

Superparamagnetic reconstituted high-density lipoprotein nanocarriers for magnetically guided drug delivery

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Abstract: Current cancer chemotherapy is frequently associated with short- and long-term side effects, affecting the quality of life of cancer survivors. Because malignant cells are known to overexpress specific surface antigens, including receptors, targeted drug delivery is often utilized to reduce or overcome side effects. The current study involves a novel targeting approach using specifically designed nanoparticles, including encapsulation of the anti-cancer drug valrubicin into superparamagnetic iron oxide nanoparticle (SPION) containing reconstituted high-density lipoprotein (rHDL) nanoparticles. Specifically, rHDL–SPION–valrubicin hybrid nanoparticles were assembled and characterized with respect to their physical and chemical properties, drug entrapment efficiency and receptor-mediated release of the drug valrubicin from the nanoparticles to prostate cancer (PC-3) cells. Prussian blue staining was used to assess nanoparticle movement in a magnetic field. Measurements of cytotoxicity toward PC-3 cells showed that rHDL–SPION–valrubicin nanoparticles were up to 4.6 and 31 times more effective at the respective valrubicin concentrations of 42.4 µg/mL and 85 µg/mL than the drug valrubicin alone. These studies showed, for the first time, that lipoprotein drug delivery enhanced via magnetic targeting could be an effective chemotherapeutic strategy for prostate cancer.

Keywords: drug delivery, nanoparticles, rHDL, magnetic nanoparticles, SPION

Background

Despite many new approaches involving gene and immunotherapy, surgery and subsequent adjuvant chemotherapy remain the standard clinical approaches for the management of most malignancies, the second highest cause of mortality worldwide.^{1–5} The efficacy of cancer chemotherapy is often limited by physical and biological barriers.^{6–9} These include solubility, bioavailability, pharmacokinetics and biological barriers such as tumor microenvironment, hypoxic core and extracellular tumor pH, interstitial fluid pressure and many other factors.^{9–15} Nanotechnology has been invoked as a means of overcoming several of these difficulties.¹⁶ More specifically, cancer chemotherapy is frequently associated with short- and long-term side effects,^{17–20} affecting the quality of life of cancer survivors.^{21,22} To advance the state of the art in cancer therapeutics, numerous new approaches have been developed, including active or passive targeting of the drug to the desired site via nanocarrier drug delivery platform designed as transporters of anti-cancer drugs with the potential application for enhanced chemotherapy.^{23–26} Other localized treatment options include magnetically guided drug delivery utilizing iron particles that can subsequently be directed to the site of the tumor.^{27,28}

The current study involves a novel targeting approach utilizing encapsulation of the anti-cancer drug into reconstituted high-density lipoprotein (rHDL) nanoparticles with

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superparamagnetic iron oxide nanoparticle (SPION). rHDLs and other lipoprotein constructs have been developed by combining apolipoprotein A-1 (Apo-A1) or its mimetic peptides with phospholipids.^{29–33} The resultant lipoprotein mimetic nanoparticles possess several favorable characteristics for drug transport, including their small size, biocompatibility, stability, monodispersity and high drug payload. In addition, the lipid and protein components of rHDL may be modified via conjugation with targeting molecules to optimize their pharmaceutical potential for clinical applications.^{33,34} Earlier studies from our laboratory demonstrated that the solubility and bioavailability of valrubicin may be enhanced by incorporation into rHDL nanoparticles.^{29,30,33,34} In addition, Shah et al³⁵ provided proof of concept via photophysical characterization of the rHDL–valrubicin complex as a potential tumor imaging agent.

These preclinical studies were designed to evaluate the effectiveness of SPIONs in combination with the drug valrubicin (AD-32) carrying rHDL nanoparticles as potential cancer therapeutic agents. Some of the physical/chemical properties of the SPION–rHDL/valrubicin hybrid nanoparticles have been evaluated, in addition to their cytotoxicity against prostate cancer cells and their interaction with the cancer cells via a scavenger receptor-mediated (scavenger receptor type B [SR-B1]) mechanism. These studies showed, for the first time, that lipoprotein drug delivery enhanced via magnetic targeting could be an effective chemotherapeutic strategy for prostate cancer.

Materials and methods

Materials

All reagents were purchased from Sigma-Aldrich (St Louis, MO, USA). Prussian blue was purchased from Acros Organics (Geel, Belgium). All organic chemicals and solvents used were of reagent grade. Bacterial protein extraction reagent and bicinchoninic acid (BCA) protein assay kits were purchased from Thermo Fisher Scientific (Rockford, IL, USA). Apo-A1 was obtained from Mc Lab (San Francisco, CA, USA). Cholesterol and phospholipid estimation kits were obtained from Wako Pure Chemical Industries Ltd (Richmond, VA, USA). Block lipid transport-1 (BLT-1; SR-B1 inhibitor) was obtained from Cambridge Corporation (San Diego, CA, USA) and prepared as 5 mg/mL stock solution in 100% dimethyl sulfoxide (DMSO). Roswell Park Memorial Institute (RPMI) 1640 media, keratinocyte media and fetal bovine serum (FBS) were obtained from Invitrogen (Carlsbad, CA, USA). A prostate cancer cell line PC-3 and an immortalized normal epithelial cell line PZ-HPV were

obtained from American Type Culture Collection (ATCC; Manassas, VA, USA).

Methods

Preparation of SPIONs

SPION nanoparticles were prepared as described by Raut et al.³⁶ Thermal decomposition was used to prepare oleic acid-coated magnetic nanoparticles (OAMNP). A total of 10 mL of octyl ether was added to 1.43 mL of oleic acid. The mixture was heated to 100°C under an inert atmosphere for 30 minutes. To this mixture, 0.28 mL of iron pentacarbonyl [$\text{Fe}(\text{CO})_5$] was added, followed by a raise in the temperature to 300°C, which was maintained until the reaction mixture turned black. The solution was then allowed to cool down to room temperature, after which ethanol was added to precipitate the nanoparticles. The mixture was then centrifuged at 8,000 rpm for 15 minutes, after which the obtained precipitate was re-dissolved in hexane. This process of washing the OAMNP with hexane was repeated three times, and the nanoparticles were stored as hexane dispersion until further use.

