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CHECKLIST

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Annotated checklist of fish cestodes from South America

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Abstract

An exhaustive literature search supplemented by a critical examination of records made it possible to present an annotated checklist of tapeworms (Cestoda) that, as adults or larvae (metacestodes), parasitize freshwater, brackish water and marine fishes, i.e. cartilaginous and bony fishes, in South America. The current knowledge of their species diversity, host associations and geographical distribution is reviewed. Taxonomic problems are discussed based on a critical evaluation of the literature and information on DNA sequences of individual taxa is provided to facilitate future taxonomic and phylogenetic studies. As expected, the current knowledge is quite uneven regarding the number of taxa and host-associations reported from the principal river basins and marine ecoregions. These differences may not only reflect the actual cestode richness but may also be due to the research effort that has been devoted to unravelling the diversity of these endoparasitic helminths in individual countries. A total of 297 valid species, 61 taxa identified to the generic level, in addition to unidentified cestodes, were recorded from 401 species of fish hosts. Among the recognized cestode orders, 13 have been recorded in South America, with the Onchoproteocephalidea displaying the highest species richness, representing *c.* 50% of all species diversity. The majority of records include teleost fish hosts (79%) that harbour larval and adult stages of cestodes, whereas stingrays (Myliobatiformes) exhibit the highest proportion of records (39%) among the elasmobranch hosts. Fish cestodes are ubiquitous in South America, being mostly recorded from the Warm Temperate Southeastern Pacific (WTSP; 31%) for marine hosts and the Amazon River basin (45%) for freshwater ones. The following problems were detected during the compilation of literary data: (i) unreliability of many records; (ii) poor taxonomic resolution, i.e. identification made only to the genus or even family level; (iii) doubtful host identification;

and (iv) the absence of voucher specimens that would enable us to verify identification. It is thus strongly recommended to always deposit representative specimens in any type of studies, including faunal surveys and ecological studies. An analysis of the proportion of three basic types of studies, i.e. surveys, taxonomic and ecological papers, has shown a considerable increase of ecological studies over the last decade.

Keywords
Biodiversity, marine ecoregions, river basins, species richness, tapeworms

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Introduction

Tapeworms (Cestoda) are a monophyletic assemblage of flatworms (Phylum Platyhelminthes) and they are obligate internal parasites of vertebrates. Their complex life-cycles include one or more intermediate hosts in a wide array of animal phyla (mostly arthropods) and they are exclusively transmitted perorally, i.e. via the food chain (Caira and Littlewood 2013; Littlewood et al. 2015). The cestodes are the second species-richest group of platyhelminths, with more than 5000 species in 751 recognized genera that have radiated through marine, freshwater and terrestrial environments (Waeschenbach et al. 2012; Caira and Littlewood 2013).

Cestodes parasitizing elasmobranchs and teleost fishes in at least one stage of development comprise one of the most diverse lineages of tapeworms (Caira et al. 2014; Caira and Jensen 2014), only comparable in species richness with cyclophyllidean cestodes, parasites of tetrapods (Caira and Littlewood 2013). Since these parasites usually exhibit a strict host specificity, they are considered suitable models for studies of host-parasite co-evolution (Caira and Jensen 2014) or even helping in circumscribing species boundaries of cestode hosts (Caira and Jensen 2015).

South America is a megadiverse continent, including at least five of the world's biodiversity 'hotspots', more than 30.000 km of coastline and two of the 10 largest freshwater drainage systems of the world, i.e. the Amazon and Paraná River basins, which is reflected in its species-rich ichthyofauna (Myers et al. 2000; Miloslavich et al. 2011; Reis 2013). Bearing that in mind, one might expect a high diversity of fish cestodes as well, even though there are no comprehensive checklists or other faunistic studies encompassing the whole continent that could provide an overview of the cestode diversity, except those with regional focus (e.g. Thatcher 2006 for Amazonia; Tantaléan and Huiza 1994 for Peru; Muñoz and Olmos 2008 for Chile).

Studies on fish cestodes from South America date back to the early 19th Century, when C. A. Rudolphi described *Anthocephalus macrourus* Rudolphi, 1819 from an unidentified sparid fish and *Anthocephalus interruptum* Rudolphi, 1819 (both cestodes of the order Trypanorhyncha) from *Trichiurus lepturus* Linnaeus off the Brazilian coast, even though these species are no longer valid (Campbell and Beveridge 1996). Subsequently, K. M. Diesing (1850, 1856) described several species that are now included in three different orders. Both workers studied cestodes collected by renowned naturalists, such as F. Sellow, I.F.W.M. Von Olfers and J. Natterer. With few exceptions, the number of descriptions and/or reports have considerable increased in the 20th Century and a large amount of information has been generated,

yet many studies are faunal surveys dispersed in regional journals that are not readily accessible.

Detailed taxonomic studies combining morphological and molecular approaches have recently expanded our knowledge at lower and higher taxonomic levels, mostly under the framework of the National Science Foundation (Planetary Biodiversity Inventory program) funded project called "A survey of the tapeworms (Cestoda: Platyhelminthes) from vertebrate bowels of the earth" (see http://tapewormdb.uconn.edu). This project funded, amongst others, intensive research on fish cestodes in South America, which were mainly undertaken by A. de Chambrier, T. Scholz and A. A. Gil de Pertierra for teleost hosts, and V. A. Ivanov, F. P. L. Marques and F. Reyda for elasmobranch hosts. The present paper aims at addressing the following objectives: (1) to provide for the first time an annotated checklist that summarizes records of cestodes in marine and freshwater fishes from South America, including detailed information on their hosts, site of infection, geographical distribution, stage of development and molecular data; (2) to critically assess some doubtful reports; and (3) to depict the problems that impede a better understanding of the diversity and host associations of cestodes in South America.

Materials and methods

Parasite-host and host-parasite checklists for fish cestodes from South America were compiled on the basis of an exhaustive search of literature published until August 2016; abstracts of meetings, theses and reports without primary data were not considered. The bibliographic search was complemented by the information gathered from *Helminthological Abstracts*, Host-Parasite Database of the Natural History Museum, London (Gibson et al. 2005), Global Cestode Database (GDC) (Caira et al. 2012), *Google Scholar*, *ScienceDirect*, *Web of Knowledge*, as well as some previously published books (e.g. Palm 2004; Thatcher 2006). The classification of cestodes proposed by Khalil et al. (1994) is basically followed, but it is updated based on revisional papers on individual cestode orders or molecular phylogenetic studies at the ordinal level, such as Kuchta et al. (2008) for bothriocephalideans and diphyllobothriideans, Olson et al. (2010) for trypanorhynchs, Caira et al. (2014) for onchoproteocephalideans, phyllobothriideans and 'tetraphyllideans', Healy et al. (2009) and Ruhnke et al. (2015) for rhinebothriideans, and Jensen et al. (2016) for lecanicephalideans.

The species are arranged according to taxonomic categories and are presented in alphabetical order followed by data on their hosts (species name, class and family), habitat, site of infection, stage of development, marine ecoregion according to Spalding et al. (2007), river basins or lakes, country and references (between parentheses). All cestodes presented herein follow the most recent taxonomic literature and the validity of individual taxa or the reliability of their records were critically assessed by the present authors, who consulted with experts for some tapeworm groups.

Host species are arranged in taxonomic and then alphabetical order. The scientific names of hosts have been updated based on Froese and Pauly (2016) and supplement-

ed by the most recent taxonomic papers for certain problematic taxa (e.g. *Cichla* Bloch & Schneider, *Pseudoplatystoma* Bleeker and *Zungaro* Bleeker).

The following abbreviations are used for collections:

BMNH The British Museum (Natural History) Collection at the Natural History Museum, London, UK;
CHIOC Coleção Helmintológica do Instituto Oswaldo Cruz, Rio de Janeiro, Brazil;
HWML Harold W. Manter Laboratory of Parasitology, University of Nebraska State Museum, Lincoln, Nebraska;
MLP Departamento de Zoología de Invertebrados (Parasitología), La Plata, Argentina;
MHNG Natural History Museum, Geneva, Switzerland;
MNHB Museum der Naturkunde für Humboldt Universität zu Berlin, Germany;
NHMW Natural History Museum, Vienna, Austria;
USNPC United States National Parasite Collection, Beltsville, Maryland, USA, which has been transferred recently to the National Museum of Natural History (**USNM**) of the Smithsonian Institution, Washington, D.C., USA

The following abbreviations are used for marine ecoregions according to Spalding et al. (2007):

JFD Juan Fernández and Desventuradas;
NBS North Brazil Shelf;
TEP Tropical Eastern Pacific;
TNA Tropical Northwestern Atlantic;
TSA Tropical Southwestern Atlantic;
WTSA Warm Temperate Southwestern Atlantic;
WTSP Warm Temperate Southeastern Pacific

The following abbreviations are used for molecular markers:

18S small subunit of the nuclear ribosomal RNA gene;
ITS1 first nuclear ribosomal internal transcribed spacer;
5.8S 5.8S ribosomal RNA gene;
ITS2 second nuclear ribosomal internal transcribed spacer;
28S large subunit of the nuclear ribosomal RNA gene;
16S large subunit of the mitochondrial ribosomal RNA gene;
***cox*1** cytochrome *c* oxidase I

The following abbreviation is used for records of metacestodes in the host-parasite list:

L larvae.
* Asterisks in the parasite-host list indicate the type species of the genus.

Parasite-Host List

Class Cestoda Rudolphi, 1808
Order Amphilinidea Poche, 1922
Family Amphilinidae Claus, 1879

Nesolecithus janickii* Poche, 1922
[Syns. *Monostoma liguloideum* Diesing, 1850 (*pro parte*); *Amphilina liguloidea* Monticelli, 1892 sensu Janicki (1808); *Schizochoerus janickii* (Poche, 1922) Bandoni & Brooks, 1987]

Arapaima gigas (Actinopterygii: Arapaimidae); freshwater; body cavity; adult; Amazon River basin; Brazil (Janicki 1908; Poche 1922; Travassos and Teixeira de Freitas 1964; Rego et al. 1974; Araújo et al. 2009).
Notes: type host. Bandoni and Brooks (1987) proposed *Schizochoerus janickii* as a new combination for this species, but we are following the classification proposed by Gibson (1994).

Schizochoerus liguloideus* (Diesing, 1850) Poche, 1922
[Syns. *Monostoma liguloideum* Diesing, 1850 (*pro parte*); *Amphilina liguloidea* Monticelli, 1892]

Arapaima gigas (Actinopterygii: Arapaimidae); freshwater; body cavity; adult; Amazon River basin; Brazil, Peru (Diesing 1850; Poche 1922; Travassos and Teixeira de Freitas 1964; Rego et al. 1974; Mariaux 1998; Olson and Caira 1999; Araújo et al. 2009; Serrano-Martínez et al. 2015).
Notes: type host. Diesing (1850) described *M. liguloideum* based on a mixture of *S. liguloideus* and *N. janickii*. Monticelli (1892) transferred the former species to *Amphilina* Wagener, 1858, whereas the latter was described by Janicki (1908) also under the name *Amphilina liguloidea*. Poche (1922) distinguished the both species and proposed the name *N. janickii*. Sequences of partial 18S (Z98305, Z98306, Z98307, AF124454) (Mariaux 1998; Olson and Caira 1999).

Order Bothriocephalidea Kuchta, Scholz, Brabec & Bray, 2008
Family Bothriocephalidae Blanchard, 1849

***Bothriocephalus timii* Gil de Pertierra, Arredondo, Kuchta & Incorvaia, 2015**

Cottoperca gobio (Actinopterygii: Bovichtidae); marine; intestine, pyloric caeca; adult; Magellanic; Argentina (Brabec et al. 2015; Gil de Pertierra et al. 2015).
Notes: type host. Sequences of 18S (KR780929), 28S (KR780885), 16S (KR780839) and *cox*1 (KR780787) (Brabec et al. 2015).

***Bothriocephalus* sp.**

Eleginops maclovinus (Actinopterygii: Eleginopsidae); marine; intestine; adult; WTSP; Chile (George-Nascimento et al. 2009).

Engraulis anchoita (Actinopterygii: Engraulidae); marine; pyloric caeca; adult; Magellanic, WTSA; Argentina (Timi 2003; Timi and Poulin 2003).

Engraulis ringens (Actinopterygii: Engraulidae); marine; intestine; adult; WTSP; Chile (Chávez et al. 2007; Valdivia et al. 2007; George-Nascimento and Moscoso 2013).

Helicolenus lengerichi (Actinopterygii: Sebastidae); marine; intestine; adult; WTSP; Chile (George-Nascimento and Iriarte 1989; Balboa and George-Nascimento 1998).

Clestobothrium crassiceps* (Rudolphi, 1819) Lühe, 1899

[Syn. *Bothriocephalus crassiceps* Rudolphi, 1819]

Aphos porosus (Actinopterygii: Batrachoididae); marine; intestine; adult (immature); WTSP; Chile (Cortés and Muñoz 2008, 2009).

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; intestine, pyloric caeca; stage of development not given; Magellanic; Falkland Islands (Brickle et al. 2006).

Note: R. Kuchta (pers. comm.) suggested that this report might be wrong.

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; intestine; adult; Magellanic; Argentina, Chile (Szidat 1961; Oliva 2001; Chávez et al. 2012).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; intestine; adult; WTSP; Chile (Carvajal et al. 1979; George-Nascimento 1996; Oliva and Ballón 2002; Chávez et al. 2012).

Note: Carvajal et al. (1979) reported the species as *Clestobothrium* sp., and R. Kuchta (pers. comm.) suggested that it belongs to *C. crassiceps*.

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; intestine; adult; WTSP; Peru (Durán and Oliva 1980; Jara 1998; Chero et al. 2014a).

Merluccius sp. (Actinopterygii: Merlucciidae); marine; intestine; adult; Magellanic, WTSP; Argentina, Chile (Oliva 1982; Gil de Pertierra et al. 2011).

Note: Gil de Pertierra et al. (2011) studied the vouchers deposited in BMNH; they reported the species as *Clestobothrium* sp., and R. Kuchta (pers. comm.) suggested that it belongs to *C. crassiceps*.

Micromesistius australis australis (Actinopterygii: Gadidae); marine; intestine; adult; Magellanic; Chile (Chávez et al. 2012).

***Clestobothrium cristinae* Gil de Pertierra, Incorvaia & Arrendondo, 2011**

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; intestine; adult; Magellanic; Argentina (Szidat 1955, 1961; MacKenzie and Longshaw 1995; Sardella and Timi 1996, 2004; Gil de Pertierra et al. 2011; Brabec et al. 2015).

Notes: type host. R. Kuchta (pers. comm.) suggested that all reports from *M. hubbsi* before the original description of *C. cristinae* were erroneously reported as *C. crassiceps*. Sequences of 18S (KR780948), 28S (KR780901), 16S (KR780862) and *cox*1 (KR7808301) (Brabec et al. 2015).

***Clestobothrium splendidum* Gil de Pertierra, Incorvaia & Arredondo, 2011**

Merluccius australis (Actinopterygii: Merlucciidae); marine; intestine; adult; Magellanic; Argentina, Chile (Fernández 1985; George-Nascimento and Arancibia 1994; González and Carvajal 1994; MacKenzie and Longshaw 1995; Gil de Pertierra et al. 2011; Chávez et al. 2012; Brabec et al. 2015).

Notes: type host. R. Kuchta (pers. comm.) suggested that all reports from *M. australis* before the original description of *C. splendidum* as well as that of Chávez et al. (2012) were erroneously reported as *C. crassiceps*. Sequences of 18S (KR780967), 28S (KR780920), 16S (KR780877) and *cox*1 KR7808271 (Brabec et al. 2015).

Schyzocotyle acheilognathi* (Yamaguti, 1934) Brabec, Waeschenbach, Scholz, Littlewood & Kuchta, 2015

[For synonyms, see Kuchta and Scholz (2007) and Brabec et al. (2015)]

Cyprinus carpio (Actinopterygii: Cyprinidae); freshwater; intestine; adult; Paraná State (fishpond), Negro River basin; Argentina, Brazil (Rego et al. 1999a; Waicheim et al. 2014).

Notes: these reports from South America are probably result of the import of common carp from Europe to Brazil (Rego et al. 1999a; Scholz et al. 2011). Waicheim et al. (2014) reported the cestode as *Bothriocephalus* sp., but it was most probably *S. acheilognathi* (R. Kuchta, pers. comm.).

Pethia conchonius (Actinopterygii: Cyprinidae); freshwater; intestine; adult; Santa Catarina State; Brazil (Piazza et al. 2006).

Notes: host reported as *Puntius conchonius*.

Poecilia reticulata (Actinopterygii: Poeciliidae); freshwater; intestine; adult; Paraná River Basin; Brazil (Moreira et al. 2014).

