure 2. Effect of ocean acidification on (a) developmental progression and (b) survival of sea urchin larvae. Mean  $+$  s.d.

flow can impede the adaptive divergence in populations of L. albus[\[20](#page-4-0)], local adaptation may still occur if phenotype-specific mortality occurs after larval dispersal [\[23\]](#page-4-0).

In a projected OA scenario, the stronger negative effects on larval growth and developmental dynamics in populations from the  $CO<sub>2</sub>$ -undersaturated region can lead to prolonged pelagic larval duration, increasing the susceptibility to predation and reducing the number of settlers due to the high mortality in the plankton [[18\]](#page-4-0). For these populations, greater shifts in skeletal morphology caused by OA may influence larval feeding and their ability to disperse [\[24](#page-4-0)], affecting the energy transfer across trophic levels and potentially influencing population dynamics and predator –prey interactions in marine food webs [14]. In conclusion, our study reinforces the importance of considering geographical differences in plasticity, tolerances and

sensitivities to increased  $pCO<sub>2</sub>$  for predicting species responses to future OA.

Ethics. This study was conducted under approval of the UACH animal ethics committee no. 056/12.

Data accessibility. Data are available as the electronic supplementary material.

Authors' contributions. J.D.G.-E. and L.D.B. conceived the study, carried out analyses and drafted the manuscript. J.D.G.-E., P.A.V. and J.M.N. developed experiments. J.L. and R.T. analysed water chemistry. All authors approved the final version of the manuscript and agree to be held accountable for the content.

Competing interests. We have no competing interests.

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### **References**

- 1. Botero C, Weissing F, Wright J, Rubenstein D. 2015 Evolutionary tipping points in the capacity to adapt to environmental change. Proc. Natl Acad. Sci. USA 112, 184– 189. [\(doi:10.1073/pnas.1408589111](http://dx.doi.org/10.1073/pnas.1408589111))
- 2. Piersma T, van Gils J. 2011 The flexible phenotype. A body-centred integration of ecology, physiology and behaviour. New York, NY: Oxford University Press.
- 3. Pigliucci M. 2001 Phenotypic plasticity. In Evolutionary ecology: concepts and case studies (eds CW Fox, DA Roff, DJ Fairbairn), pp. 58 – 69. Oxford, UK: Oxford University Press.
- 4. Gabriel W, Luttbeg B, Sih A, Tollrian R, Gabriel W, Luttbeg B, Sih A, Tollrian R. 2005 Environmental tolerance, heterogeneity, and the evolution of reversible plastic responses. Am. Nat. 166, 339– 353. ([doi:10.1086/432558](http://dx.doi.org/10.1086/432558))
- 5. Molina-Montenegro M, Naya D. 2012 Latitudinal patterns in phenotypic plasticity and fitness-related traits: assessing the climatic variability hypothesis (CVH) with an invasive plant species. PLoS ONE 7, 23 – 28. ([doi:10.1371/journal.pone.0047620](http://dx.doi.org/10.1371/journal.pone.0047620))
- 6. Gaita´n-Espitia JD, Marshall D, Dupont S, Bacigalupe LD, Bodrossy L, Hobday AJ. 2017 Geographical gradients in selection can reveal genetic constraints for evolutionary responses to ocean acidification. Biol. Lett. 13, 20160784. [\(doi:10.1098/rsbl.2016.0784\)](http://dx.doi.org/10.1098/rsbl.2016.0784)
- 7. Janzen D. 1967 Why mountain passes are higher in the tropics. Am. Nat. 101, 233 – 249. [\(doi:10.1086/](http://dx.doi.org/10.1086/282487) [282487](http://dx.doi.org/10.1086/282487))
- 8. Vargas C, Contreras P, Perez C, Sobarzo M, Saldias G, Salisbury J. 2016 Influences of riverine and upwelling waters on the coastal carbonate system off Central Chile and their ocean acidification implications. J. Geophys. Res. 121, 1468– 1483. [\(doi:10.1002/2015JG003213\)](http://dx.doi.org/10.1002/2015JG003213)
- 9. Torres R et al. 2011 Air-sea  $CO<sub>2</sub>$  fluxes along the coast of Chile: from  $CO<sub>2</sub>$  outgassing in central northern upwelling waters to  $CO<sub>2</sub>$  uptake in southern Patagonian fjords. J. Geophys. Res. 116, C09006. ([doi:10.1029/2010JC006344\)](http://dx.doi.org/10.1029/2010JC006344)
- 10. Hofmann G et al. 2011 High-frequency dynamics of ocean pH: a multi-ecosystem comparison. PLoS ONE 6, e28983. ([doi:10.1371/journal.pone.](http://dx.doi.org/10.1371/journal.pone.0028983) [0028983\)](http://dx.doi.org/10.1371/journal.pone.0028983)
- 11. Hendriks I, Duarte C, Olsen Y, Steckbauer A, Ramajo L, Moore T, Trotter J, McCulloch M. 2015 Biological mechanisms supporting adaptation to ocean acidification in coastal ecosystems. Estuar. Coast. Shelf Sci. 152, A1–A8. [\(doi:10.1016/j.ecss.2014.](http://dx.doi.org/10.1016/j.ecss.2014.07.019) [07.019](http://dx.doi.org/10.1016/j.ecss.2014.07.019))
- 12. Padilla-Gamiño J, Gaitán-Espitia J, Kelly M, Hofmann G. 2016 Physiological plasticity and local

adaptation to elevated  $pCO<sub>2</sub>$  in calcareous algae: an ontogenetic and geographic approach. Evol. Appl. 9, 1043– 1053. ([doi:10.1111/eva.12411\)](http://dx.doi.org/10.1111/eva.12411)