Preparation of rHDL nanoparticles

The rHDL nanoparticles containing drug and SPIONs were prepared via the cholates dialysis procedure with some modifications.^{33,34} To a glass vial, phosphatidylcholine 7.5 mg/mL, free cholesterol 0.175 mg/mL and cholesteryl oleate 0.075 mg/mL, all from previously prepared stocks, were added and mixed thoroughly. This mixture was then dried under nitrogen. To this mixture, 1 mg/mL anti-cancer drug (valrubicin) and 30 μL /mL DMSO were added, followed by 2.5 mg/mL Apo-A1 and 1 mg/mL SPIONs (wet weight). The mixture was then vortexed. After adding 7.5 mg/mL sodium cholate, buffer (10 mM Tris, 0.1 M potassium chloride, 1 mM ethylenediaminetetraacetic acid [EDTA] pH 8.0) was used to make up the desired volume. The mixture was mixed thoroughly, frozen and then lyophilized overnight. It was then dialyzed against 2 L of 1 \times phosphate-buffered saline (PBS) for 48 hours, with a change of buffer three times every 2 hours on the 1st day, and later kept overnight. The mixture was centrifuged at 1,500 rpm for 1 minute and then filtered through a 0.45- μM filter and stored at 4°C until used (Figure 1).

Estimation of chemical composition of nanoparticles

Chemical composition estimations were determined using respective chemical assays, which were conducted using 96-well microtiter plates. Measurements of absorbance were conducted at different wavelengths (as outlined in each assay procedure) using a PowerWave 340 spectrophotometer

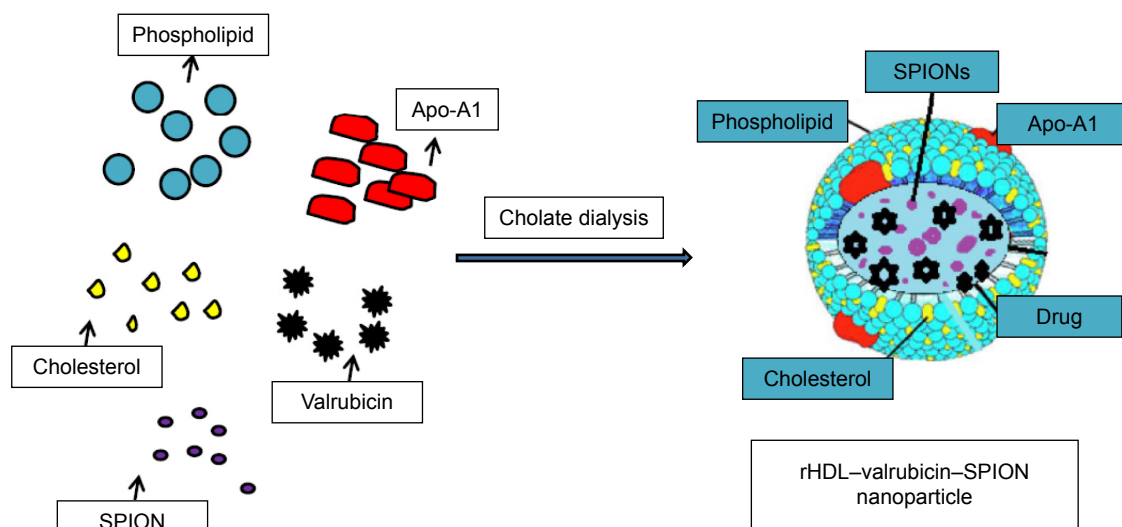


Figure 1 Schematic representation of formulation of rHDL-valrubicin-SPION nanoparticles – a conceptual model.

Abbreviations: rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle; Apo-A1, apolipoprotein A-1.

(Bio-Tek, Winooski, VT, USA). Standard graphs were then plotted with the known concentrations of respective standards, and the concentrations of each component were calculated. Each assay was carried out at least twice, and estimations for samples were carried out in triplicates. The following chemical composition estimations were conducted:

1. Cholesterol and phospholipid contents were determined by their respective enzymatic reagent kits (Cholesterol E and Phospholipid C; Waco Chemicals, Richmond, VA, USA) as per manufacturer's suggestions.
2. Protein estimations were conducted using a BCA protein assay kit as per manufacturer's instructions. A total of 10 μL of each sample was placed in triplicate, and 200 μL of BCA reagent was added and mixed thoroughly. The plates were incubated at 37°C for 30 minutes and then read at 540 nm.
3. Drug concentration was monitored by measuring the absorbance at 490 nm. The concentrations were extrapolated from a standard curve using different concentrations of valrubicin. Three individual readings were taken, and average and standard deviation (SD) were calculated.
4. Iron incorporation into the rHDL nanoparticles was assessed by using the phenanthroline reagent with some modifications adapting the assay to be carried out in a 96-well plate.³⁷ Ferrous ammonium sulfate hexahydrate solution (7 mg/mL) was used to prepare the standard curve for the iron determinations. Briefly, aliquots of the standard stock solution ranging from 1 μL to 20 μL were placed. To this solution, 1 μL of hydroxyl amine from stock solution (100 mg/mL), 10 μL of 1,10-phenanthroline

solution (from a stock of 1 mg/mL) and 8 μL of sodium acetate (from a stock of 1.2 M) was sequentially added. Finally, the samples were mixed and incubated at room temperature for 10 minutes, before measuring their absorbance at 450 nm.

Entrapment efficiency of iron and valrubicin

The drug or iron entrapment efficiency (EE) was determined using the equation:

$$EE = \frac{\text{Final Drug or Iron Concentration}}{\text{Initial Drug or Iron Concentration}} * 100$$

Measurement of size and zeta potential of the nanoparticles

The hydrodynamic diameter of the nanoparticles was estimated by dynamic laser scattering with a Delsa Nanoparticle Size Analyzer (Malvern Instruments, Herrenberg, Germany). An average of 70 runs was captured by the machine and is indicated by the average particle size. Zeta potential was measured using Zetasizer Nano ZS (Malvern Instruments). The zeta potential measurements were performed by dispersing the particles in an aqueous solution at 25°C with a scattering angle of 90°. The experiment was repeated three times, and the results were averaged.