Note: tapeworms reported as 'Pseudophyllidea', but considered as *S. acheilognathi* by R. Kuchta (pers. comm.).

Xiphophorus hellerii (Actinopterygii: Poeciliidae); freshwater; intestine; adult; Santa Catarina State; Brazil (Piazza et al. 2006).

Xiphophorus maculatus (Actinopterygii: Poeciliidae); freshwater; intestine; adult; Santa Catarina State; Brazil (Piazza et al. 2006)

Notes: *P. conchonius*, *X. hellerii* and *X. maculatus* are ornamental fish imported to South America (Froese and Pauly 2016); their tapeworms were reported as 'cestodes', but they were most probably conspecific with *S. acheilognathi* (R. Kuchta, pers. comm.).

***Senga* sp.**

Astyanax altiparanae (Actinopterygii: Characidae); freshwater; pyloric caeca; adult; Rio das Pedras Farm (lakes); Brazil (Azevedo et al. 2007).

Astyanax scabripinnis (Actinopterygii: Characidae); freshwater; intestine; adult; São Paulo State; Brazil (Rego 1997).

Unidentified bothriocephalid cestode

Girella laevifrons (Actinopterygii: Kyphosidae); marine; site of infection not given; adult; WTSP; Chile (Muñoz and Delorme 2011).

Family Echinophallidae Schumacher, 1914

Neobothriocephalus aspinosus* Mateo & Bullock, 1966

Seriolella violacea (Actinopterygii: Centrolophidae); marine; intestine, stomach; adult; WTSP; Chile, Peru (Mateo and Bullock 1966; Soto and Carvajal 1979; Oliva 1982; Iannacone 2003; Brabec et al. 2015).

Notes: type host; it was originally reported as *Neptomenus crassus*. Sequences of 18S (KR780944), 28S (KR780897), 16S (KR780857) and *cox*1 (KR780805) (Brabec et al. 2015).

***Neobothriocephalus* sp.**

Hippoglossina macrops (Actinopterygii: Paralichthyidae); marine; intestine; adult; WTSP; Chile (Riffo 1991; González et al. 2001; Oliva et al. 2004; González et al. 2008).

Note: all but one authors reported the cestode as *N. aspinosus*. Kuchta et al. (2008) stated that further analyses should be performed to confirm these records since there is no material deposited in any helminthological collection and cestodes from this fish host may represent a new species.

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; intestine; adult; WTSP; Chile (Riffo 1995).

Paralichthys microps (Actinopterygii: Paralichthyidae); marine; intestine; adult; WTSP; Chile (Riffo 1995).

***Parabothriocephalus* sp.**

Macrourus holotrachys (Actinopterygii: Macrouridae); marine; intestine, pyloric caeca; adult; WTSP: Chile (Ñacari and Oliva 2016).

Family Triaenophoridae Lönnberg, 1899

Ailinella mirabilis* Gil de Pertierra & Semenas, 2006

Aplochiton zebra (Actinopterygii: Galaxiidae); amphidromous; intestine; adult; Patagonian lakes; Argentina (Ortubay et al. 1994; Fernández et al. 2012).

Note: Ortubay et al. (1994) misidentified the parasite as *Nippotaenia* sp. (Gil de Pertierra and Semenas 2006; Fernández et al. 2012).

Galaxias maculatus (Actinopterygii: Galaxiidae); amphidromous; intestine; adult; Moreno and Nahuel Huapi Lake systems (Andean-Patagonian region); Argentina

(Ortubay et al. 1994; Rauque et al. 2003; Revenga et al. 2005; Gil de Pertierra and Semenas 2006; Viozzi et al. 2009; Fernández et al. 2010).
Notes: type host. Ortubay et al. (1994), Rauque et al. (2003) and Revenga et al. (2005) misidentified the parasite as *Nippotaenia* sp. (Gil de Pertierra and Semanas 2006).

Anchistrocephalus microcephalus* (Rudolphi, 1819) Monticelli, 1890
[For synonyms, see Kuchta and Scholz (2007)].
Mola mola (Actinopterygii: Molidae); marine; intestine; adult; WTSA; Brazil (Mendes 1944).
Notes: type host. The tapeworms were reported as *Amphigonophorus carvalhoi* Mendes, 1944. Kennedy and Andersen (1982) synonymized *Amphigonophorus* with *Anchistrocephalus*, which was rejected by Bray et al. (1994), but accepted by Kuchta and Scholz (2007). We are following the recent revision of bothriocephalideans by Kuchta et al. (2008).
Mola ramsayi (Actinopterygii: Molidae); marine; intestine; adult; WTSP; Chile (Villalba and Fernández 1985).

***Anonchocephalus argentinensis* Szidat, 1961**
Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; intestine; adult; WTSA; Argentina (Szidat 1961; Alarcos and Timi 2012, 2013).

Anonchocephalus chilensis* (Riggenbach, 1896) Lühe, 1902
[Syn. *Bothriotaenia chilensis* Riggenbach, 1896]
Genypterus blacodes (Actinopterygii: Ophidiidae); marine; intestine; adult; WTSA, WTSP; Argentina, Chile (Riffo 1994; Sardella et al. 1998; Suriano and Labriola 1998; Brabec et al. 2015).
Notes: Suriano and Labriola (1998) redescribed this species. Sequence of *cox*1 (KR780782) (Brabec et al. 2015).
Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; intestine; adult; WTSA; Argentina, Brazil (Sardella et al. 1998; Pereira 2000).
Note: Pereira (2000) redescribed this species.
Genypterus chilensis (Actinopterygii: Ophidiidae); marine; intestine, pyloric caeca; adult; WTSP; Chile (Riggenbach 1896a; Vergara and George-Nascimento 1982).
Note: type host.
Genypterus maculatus (Actinopterygii: Ophidiidae); marine; intestine; adult; WTSP; Chile (George-Nascimento and Huet 1984).

***Anonchocephalus patagonicus* Suriano & Labriola, 1998**
Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; intestine; adult; Magellanic; Argentina (Suriano and Labriola, 1998).
Note: type host.

***Anonchocephalus* sp.**

Pinguipes brasilianus (Actinopterygii: Pinguipedidae); marine; intestine; adult; WTSA; Argentina (Timi et al. 2009, 2010a).

Galaxitaenia toloi* Gil de Pertierra & Semenas, 2005

Galaxias platei (Actinopterygii: Galaxiidae); amphidromous; intestine; adult; Moreno Lake system (Patagonian region); Argentina (Ortubay et al. 1994; Rauque et al. 2003; Gil de Pertierra and Semenas 2005).

Notes: type host. Ortubay et al. (1994) and Rauque et al. (2003) misidentified the parasite as *Nippotaenia* sp. (Gil de Pertierra and Semenas 2006).

Unidentified bothriocephalideans

Aplochiton taeniatus (Actinopterygii: Galaxiidae); amphidromous; intestine; adult; Patagonian lakes; Argentina (Ortubay et al. 1994).

Note: reported as *Nippotaenia* sp. but the nippotaeniids are not found in the Americas (Bray 1994).

Cichla monoculus (Actinopterygii: Cichlidae); freshwater; intestine; adult; Rio das Pedras Farm (lakes); Brazil (Müller et al. 2008).

Note: reported as *B. cuspidatus* and considered as misidentification by R. Kuchta (pers. comm.).

Odontesthes smitti (Actinopterygii: Atherinopsidae); marine; intestine; metacestode; Magellanic; Argentina (Carballo et al. 2012).

Oncorhynchus mykiss (Actinopterygii: Salmonidae); anadromous; intestine; adult; Moreno and Nahuel Huapi lakes (Patagonian region); Argentina (Rauque et al. 2003).

Note: reported as *Nippotaenia* sp.

Paralabrax humeralis (Actinopterygii: Serranidae); marine; intestine; adult; WTSP; Chile (Henríquez and González 2014).

Percichthys trucha (Actinopterygii: Percichthyidae); freshwater; intestine; adult; Moreno and Nahuel Huapi lakes (Patagonian region); Argentina (Ortubay et al. 1994; Rauque et al. 2003).

Note: reported as *Nippotaenia* sp.

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina (Braicovich and Timi 2010).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1935c).

Note: host reported as *P. squamosissima* and its tapeworms as an unidentified ptychobothriid (Woodland 1935c), but considered only as Bothriocephalidea by Kuchta and Scholz (2007).

Salvelinus fontinalis (Actinopterygii: Salmonidae); anadromous; intestine; adult; Moreno and Nahuel Huapi lakes (Patagonian region); Argentina (Rauque et al. 2003).

Note: reported as *Nippotaenia* sp.

Unidentified bothriocephalideans (identified as 'Pseudophyllidea')

Eleginops maclovinus (Actinopterygii: Eleginopsidae); marine; intestine; adult; Magellanic; Falkland Islands (Brickle and MacKenzie 2007).

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; intestine; adult; WTSP; Chile (Oliva et al. 1996).

Prolatilus jugularis (Actinopterygii: Pinguipedidae); marine; intestine; adult; Magellanic; Chile (Sepúlveda et al. 2004).

Order Caryophyllidea van Beneden in Carus, 1863

[Caryophyllidean tapeworms do not occur in the Neotropical region, where their common hosts, i.e. cyprinid and catostomid fishes, are absent; therefore, these reports need verification]

Unidentified caryophyllideans

Cyprinus carpio (Actinopterygii: Cyprinidae); freshwater; intestine; adult; Paraná State; Brazil (Rego 2004).
Note: introduced fish host (Froese and Pauly 2016).

Geophagus brasiliensis (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná State (dams); Brazil (Bellay et al. 2012).

Order Cathetocephalidea Schmidt & Beveridge, 1990
Family Cathetocephalidae Dailey & Overstreet 1973

***Cathetocephalus australis* Schmidt & Beveridge, 1990**

Carcharhinus brachyurus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Argentina (Suriano and Labriola 2001a).
Note: type host.

Cathetocephalus thatcheri* Dailey & Overstreet, 1973

Carcharhinus leucas (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSP; Peru (Rivera and Sarmiento 1990).
Note: type host.

Family Disculicepitidae Joyeux & Baer, 1935

***Disculiceps galapagoensis* Nock & Caira, 1988**

Carcharhinus longimanus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; Galapagos; Ecuador (Nock and Caira 1988).
Note: type host.

Disculiceps pileatus* (Linton, 1890) Joyeux & Baer, 1936
[Syn. *Discocephalum pileatum* Linton, 1890]
Carcharhinus porosus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSP; Peru (Tantaleán 1991).
Note: tapeworms reported as *Discocephalum pileatum*.

***Disculiceps* sp.**
Aetobatus narinari (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: these specimens probably belong to *Tylocephalum* (Lecanicephalidea), according to Koch et al. (2012).

Order Cyclophyllidea van Beneden in Braun, 1900
Family Gryporhynchidae Spassky & Spasskaya, 1973

***Glossocercus auritus* (Rudolphi, 1819) Bona, 1994**
[For synonyms, see Scholz et al. (2004)]
Poecilia reticulata (Actinopterygii: Poeciliidae); freshwater; mesentery; metacestode; Pampulha Dam, Minas Gerais State; Brazil (Pinto and Melo 2011a).

***Parvitaenia macropeos* (Wedl, 1855) Baer & Bona, 1960**
[For synonyms, see Scholz et al. (2004)]
Australoheros facetus (Actinopterygii: Cichlidae); freshwater; intestine; metacestode; Pampulha Dam, Minas Gerais State; Brazil (Pinto and Melo 2011b).

***Valipora campylancristrota* (Wedl, 1855) Baer & Bona, 1960**
[For synonyms, see Scholz et al. (2004)]
Geophagus brasiliensis (Actinopterygii: Cichlidae); freshwater; gallbladder; metacestode; Paraná State (dams); Brazil (Bellay et al. 2012).
Hoplosternum littorale (Actinopterygii: Callichthyidae); freshwater; gallbladder; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).
Prochilodus lineatus (Actinopterygii: Prochilodontidae); freshwater; gallbladder; metacestode; Paraná River basin; Brazil (Lizama et al. 2005, 2006).

***Valipora* sp.**
Crenicichla britskii (Actinopterygii: Cichlidae); freshwater; gallbladder; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).
Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; gallbladder; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).
Prochilodus argenteus (Actinopterygii: Prochilodontidae); freshwater; gallbladder; metacestode; São Francisco River basin; Brazil (Monteiro et al. 2009).

Unidentified cyclophyllideans

Dormitator maculatus (Actinopterygii: Eleotridae); amphidromous; liver, intestine, gonads; metacestode; TNA; Venezuela (Moreno et al. 2008).

Percichthys trucha (Actinopterygii: Percichthyidae); freshwater; body cavity; metacestode; Negro River basin (Patagonian region); Argentina (Ortubay et al. 1994).

Satanoperca pappaterra (Actinopterygii: Cichlidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Kohn et al. 2011).

Order Diphyllidea van Beneden in Carus, 1863
Family Echinobothriidae Perrier, 1897

Ahamulina catarina* Marques, Jensen & Caira, 2012

Scyliorhinus besnardi (Elasmobranchii: Scyliorhinidae); marine; spiral valve; adult; WTSA; Brazil (Marques et al. 2012; Caira et al. 2013).

Notes: type host. Sequences of partial 18S (KC860176–KC860180), 28S (KC860128–KC860132) and *cox*1 (KC860220–KC860224) (Caira et al. 2013).

***Coronocestus notoguidoi* (Ivanov, 1997) Caira, Marques, Jensen, Kuchta & Ivanov, 2013**

[Syn. *Echinobothrium notoguidoi* Ivanov, 1997]

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WSTA; Argentina (Ivanov 1997; Alarcos et al. 2006; Tyler 2006).

Notes: type host. Tyler (2006) provided new morphological data based on examination of the type specimens deposited in MLP and USNPC.

***Halysioncum euzeti* (Campbell & Carvajal, 1980) Caira, Marques, Jensen, Kuchta & Ivanov, 2013**

[Syn. *Echinobothrium euzeti* Campbell & Carvajal, 1980]

Sympterygia lima (Elasmobranchii: Arhynchobatidae) marine; spiral valve; adult; WTSP; Chile (Campbell and Carvajal 1980; Tyler 2006).

Notes: type host; it was originally reported as *Psammobatis lima*. Tyler (2006) provided new morphological data based on examination of the type specimens deposited in USNPC.

***Halysioncum megacanthum* (Ivanov & Campbell, 1998) Caira, Marques, Jensen, Kuchta & Ivanov, 2013**

[Syn. *Echinobothrium megacanthum* Ivanov & Campbell, 1998]

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; Magellanic; Argentina (Ivanov and Campbell 1998b; Tyler 2006).

Notes: type host. Tyler (2006) provided new morphological data based on examination of the type specimens deposited in MLP and USNPC.

***Halysioncum pigmentatum* (Ostrowski de Núñez, 1971) Caira, Marques, Jensen, Kuchta & Ivanov, 2013**

[Syn. *Echinobothrium pigmentatum* Ostrowski de Núñez, 1971]

Zapteryx brevirostris (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1971; Tyler 2006).

Note: type host. Tyler (2006) provided new morphological data based on examination of the type specimens deposited in the Ostrowski de Núñez's collection.

Unidentified diphyllideans

Notothenia cf. *angustata* (Actinopterygii: Nototheniidae); marine; intestine; metacestode; WTSP; Chile (Muñoz et al. 2001).

Sebastes capensis (Actinopterygii: Sebastidae); marine; unspecified site of infection; metacestode; WTSP; Chile (González and Poulin 2005a, b; González et al. 2006).

Note: two morphotypes were distinguished by González et al. (2006).

Order Diphyllobothriidea Kuchta, Scholz, Brabec & Bray, 2008
Family Diphyllobothriidae Lühe, 1910

***Adenocephalus pacificus* Nybelin, 1931**

[For synonyms, see Hernández-Orts et al. (2015)]

Anisotremus scapularis (Actinopterygii: Haemulidae); marine; body cavity, viscera; metacestode; WTSP; Peru (Luque 1991).

Ariopsis seemanni (Actinopterygii: Ariidae); brackish, marine; peritoneum; metacestode; WTSP; Peru (Escalante and Miranda 1986).

Note: Escalante and Miranda (1986) performed experimental infection in dogs; they reported the host as *Galeichthys jordani*.

Cilus gilberti (Actinopterygii: Sciaenidae); marine; viscera; metacestode; WTSP; Peru (Chero et al. 2014b).

Cynoscion analis (Actinopterygii: Sciaenidae); marine; viscera; metacestode; WTSP; Peru (Escalante 1983).

Galeichthys peruvianus (Actinopterygii: Ariidae); marine; viscera, peritoneum; metacestode; WTSP; Peru (Kuchta et al. 2015).

Note: Kuchta et al. (2015) provided a list of records gathered from published data in an appendix.

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; viscera; metacestode; WTSP; Peru (Escalante 1983).