- 13. Lawrence J. 2013 Sea urchins: biology and ecology, 3rd edn. Amsterdam, The Netherlands: Academic Press.
- 14. Byrne M. 2011 Impact of ocean warming and ocean acidification on marine invertebrate life history stages: vulnerabilities and potential for persistence in a changing ocean. Oceanogr. Mar. Biol. Annu. Rev. 49, 1 – 42. [\(doi:10.1201/b11009-6](http://dx.doi.org/10.1201/b11009-6))
- 15. Torres R, Manriquez PH, Duarte C, Navarro JM, Lagos NA, Vargas CA, Lardies MA. 2013 Evaluation of a semiautomatic system for long-term seawater carbonate chemistry manipulation. Rev. Chil. Hist. Nat. 86, 443 -451. [\(doi:10.4067/S0716-078X2013000400006](http://dx.doi.org/10.4067/S0716-078X2013000400006))
- 16. Padilla-Gamiño J, Kelly M, Evans T, Hofmann G. 2013 Temperature and  $CO<sub>2</sub>$  additively regulate physiology, morphology and genomic responses of larval sea urchins, Strongylocentrotus purpuratus. Proc. R. Soc. B 280, 20130155. ([doi:10.1098/rspb.](http://dx.doi.org/10.1098/rspb.2013.0155) [2013.0155](http://dx.doi.org/10.1098/rspb.2013.0155))
- 17. Dupont S, Ortega-Martínez O, Thorndyke M. 2010 Impact of near-future ocean acidification on echinoderms. Ecotoxicology 19, 449-462. ([doi:10.](http://dx.doi.org/10.1007/s10646-010-0463-6) [1007/s10646-010-0463-6](http://dx.doi.org/10.1007/s10646-010-0463-6))
- <span id="page-4-0"></span>18. Byrne M, Lamare M, Winter D, Dworjanyn S, Uthicke S. 2013 The stunting effect of a high  $CO<sub>2</sub>$ ocean on calcification and development in sea urchin larvae, a synthesis from the tropics to the poles. Phil. Trans. R. Soc. B 368, 20120439. ([doi:10.](http://dx.doi.org/10.1098/rstb.2012.0439) [1098/rstb.2012.0439\)](http://dx.doi.org/10.1098/rstb.2012.0439)
- 19. Stumpp M, Wren J, Melzner F, Thorndyke M, Dupont S. 2011  $CO<sub>2</sub>$  induced seawater acidification impacts sea urchin larval development I: elevated metabolic rates decrease scope for growth and induce developmental delay. Comp. Biochem. Physiol. A Mol.

Integr. Physiol. 160, 331–340. [\(doi:10.1016/j.cbpa.](http://dx.doi.org/10.1016/j.cbpa.2011.06.022) [2011.06.022\)](http://dx.doi.org/10.1016/j.cbpa.2011.06.022)

- 20. Futuyma D, Moreno G. 1988 The evolution of ecological specialization. Annu. Rev. Ecol. Syst. 19, 207–233. [\(doi:10.1146/annurev.es.19.110188.001231\)](http://dx.doi.org/10.1146/annurev.es.19.110188.001231)
- 21. Pespeni M et al. 2013 Evolutionary change during experimental ocean acidification. Proc. Natl Acad. Sci. USA 110, 6937-6942[. \(doi:10.1073/pnas.](http://dx.doi.org/10.1073/pnas.1220673110) [1220673110\)](http://dx.doi.org/10.1073/pnas.1220673110)
- 22. Kelly M, Padilla-Gamiño J, Hofmann G. 2013 Natural variation and the capacity to adapt to ocean

acidification in the keystone sea urchin Strongylocentrotus purpuratus. Glob. Chang. Biol. 19, 2536– 2546. ([doi:10.1111/gcb.12251\)](http://dx.doi.org/10.1111/gcb.12251)

- 23. Marshall D, Monro K, Bode M, Keough M, Swearer S. 2010 Phenotype – environment mismatches reduce connectivity in the sea. Ecol. Lett. 13, 128-140. ([doi:10.1111/j.1461-0248.2009.01408.x\)](http://dx.doi.org/10.1111/j.1461-0248.2009.01408.x)
- 24. Chan K, Grünbaum D, O'Donnell M. 2011 Effects of ocean-acidification-induced morphological changes on larval swimming and feeding. J. Exp. Biol. 214, 3857– 3867. ([doi:10.1242/jeb.054809\)](http://dx.doi.org/10.1242/jeb.054809)