Demonstration of morphology of rHDL-SPION particles using transmission electron microscopy (TEM)

Nanoparticles were imaged using TEM by depositing nanoparticles on a TEM grid from a PBS dispersion. After drying,

1% phosphotungstic acid as a negative stain was applied and TEM grids were left to dry at room temperature. The particles were then imaged using a JEM 2100 transmission electron microscope (JEOL, Peabody, MA, USA).

Uptake of iron and valrubicin in presence of SR-B1 inhibitors

Prostate cancer cells (PC-3) were plated in two 24-well plates (100,000 cells/well) in the RPMI medium with 10% FBS and incubated at 37°C with 5% CO₂ for 24 hours. The monolayers were washed with PBS, pH 7.4, and then incubated at 37°C with serum-free medium for 90 minutes. The cells were washed with PBS, and serum-free medium was again added. To one of the plates, 5 μM of BLT-1 was added in each well, and to another plate, 10 μg/mL of human high-density lipoprotein (HDL) was added. The plates were incubated for 30 minutes at 37°C. Two wells were kept as no inhibitor control. To one of the plates, 20 μL of the rHDL–SPION was added to each well, and to the other plate, 20 μL of rHDL–SPION–valrubicin nanoparticles were added. The plates were incubated at 37°C for 90 minutes. The wells were washed once with 1× PBS, pH 3.0, and subsequently with 1× PBS, pH 7.4, respectively. The cells were then lysed with lysis buffer (50 mM Tris-HCl [pH 8.0], 150 mM NaCl, 0.02% sodium azide, 100 mg/mL phenylmethylsulfonyl fluoride, 1 mg/mL aprotinin and 1% Triton X-100). The lysates were centrifuged at 10,000 rpm for 5 minutes. The protein content of the lysate was determined by BCA assays. The iron content and valrubicin content were measured by an iron estimation assay and spectrophotometric measurements, respectively.

Monitoring the movement of the rHDL–SPION particles

PC-3 cells were grown on a Petri dish. The cells were treated with 20 μL of rHDL–SPION particles. A magnetic force was applied at one end of the Petri dish to attract SPIONs. The plates were incubated at 37°C for 5 minutes. The cells were then washed with PBS (pH 7.4), keeping the magnet attached at the end. Equal parts of 20% HCl and 10% potassium ferrocyanide (Prussian blue) were prepared immediately before use.³⁸ The plate was covered in this solution for 20 minutes, washed three times with distilled water, counterstained with nuclear fast red for 5 minutes and rinsed twice with distilled water. Images were taken at areas near and away from the magnetic field using a TMS microscope (Nikon Instruments Inc., Melville, NY, USA) with camera.

Stability of rHDL–SPION and rHDL–SPION–valrubicin nanoparticles

Stability studies were conducted by the method described earlier.³⁹ A total of 1 mL of rHDL–SPION and rHDL–SPION–valrubicin formulation was placed in a dialysis bag (6–8 kD molecular weight cut off). The bags were placed in 50 mL of 1× PBS and incubated at 37°C for 48 hours. A total of 100 μL of the buffer was withdrawn at 0-, 2-, 4-, 6-, 8-, 24- and 48-hour intervals. Each time the buffer was withdrawn, a fresh solution of 100 μL of PBS was transferred in the outer chamber. A sample was saved before starting the dialysis, and a sample inside the dialysis bag was collected after the incubation period was over. Iron and valrubicin concentrations of the respective samples were determined by appropriate assay methods.

Cytotoxicity studies using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay method

A standard protocol using 5,000 cells/well of a 96-well plate was used.⁴⁰ PC-3 and PZ-HPV cells were incubated at 37°C in RPMI-1640 and keratinocyte media, respectively, and treated with increasing drug concentrations. The plate was divided into two parts: one part was treated with free valrubicin at concentrations of 0.85 μg/mL, 8.5 μg/mL, 17 μg/mL, 42.5 μg/mL and 85 μg/mL and the other part was treated with the same concentration of rHDL–SPION–valrubicin. For each concentration, four replicates were used; for control, cells without drug were used as a control. In a separate plate, rHDL and rHDL–SPION were each tested with corresponding varying protein concentrations. A control with rHDL–valrubicin was also kept. The plates were incubated for 48 hours at 37°C. A total of 25 μL of MTT reagent was added to each well, and plates were incubated at 37°C for 3 hours. The plates were centrifuged at 1,500 rpm for 5 minutes, and the supernatant was then discarded. The purple-colored compound in the resulting pellet was dissolved in DMSO, and the absorbance was measured at 570 nm. Percentage survival of cells was calculated and plotted against concentration of valrubicin.

Statistical analysis

Data of EE, mean diameter and zeta potential are reported as mean ± SD. Percentage iron uptake, percentage iron retained and percentage cell survival are reported as mean ± standard error of the mean. The half maximal inhibitory concentration (IC₅₀) values for the different drug formulations were calculated

using CalcuSyn software (Cambridge, UK). The results were analyzed using analysis of variance (ANOVA), and the differences were considered significant at a level of $P < 0.05$.

Results

Compositional analysis

Chemical composition of empty nanoparticles (Figure 2A), rHDL–SPION nanoparticles (Figure 2B), rHDL–valrubicin nanoparticles (Figure 2C) and rHDL–SPION–valrubicin nanoparticles (Figure 2D) are shown. The percentage distribution of phospholipid, protein and cholesterol of the rHDL, rHDL–SPION, rHDL–valrubicin and rHDL–SPION–valrubicin complexes ranged from 41.11% to 53.72%, 32.56% to 46.83% and 5.54% to 8.53%, respectively, indicating that the incorporation of SPION

does not substantially alter the chemical composition of the rHDL/drug formulation. The amounts of iron in rHDL–SPION and rHDL–SPION–valrubicin nanoparticles were 0.75% and 0.61%, respectively, whereas the valrubicin content was ~3% lower in the rHDL–SPION–valrubicin nanoparticles compared to the rHDL–valrubicin nanoparticles (Figure 2B–D).