Menticirrhus ophicephalus (Actinopterygii: Sciaenidae); marine; body cavity, viscera; metacestode; WTSP; Peru (Luque 1991).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; peritoneum, viscera, stomach surface; metacestode; WTSP; Peru (Escalante 1983; Chero et al. 2014a).

Note: Chero et al. (2014a) also reported *D. arctocephalinum* Johnston, 1937 from this host, but this species is a junior synonym of *A. pacificus* (see Hernández-Orts et al. 2015).

Paralabrax humeralis (Actinopterygii: Serranidae); marine; site of infection not given; metacestode; WTSP; Peru (Iannacone and Alvariño 2009).

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; stomach surface; metacestode; WTSP; Peru (Escalante and Miranda 1986).

Paralonchurus peruanus (Actinopterygii: Sciaenidae); marine; intestinal surface; metacestode; WTSP; Peru (Tantaleán 1975).

Note: the author reported the host as *Polyclemus peruanus*.

Sarda chiliensis (Actinopterygii: Scombridae); marine; body cavity; metacestode; WTSP; Peru (Baer 1969; Hernández-Orts et al. 2015; Kuchta et al. 2015).

Notes: the genus *Adenocephalus* Nybelin, 1931 was resurrected by Hernández-Orts et al. (2015), based on morphological and molecular data. Sequences of partial 28S (KR269760) and *cox*1 (KR269747) (Hernández-Orts et al. 2015).

Sciaena callaensis (Actinopterygii: Sciaenidae); marine; peritoneum, stomach surface; metacestode; WTSP; Peru (Tantaleán 1975).

Sciaena deliciosa (Actinopterygii: Sciaenidae); marine; peritoneum, stomach surface, viscera; metacestode; WTSP; Peru (Tantaleán 1975; Escalante and Miranda 1986; Llerena et al. 2001; Wicht et al. 2010; Chero et al. 2014c; Kuchta et al. 2015).

Note: Chero et al. (2014a) also reported *D. arctocephalinum* (syn. of *A. pacificus*) from this host.

Scomberomorus sierra (Actinopterygii: Scombridae); marine; body cavity; metacestode; WTSP; Peru (Baer 1969).

Note: host reported as *S. maculatus*.

Seriolella violacea (Actinopterygii: Centrolophidae); marine; peritoneum; metacestode; WTSP; Peru (Escalante and Miranda 1986).

Trachinotus paitensis (Actinopterygii: Carangidae); marine; peritoneum; metacestode; WTSP; Peru (Escalante and Miranda 1986).

Trachurus murphyi (Actinopterygii: Carangidae); marine; body cavity; metacestode; WTSP; Peru (Kuchta et al. 2015).

***Diphyllobothrium dendriticum* (Nitzsch, 1824) Lühe, 1910**

[For synonyms, see Scholz et al. (2009)]

Basilichthys australis (Actinopterygii: Atherinopsidae); freshwater; mesentery, liver, muscles; metacestode; Riñihue and Panguipulli Lakes; Chile (Torres et al. 1998, 2004, 2012).

Galaxias maculatus (Actinopterygii: Galaxiidae); amphidromous; liver, body cavity, intestinal surface; metacestode; Nahuel Huapi (Patagonian region), Riñihue Lakes; Argentina, Chile (Ortubay 1994; Torres et al. 1998; Viozzi et al. 2009).

Odontesthes mauleanum (Actinopterygii: Atherinopsidae); freshwater; mesentery; metacestode; Panguipulli Lake; Chile (Torres et al. 2004, 2012).

Oncorhynchus kisutch (Actinopterygii: Salmonidae); anadromous; intestinal surface, mesentery, spleen; metacestode; Llanquihue Lake; Chile (Torres 1990).

Oncorhynchus mykiss (Actinopterygii: Salmonidae); anadromous; body cavity, mesentery, internal organs, muscles; metacestode; lakes of Valdivia River basin, Huechulaufquen, Rosario, Moreno and Nahuel Huapi Lakes (Patagonian region), lakes of Chiloé Island; Argentina, Chile (Szidat and Soria 1957; Szidat 1964; Torres et al. 1981, 1983, 1989a, b, 1991, 2004, 2012; Revenga 1993; Semenas and Kreiter 1995; Rozas et al. 2012).

Notes: tapeworms described as *D. microcordiceps* by Szidat and Soria (1957). Host reported as *Salmo gairdneri* by some authors. Sequences of partial 18S +ITS1 + 5.8S + ITS2 (JN153006–JN153018) and partial *cox*1 (JN152993–JN153005) (Rozas et al. 2012).

Percichthys trucha (Actinopterygii: Percichthyidae); freshwater; body cavity, mesentery, internal organs, muscles; metacestode; Riñihue Lake; Chile (Torres et al. 1998).

Percilia gillissi (Actinopterygii: Perciliidae); freshwater; body cavity, mesentery, internal organs, muscles; metacestode; Riñihue Lake; Chile (Torres et al. 1989a).

Salmo salar (Actinopterygii: Salmonidae); anadromous; muscles; metacestode; Nahuel Huapi Lake (Patagonian region); Argentina (Szidat and Soria 1957; Szidat 1964).

Salmo trutta (Actinopterygii: Salmonidae); anadromous; body cavity; metacestode; Huechulaufquen and Rosario Lakes (Patagonian region), Valdivia River basin; Argentina, Chile (Torres et al. 1989a, b, 1991; Semenas and Kreiter 1995).

Salvelinus fontinalis (Actinopterygii: Salmonidae); anadromous; body cavity, muscles; metacestode; Huechulaufquen, Rosario, Moreno, Nahuel Huapi Lakes (Patagonian region); Argentina (Szidat and Soria 1957; Szidat 1964; Revenga 1993; Semenas and Kreiter 1995).

***Diphyllobothrium latum* (Linnaeus, 1758) Lühe, 1910**

[For synonyms see Scholz et al. (2009) and references therein]

Basilichthys australis (Actinopterygii: Atherinopsidae); freshwater; muscles; metacestode; Panguipulli Lake; Chile (Torres et al. 2004, 2012).

Diplomystes camposensis (Actinopterygii: Diplomystidae); freshwater; liver; metacestode; Riñihue Lake; Chile (Torres et al. 1989a).

Note: the authors reported the tapeworms as *Diphyllobothrium* sp., but Muñoz and Olmos (2008) and R. Kuchta (pers. comm.) suggested that they belong to *D. latum*.

Galaxias maculatus (Actinopterygii: Galaxiidae); amphidromous; body cavity, muscles; metacestode; Panguipulli and Riñihue Lakes; Chile (Torres et al. 1989a, 1998, 2004).

Galaxias platei (Actinopterygii: Galaxiidae); amphidromous; body cavity; metacestode; Valdivia River Basin; Chile (Torres et al. 1989a).

Odontesthes mauleanum (Actinopterygii: Atherinopsidae); freshwater; liver, gonads, mesentery, muscles; metacestode; Panguipulli Lake; Chile (Torres et al. 2004, 2012).

Oncorhynchus mykiss (Actinopterygii: Salmonidae); anadromous; body cavity, internal organs, muscles; metacestode; lakes of Valdivia River basin, Huechulaufquen, Rosario, Moreno and Nahuel Huapi Lakes (Patagonian region); Argentina, Chile (Neghme et al. 1950; Neghme and Bertín 1951; Torres et al. 1989a, 1991, 2004, 2012; Revenga 1993; Semenas and Kreiter 1995).

Note: host reported as *Salmo gairdneri* by some authors.

Percichthys trucha (Actinopterygii: Percichthyidae); freshwater; body cavity, muscles; metacestode; lakes of Valdivia River basin; Chile (Torres et al. 1989a, 1998, 2004, 2012).

Percichthys sp. (Actinopterygii: Percichthyidae); freshwater; body cavity, pyloric caeca, liver, stomach, gonads, muscles; metacestode; Moreno and Nahuel Huapi Lakes (Patagonian region); Argentina (Revenga 1993).

Salmo trutta (Actinopterygii: Salmonidae); anadromous; body cavity; metacestode; lakes of Valdivia River basin, Huechulaufquen and Rosario Lakes (Patagonian region); Argentina, Chile (Neghme et al. 1950; Neghme and Bertín 1951; Torres et al. 1989a, b, 1991; Semenas and Kreiter 1995).

Note: host reported as *Salmo trutta trutta* and *Salmo trutta fario* by some authors.

Salvelinus fontinalis (Actinopterygii: Salmonidae); anadromous; liver; metacestode; Huechulaufquen, Moreno, Nahuel Huapi and Rosario Lakes (Patagonian region); Argentina (Revenga 1993; Semenas and Kreiter 1995).

***Diphyllobothrium* sp.**

[Reports from freshwater fishes most likely correspond to *D. dendriticum* or *D. latum* (R. Kuchta, pers. comm.). All reports of unidentified diphyllobothrideans are included in this section]

Basilichthys australis (Actinopterygii: Atherinopsidae); freshwater; liver; metacestode; Riñihue Lake; Chile (Torres et al. 1989a, 1998).

Cilus gilberti (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSP; Chile (García et al. 2001).

Cynoscion analis (Actinopterygii: Sciaenidae); marine; body cavity, peritoneum, stomach surface; metacestode; WTSP; Peru (Escalante 1983).

Engraulis ringens (Actinopterygii: Engraulidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Moscoso 2013).

Galaxias maculatus (Actinopterygii: Galaxiidae); amphidromous; body cavity, liver; metacestode; Moreno Lake (Patagonian region); Argentina (Ortubay 1994; Viozzi et al. 2009).

Galaxias platei (Actinopterygii: Galaxiidae); amphidromous; liver; metacestode; Riñihue Lake; Chile (Torres et al. 1989a).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; body cavity, intestinal serosa, intestine, muscles; metacestode; WTSA; Brazil (Knoff et al. 2008).

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; body cavity, peritoneum, stomach surface; metacestode; WTSP; Chile, Peru (Escalante 1983, George-Nascimento and Huet 1984).

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; body cavity, intestinal serosa; metacestode; WTSA; Brazil (Knoff et al. 2011).

Merluccius australis (Actinopterygii: Merlucciidae); marine; stomach wall; metacestode; Magellanic, WTSP; Chile, Falkland Islands (MacKenzie and Longshaw 1995).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; body cavity, mesentery, peritoneum, stomach surface; metacestode: WTSP; Peru (Escalante 1983; Jara 1998).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; stomach wall; metacestode; Magellanic; Argentina, Falkland Islands (MacKenzie and Longshaw 1995).

Micromesistius australis australis (Actinopterygii: Gadidae); marine; site of infection not given; metacestode; WTSP; Chile (Niklitschek et al. 2010; George-Nascimento et al. 2011; Chávez et al. 2012).

Odontesthes regia (Actinopterygii: Atherinopsidae); freshwater; liver, gonads; metacestode; Riñihue Lake; Chile (Torres et al. 1989a, 1998).

Oncorhynchus kisutch (Actinopterygii: Salmonidae); anadromous; stomach, spleen, liver, mesentery, gonads; metacestode; Aisén River basin; Chile (Torres et al. 1995, 2000).

Oncorhynchus mykiss (Actinopterygii: Salmonidae); anadromous; body cavity, internal organs, mesentery, muscles; metacestode; lakes of Valdivia River basin, Moreno and Nahuel Huapi Lakes (Patagonian region), Tarahuin Lake (Chiloe Island); Argentina, Chile (Wolffhügel 1949; Torres et al. 1977, 1980, 1982, 1989a, 2002, 2010; González et al. 1978, 1980; Revenga and Semenas 1991; Revenga et al. 1995; Torres and Puga 2011).

Notes: host reported as *S. gairdneri* or *S. gairdneri irideus* by some authors. After experimental infections of small rodents with metacestodes, González et al. (1980) recovered tapeworms morphologically similar with *D. dentriticum*.

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; body cavity, mesentery, liver, ovary, stomach; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Note: Felizardo et al. (2010) distinguished two morphotypes.

Paralonchurus peruanus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSP; Peru (Tantaleán 1975).

Note: host reported as *Polyclemus peruanus*.

Salmo trutta (Actinopterygii: Salmonidae); anadromous; body cavity, peritoneum, liver, mesentery, muscles; metacestode; Rupanco and Calafquén Lakes, Moreno and Nahuel Huapi Lakes (Patagonian region); Argentina, Chile (González et al. 1978, 1980; Torres et al. 1980; Revenga and Semenas 1991).

Note: host reported as *S. trutta fario* or *S. trutta trutta* by some authors.

Salvelinus fontinalis (Actinopterygii: Salmonidae); anadromous; body cavity, muscles; metacestode; Moreno and Nahuel Huapi Lakes (Patagonian region); Argentina (Revenga and Semenas 1991; Revenga et al. 1995).

Sciaena callaensis (Actinopterygii: Sciaenidae); marine; peritoneum, stomach surface, body cavity; metacestode; WTSP; Peru (Escalante 1983).

Sciaena deliciosa (Actinopterygii: Sciaenidae); marine; body cavity, intestinal surface; metacestode; WTSP; Peru (Tantaleán 1975; Llerena et al. 2001).

Scomber japonicus (Actinopterygii: Scombridae); marine; site of infection not given; metacestode; WTSP; Peru (Oliva et al. 2008b).

Sebastes capensis (Actinopterygii: Sebastidae); marine; site of infection not given; metacestode; Magellanic; Chile (González and Poulin 2005a, b; González et al. 2006).

Trachurus murphyi (Actinopterygii: Carangidae); marine; liver; metacestode; WTSP; Chile, Peru (Pérez et al. 1999; George-Nascimento and Oliva 2015).

Unidentified 'Pseudophyllidea' (larval stages)

[Larval stages found in the body cavity and mesentery are most likely species of *Diphyllobothrium* (R. Kuchta, pers. comm.)]

Aphos porosus (Actinopterygii: Batrachoididae); marine; body cavity; metacestode; WTSP; Chile (Cortés and Muñoz 2009).

Balistes capriscus (Actinopterygii: Balistidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves et al. 2005).

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; anterior intestine; metacestode; Magellanic; Chile, Falkland Islands (Brickle et al. 2006; Oliva et al. 2008a; Brown et al. 2013).

Engraulis anchoita (Actinopterygii: Engraulidae); marine; mesentery; metacestode; WTSA; Argentina (Timi 2003; Timi and Poulin 2003; Timi et al. 2010b).

Gobiesox marmoratus (Actinopterygii: Gobiesocidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz 2014).

Helcogrammoides chilensis (Actinopterygii: Tripterygiidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz and Delorme 2011).

Hypsoblennius sordidus (Actinopterygii: Blenniidae); marine; intestine; metacestode; Magellanic; Chile (Sepúlveda et al. 2004).

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; body cavity; metacestode; Magellanic; Argentina, Chile (Oliva 2001; MacKenzie et al. 2013).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; stomach wall; metacestode; WTSP; Chile (Oliva and Ballón 2002).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; Magellanic; Argentina (Sardella and Timi 2004).

Note: Sardella and Timi (2004) distinguished two morphotypes.

Micromesistius australis australis (Actinopterygii: Gadidae); marine; unspecified site of infection; metacestode; Magellanic; Chile (Niklitschek et al. 2010; George-Nascimento et al. 2011).

Odontesthes regia (Actinopterygii: Atherinopsidae); marine; intestine; metacestode; Magellanic; Chile (Sepúlveda et al. 2004).

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina, Uruguay (Braicovich and Timi 2008).

Prolatilus jugularis (Actinopterygii: Pinguipedidae); marine; intestine; metacestode; Magellanic; Chile (Sepúlveda et al. 2004).

Scartichthys viridis (Actinopterygii: Bleniidae); marine; site of infection not given; metacestode; WTSA; Chile (Muñoz and Delorme 2011; Muñoz and Randhawa 2011).

Sicyases sanguineus (Actinopterygii: Gobiesocidae); marine; site of infection not given; metacestode; WTSA; Chile (Muñoz and Delorme 2011; Muñoz and Randhawa 2011).

Trachurus lathami (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Braicovich et al. 2012).

Trachurus murphyi (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Arancibia 1992; George-Nascimento 2000; George-Nascimento and Oliva 2015).

Order Gyrocotylidea Poche, 1926
Family Gyrocotylidae Benham, 1901

***Gyrocotyle maxima* MacDonagh, 1927**

[Syns. *Gyrocotyle meandrica* Mendívil-Herrera, 1946; *G. urna* sensu Manter, 1951; *Amphiptyches urna* Spencer, 1889]

Callorhinchus callorynchus (Holocephali: Callorhinchidae); marine; spiral valve; adult; WTSA, WTSP; Brazil, Chile, Peru, Uruguay (Mendívil-Herrera 1946; Rego et al. 1974; Fernández et al. 1986; Tantaleán 1991).

Note: tapeworms reported as *G. meandrica* by Mendívil-Herrera (1946) and Rego et al. (1974).

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Argentina (MacDonagh 1927).