Iron and valrubicin EE

The entrapment of valrubicin in the rHDL–valrubicin nanoparticles vs rHDL–SPION–valrubicin nanoparticles was 89% and 55%, respectively (Figure 3A). The EE of iron was 44% in rHDL–SPION nanoparticles and 30% in rHDL–SPION–valrubicin nanoparticles (Figure 3B). As may be seen in the following data (Figure 3A and B).

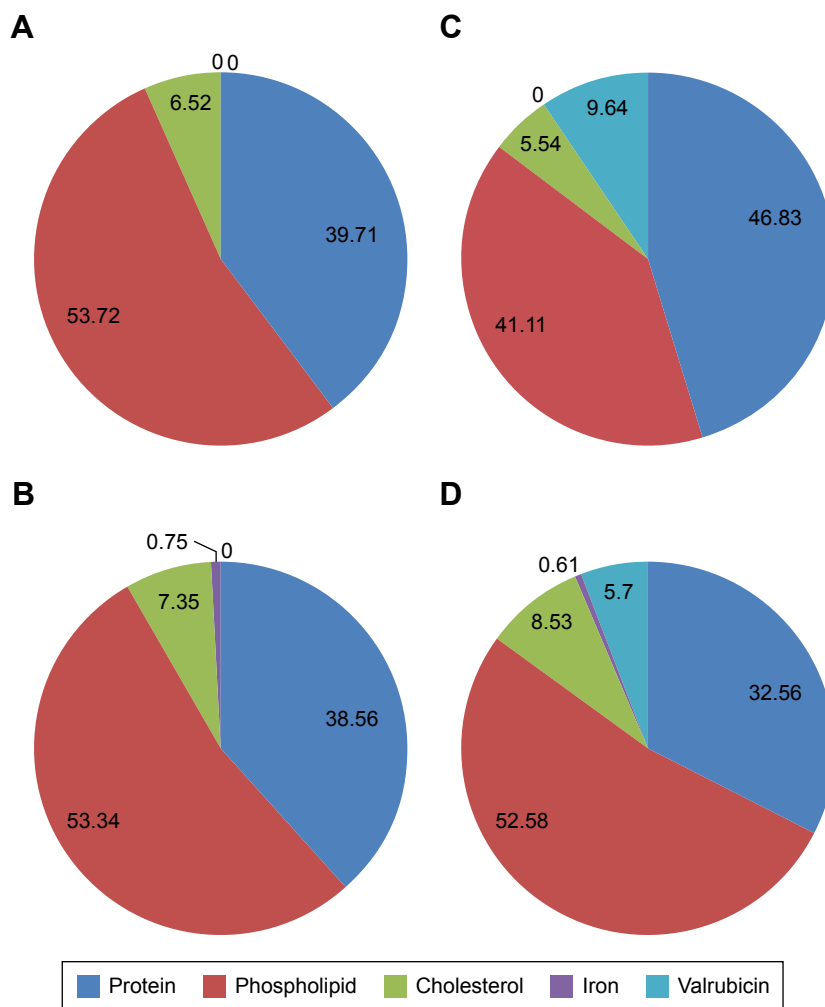


Figure 2 Chemical compositional analysis of nanoparticles.

Notes: (A) rHDL only; (B) rHDL–SPION; (C) rHDL–Valrubicin; (D) rHDL + SPION + valrubicin. Protein (dark blue), phospholipid (red), cholesterol (green), iron (purple) and valrubicin (light blue) were estimated by standard procedures.

Abbreviations: rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle.

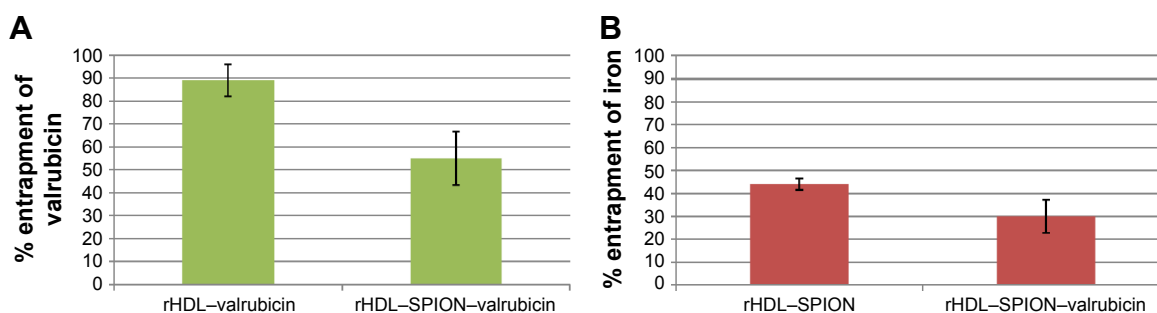


Figure 3 EE in rHDL-valrubicin, rHDL-SPION and rHDL-SPION-valrubicin: (A) valrubicin and (B) iron.

Note: The bars represent the amount of material incorporated relative to the respective initial amounts present.

Abbreviations: EE, entrapment efficiency; rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle.

Physical properties of the nanoparticles

The particle diameter analysis of the four types of nanoparticles showed them to be reasonably homogeneous, while the average diameter increased with the addition of SPIONs and/or valrubicin (presumably located) in the core of the nanoparticles (Table 1 and Figure 4A–D). The average diameter of rHDL, rHDL-SPION, rHDL-valrubicin and rHDL-SPION-valrubicin was 7.1 ± 1.3 nm, 67.6 ± 19.4 nm, 48.1 ± 13.9 nm and 96.9 ± 18.7 nm, respectively. The polydispersity index (PDI) of these particles was 0.47, 0.352, 0.350 and 0.378, respectively.