Notes: type host; it was reported as *Mustelus asterias*, but most likely MacDonagh (1927) misidentified the elephant fish *C. callorynchus* (see Caira et al. 2012), common definitive host of *G. maxima*.

Gyrocotyle rugosa* Diesing, 1850

[Syn. *Gyrocotyle plana* Linton, 1924]

Callorhinchus callorynchus (Holocephali: Callorhinchidae); marine; spiral valve; adult; WTSA, WTSP; Argentina, Chile (MacDonagh 1927; Fernández et al. 1986).

Note: type host.

Unidentified gyrocotylidean

Callorhinchus callorynchus (Holocephali: Callorhinchidae); marine; spiral valve; adult; WTSA; Uruguay (Mendívil-Herrera 1946).

Order Lecanicephalidea Wardle & McLeod, 1952
Family Aberrapecidae Jensen, Caira, Cielocha, Littlewood & Waeschenbach, 2016

***Aberrapex arrhynchum* (Brooks, Mayes & Thorson, 1981) Jensen, 2001**
[Syn. *Discobothrium arrhynchum* Brooks, Mayes & Thorson, 1981]
Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (estuary of the La Plata River); Uruguay (Brooks et al. 1981a; Jensen 2001).
Notes: type host.

Family Cephalobothriidae Pintner, 1928

***Tylocephalum brooksi* Ivanov & Campbell, 2000**
Rhinoptera bonasus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Ivanov and Campbell 2000).
Note: type host.

***Tylocephalum* sp.**
Rhinoptera bonasus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Family Lecanicephalidae Braun, 1900

Lecanicephalum peltatum* Linton, 1890
Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TEP; Colombia (Brooks and Mayes 1980).

Family Paraberrapecidae Jensen, Caira, Cielocha, Littlewood & Waeschenbach, 2016

***Paraberrapex atlanticus* Mutti & Ivanov, 2016**
Squatina guggenheim (Elasmobranchii: Squatinidae); marine; spiral valve; adult; Magellanic, WTSA; Argentina (Mutti and Ivanov 2016).
Note: type host.

Family Polypocephalidae Meggitt, 1924

***Polypocephalus medusia* (Linton, 1890) Southwell, 1925**
[Syn. *Parataenia medusia* Linton, 1890]
Dasyatis americana (Elasmobranchii: Dasyatidae) marine; spiral valve; adult; TEP; Colombia (Brooks and Mayes 1980).

Order Onchoproteocephalidea Caira, Jensen, Waeschenbach, Olson & Littlewood, 2014
(Syns. Proteocephalidea Mola, 1928; Tetraphyllidea Carus, 1863 *pro parte*)

Family Onchobothriidae Braun, 1900

***Acanthobothrium amazonense* Mayes, Brooks & Thorson, 1978**
Potamotrygon constellata (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Mayes et al. 1978; Brooks et al. 1981b).
Notes: type host; it was reported as *P. circularis*. Brooks et al. (1981b) studied the type specimens deposited in USNM.

***Acanthobothrium americanum* Campbell, 1969**
Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: type host.

***Acanthobothrium annapinkiense* Carvajal & Goldstein, 1971**
Zearaja chilensis (Elasmobranchii: Rajidae); marine; spiral valve; adult; Magellanic; Chile (Carvajal and Goldstein 1971).
Note: type host; it was reported as *Raja chilensis*.

***Acanthobothrium atahualpai* Marques, Brooks & Barriga, 1997**
Gymnura afuerae (Elasmobranchii: Gymnuridae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).
Note: type host.

***Acanthobothrium batailloni* Euzet, 1955**
Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal and Jeges 1980; Oliva 1982; Escalante 1986).

***Acanthobothrium brevissime* Linton, 1908**
Myliobatis peruvianus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Tantaleán 1991).

***Acanthobothrium campbelli* Marques, Brooks & Monks, 1995**
Dasyatis longa (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).

***Acanthobothrium cartagenense* Brooks & Mayes, 1980**
Urobatis jamaicensis (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).
Note: type host; it was reported as *Urolophus jamaicensis*.

***Acanthobothrium chilense* Rego, Vicente & Herrera, 1968**

Sarda chiliensis (Actinopterygii: Scombridae); marine; intestine; adult; WTSP; Peru (Rego et al. 1968).

Notes: type host. Elasmobranchs are the typical definitive host for *Acanthobothrium* species (Campbell and Beveridge 2002); therefore, the record of adult specimens from a bony fish host needs verification.

***Acanthobothrium colombianum* Brooks & Mayes, 1980**

Aetobatus narinari (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).

Note: type host.

***Acanthobothrium coquimbense* Carvajal & Jeges, 1980**

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Jeges 1980).

Note: type host.

***Acanthobothrium costarricense* Marques, Brooks & Monks, 1995**

Dasyatis longa (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).

Note: type host.

***Acanthobothrium dasybati* Yamaguti, 1934**

Unidentified ray host (Elasmobranchii); marine; spiral valve; adult (only one immature specimen; WTSA; Brazil (Rego et al. 1974).

Note: this species was described from *Dasyatis akajei* in the Western Pacific (Japanese Sea) and its report from the Brazilian coast needs verification.

***Acanthobothrium electricolum* Brooks & Mayes, 1978**

Narcine brasiliensis (Elasmobranchii: Narcinidae); marine; spiral valve; adult; TNA; Colombia, Venezuela (Brooks and Mayes 1978; Mayes and Brooks 1981).

Note: type host.

***Acanthobothrium fogeli* Goldstein, 1964**

Gymnura micrura (Elasmobranchii: Gymnuridae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Note: type host.

***Acanthobothrium gonzalesmugaburoi* Severino & Sarmiento, 1979**

Myliobatis peruvianus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Severino and Sarmiento 1979).

Note: type host.

***Acanthobothrium himanturi* Brooks, 1977**

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks 1977).
Note: type host.

***Acanthobothrium holorhini* Alexander, 1953**

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Rodriguez and Tantaleán 1980)

***Acanthobothrium lintoni* Goldstein, Henson & Schlicht, 1968**

Narcine brasiliensis (Elasmobranchii: Narcinidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1978).
Note: type host.

***Acanthobothrium lusarmientoi* Severino & Verano, 1980**

Sympterygia brevicaudata (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Peru (Severino and Verano 1980).
Note: type host; it was reported as *Psammobatis caudispina*.

***Acanthobothrium marplatense* Ivanov & Campbell, 1998**

Atlantoraja castelnaui (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov and Campbell 1998a).
Note: type host; it was reported as *Rioraja castelnaui*.

***Acanthobothrium minusculum* Marques, Brooks & Barriga, 1997**

Urobatis tumbesensis (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).
Note: type host; it was reported as *Urolophus tumbesensis*.

***Acanthobothrium monksi* Marques, Brooks & Barriga, 1997**

Aetobatus narinari (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).
Note: type host.

***Acanthobothrium obuncum* Marques, Brooks & Barriga, 1997**

Dasyatis longa (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TEP; Ecuador (Marques et al. 1997).
Note: type host.

***Acanthobothrium olseni* Dailey & Mudry, 1968**

Rhinobatos planiceps (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Dailey and Carvajal 1976; Iannacone et al. 2011).

***Acanthobothrium peruviense* Reyda, 2008**

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda 2008).
Note: host reported as *Potamotrygon* cf. *castexi* and the tapeworms as *Acanthobothrium* cf. *peruviense*.

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda 2008).
Note: type host.

***Acanthobothrium psammobati* Carvajal & Goldstein, 1969**

Psammobatis scobina (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal and Goldstein 1969; Carvajal et al. 1985; Tantaleán 1991).
Note: type host.

Sympterygia brevicaudata (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Ruíz 1987).

***Acanthobothrium quinonese* Mayes, Brooks & Thorson, 1978**

Potamotrygon magdalenae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Magdalena River basin; Colombia (Mayes et al. 1978; Brooks et al. 1981b).
Notes: type host. Brooks et al. (1981b) studied the type specimens deposited in USNM.

Potamotrygon yepezi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Maracaibo basin; Venezuela (Brooks et al. 1981b).

***Acanthobothrium ramiroi* Ivanov, 2005**

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Argentina, Peru (Ivanov 2005; Reyda 2008).
Notes: type host. Reyda (2008) reported the tapeworms as *Acanthobothrium* cf. *ramiroi*.

***Acanthobothrium regoi* Brooks, Mayes & Thorson, 1981**

Potamotrygon falkneri (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Lacerda et al. 2008, 2009).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Brooks and Amato 1992).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Orinoco River basin; Venezuela (Brooks et al. 1981b).
Note: type host; it was reported as *P. hystrix*.

***Acanthobothrium robustum* Alexander, 1953**

Rhinobatos planiceps (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSP; Peru (Escalante 1986).

***Acanthobothrium tasajerasi* Brooks, 1977**

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; Maracaibo basin; Venezuela (Mayes and Brooks 1981).

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks 1977).
Note: type host.

***Acanthobothrium terezae* Rego & Dias, 1976**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).
Notes: Reyda and Marques (2011) reported the tapeworms as *Acanthobothrium* cf. *terezae*. Sequence of partial *cox*1 (JF803661) (Reyda and Marques 2011).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Rego and Dias 1976).
Note: type host; it was reported as *Paratrygon motoro* and *Elipesurus* sp.

***Acanthobothrium tortum* (Linton, 1916) Baer & Euzet, 1962**

Aetobatus narinari (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: type host.

***Acanthobothrium urotrygoni* Brooks & Mayes, 1980**

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Urotrygon venezuelae (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).
Note: type host.

***Acanthobothrium zapterycum* Ostrowski de Núñez, 1971**

Zapteryx brevirostris (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1971).
Note: type host.

***Acanthobothrium* sp.**

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Tresierra et al. 1986).

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (La Plata River estuary); Uruguay (Brooks et al. 1981a).
Note: the authors distinguished two morphotypes.

Sympterygia brevicaudata (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Ruíz 1987).

Zapteryx brevirostris (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1971).

Acanthobothroides thorsoni* Brooks, 1977

Dasyatis dipterura (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; WTSP; Peru (Tantaleán and Rodríguez 1987).
Note: host reported as *D. brevis*.

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks 1977).
Note: type host.

***Platybothrium auriculatum* Yamaguti, 1952**

[Syns. *Platybothrium baeri* Euzet, 1952; *Cylindrophorus posteroporus* Riser, 1955]

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; TSA, WTSP; Brazil, Chile, Peru (Carvajal 1974; Rego and Mayer 1976; Escalante 1986).
Note: Healy (2003) revised the genus *Platybothrium* Linton, 1890.

***Platybothrium* sp.**

Sphyrna zygaena (Elasmobranchii: Sphyrnidae); marine; spiral valve; adult; WTSP; Peru (López de McDonald and Tantaleán 1985).

***Potamotrygonocestus amazonensis* Mayes, Brooks & Thorson, 1981**

Potamotrygon constellata (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Brooks et al. 1981b; Mayes et al. 1981; Marques et al. 2003).
Notes: type host; it was reported as *P. circularis*. Marques et al. (2003) reported this host as *P. orbignyi*, but according to Brooks and Amato (1992), all potamotrygonids collected near Leticia, Colombia should be considered as *P. constellata*.

Potamotrygon falkneri (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Marques et al. 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil (Marques et al. 2003).
Note: Marques et al. (2003) redescribed this species.

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Orinoco River basins; Brazil, Venezuela (Brooks et al. 1981b; Marques et al. 2003).
Note: host reported as *P. reticulatus* by Brooks et al. (1981b).

Potamotrygon scobina (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).

Potamotrygon yepezi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Maracaibo basin; Venezuela (Brooks et al. 1981b).

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).

Potamotrygonocestus chaoi **Marques, Brooks & Araujo, 2003**

Plesiotrygon iwamae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003; Luchetti et al. 2008).
Notes: type host. Luchetti et al. (2008) redescribed this species.

Potamotrygonocestus fitzgeraldae **Marques, Brooks & Araujo, 2003**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil, Peru (Marques et al. 2003; Reyda 2008).
Notes: type host. Reyda (2008) reported this species as *P*. cf. *fitzgeraldae*.

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Caira et al. 2014).
Notes: host reported as *P. castexi* and the tapeworms as *P*. cf. *fitzgeraldae*. Sequences of partial 18S (KF685832) and 28S (KF685773) (Caira et al. 2014).

Potamotrygon leopoldi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina (Marques et al. 2003).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Marques et al. 2003).

Potamotrygonocestus magdalenensis **Brooks & Thorson, 1976***

Potamotrygon magdalenae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Magdalena River basin; Colombia (Brooks and Thorson 1976; Brooks et al. 1981b; Caira and Orringer 1995; Marques et al. 2003).
Note: type host.

Potamotrygonocestus marajoara **Luchetti, Marques & Charvet-Almeida, 2008**

Plesiotrygon iwamae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin (estuary); Brazil (Luchetti et al. 2008).
Note: type host.

Potamotrygonocestus maurae **Marques, Brooks & Araujo, 2003**

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).
Note: type host.

Potamotrygonocestus travassosi **Rego, 1979**

[Syn. *Potamotrygonocestus orinocoensis* Brooks, Mayes & Thorson, 1981]

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).

Potamotrygon constellata (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).

Potamotrygon falkneri (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Lacerda et al. 2008, 2009).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Brooks and Amato 1992).
Note: tapeworms reported as *Potamotrygonocestus orinocoensis*.

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Orinoco River basins; Brazil, Venezuela (Rego 1979; Brooks et al. 1981b; Brooks and Amato 1992; Marques et al. 2003).
Notes: type host; it was reported as *P. reticulatus* and *P. hystrix*. The taxon was considered a *species inquirenda* by Brooks et al. (1981b) and Brooks and Amato (1992), but its validity was confirmed by Marques et al. (2003) who also considered *Potamotrygonocestus orinocoensis*, both described from this host, a junior synonym of *P. travassosi*.

***Potamotrygonocestus* sp.**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda and Olson 2003).
Note: Reyda and Olson (2003) reported hyperparasitism caused by metacestodes of proteocephalids.

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda 2008).
Note: host reported as *Potamotrygon* cf. *castexi*.

Potamotrygon henlei (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Tocantins-Araguaia River basin; Brazil (Marques et al. 2003).
Note: these cestodes may represent an undescribed species of the genus (Marques et al. 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda 2008).

Potamotrygon schroederi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2003).
Note: these cestodes may represent an undescribed species of the genus (Marques et al. 2003).

Family Prosobothriidae Baer & Euzet, 1955

Prosobothrium armigerum* Cohn, 1902

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSP; Peru (Rivera and Sarmiento 1990).

Family Proteocephalidae La Rue, 1911

[Even though recent molecular data suggest that most of the traditionally recognized subfamilies are artificial, i.e. non-monophyletic, we are following Woodland's subfamilial classification for practical reasons]

Subfamily Corallobothriinae Freze, 1965

***Corallotaenia* sp.**

Ageneiosus pardalis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult (immature specimens); Magdalena River basin; Colombia (Brooks and Deardorff 1980).

Notes: host reported as *A. caucanus*. This is the first record of the genus in South America (Brooks and Deardorff 1980), but since only immature specimens were found, this needs verification.

Megathylacus jandia* Woodland, 1934

[Syn. *Megathylacus brooksi* Rego & Pavanelli, 1985]

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Gibson 1989; Rego and Pavanelli 1985; Eiras et al. 1986; Kohn and Fernandes 1987; Pavanelli and Machado 1991; Takemoto and Pavanelli 1994; Kohn et al. 2011; de Chambrier et al. 2014).

Notes: host reported as *Paulicea luetkeni* or *Z. zungaro* (for details on the host taxonomic status, see Boni et al. 2011). Rego and Gibson (1989) and Rego and Pavanelli (1985) reported hyperparasitism caused by metacestodes of proteocephalids.

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1934c; Zehnder and Mariaux 1999; Hypša et al. 2005; de Chambrier et al. 2014, 2015a).

Notes: type host; it was originally reported as *Rhamdia* sp., but it is most likely *Z. zungaro* as discussed by de Chambrier et al. (2014), who reassessed the taxonomic status of this cestode species. Sequences of partial 18S (AY551111), complete ITS2 (AY551147), partial 28S (AJ388596) and 16S (AJ389515) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Megathylacus* sp.**

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego 1990).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult (also immature specimens); Amazon River basin; Peru (de Chambrier et al. 2014, 2015a).

Note: host reported as *P. punctifer* by de Chambrier et al. (2014), however, it falls within the range of genetic variability of *P. fasciatum* (*sensu lato*), according to Carvalho-Costa et al. (2011).

***Megathylacus travassosi* Pavanelli & Rego, 1992**

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1991; Pavanelli and Rego 1992; Machado et al. 1994, 1995, 1996; Rego 2002; de Chambrier et al. 2014; Ribeiro et al. 2014).

Notes: type host. Pavanelli and Machado (1991) referred to *M. travassosi* prior its formal publication one year later, which could render the name a *nomen nudum* but which was neglected in subsequent works.

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Campos et al. 2008, 2009a, b).

Sciadocephalus megalodiscus* Diesing, 1850

Cichla kelberi (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná River basin; Brazil (Yamada and Takemoto 2013).
Note: this host should be synonymized with *C. ocellaris* based on molecular data (Willis et al. 2012).

Cichla monoculus (Actinopterygii: Cichlidae); freshwater; intestine, stomach; adult; Amazon and Paraná River basins; Brazil, Peru (Diesing 1850; Woodland 1933b; Rego et al. 1999b; Machado et al. 2000; de Chambrier et al. 2006a, 2015b).
Notes: type host. *C. monoculus* should be synonymized with *C. ocellaris* based on molecular data (Willis et al. 2012), but it is accepted by morphology-based studies (Kullander and Ferreira 2006). Sequence of partial 28S (KP729403) (de Chambrier et al. 2015b).

Cichla piquiti (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná and Tocantins-Araguaia River basins; Brazil (Franceschini et al. 2013; Yamada and Takemoto 2013).

Subfamily Endorchiinae Woodland, 1934

***Endorchis auchenipteri* de Chambrier & Vaucher, 1999**

Auchenipterus osteomystax (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: type host.

Endorchis piraeeba* Woodland, 1934

Brachyplatystoma filamentosum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil; (Woodland 1934b; Rego 1991; de Chambrier and Vaucher 1997; Zehnder and Mariaux 1999; Zehnder et al. 2000; Hypša et al. 2005).
Notes: type host. In type material, *Nominoscolex piraeeba* and *E. piraeeba* are mixed on the same slide (de Chambrier and Vaucher 1997); Rego (1991) synomymized *E. piraeeba* with the former species, but de Chambrier and Vaucher (1997) re-validated it. Sequences of partial 18S (AY551107), complete ITS2 (AY551142), partial 28S (AJ388603) and 16S (AJ389522) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Brachyplatystoma cf. *filamentosum* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

***Endorchis* sp.**

Pimelodus altissimus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Pimelodus cf. *maculatus* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Trachelyopterus striatulus (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Subfamily Ephedrocephalinae Mola, 1929

Ephedrocephalus microcephalus* Diesing, 1850

[Syn. *Rudolphiella microcephalus* (Diesing, 1850) Brooks, 1995]

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Diesing 1850, 1855; Mola 1906; Woodland 1933b; Fuhrmann 1934; Rego 1984b; Zehnder and Mariaux 1999; Hypša et al. 2005; Ruedi and de Chambrier 2012; Scholz et al. 2013).

Notes: type host. Sequences of complete and partial 18S (KC786007, AY551108), respectively; complete ITS2 (AY551143), partial 28S (KC786017, AJ388605), partial 16S (KC785994, AJ389509) and partial *cox*1 (KC785982) (Zehnder and Mariaux 1999; Hypša et al. 2005; Scholz et al. 2013).

Subfamily Monticelliinae Mola, 1929

Ageneiella brevifilis* de Chambrier & Vaucher, 1999

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Argentina, Paraguay (de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Gil de Pertierra 2009).

Notes: type host; it was reported as *A. brevifilis*. Sequences of partial 18S (AY551102), complete ITS2 (AY551138), partial 28S (AJ388600) and 16S (AJ389495) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Ageneiosus militaris (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2009).

***Ageneiella* sp.**

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Chambriella agostinhoi* (Pavanelli & Machado, 1992) Rego, Chubb & Pavanelli, 1999

[Syn. *Goezeella agostinhoi* Pavanelli & Machado, 1992]

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Bachmann et al. 2007).
Notes: tapeworms reported as *Goezeella agostinhoi*. This report needs verification, but apparently, there are no vouchers deposited in any museum collection.

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1991, 1992; Takemoto and Pavanelli 1994; Ceccarelli et al. 2006; Kohn et al. 2011).
Notes: type host; it was reported as *Z. zungaro* or *P. luetkeni* and the tapeworms as *Goezeella agostinhoi* by some authors. Before formal description of the species, Pavanelli and Machado (1991) had used the name *G. agostinhoi*. Since *Robertiella agostinhoi* and *R. paranaensis* sensu Rego (1999) were not formally described, we considered them as *nomina nuda*.

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).

***Chambriella paranaensis* (Pavanelli & Rego, 1989) Rego, Chubb & Pavaneli, 1999**
[Syns. *Goezeella paranaensis* Pavanelli & Rego, 1989; *Spatulifer paranensis* (sic!) (Pavanelli & Rego, 1989) Brooks, 1995]

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (Pavanelli and Rego 1989; Pavanelli and Machado 1991; de Chambrier and Vaucher 1999; Guidelli et al. 2003; de Chambrier et al. 2006a, 2015a).

***Chambriella* sp.**

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).
Note: *Chambriella* sp 1. sensu de Chambrier et al. (2015a).

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (de Chambrier et al. 2006a, 2015a; Ruedi and de Chambrier 2012).
Note: *Chambriella* sp. 2 sensu de Chambrier et al. (2015a).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).
Note: *Chambriella* sp. 3 sensu de Chambrier et al. (2015a).

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (de Chambrier and Scholz 2008; de Chambrier et al. 2015a).
Note: *Chambriella* sp. 4 sensu de Chambrier et al. (2015a).

Choanoscolex abscisus* (Riggenbach, 1895) La Rue, 1911
[Syns. *Ichthyotaenia abscisa* Riggenbach, 1895; *Corallobothrium abscissus* (sic!) (Riggenbach, 1895) Meggitt, 1927; *Proteocephalus abscissus* (sic!) (Riggenbach, 1895) Fuhrmann, 1933; *Spatulifer abscissus* (sic!) (Riggenbach, 1895) Brooks, 1995]

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná and São Francisco River basins; Brazil, Paraguay (Riggenbach 1895, 1896b; La Rue 1911, 1914; Rego and Gibson 1989; Rego 1990, 2002; Pavanelli and Machado 1991; Machado et al. 1994, 1995, 1996; Mariaux 1998; de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Ceccarelli et al. 2006; Kohn et al. 2011; Ribeiro and Takemoto 2014; Ribeiro et al. 2014).

Notes: type host. Riggenbach (1895, 1896b) reported the host as *Silurus* sp. and proposed the parasite name twice. Rego and Gibson (1989) reported hyperparasitism caused by metacestodes of proteocephalids. Sequences of partial 18S (AY551105, Z98382, Z98381, Z98380), complete ITS2 (AY551141), partial 28S (AJ388630) and 16S (AJ389501) (Mariaux 1998; Zehnder and Mariaux 1999; Hypša et al. 2005).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon, Orinoco, Paraná and São Francisco River basins; Brazil, Peru, Venezuela (Brooks and Rasmussen 1984; Rego 1990, 2002; Zehnder et al. 2000; de Chambrier et al. 2004b, 2015a; Ceccarelli et al. 2006; Campos et al. 2008, 2009a, b; Jerônimo et al. 2013).

Notes: tapeworms reported as *Choanoscolex* cf. *abscisus* by Zehnder et al. (2000) and de Chambrier et al. (2004b). Host reported as *P. reticulatum* by Jerônimo et al. (2013), but it falls within the range of genetic variability of *P. fasciatum* (*sensu lato*), according to Carvalho-Costa et al. (2011). Sequence of partial 28S (AJ275064) (Zehnder et al. 2000).

Rhaphiodon vulpinus (Actinopterygii: Cynodontidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Pavanelli 1990).

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1991; Ceccarelli et al. 2006).

Notes: host reported as *P. luetkeni* or *Z. zungaro*; it was considered an accidental host by de Chambrier and Vaucher (1999).

***Choanoscolex* sp.**

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Paraguay, Peru (de Chambrier and Vaucher 1999; de Chambrier et al. 2006a).

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2006a).

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult (mostly immature specimens); Amazon River basin; Brazil, Peru (de Chambrier and Scholz 2008; de Chambrier et al. 2015a).

***Goezeella danbrooksi* de Chambrier, Rego & Mariaux, 2004**

[Syn. *Goezeella siluri* sensu Brooks & Deardorff, 1980]

Ageneiosus pardalis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Magdalena River Basin; Colombia (Brooks and Deardorff 1980; de Chambrier et al. 2004a).

Note: type host; it was originally reported as *A. caucanus*, whereas the tapeworms have been reported as *Goezeella siluri* following Brooks and Deardorff's (1980) description.

Goezeella siluri* Fuhrmann, 1916

[Syns. *Goezeella piramutab* Woodland, 1933; *Monticellia piramutab* (Woodland, 1933) Woodland, 1935; *M. siluri* (Fuhrmann, 1916) Woodland, 1935; *Corallobothrium siluri* (Fuhrmann, 1916) Harwood, 1933; *Spatulifer piramutab* (Woodland, 1933) Brooks & Deardorff, 1980; *S. siluri* (Fuhrmann, 1916) Brooks, 1995]

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature and mature specimens); Amazon and Orinoco River basins; Brazil, Venezuela (Woodland 1933c; Brooks and Rasmussen 1984; de Chambrier et al. 2004a).

Note: the specimens studied by Woodland (1933c) were described as *G. piramutab* and it corresponds in fact to a mixed infection of the present species and *Brooksiella praeputialis*, according to de Chambrier et al. (2004a).

Cetopsis coecutiens (Actinopterygii: Cetopsidae); freshwater; intestine; adult; Amazon River basin; Brazil (Fuhrmann 1916; Rego et al. 1974; Rego 1975; de Chambrier and Vaucher 1999; de Chambrier et al. 2004a).

Note: type host.

Cetopsis othonops (Actinopterygii: Cetopsidae); freshwater; intestine; adult; Orinoco River basin; Venezuela (Brooks and Rasmussen 1984).

Note: host reported as *Pseudocetopsis othonops*.

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder and Mariaux 1999; de Chambrier et al. 2004a; Hypša et al. 2005).

Note: sequences of partial 18S (AY551110), complete ITS2 (AY551146), partial 28S (AJ388612) and 16S (AJ389518) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Lenhataenia megacephala* (Woodland, 1934) de Chambrier & Scholz, 2008

[Syn. *Monticellia megacephala* Woodland, 1934]

Rhamdia quelen (Actinopterygii: Heptapteridae); freshwater; intestine, stomach; adult; Chascomus lagoon (Salado River basin); Argentina (Rabey 1973).

Note: host reported as *Rhamdia sapo*.

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1934c; Rego 1975; de Chambrier and Scholz 2008; de Chambrier et al. 2015a).

Notes: type host; it was originally reported as *Platystomatichthys sturio*.

Manaosia bracodemoca* Woodland, 1935

[Syns. *Paramonticellia itaipuensis* Pavanelli & Rego, 1991; *Goezeella nupeliensis* Pavanelli & Rego, 1991; *Spatulifer nupeliensis* (Pavanelli & Rego, 1991) Brooks, 1995]

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1991).

Sorubim lima (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (Woodland 1935b; Pavanelli and Machado 1991; Pavanelli and Rego 1991; de Chambrier and Vaucher 1999; Pavanelli and Takemoto 2000; Takemoto and Pavanelli 2000; Rego 2002; de Chambrier 2003; Kohn et al. 2011; de Chambrier et al. 2015a, b).
Notes: type host. Woodland (1935b) reported the host as *Platystoma* sp., but it is supposed to be *Sorubim lima*, locally known as 'braço-de-moça' (de Chambrier 2003). Sequence of partial 28S (KP729414) (de Chambrier et al. 2015b).

***Monticellia amazonica* de Chambrier & Vaucher, 1997**

[Syns. *Nomimoscolex piracatinga* Woodland, 1935; *Monticellia rugata* Rego, 1975 (*pro parte*); *Spatulifer rugata* (Rego, 1975) Brooks & Deardorff, 1980; *Paramonticellia piracatinga* (Woodland, 1935) Brooks, 1995]

Calophysus macropterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1935b; Rego 1975; de Chambrier and Vaucher 1997; de Chambrier et al. 2006a, 2015a; Scholz et al. 2008).
Note: type host; it was originally reported as *Pimelodus pati* (syn. of *Luciopimelodus pati* according to Froese and Pauly 2016), but this host does not occur in the Amazon River basin; 'piracatinga' is also the vernacular name of *C. macropterus*, which is endemic to the Amazon and Orinoco River basins (for details, see Scholz et al. 2008).

***Monticellia belavistensis* Pavanelli, Machado, Takemoto & dos Santos, 1994**

Pterodoras granulosus (Actinopterygii: Doradidae); freshwater; intestine; adult; Amazon and Paraná River basins; Argentina, Brazil, Paraguay, Peru (Pavanelli et al. 1994; de Chambrier and Vaucher 1999; Gil de Pertierra 2005; Kohn et al. 2011; de Chambrier et al. 2015a).
Note: type host.

Monticellia coryphicephala* (Monticelli, 1891) La Rue, 1911

[Syns. *Taenia coryphicephala* Monticelli, 1891; *Tetracotylus coryphicephala* Monticelli, 1891; *Ichthyotaenia coryphicephala* (Monticelli, 1891) Lönnberg, 1894; *Proteocephalus* (*Proteocephalus*) *coryphicephala* (Monticelli, 1891) Harwood, 1933]

Salminus brasiliensis (Actinopterygii: Bryconidae); freshwater; intestine; adult; Paraná and São Francisco River basins; Brazil, Paraguay (Monticelli 1891; La Rue 1911, 1914; Rego 1975; Rego and Pavanelli 1990; Pavanelli and Machado 1991; de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000; Brasil-Sato 2003; Mesquita et al. 2012; Karling et al. 2013a, b).

Notes: type host; it was originally reported as *Silurus* sp., but this genus only occurs in the Palaearctic region. Sequences of complete ITS2 (AJ238839), partial 28S (AJ238832) and 16S (AJ238831) (Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000).

Salminus franciscanus (Actinopterygii: Bryconidae); freshwater; intestine; adult; São Francisco River basin; Brazil (Rego and Pavanelli 1990).
Note: host reported as *S. brevidens*.

***Monticellia dlouhyi* de Chambrier & Vaucher, 1999**

Acestrorhynchus altus (Actinopterygii: Acestrorhynchidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: type host.

***Monticellia magna* (Rego, Santos & Silva, 1974) de Chambrier & Vaucher, 1997**

[Syns. *Nomimoscolex magna* Rego Santos & Silva, 1974 (*pro parte*); *Monticellia loyolai* Pavanelli & Machado, 1992]

Pimelodus albicans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2004).

Pimelodus argenteus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2004).

Pimelodus cf. *blochii* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná and São Francisco River basins; Brazil, Argentina (Rego et al. 1974; Pavanelli and Machado 1991, 1992; de Chambrier and Vaucher 1997, 1999; Brasil-Sato 2003; Gil de Pertierra 2004; Kohn et al. 2011).
Notes: type host; it was reported as *Pimelodus clarias*. There is a mixture of two species in the type material, originally described as *N. magna*, which can be differentiated by the position of internal organs (see p. 255 in de Chambrier and Vaucher (1997); Rego et al. (1999a) proposed the name *Proteocephalus magna* for those specimens considered as *Proteocephalus* sp. by the former authors, but they superficially circumscribed the new taxon.

Pimelodus cf. *maculatus* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: they reported the tapeworms as *Monticellia* cf. *magna*.

***Monticellia santafesina* Arredondo & Gil de Pertierra, 2010**

Megalonema platanum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Arredondo and Gil de Pertierra 2010).
Note: type host.

Megalonema platycephalum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Monticellia ventrei **de Chambrier & Vaucher, 1999**

[Syn. *Myzophorus admonticellia* Woodland 1934 (*pro parte*)]

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2005).

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (Woodland 1934a; de Chambrier and Vaucher 1999; de Chambrier et al. 2015a).

Notes: this host is assumed to be the type, because all possible fish hosts cited by Woodland (1934a), i.e. *Pimelodus pinarampu*, *Pinirampus pirinampus* (sic!) and *P. typus* are junior synonym of *P. pirinampu* (see Froese and Pauly 2016). *Nomimoscolex admonticellia* and *M. ventrei* are mixed in the type material of *Myzophorus admonticellia*, according to de Chambrier and Vaucher (1999).

Monticellia **sp.**

Brycon orbignyanus (Actinopterygii: Bryconidae); freshwater; intestine; adult (immature specimens); Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; São Francisco River basin; Brazil (Brasil-Sato 2003; Santos et al. 2003).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999; Santos et al. 2003).

Regoella brevis **Arredondo, Gil de Pertierra & de Chambrier, 2013***

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Arredondo et al. 2013; de Chambrier et al. 2015b).

Notes: type host. Sequence of partial 28S (KP729389) (de Chambrier et al. 2015b). Host recorded as *Pseudoplatystoma reticulatum* in the GenBank database.