Zeta potential was also assessed as an indication of the electrostatic or charge repulsion/attraction between particles and as a potential barrier to cell/nanoparticle interactions.^{40,41} The zeta potential values for these particles are shown in Table 1. Insertion of the drug (valrubicin) into the core of the respective nanoparticles³⁵ changed the zeta potential from -10.57 ± 0.157 for the rHDL nanoparticles to -7.93 ± 0.548 for the rHDL-SPION-valrubicin nanoparticles.

Receptor-mediated uptake of the iron payload from the nanoparticles

The uptake of SPIONs and the drug from rHDL-SPION and rHDL-SPION-valrubicin nanoparticles was anticipated to proceed via the SR-B1 receptors.^{32,42} To investigate this

uptake mechanism, two inhibitors of SR-B1 were selected (BLT-1 and human HDL) for the iron uptake studies. Incubations in the absence of the SR-B1 inhibitor were considered as 100% iron uptake. The average iron uptake from the rHDL-SPION nanoparticles was reduced to 12.2% and 15.4% (of the control), respectively, in the presence of $5 \mu\text{M}$ BLT-1 and $10 \mu\text{g/mL}$ HDL (Figure 5A). The same trend was observed with rHDL-SPION-valrubicin nanoparticles in the presence of BLT-1 and HDL, resulting in decreased iron uptakes to 20.1% and 18.2% of the (uninhibited) control, respectively (Figure 5B).

Movement of SPIONs in response to a magnetic field

This experiment was designed to determine whether the SPIONs containing nanoparticles move in the direction of the magnetic field (Figure 6A). When the SPIONs containing rHDL nanoparticles were incubated on a plate grown with prostate cancer cells (PC-3), the cells in the vicinity of the applied magnetic field (Figure 6B) showed high iron particle concentration, as shown by the deep blue color due to Prussian blue dye staining. The cells on the opposite end of the magnetic field did not incorporate the SPIONs (Figure 6C), as evidenced by the lack of blue color, indicating that only the SPIONs containing nanoparticles may be maneuvered to a desired location via a magnetic field.

Stability of rHDL-SPION and rHDL-SPION-valrubicin

The stability of rHDL-SPION and rHDL-SPION-valrubicin was assessed under conditions resembling a physiological milieu (pH 7.4, 37°C). The initial iron content was designated as 100%. After 48 hours, rHDL-SPION had 67.07% of iron remaining, while rHDL-SPION-valrubicin had 65.5% of iron remaining (Figure 7).

Table 1 Comparative physical characteristics of rHDL nanoparticles

Type of particles	Average diameter \pm SD	PDI	Zeta potential \pm SD
rHDL only	7.1 ± 1.3	0.470	-10.57 ± 0.153
rHDL-SPION	67.6 ± 19.4	0.352	-8.61 ± 0.726
rHDL-valrubicin	48.1 ± 13.9	0.350	-8.27 ± 0.724
rHDL-SPION-valrubicin	96.9 ± 18.7	0.378	-7.93 ± 0.548

Note: Average particle size, PDI and zeta potential were measured.

Abbreviations: rHDL, reconstituted high-density lipoprotein; SD, standard deviation; PDI, polydispersity index; SPION, superparamagnetic iron oxide nanoparticle.

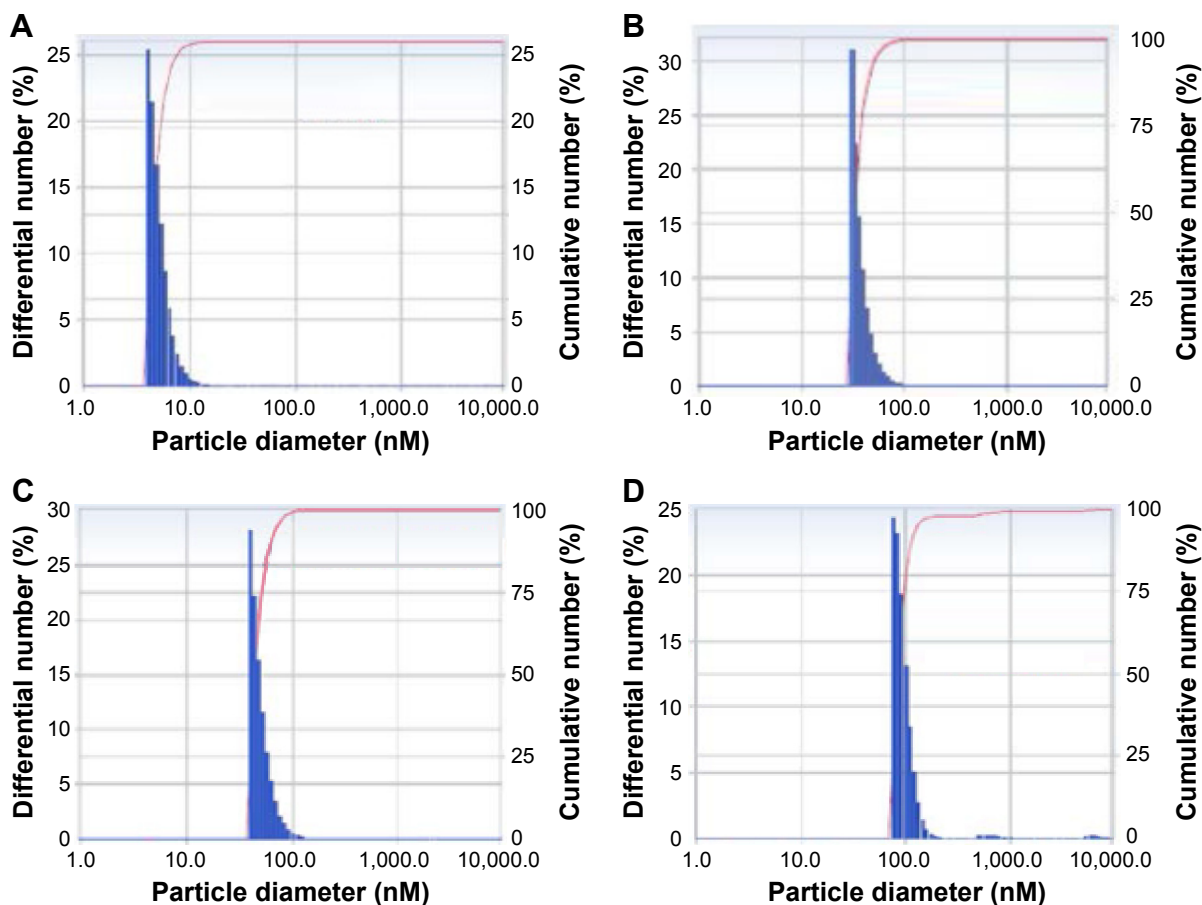


Figure 4 Comparative size of rHDL nanoparticles.

Notes: (A) rHDL only, (B) rHDL-SPION, (C) rHDL-valrubicin and (D) rHDL-SPION-valrubicin. Measurements were taken using Delsa Nanoparticle Size Analyzer.

Abbreviations: rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle.

Cytotoxicity of free valrubicin and rHDL-SPION-valrubicin against (malignant) prostate cancer cells (PC-3) and (nonmalignant) prostate epithelial cells (PZ-HPV)

The cytotoxicity of valrubicin delivered via rHDL-SPION nanoparticles was compared to free (unencapsulated)

valrubicin. A concentration-dependent decrease in the survival of PC-3 and PZ-HPV cells was observed with both free and rHDL-encapsulated valrubicin preparations (Figure 8A and B). The effect of rHDL-SPION-valrubicin preparation on PC-3 cell survival was most pronounced at higher valrubicin concentrations, producing 92% and 100% killing, respectively, at 42.4 $\mu\text{g/mL}$ (58.5 μM) and 85 $\mu\text{g/mL}$ (117.5 μM ;

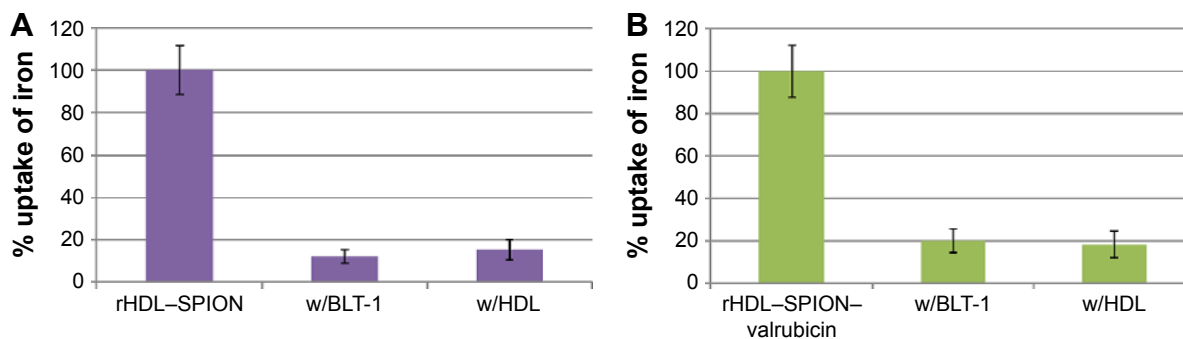


Figure 5 Inhibition of iron uptake from rHDL-SPION (A) and rHDL-SPION-valrubicin (B) nanoparticles by PC-3 cells, induced by a BLT-1 inhibitor and an excess of human HDL.

Abbreviations: rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle; BLT-1, block lipid transport-1; HDL, high-density lipoprotein; w/, with.

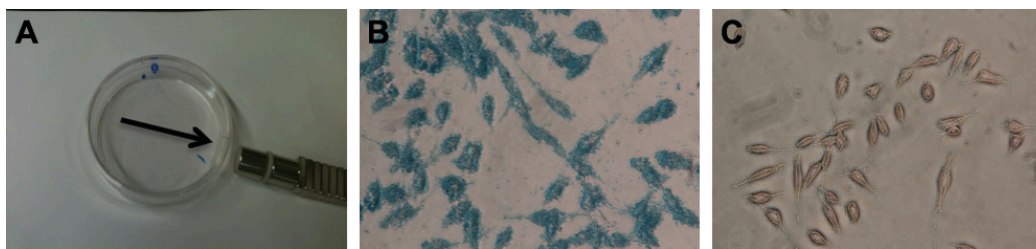


Figure 6 Movement of SPIONs in response to magnetic field.
Notes: Prussian blue staining shows location of iron particles with respect to cancer cells. (A) Petri dish with prostate cancer cells treated with rHDL-SPION nanoparticles to which magnetic field is applied. (B) Cells closer to magnetic field. (C) Cells away from magnetic field (did not turn blue with stain). Magnification x20.
Abbreviations: SPION, superparamagnetic iron oxide nanoparticle; rHDL, reconstituted high-density lipoprotein.

Figure 8A). On the other hand, the free drug administered at the same concentrations inhibited PC-3 survival by only 60% and 79%, respectively. (The control rHDL and rHDL-SPION nanoparticles with the same amounts of protein showed <10% inhibition at these higher concentrations, data not shown). The IC₅₀ values for free valrubicin and rHDL-SPION-valrubicin were found to be 16 µg/mL and 12 µg/mL (22.1 µM and 16.5 µM), respectively (*P*-value <0.05). On the other hand, an opposite trend was observed regarding the survival of nonmalignant PZ-HPV cells in the presence of free valrubicin vs rHDL-SPION-valrubicin. At the respective valrubicin concentrations of 42.4 µg/mL and 85 µg/mL, the percentage survival of cells treated with rHDL-SPION-valrubicin nanoparticles was twice as compared to those

treated with free valrubicin (Figure 8B). Accordingly, the IC₅₀ values of free valrubicin and rHDL-SPION-valrubicin for PZ-HPV were found to be 22 µg/mL and 115 µg/mL (30.4 µM and 158.9 µM), respectively (*P*-value <0.05). The potential therapeutic enhancement of the encapsulation of the therapeutic agent (valrubicin) was calculated using to the following formula:

$$\frac{\text{Valrubicin IC}_{50} \text{ of PC-3 cells}}{\text{Valrubicin IC}_{50} \text{ of prostate epithelial cells}} \bigg/ \frac{\text{rHDL-valrubicin IC}_{50} \text{ of PC-3 cells}}{\text{rHDL-valrubicin IC}_{50} \text{ of prostate epithelial cells}}$$

Accordingly, the therapeutic enhancement via the rHDL-valrubicin-SPION formulation (vs free valrubicin) is indicated to be approximately sevenfold. These findings indicate substantial potential benefits during chemotherapy by limiting or avoiding off target toxicity of the rHDL/drug formulations.