Spasskyellina lenha **(Woodland, 1933) Freze, 1965***

[Syn. *Monticellia lenha* Woodland, 1933]

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1933a; de Chambrier and Scholz 2008; de Chambrier et al. 2015a, b).

Notes: type host. Sequence of 28S (KP729413) under the name *Lenhataenia megacephala* in the GenBank database – see de Chambrier et al. (2015b).

Spasskyellina mandi **Pavanelli & Takemoto, 1996**

[Syn. *Monticellia mandi* (Pavanelli & Takemoto, 1996) de Chambrier & Vaucher, 1999]

Pimelodus ornatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Takemoto 1996).

Note: type host.

***Spasskyellina spinulifera* (Woodland, 1935) Freze, 1965**

[Syns. *Monticellia spinulifera* (Woodland, 1935); *M. spinulifer* (sic!) of Brooks (1995)]

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil, Paraguay (Rego 1990, 2002; Pavanelli and Machado 1991; Machado et al. 1994, 1995, 1996; de Chambrier and Vaucher 1999; Santos et al. 2003; Ceccarrelli et al. 2006; Kohn et al. 2011; Ribeiro et al. 2014; de Chambrier et al. 2015b).

Note: sequence of 28S (KP729417) (de Chambrier et al. 2015b).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Woodland 1935a; Rego 1975, 2002; de Chambrier and Vaucher 1999; Santos et al. 2003; de Chambrier et al. 2006a, 2015a; Ceccarrelli et al. 2006).

Note: type host.

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).

Sorubim lima (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Takemoto 2000; Takemoto and Pavanelli 2000).

***Spasskyellina* sp.**

Pimelodus ornatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Takemoto et al. 2009).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Santos et al. 2003).

***Spatulifer maringaensis* Pavanelli & Rego, 1989**

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; intestine, stomach; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (Pavanelli and Rego 1989; Pavanelli and Machado 1991; de Chambrier and Vaucher 1999; Guidelli et al. 2003; de Chambrier et al. 2006a, 2015a).

Note: type host.

Sorubim lima (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Argentina, Brazil, Paraguay, Peru (Pavanelli and Rego 1989; Pavanelli and Machado 1991; Mariaux 1998; de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Pavanelli and Takemoto 2000; Takemoto and Pavanelli 2000; Hypša et al. 2005; Arrendondo and Gil de Pertierra 2008; de Chambrier et al. 2015a).

Notes: Arredondo and Gil de Pertierra (2008) suggested, based on ecological data, that *Sorubim lima* is the principal final host. Tapeworms reported as *Spatulifer* cf. *maringaensis* by de Chambrier and Vaucher (1999) and confirmed to be *S. maringaensis* by the former authors, who evaluated the vouchers deposited in MHNG. Sequences of partial 18S (AY551136, Z98385, Z98384, Z98383), complete ITS2 (AY551176), partial 28S (AJ388634) and 16S (AJ389507) (Mariaux 1998; Zehnder and Mariaux 1999; Hypša et al. 2005).

***Spatulifer rugosa* (Woodland, 1935) Brooks & Deardorff, 1980**

[Syn. *Monticellia rugosa* Woodland, 1935]

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Argentina, Brazil, Peru (Woodland 1935b; Rego 1975, 1989, 2002; de Chambrier et al. 2006a, 2015a; Arrendondo and Gil de Pertierra 2008; Campos et al. 2008, 2009a, b; Lopes et al. 2009).
Notes: type host; it was reported as *P. punctifer* by de Chambrier et al. (2006a) and Lopes et al. (2009).

Spatulifer surubim* Woodland, 1934

[Syns. *Peltidocotyle rugosa* sensu Woodland, 1933b *nec* Diesing, 1850; *Spatulifer surubim* Woodland, 1934; *Monticellia surubim* (Woodland, 1934) Woodland, 1935]

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1933b, 1934a; Rego 1975, 2002).
Note: type host.

***Spatulifer* sp.**

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).
Note: probably *Spatulifer surubim* according to de Chambrier et al. (2006a).

Monticelliinae gen. sp.

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Subfamily Nupeliinae Pavanelli & Rego, 1991

Nupelia portoriquensis* Pavanelli & Rego, 1991

Sorubim lima (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil, Paraguay (Pavanelli and Machado 1991; Pavanelli and Rego 1991; de Chambrier and Vaucher 1999; Pavanelli and Takemoto 2000; Takemoto and Pavanelli 2000; de Chambrier et al. 2015b).
Notes: type host. Sequence of partial 28S (KP729401) (de Chambrier et al. 2015b).

***Nupelia tomasi* de Chambrier & Vaucher, 1999**

Trachelyopterus galeatus (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: type host.

Trachelyopterus cf. *striatulus* (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

***Nupelia* sp.**

Goeldiella eques (Actinopterygii: Heptapteridae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).

Subfamily Peltidocotylinae Woodland, 1934

Amazotaenia yvettae* de Chambrier, 2001

Brachyplatystoma capapretum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (de Chambrier 2001).
Notes: type host; it was reported as *B. filamentosum* and re-identified by J. Lundberg (pers. comm.).

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (de Chambrier 2001).

Jauella glandicephalus* Rego & Pavanelli, 1985

[Syn. *Spatulifer glandicephala* (Rego & Pavanelli, 1985) Brooks, 1995]

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil, Paraguay (Rego and Pavanelli 1985; Eiras et al. 1986; Rego and Gibson 1989; Pavanelli and Machado 1991; Takemoto and Pavanelli 1994; de Chambrier and Vaucher 1999; Gil de Pertierra 2009; de Chambrier et al. 2015b).
Notes: type host; it was reported as *P. luetkeni* or *Z. zungaro*. Rego and Pavanelli (1985) proposed the subfamily Jauellinae Rego and Pavanelli, 1985, which was not followed by other workers. Rego and Gibson (1989) and Rego and Pavanelli (1985) reported hyperparasitism caused by metacestodes of proteocephalids. Sequence of partial 28S (KP729399) (de Chambrier et al. 2015b).

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Luciaella ivanovae* Gil de Pertierra, 2009

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2009).
Note: type host.

Mariauxiella pimelodi* de Chambrier & Rego, 1995

Pimelodus ornatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil, Paraguay (de Chambrier and Rego 1995; de Chambrier and Vaucher 1999).

Pimelodus sp. (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (de Chambrier and Rego 1995; de Chambrier and Vaucher 1999).
Note: type host.

***Mariauxiella piscatorum* de Chambrier & Vaucher, 1999**

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (de Chambrier and Vaucher 1999; Guidelli et al. 2003; de Chambrier et al. 2006a, 2015a).
Note: type host.

***Peltidocotyle lenha* (Woodland, 1933) Woodland, 1934**

[Syns. *Othinoscolex lenha* Woodland, 1933; *Othinoscolex myzofer* Woodland, 1933; *Woodlandiella myzofera* (Woodland, 1933) Freze, 1965; *Peltidocotyle rugosa* of Schmidt, 1986; *Rudolphiella lenha* (Woodland, 1933) Brooks, 1995]

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1933a, 1934b; Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000; Hypša et al. 2005; de Chambrier and Scholz 2008; de Chambrier et al. 2015a).
Notes: type host; it was originally reported as *Platystomatichthys sturio*. Sequences of partial 18S (AY551122), complete IT2 (AJ238842), partial 28S (AJ238836) and 16S (AJ238827) (Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000; Hypša et al. 2005).

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil (Rego and Pavanelli 1987; Zehnder and de Chambrier 2000; Gil de Pertierra 2009).
Notes: host reported as *Zungaro zungaro* or *Paulicea luetkeni*. Rego and Pavanelli (1987) mistakenly reported the tapeworm as *Peltidocotyle rugosa*, according to Zehnder and de Chambrier (2000); the former workers also reported hyperparasitism caused by larval cestodes.

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Zehnder and Mariaux 1999; de Chambrier et al. 2006a).
Notes: sequences of complete IT2 (AJ238840, AJ238843), partial 28S (AJ238834, AJ238837) and 16S (AJ238826, AJ238829) (Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000).

Peltidocotyle rugosa* Diesing, 1850

Pseudopimelodus mangurus (Actinopterygii: Pseudopimelodidae); freshwater; intestine; adult; locality not given; Argentina (Rego and Pavanelli 1987).
Note: host reported as *Zungaro mangurus*.

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil (Diesing 1850; Fuhrmann 1934; Rego 1990, 2002; de Chambrier and Vaucher 1999; Zehnder and de Chambrier 2000; Gil de Pertierra 2009).
Note: type host; it was originally reported as *Platystoma tigrinum* (syn. of *Pseudoplatystoma tigrinum*), but it does not occur in the Paraná River basin, thus the fish host is assumed to be *P. corruscans* (see de Chambrier and Vaucher 1999).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Paraguay, Peru (Rego 1989, 1990; de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000; Olson et al. 2001; Rego 2002; Ceccarrelli et al. 2006; Campos et al. 2008, 2009a, b; de Chambrier et al. 2015a).
Note: sequence of complete 18S (AF286989) and ITS2 (AJ238841), partial 28S (AJ238835, AF286937) and 16S (AJ238828) (Zehnder and Mariaux 1999; Zehnder and de Chambrier 2000; Olson et al. 2001).

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Gibson 1989; Pavanelli and Machado 1991; Takemoto and Pavanelli 1994; Ceccarrelli et al. 2006; Kohn et al. 2011).
Notes: host reported as *P. luetkeni* or *Z. zungaro*. Rego and Gibson (1989) reported hyperparasitism caused by metacestodes of proteocephalids.

***Peltidocotyle* sp.**

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: host reported as *Paulicea luetkeni*.

Subfamily Proteocephalinae La Rue, 1911

Brayela karuatayi* (Woodland, 1934) Rego, 1984
[Syn. *Anthobothrium karuatayi* Woodland, 1934]

Platynematichthys notatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult (also immature specimens); Amazon River basin; Brazil, Peru (Woodland 1934c; Rego 1984a; de Chambrier et al. 2014, 2015a, b).
Notes: type host; it was originally reported as *Glanidium* sp.; Rego (1984a) erected the new subfamily Brayelainae, which was not accepted by other authors. Sequence of partial 28S (KP729406) (de Chambrier et al. 2015b).

Cangatiella arandasi* Pavanelli & Machado, 1991

Trachelyopterus galeatus (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1990, 1991; Kohn et al. 2011; de Chambrier et al. 2015b).
Notes: host also reported as *Parauchenipterus galeatus*. Sequence of 28S (KP729411) (de Chambrier et al. 2015b).
Note: type host.

***Cangatiella macdonaghi* (Szidat & Nani, 1951) Gil de Pertierra & Viozzi, 1999**
[Syns. *Ichthyotaenia macdonaghi* Szidat & Nani, 1951; *Proteocephalus macdonaghi* (Szidat & Nani, 1951) Yamaguti, 1959]

Odontesthes bonariensis (Actinopterygii: Atherinopsidae); freshwater; intestine; adult; lakes in Buenos Aires and Córdoba Provinces; Argentina (MacDonagh 1932; Ringuelet 1943; Fuster de Plaza and Boschi 1957; Ortubay et al. 1994; Mancini and Grosman 1998; Mancini et al. 2008; Drago 2012; Bethular et al. 2014).
Notes: Mancini and Grosman (1998) reported the tapeworms as *Proteocephalus* sp. (Mancini et al. 2008), whereas MacDonagh (1932) and Ringuelet (1943) reported it as *Ichthyotaenia* sp.

Odontesthes hatcheri (Actinopterygii: Atherinopsidae); freshwater; intestine; adult; Peligrini lake; Argentina (Szidat and Nani 1951; Gil de Pertierra and Viozzi 1999).
Notes: type host; it was originally reported as *Basilichthys microlepidotus*.

Euzetiella tetraphylliformis* de Chambrier, Rego & Vaucher, 1999

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature); Amazon River basin; Peru (de Chambrier et al. 2015a).

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil, Paraguay (de Chambrier et al. 1999).

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (de Chambrier et al. 1999, 2006a, 2015a).
Notes: host originally reported as *Paulicea luetkeni* (syn. of *Z. jahu* and *Z. zungaro*); since the holotype was described from a fish collected in the Amazon River, *Z. zungaro* should be considered the type host.

Frezella vaucheri* Alves, de Chambrier, Scholz & Luque, 2015

Tocantinsia piresi (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Amazon River basin; Brazil (Alves et al. 2015).
Notes: type host. Sequence of partial 28S (KM387399) (Alves et al. 2015).

Margaritaella gracilis* Arredondo & Gil de Pertierra, 2012

Callichthys callichthys (Actinopterygii: Callichthyidae); freshwater; intestine; adult; Paraná River basin; Argentina (Arredondo and Gil de Pertierra 2012).
Note: type host.

***Proteocephalus bagri* Holcman-Spector & Mañé-Garzón, 1988**

Rhamdia quelen (Actinopterygii: Heptapteridae); freshwater; intestine; Chis-Chis, Chascomús, Sauce, Diario and Dos Patos lagoons; Argentina, Brazil, Uruguay (Holcman-Spector and Mañé-Garzón 1988; Gil de Pertierra 2002b).
Note: type host; it was originally reported as *R. sapo*.

***Proteocephalus fossatus* (Riggenbach, 1895) La Rue, 1911**

[Syn. *Ichthyotaenia fossata* Riggenbach, 1895]

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (Riggenbach 1895, 1896; La Rue 1911, 1914).
Note: type host; it was originally reported as *Pimelodus pati*.

***Proteocephalus gibsoni* Rego & Pavanelli, 1991**

[Syn. *Proteocephalus ocellatus* sensu Rego & Pavanelli, 1990 *nec Proteocephalus ocellatus* (Rudolphi, 1802)]

Astronotus ocellatus (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon River basin; Peru (Rego and Pavanelli 1990; de Chambrier et al. 2006a, 2015a; Bittencourt et al. 2014).

Notes: type host. Rego and Pavanelli (1991) proposed the name *Proteocephalus gibsoni* one year later of its original description in order to avoid the homonym with *P. ocellatus* (syn. of *P. percae* [Müller, 1780]), a parasite of percids in Europe (Scholz 1989).

Astronotus sp. (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon River basin; Brazil (Rego and Pavanelli 1990).

Geophagus brasiliensis (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná and Doce River basins; Brazil (Rego and Pavanelli 1990; Bellay et al. 2012).

***Proteocephalus hemioliopteri* de Chambrier & Vaucher, 1997**

[Syns. *Myzophorus woodlandi* Rego, 1984; *Nomimoscolex woodlandi* (Rego, 1984) Rego & Pavanelli, 1992]

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Rego 1984b; de Chambrier and Vaucher 1997; Zehnder and Mariaux 1999; de Chambrier et al. 2005, 2015a; Hypša et al. 2005; Ruedi and de Chambrier 2012).

Notes: type host. Sequences of partial 18S (AY551129), complete ITS2 (AY551165) and partial 28S (AJ388622) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Proteocephalus hobergi* de Chambrier & Vaucher, 1999**

Oxydoras kneri (Actinopterygii: Doradidae); freshwater; intestine; adult; Amazon and Paraná River basins; Paraguay, Peru (de Chambrier and Vaucher 1999; de Chambrier et al. 2004b; de Chambrier et al. 2015a).

Notes: type host. Sequence of partial 28S (AJ275062) (de Chambrier et al. 2004b).

Oxydoras niger (Actinopterygii: Doradidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015).

***Proteocephalus kuyukuyu* Woodland, 1935**

Megalodoras uranoscopus (Actinopterygii: Doradidae); freshwater; intestine; adult (immature specimens); Amazon and Orinoco River basins; Peru, Venezuela (Brooks and Rasmussen 1984; de Chambrier et al. 2015a).

Note: Brooks and Rasmussen (1984) reported the host as *M. irwini* and the tapeworms as *P.* cf. *kuyukuyu*.

Oxydoras niger (Actinopterygii: Doradidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Brazil (Woodland 1935c; Santos and Tavares-Dias 2010; Silva et al. 2011).

Notes: type host; it was reported as *Pseudodoras niger*, but Woodland (1935c) also found some specimens in other doradid fish, assumed to be *Pseudodoras brunnescens* (syn. of *Acanthodoras spinosissimus*). It is argued that the presence of only immature proglottids in the adults is due to a hyperapolytic development (see de Chambrier et al. 2015a); considered *species inquirenda* by some authors, e.g. Freze (1965), Rego (1987b) and Rego et al. (1999a).

Pterodoras granulosus (Actinopterygii: Doradidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2015a, b). Note: sequence of partial 28S (KP729388) (de Chambrier et al. 2015b).

Pterodoras sp. (Actinopterygii: Doradidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2015a).