The TEM data on rHDL-SPION nanoparticles before and after formulation indicated that these particles are somewhat spherical with a diameter ranging from 15 nm to 22 nm with the mean diameter of 17.9±5.1 nm (Figure 9A). The SPIONs before going in the formulation showed the diameter ranging from 2 nm to 4 nm with a mean diameter of 3.3 nm (Figure 9B).

Discussion

The present study evaluated the feasibility of preparing SPIONs in combination with rHDL nanoparticles, potentially for cancer therapeutics. The findings, reported earlier, provide additional substantial proof of concept for the utility of rHDL-SPION-valrubicin nanoparticles against prostate cancer (PC-3) cells vs nonmalignant cells and thus suggest broader application in cancer chemotherapy. Accordingly, we have shown that iron can be incorporated into the rHDL complex and that the chemical composition of rHDL-SPION

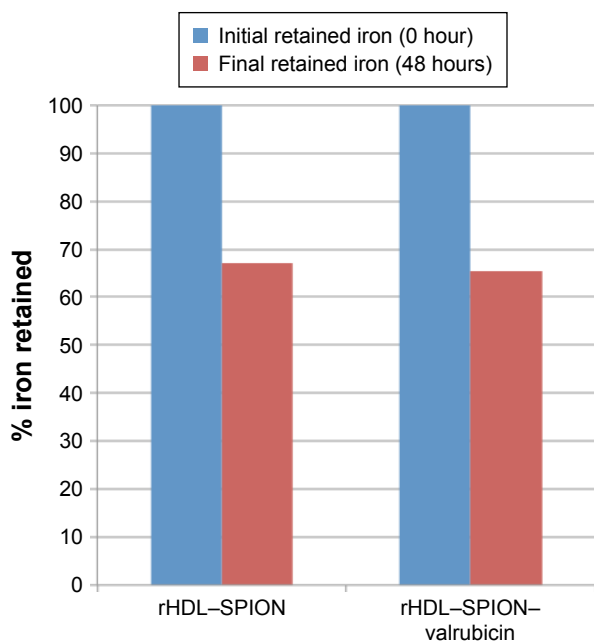


Figure 7 Stability of rHDL-SPION-valrubicin nanoparticles.
Notes: Samples were dialyzed against PBS at pH 7.4, 37°C for 48 hours. The iron retained in the nanoparticles after 48 hours of dialysis is expressed as percentage of the amount present initially.
Abbreviations: rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle; PBS, phosphate-buffered saline.

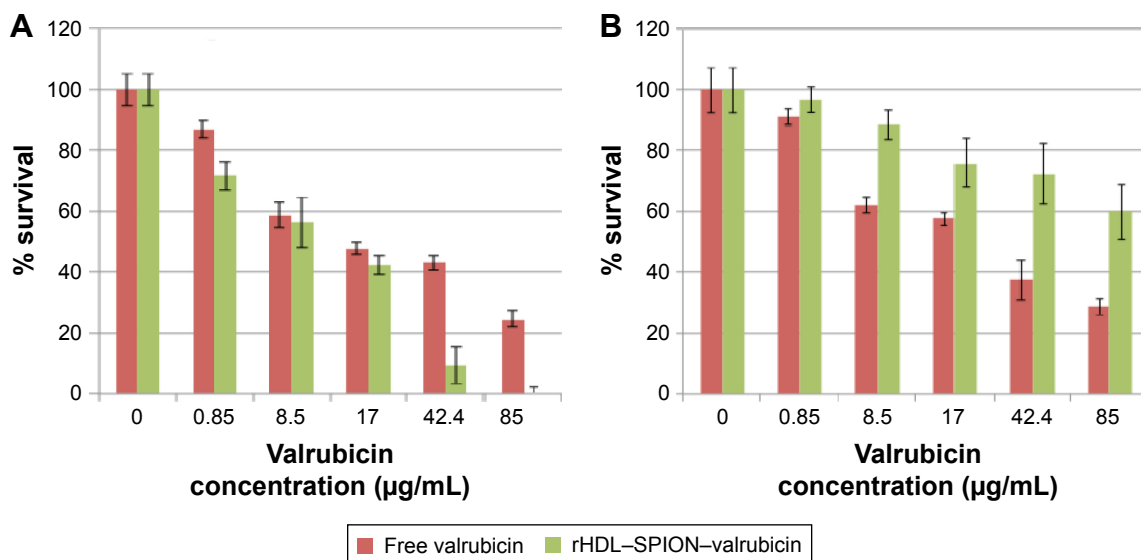


Figure 8 Survival of PC-3 cells (A) and PZ-HPV cells (B) upon treatment with rHDL-SPION-valrubicin vs free valrubicin and incubated for 48 hours as determined by MTT assay. **Abbreviations:** rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide.

nanoparticles was comparable to the rHDL-drug particles reported earlier.^{33,34} The protein and phospholipid contents of the rHDL-SPION valrubicin complex were comparable to those of empty rHDL and to those of the rHDL-valrubicin nanoparticles, which have been shown to be quite stable.^{33,34} These findings further support the feasibility of the eventual utilization of the rHDL-SPION-valrubicin nanoparticles for clinical applications. As expected, the EE of iron and valrubicin for the rHDL-SPION-valrubicin complexes was lower than the entrapment of both rHDL-SPION and rHDL-valrubicin due to the iron particles being incorporated into the nanoparticle complex.

The size and surface properties play an important role in targeting therapeutic nanocarriers. Smaller size facilitates the nanoparticles to penetrate the tumor microenvironment and stroma more efficiently than larger structures.⁴³⁻⁴⁵ The

estimated diameter of rHDL-SPION particles by dynamic light scattering was $\sim 67.6 \pm 19.4$ nm, whereas with TEM, the diameter was found to be 17.9 ± 5.1 nm. The discrepancy between the two methods could be due to the differences in the measurements; however, the size was observed to be within the desirable range of the standard nanoparticles.⁴⁵⁻⁴⁸ Nevertheless, the size of the nanoparticles could likely be further modulated (decreased) by increasing their protein content or modifying the procedure for preparing the magnetic valrubicin rHDL formulations. The PDI for empty rHDL particles was 0.470 and for the rHDL-SPION-valrubicin nanoparticles was 0.378 that are considered to be in excess of a desirable homogeneity. This parameter could also likely be improved by utilizing alternate preparatory procedures for assembling the respective nanoparticles.^{49,50} More careful analyses of particle size and diameter will also have to be conducted to accurately assess the size and the heterogeneity of these nanoformulations. The zeta potential of rHDL-SPION-valrubicin nanoparticles (-7.93 mV) was closer to 0 than the empty rHDL particles (-10.57 mV), indicating that their interaction with cells may be more intense than that of empty rHDL.

BLT-1 has been shown to inhibit the function of the SR-B1 receptor, while circulating HDL competes with the rHDL nanoparticles and thus has been shown to interfere with the drug uptake from the rHDL nanoparticles.^{34,50-52} When the PC-3 cells were pre-incubated with BLT-1 and human HDL, respectively, and subsequently treated with rHDL-SPION and rHDL-SPION-valrubicin nanoparticles, the uptake of

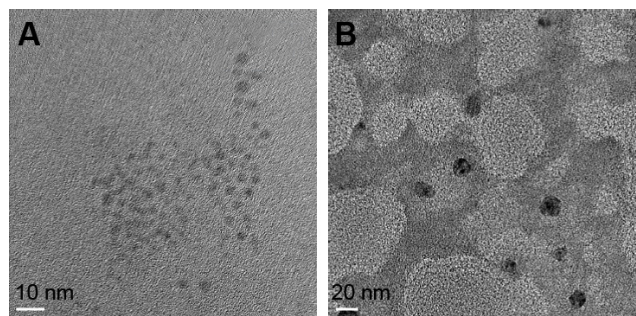


Figure 9 TEM image of rHDL-SPION nanoparticles: (A) iron oxide particles and (B) rHDL-SPION nanoparticles.

Abbreviations: TEM, transmission electron microscopy; rHDL, reconstituted high-density lipoprotein; SPION, superparamagnetic iron oxide nanoparticle.

iron was reduced by almost 90%, indicating that the mechanism of iron uptake was specific and likely dependent on the SR-B1 receptors. These data are also consistent with earlier findings regarding the release of the drug payload to cancer cells and tumors from the rHDL nanoparticles.^{34,42,52} Stability studies showed that 60%–65% of the iron remained within the nanoparticles after 48 hours of incubation, indicating that encasement of SPIONs and valrubicin within rHDL provided a stable environment that has the potential to remain longer in the circulation than conventional nonencapsulated drugs. Free drugs generally exit the circulation within a few hours.⁴² Prussian blue staining provided evidence of movement for the majority of SPIONs in the direction of the magnetic field; the accumulation of particles toward PC-3 suggest that the SPIONs inside the rHDL–drug nanoparticles may help to maneuver them toward the target site, thus facilitating the subsequent drug delivery to the cells of interest. This approach could minimize the wastage of drugs during therapy by avoiding normal healthy cells and tissues and thus reduce off-target effects.

Finally, the cytotoxicity studies support the effectiveness of rHDL–SPION–valrubicin against PC-3 over the traditional method of administering the drug in the free form. These studies are consistent with our previous observations with rHDL–valrubicin nanoparticles with PC-3 cells.³⁴ The SPION particles are permitted to move when a magnetic field is applied, so localizing the site of attack could become even more efficient than the rHDL-encapsulated delivery. An increase in the IC_{50} values of rHDL–SPION–valrubicin as compared to the free valrubicin in nonmalignant PZ-HPV cells indicates that there is a protective effect of rHDL-encapsulated drug. In vivo, this may suggest reduced side effects of the drug when delivered via the rHDL vehicle.

Although rHDL–SPION–valrubicin drug delivery model has shown a strong potential for application in cancer therapeutics, it will require significant additional work (tumor suppression, toxicology, etc) before its translation toward the clinic. One of the concerns to be addressed is the expression of SR-B1 receptor, the gateway for delivering drugs to tumors by the rHDL nanoparticles, that has been shown to be variable among cancer cells and tumors.⁵³ This feature of tumors actually may be taken advantage of and extended toward personalized or precision therapy by pre-screening the patients' tumors for expression of the SR-B1 receptor prior to therapy to determine their susceptibility to drugs, carried by the rHDL nanoparticles. Ultimately, another concern may arise due to additional costs of the equipment and personnel to generate and monitor the magnetic field applied during therapy.

Conclusion

While the technology involving magnetic nanoparticles has been primarily investigated for the purposes of imaging,^{53–55} the use of rHDL–SPION–valrubicin for targeted drug delivery against malignant cells and tissues had not previously been explored. The findings of the present study show that it is feasible to formulate rHDL–SPION–drug nanoparticles for the purpose of targeted delivery of the drug payload to a desired site. Additionally, due to their properties of contrast enhancement, SPION containing rHDL particles could also be utilized as magnetic resonance imaging contrast agents, providing the rHDL–SPION–valrubicin nanoparticles with theranostic potential.

The major advantages of this technology are projected to be:

1. less toxic effects to normal cells and tissues, thus reducing the off-target effects of chemotherapy;
2. lower doses of the drug may be required to treat patients due to the more effective targeting strategy and
3. the drug payload is expected to be in circulation for a longer time, resulting in increased therapeutic effectiveness.

Subsequent to successful tumor suppression, toxicology studies and clinical trials, the inclusion of the rHDL–SPION–valrubicin nanoparticles could provide a major advance for the chemotherapy of cancers in general and prostate cancer in particular.

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Disclosure

The authors report no conflicts of interest in this work.

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