***Proteocephalus macrophallus* (Diesing, 1850) La Rue, 1914**

[Syns. *Taenia macrophalla* Diesing, 1850; *Ichthyotaenia macrophalla* (Diesing, 1850) Riggenbach, 1896]

Cichla kelberi (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná and São Francisco River basin; Brazil (Yamada and Takemoto 2013; Santos-Clapp and Brasil-Sato 2014).

Cichla monoculus (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Diesing 1850; La Rue 1914; Takemoto and Pavanelli 1996; de Chambrier et al. 2006a, 2015a, b; Machado et al. 2000; Kohn et al. 2011).

Notes: type host; some authors assumed *C. ocellaris* as the type host, but the taxonomic status of these cichlids is unclear (see notes on p. 33). Sequence of partial 28S (KP729394) (de Chambrier et al. 2015b).

Cichla ocellaris (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon, Orinoco, Paraíba do Sul and Paraná River basins; Brazil, Venezuela (Woodland 1933b; Scholz et al. 1996; Azevedo et al. 2010, 2011).

Cichla piquiti (Actinopterygii: Cichlidae); freshwater; intestine, stomach; adult; Paraná and Tocantins-Araguaia River basins; Brazil (Martins et al. 2009, 2011; Franceschini et al. 2013; Lacerda et al. 2013; Yamada and Takemoto 2013).

Cichla sp. (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná River basin; Brazil (Santos et al. 2011).

***Proteocephalus mahnerti* de Chambrier & Vaucher, 1999**

Hoplerythrinus unitaeniatus (Actinopterygii: Erythrinidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999). Note: type host.

***Proteocephalus microscopicus* Woodland, 1935**

Cichla kelberi (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná and São Francisco River basins; Brazil (Yamada and Takemoto 2013; Santos-Clapp and Brasil-Sato 2014).

Cichla monoculus (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Takemoto and Pavanelli 1996; Machado et al. 2000; de Chambrier et al. 2006a, 2015a; Müller et al. 2008; Kohn et al. 2011).

Cichla ocellaris (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1935c).
Note: type host.

Cichla piquiti (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná and Tocantins-Araguaia River basins; Brazil (Martins et al. 2009, 2011; Franceschini et al. 2013; Lacerda et al. 2013; Yamada and Takemoto 2013).

Cichla sp. (Actinopterygii: Cichlidae); freshwater; intestine; adult; Paraná River basin; Brazil (Santos et al. 2011).

***Proteocephalus pilarensis* de Chambrier & Vaucher, 1999**

Paraloricaria sp. (Actinopterygii: Loricariidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: type host.

***Proteocephalus pimelodi* (Gil de Pertierra, 1995) de Chambrier & Vaucher, 1997**

[Syn. *Nomimoscolex pimelodi* Gil de Pertierra, 1995]

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 1995; de Chambrier and Vaucher 1997).
Note: type host.

***Proteocephalus platystomi* Lynsdale, 1959**

Pseudoplatystoma sp. (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Lynsdale 1959; Rego 2002).
Note: host originally reported as *Platystoma* sp.; the specimens collected by Woodland in 1937 were deposited without any identification in BMNH (Lynsdale 1959).
Note: type host.

***Proteocephalus regoi* de Chambrier, Scholz and Vaucher, 1996**

Hoplias malabaricus (Actinopterygii: Erythrinidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier et al. 1996; de Chambrier and Vaucher 1999).
Note: type host.

***Proteocephalus renaudi* de Chambrier & Vaucher, 1994**

Franciscodoras marmoratus (Actinopterygii: Doradidae); freshwater; intestine; adult; São Francisco River basin; Brazil (Santos and Brasil-Sato 2006).

Platydoras costatus (Actinopterygii: Doradidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1994, 1999; Zehnder and Mariaux 1999).

Notes: type host; it does not occur in the Paraná River basin (Ferraris 2007; Piorski et al. 2008). Therefore, *P. armatulus* (Valenciennes), which is the only known *Platydoras* species from this river basin, is most probably the true host of *Proteocephalus renaudi*. Sequences of partial 28S (AJ388638) and 16S (AJ389503) (Zehnder and Mariaux 1999).

***Proteocephalus rhamdiae* Holcman-Spector & Mañé-Garzón, 1988**

Rhamdia quelen (Actinopterygii: Heptapteridae); freshwater; intestine; adult; Chis-Chis, Chascomús, Sauce, Diario and Dos Patos lagoons and Paraná River basin; Argentina, Brazil, Paraguay, Uruguay (Holcman-Spector and Mañé-Garzón 1988; de Chambrier and Vaucher 1999; Gil de Pertierra 2002b).

Note: type host; it was originally reported as *R. sapo*.

***Proteocephalus serrasalmus* Rego & Pavanelli, 1990**

Pygocentrus nattereri (Actinopterygii: Serrasalmidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Pavanelli 1990).

Note: host originally reported as *Serrasalmus nattereri*.

Serrasalmus maculatus (Actinopterygii: Serrasalmidae); freshwater; intestine; adult; Paraná River basin; Brazil, Paraguay (Rego and Pavanelli 1990; Pavanelli and Machado 1991; de Chambrier and Vaucher 1999).

Note: type host; it was originally reported as *S. spilopleura*.

***Proteocephalus soniae* de Chambrier & Vaucher, 1994**

Platydoras costatus (Actinopterygii: Doradidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1994, 1999).

Note: type host; it was most likely mistaken (see notes for *P. renaudi* on p. 50 for more details).

***Proteocephalus sophiae* de Chambrier & Rego, 1994**

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (de Chambrier and Rego 1994; de Chambrier et al. 2015a).

Note: type host; it was originally reported as *P. luetkeni*.

***Proteocephalus vazzolerae* Pavanelli & Takemoto, 1995**

Leporinus friderici (Actinopterygii: Anostomidae); freshwater; caeca, intestine; adult; Paraná River basin; Brazil (Guidelli et al. 2006, 2011).

Leporinus lacustris (Actinopterygii: Anostomidae); freshwater; caeca, intestine; adult; Paraná River basin; Brazil (Guidelli et al. 2006, 2011).

Piaractus mesopotamicus (Actinopterygii: Serrasalmidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Takemoto 1995).
Note: type host.

***Proteocephalus vladimirae* de Chambrier & Vaucher, 1999**

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: type host.

***Proteocephalus* sp.**

Franciscodoras marmoratus (Actinopterygii: Doradidae); freshwater; intestine; adult; São Francisco River basin; Brazil (Santos and Brasil-Sato 2004).

Gymnotus carapo (Actinopterygii: Gymnotidae); freshwater; intestine; adult; Paraiba do Sul River basin; Brasil (Azevedo et al. 2010, 2011).

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).
Note: they named this morphotype as *Proteocephalus* sp. 1 and it is probably a new species (de Chambrier et al. 2006a).

Pimelodus blochii (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2015a).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego et al. 1974; de Chambrier and Vaucher 1997).
Note: Rego et al. (1999a) named part of the material described by Rego et al. (1974) as *Proteocephalus magna* (Rego, Santos and Silva, 1974), but a formal description was not provided.

Platydoras costatus (Actinopterygii: Doradidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1994, 1999).
Note: the host species was most likely misidentified (see notes on p. 50 for more details).

Pterodoras granulosus (Actinopterygii: Doradidae); freshwater; intestine; adult (immature specimens); Amazon River basin; Peru (de Chambrier et al. 2006a, 2015a).
Note: they named this morphotype as *Proteocephalus* sp. 2 and it is probably a new species (de Chambrier et al. 2006a).

***Pseudocrepidobothrium chanaorum* Arredondo, Gil de Pertierra & de Chambrier, 2014**

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Arredondo et al. 2014).
Note: type host; it was reported as *P. reticulatum* (see the note on p. 36).

Pseudocrepidobothrium eirasi **(Rego & de Chambrier, 1995) Rego & Ivanov, 2001***

[Syn. *Crepidobothrium eirasi* Rego & de Chambrier, 1995]

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Rego and de Chambrier 1995; Zehnder and Mariaux 1999; Rego and Ivanov 2001; Hypša et al. 2005; Ruedi and de Chambrier 2012).

Notes: type host. Sequences of partial 18S (AY551106), complete ITS2 (AY551179), partial 28S (AJ388623) and 16S (AJ389494) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Pseudocrepidobothrium ludovici **Ruedi & de Chambrier, 2012**

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Ruedi and de Chambrier 2012).

Note: type host.

Pseudocrepidobothrium **sp.**

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder and de Chambrier 2000; de Chambrier et al. 2004b).

Notes: tapeworms reported as *Crepidobothrium* sp. Sequence of complete ITS2 (AJ238838), partial 28S (AJ238833, AJ275063) and 16S (AJ238830) (Zehnder and de Chambrier 2000; de Chambrier et al. 2004b).

Scholzia emarginata **(Diesing, 1850) de Chambrier, Rego & Gil de Pertierra, 2005***

[Syns. *Tetrabothrium emarginatum* Diesing, 1850; *Tetrabothrium* (*Eutetrabothrium*) *emarginatum* Diesing, 1856; *Nomimoscolex emarginatum* (Diesing, 1850) Rego, Chubb & Pavaneli, 1999; *Myzophorus pirarara* Woodland, 1935; *Nomimoscolex pirarara* (Woodland, 1935) Rego & Pavanelli, 1992; *Proteocephalus pirarara* (Woodland, 1935) de Chambrier & Vaucher, 1997].

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Diesing 1850; Woodland 1935a; Rego 1984b; de Chambrier and Vaucher 1997; Zehnder et al. 2000; de Chambrier et al. 2005, 2006a, 2015a; Ruedi and de Chambrier 2012; Scholz et al. 2013).

Notes: type host. Sequences of partial 18S (AY551131, AY551112, KC786006), complete ITS2 (AY551170, AY551148), partial 28S (AJ388616, KC786016), partial 16S (AJ389513, KC785993) and partial *cox*1 (KC785981) (Zehnder and Mariaux 1999; Hypša et al. 2005; Scholz et al. 2013). Hypša et al. (2005) deposited two sequences because they considered *Myzophorus pirarara* and *Proteocephalus pirarara* as different species.

Subfamily Rudolphiellinae Woodland, 1935

Rudolphiella lobosa* (Riggenbach, 1895) Fuhrmann, 1916
[Syns. *Corallobothrium lobosum* Riggenbach, 1895; *Ephedrocephalus lobosum* (Riggenbach, 1895) Mola, 1906]

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (Riggenbach 1895, 1896; Fuhrmann 1916; Rego and Gibson 1989; de Chambrier and Vaucher 1999; Gil de Pertierra and de Chambrier 2000).

Notes: type host; it was originally reported as *Pimelodus pati*, but Gil de Pertierra and de Chambrier (2000) suspected that *Megalonema platanum* is the true host, since they share the same vernacular name 'patí' and similar tapeworms were found in the latter fish host (de Chambrier and Vaucher 1999). Rego and Gibson (1989) reported hyperparasitism caused by metacestodes of proteocephalids.

Megalonema platanum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999; Hypša et al. 2005).

Note: de Chambrier and Vaucher (1999) reported the tapeworms as *Rudolphiella* cf. *lobosa*. Sequences of partial 18S (AY551134) and complete ITS2 (AY551173) (Hypša et al. 2005).

***Rudolphiella myoides* (Woodland, 1934) Woodland, 1935**
[Syn. *Amphilaphorchis myoides* Woodland, 1934]

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1934a, 1935b; Gil de Pertierra and de Chambrier 2000).

Note: type host.

***Rudolphiella piracatinga* (Woodland, 1935) Gil de Pertierra & de Chambrier, 2000**
[Syns. *Monticellia piracatinga* Woodland, 1935; *M. rugata* Rego, 1975 (*pro parte*); *Rudolphiella rugata* (Rego 1975) Rego, Chubb & Pavanelli, 1999; *Spatulifer piracatinga* (Woodland, 1935) Brooks & Deardorff, 1980]

Calophysus macropterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1935b; Rego 1975; Mariaux 1998; Zehnder and Mariaux 1999; Gil de Pertierra and de Chambrier 2000; de Chambrier et al. 2006a, 2015a).

Notes: host originally described as *Pimelodus pati* (for details, see Gil de Pertierra and de Chambrier 2000). Rego (1975) described *M. rugata*, based on a mixture of two different species, *Nomimoscolex piracatinga* (syn. of *M. amazonica*) and *M. piragatinga* (syn. of *R. piracatinga*) (Gil de Pertierra and de Chambrier 2000). Sequences of partial 18S (Z98391, Z98390, Z98389), 28S (AJ388627) and 16S (AJ389504) (Mariaux 1998; Zehnder and Mariaux 1999).

Note: type host.

***Rudolphiella piranabu* (Woodland, 1934) Woodland, 1935**

[Syn. *Amphilaphorchis piranabu* Woodland, 1934]

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil (Woodland 1934a, 1935b; Pavanelli and Machado 1991; Gil de Pertierra and de Chambrier 2000; Kohn et al. 2011). Note: type host.

***Rudolphiella szidati* Gil de Pertierra & de Chambrier, 2000**

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Zehnder and Mariaux 1999; Gil de Pertierra and de Chambrier 2000; Hypša et al. 2005).

Notes: type host. Sequences of partial and complete 18S (AY551135, AF286990), respectively, complete ITS2 (AY551174), partial 28S (AJ388617, AF286938) and 16S (AJ389517) (Zehnder and Mariaux 1999; Olson et al. 2001; Hypša et al. 2005).

***Rudolphiella* sp.**

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult (including immature specimens); Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Amazon and Paraná River basins; Paraguay (de Chambrier and Vaucher 1999; de Chambrier et al. 2015a).

Subfamily Zygobothriinae Woodland, 1933

***Amphoteromorphus ninoi* Carfora, de Chambrier & Vaucher, 2003**

Brachyplatystoma filamentosum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Carfora et al. 2003; Chambrier et al. 2004b). Note: sequence of 28S (AJ388624) (Chambrier et al. 2004b).

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder and Mariaux 1999; Carfora et al. 2003; Chambrier et al. 2004b).

Notes: type host. Tapeworms reported as *Amphoteromorphus piraeeba* by Zehnder and Mariaux (1999). Sequence of partial 28S (AJ275066) (Zehnder and Mariaux 1999).

***Amphoteromorphus ovalis* Carfora, de Chambrier & Vaucher, 2003**

Brachyplatystoma cf. *filamentosum* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Brachyplatystoma sp. (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Carfora et al. 2003).

Note: type host.

***Amphoteromorphus parkamoo* Woodland, 1935**

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1935b; Zehnder and Mariaux 1999; Zehnder et al. 2000; Carfora et al. 2003; Hypša et al. 2005; de Chambrier et al. 2006a, 2015a).

Notes: type host; it was originally reported as *Pseudopimelodus zungaro*, but also as *Paulicea luetkeni* in additional studies. Sequences of partial 18S (AY551103), complete ITS2 (AY551139), partial 28S (AJ388595) and 16S (AJ389523) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Amphoteromorphus peniculus* Diesing, 1850

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine: adult; Amazon River basin; Brazil, Peru (Diesing 1850; Woodland 1933b; Fuhrmann 1934; Carfora et al. 2003; de Chambrier et al. 2015a, b).

Notes: type host; it was originally reported as *Bagrus goliath*, but also as *Brachyplatystoma flavicans* in additional studies. Sequence of partial 28S (KP729410) (de Chambrier et al. 2015b).

***Amphoteromorphus piraeeba* Woodland, 1934**

Brachyplatystoma filamentosum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1933a, 1934b; de Chambrier and Vaucher 1997; Zehnder and Mariaux 1999; Carfora et al. 2003; Hypša et al. 2005; de Chambrier et al. 2015b).

Notes: type host; tapeworm reported as *Amphoteromorphus peniculus* by Woodland (1933b). Sequences of partial 18S (AY551104), complete ITS2 (AY551140), partial 28S (KP729407) and 16S (AJ389510) (Zehnder and Mariaux, 1999; Hypša et al. 2005; de Chambrier et al. 2015b).

***Amphoteromorphus piriformis* Carfora, de Chambrier & Vaucher, 2003**

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine; adult (also immature specimens); Amazon River basin; Brazil, Peru (Carfora et al. 2003; de Chambrier et al. 2004b, 2006a, 2015a).

Notes: type host; it was originally reported as *Brachyplatystoma flavicans*; de Chambrier et al. (2006a) reported the tapeworms as *Amphoteromorphus* cf. *piriformis*. Sequence of partial 28S (AJ275231) (de Chambrier et al. 2004b).

Brooksiella praeputialis* (Rego, Santos & Silva, 1974) Rego, Chubb & Pavanelli, 1999

[Syn. *Amphoteromorphus praeputialis* Rego, dos Santos & Silva, 1974]

Cetopsis coecutiens (Actinopterygii: Cetopsidae); freshwater; intestine; adult; Amazon River basin; Brazil (Rego et al. 1974; de Chambrier et al. 2004a, b).

Notes: type host; de Chambrier et al. (2004a) redescribed this species. Sequence of partial 28S (AJ275229) (de Chambrier et al. 2004b).

Cetopsis othonops (Actinopterygii: Cetopsidae); freshwater; intestine; adult; Orinoco River basin; Venezuela (Brooks and Rasmussen 1984).
Note: host reported as *Pseudocetopsis othonops*.

Gibsoniela mandube* (Woodland, 1935) Rego, 1984
[Syns. *Anthobothrium mandube* Woodland, 1935; *Endorchis* (*Pseudendorchis*) *mandube* (Woodland, 1935) Yamaguti, 1959; *Nomimoscolex mandube* (Woodland, 1935) Brooks, 1995]

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1935b; de Chambrier and Vaucher 1999; de Chambrier et al. 2015a).
Notes: type host; it was reported as *A. brevifilis* or *Pseudoageneiosus brevifilis*.

Ageneiosus sp. (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a, b).
Note: sequence of partial 28S (KP729412) (de Chambrier et al. 2015b).

***Gibsoniela meursaulti* de Chambrier & Vaucher, 1999**
[Syn. *Endorchis mandube* Woodland, 1935]

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Amazon and Paraná River basins; Argentina, Brazil, Paraguay (Woodland 1935b; Rego 1992; de Chambrier and Vaucher 1999; Zehnder and Mariaux 1999; Zehnder et al. 2000; Hypša et al. 2005; Gil de Pertierra 2009).
Notes: type host; it was reported as *Ageneiosus brevifilis* and *Pseudoageneiosus brevifilis*. Rego (1992) redescribed *Gibsoniela mandube* and considered this species as senior synonym of *Endorchis mandube*, which was previously corroborated by de Chambrier (1990); after re-examination of the type (both species) and newly collected material, de Chambrier and Vaucher (1999) assumed that they represent two distinct species of the genus *Gibsoniela*, thus they proposed *G. meursaulti* to avoid the homonomy with the specimens tentatively identified as *E. mandube*. Sequences of partial 18S (AY551109), complete ITS2 (AY551145), partial 28S (AJ388631) and 16S (AJ389497) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Ageneiosus militaris (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2009).

Harriscolex kaparari* (Woodland, 1935) Rego, 1987
[Syns. *Nomimoscolex karapari* Woodland, 1935; *Houssayela karapari* (Woodland, 1935) Brooks, 1995]

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Orinoco River basin; Venezuela (Brooks and Rasmussen 1984).

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná and São Francisco River basins; Brazil (Rego 1990, 2002; Machado et al. 1994, 1995, 1996; Kohn et al. 2011; Ribeiro et al. 2014)

Note: records of *Harriscolex kaparari* from the Paraná River basin need verification, since *H. nathaliae* was described from the same river basin and fish host.

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Pavanelli and Machado 1991; Campos et al. 2008, 2009a, b; de Chambrier et al. 2015a).

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1935b; Rego 1987b, 2002; de Chambrier and Vaucher 1999; Zehnder et al. 2000; de Chambrier et al. 2004b).

Notes: type host. Sequence of partial 28S (AJ275227) and 16S (AJ275223) (de Chambrier et al. 2004b; Zehnder et al. 2000).

***Harriscolex nathaliae* Gil de Pertierra & de Chambrier, 2013**

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Paraguay (de Chambrier and Vaucher 1999; Gil de Pertierra and de Chambrier 2013).

Note: type host; de Chambrier and Vaucher (1999) reported the tapeworms as *Harriscolex* cf. *kaparari*; they observed morphological differences between the type material from the Amazon River and their specimens, which were posteriorly described as *H. nathaliae*.

***Harriscolex piramutab* (Woodland, 1933) de Chambrier, Kuchta & Scholz, 2015**

[Syns. *Anthobothrium piramutab* Woodland, 1933; *Proteocephalus piramutab* (Woodland, 1933) Rego, 1984]

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1933c; Rego 1984a, 1987b; de Chambrier et al. 2006a, 2015a).

Note: type host.

Houssayela sudobim* (Woodland, 1935) Rego, 1987

[Syns. *Myzophorus sudobim* Woodland, 1935; *Nomimoscolex woodlandi* Freze, 1965 *nec N. woodlandi* Rego & Pavanelli, 1992]

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1935a; Rego 1987b, 2002; de Chambrier and Scholz 2005; de Chambrier et al. 2006a, 2015a, b).

Notes: type host. Rego (1999) presented a scanning electron micrograph of the tapeworm scolex, but he did not mention the host and locality; in fact, it corresponds to the scolex of *Choanoscolex abscisus* – see de Chambrier et al. (2006a). de Chambrier et al. (2015b) erroneously reported *Sorubimichthys planiceps* as the host in their Table I. Sequence of partial 28S (KP729404) (de Chambrier et al. 2015b).

***Nomimoscolex admonticellia* (Woodland, 1934) Rego & Pavanelli, 1992**

[Syns. *Myzophorus admonticellia* Woodland, 1934 (*pro parte*); *Paramonticellia admonticellia* (Woodland, 1934) Brooks, 1995; *Myzophorus schaefferi* Pavanelli and Machado, 1991 (*nomen nudum*)]

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Woodland 1934a; Pavanelli and Machado 1991; Rego and Pavanelli 1992; Zehnder et al. 2000; Hypša et al. 2005; de Chambrier et al. 2015a).

Notes: see p. 40 in the section of *M. ventrei* for notes on the type host. Rego and Pavanelli (1992) redescribed this species based on a mixture of *N. admonticellia* and *M. ventrei* (see de Chambrier and Vaucher 1999). Sequences of partial 18S (AY551113), complete ITS2 (AY551149), partial 28S (AJ388628) and 16S (AJ389512) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Nomimoscolex alovarius* Brooks & Deardorff, 1980**

Pimelodus blochii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Magdalena River basin; Colombia (Brooks and Deardorff 1980).

Note: type host; it was reported as *Pimelodus clarias*.

***Nomimoscolex chubbi* (Pavanelli & Takemoto, 1995) de Chambrier & Vaucher, 1997**

[Syn. *Proteocephalus chubbi* Pavanelli & Takemoto, 1995]

Gymnotus carapo (Actinopterygii: Gymnotidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil, Paraguay (Pavanelli and Takemoto 1995; de Chambrier and Vaucher 1997, 1999; Zehnder and Mariaux 1999; Zehnder et al. 2000; Gil de Pertierra 2003, 2005).

Notes: type host. Sequences of partial 28S (AJ388625) and 16S (AJ389524) (Zehnder and Mariaux 1999).

Gymnotus sp. (Actinopterygii: Gymnotidae); freshwater; intestine; adult; Paraná River basin; Brazil (Isaac et al. 2004).

***Nomimoscolex dechambrieri* Gil de Pertierra, 2003**

Gymnotus carapo (Actinopterygii: Gymnotidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2003).

Note: type host.

***Nomimoscolex dorad* (Woodland, 1935) Freze, 1965**

[Syn. *Myzophorus dorad* Woodland, 1935]

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1935a; Freze 1965; Zehnder and Mariaux 1999; Zehnder et al. 2000; Hypša et al. 2005).

Notes: type host; it was reported as *Brachyplatystoma flavicans* in some studies. Sequences of partial 18S (AY551114), complete ITS2 (AY551150), partial 28S (AJ388613) and 16S (AJ389498) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Nomimoscolex guillermoi* Gil de Pertierra, 2003**

Gymnotus carapo (Actinopterygii: Gymnotidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 2003).
Note: type host.

***Nomimoscolex lenha* (Woodland, 1933) Woodland, 1935**

[Syns. *Proteocephalus lenha* Woodland, 1933; *Paramonticellia lenha* (Woodland, 1933) Brooks, 1995]

Sorubimichthys planiceps (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1933a, 1935b; de Chambrier and Vaucher 1997; Zehnder and Mariaux 1999; Zehnder et al. 2000; Hypša et al. 2005; de Chambrier and Scholz 2008; de Chambrier et al. 2015a).
Notes: type host; it was originally reported as *Platystomatichthys sturio*. Sequences of partial 18S (AY551115), complete ITS2 (AY551151), partial 28S (AJ388611) and 16S (AJ389499) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Nomimoscolex lopesi* Rego, 1989**

[Syn. *Paramonticellia lopesi* (Rego, 1989) Brooks, 1995]

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Argentina, Brazil, Paraguay, Peru (Rego 1989, 2002; de Chambrier and Vaucher 1999; Zehnder et al. 2000; Gil de Pertierra 2005; Hypša et al. 2005; de Chambrier et al. 2006a, 2015a).
Note: type host. Sequences of partial 18S (AY551116), complete ITS2 (AY551152), partial 28S (AJ388618) and 16S (AJ389521) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Nomimoscolex matogrossensis* Rego & Pavanelli, 1990**

Hoplerythrinus unitaeniatus (Actinopterygii: Erythrinidae); freshwater; body cavity, intestine, stomach; metacestode; Amazon River basin; Brazil (Alcântara and Tavares-Dias 2015; Gonçalves et al. 2016).
Note: the authors did not provide any molecular data on these larvae; therefore, this report needs verification.

Hoplias malabaricus (Actinopterygii: Erythrinidae); freshwater; intestine; adult and metacestode; Amazon and Paraná River basin; Brazil, Paraguay (Rego and Pavanelli 1990; de Chambrier et al. 1996; de Chambrier and Vaucher 1999; Zehnder et al. 2000; Alcântara and Tavares-Dias 2015; Gonçalves et al. 2016).
Notes: type host. Sequences of partial 18S (Z98387, Z98386, Z98388), 28S (AJ388614) and 16S (AJ389500) (Mariaux 1998; Zehnder and Mariaux 1999).

***Nomimoscolex microacetabula* Gil de Pertierra, 1995**

Pimelodus albicans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 1995).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina (Gil de Pertierra 1995).
Note: type host.

Pimelodus ornatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).
Note: they reported this species as *Nomimoscolex* cf. *microacetabula*.

***Nomimoscolex pertierrae* de Chambrier, Takemoto & Pavanelli, 2006**

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná river basin; Brazil (Pavanelli and Rego 1992; de Chambrier et al. 2006b; Ribeiro and Takemoto 2014; Ribeiro et al. 2014).
Note: type host. Pavanelli and Rego (1992) reported this species as *Nomimoscolex sudobim*, according to de Chambrier et al. (2006b).

Nomimoscolex piraeeba* Woodland, 1934

Brachyplatystoma capapretum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1934b; de Chambrier and Vaucher 1997; Rego 1991; Rego and Schaeffer 1999; Zehnder and Mariaux 1999; Zehnder et al. 2000; Olson et al. 2001; Hypša et al. 2005).
Notes: type host; it was identified as *B. filamentosum*. *Nomimoscolex piraeeba*, *N. dorad* and *N. suspectus* were defined as *Nomimoscolex* (*sensu stricto*) by Zehnder et al. (2000). Sequences of complete 18S (AF286988) and ITS2 (AY551153), partial (28S AJ388608, AF286936) and 16S (AJ389502) (Zehnder and Mariaux 1999; Olson et al. 2001; Hypša et al. 2005).

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Rego 1991; de Chambrier and Vaucher 1997; Rego and Schaeffer 1999).
Note: host reported as *Brachyplatystoma flavicans*.

***Nomimoscolex semenasae* Gil de Pertierra, 2002**

Olivaichthys viedmensis (Actinopterygii: Diplomystidae); freshwater; intestine; adult; Moreno and Nahuel Huapi Lakes; Argentina (Gil de Pertierra 2002a; Rauque et al. 2003).
Note: type host; it was originally reported as *Diplomystes viedmensis*.

***Nomimoscolex sudobim* Woodland, 1935**

[Syn. *Paramonticellia sudobim* (Woodland, 1935) Brooks, 1995]

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Pavanelli and Machado 1991; Machado et al. 1994, 1995, 1996; Rego 2002).
Note: records of this species from the Paraná River basin need verification, because *N. pertierrae* was described from the same river basin and host.

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon and Paraná River basins; Brazil, Peru (Woodland 1935a;

Zehnder and Mariaux 1999; Zehnder et al. 2000; Rego 2002; Santos et al. 2003; Hypša et al. 2005; Ceccarrelli et al. 2006; de Chambrier et al. 2006b, 2015a; Campos et al. 2008, 2009a, b; Jerônimo et al. 2013).
Notes: type host. Host reported as *P. reticulatum* by Jerônimo et al. (2013) (see the note on p. 36). Sequences of partial 18S (AY551117), complete ITS2 (AY551154), partial 28S (AJ388597) and 16S (AJ389496) (Zehnder and Mariaux 1999; Hypša et al. 2005).

Pseudoplatystoma tigrinum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2006a).
Note: they reported this species as *Nomimoscolex* cf. *sudobim*.

***Nomimoscolex suspectus* Zehnder, de Chambrier, Vaucher & Mariaux, 2000**

Brachyplatystoma filamentosum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder et al. 2000).
Notes: type host. Sequences of partial 28S (AJ275067) (Zehnder et al. 2000).

Brachyplatystoma cf. *filamentosum* (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder et al. 2000).
Notes: host reported as *Brachyplatystoma flavicans*. Sequence of partial 28S (AJ275068) (Zehnder et al. 2000).

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Zehnder and Mariaux 1999; Zehnder et al. 2000).
Notes: tapeworms reported as *Nomimoscolex* sp. by Zehnder and Mariaux (1999). Sequences of partial 18S (AY551118), complete ITS2 (AY551155), partial 28S (AJ388602) and 16S (AJ389519) (Zehnder and Mariaux 1999; Hypša et al. 2005).

***Nomimoscolex* sp.**

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin (estuary, Marajó Island); Brazil (Rocha et al. 2016).

Luciopimelodus pati (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Paraguay (de Chambrier and Vaucher 1999).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraíba do Sul, Paraná and São Francisco River basins; Brazil (Brasil-Sato 2003; Santos et al. 2007; Albuquerque et al. 2008; Azevedo et al. 2010, 2011).

Pimelodus ornatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; São Francisco River basin; Brazil (Brasil-Sato 2003).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River Basin; Brazil (Lopes et al. 2009).
Note: host reported as *P. punctifer* (see the note on p. 32).

Travassiella jandia* (Woodland, 1934) de Chambrier, Scholz & Kuchta, 2014

[Syns. *Proteocephalus jandia* Woodland, 1934; *Travassiella avitellina* Rego & Pavanellli, 1987]

Rhamdia quelen (Actinopterygii: Heptapteridae); freshwater; intestine; adult; Chis-Chis lagoon (Buenos Aires); Argentina (Gil de Pertierra and Ostrowski de Núñez 1990). Note: host reported as *Rhamdia sapo*.

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Argentina, Brazil, Paraguay (Rego and Pavanelli 1987; Rego and Gibson 1989; Pavanelli and Machado 1991; Takemoto and Pavanelli 1994; de Chambrier and Vaucher 1999; de Chambrier and Gil de Pertierra 2002; Kohn et al. 2011; de Chambrier et al. 2014, 2015b).

Notes: host also reported as *Zunguro zungaro* or *Paulicea luetkeni*. Rego and Gibson (1989) and Rego and Pavanelli (1987) reported hyperparasitism caused by metacestodes of proteocephalids. Sequence of 28S (KP729400) (de Chambrier et al. 2015b).

Zungaro zungaro (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Woodland 1934c; de Chambrier and Gil de Pertierra 2002; de Chambrier et al. 2006a, 2014, 2015a).

Note: type host; it was originally reported as *Rhamdia* sp.

Unidentified fish (Actinopterygii); freshwater; intestine; adult; Amazon River basin; Brazil (Rego et al. 1974).

Zygobothrium megacephalum* Diesing, 1850

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil, Peru (Diesing 1850, 1855; Woodland 1933b; Fuhrmann 1934; Rego 1984b; Zehnder and Mariaux 1999; Olson et al. 2001; Hypša et al. 2005; de Chambrier et al. 2006a, 2015a; Ruedi and de Chambrier 2012).

Notes: type host. Sequences of complete 18S (AF286991) and ITS2 (AY551177), partial 28S (AF286939, AJ388621) and 16S (AJ389508) (Zehnder and Mariaux 1999; Olson et al. 2001; Hypša et al. 2005).

Unidentified proteocephalids

Aequidens tetramerus (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; Amazon River basin; Brazil (Bittencourt et al. 2014).

Aphyocharax anisitsi (Actinopterygii: Characidae); freshwater; site of infection not given; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).

Astronotus ocellatus (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; fish farms in Northeast (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).

Astyanax altiparanae (Actinopterygii: Characidae); freshwater; body cavity; metacestode; Rio das Pedras Farm (lakes); Brazil (Azevedo et al. 2007).

Auchenipterus nigripinnis (Actinopterygii: Auchenipteridae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Brachyplatystoma filamentosum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Woodland 1935c).

Brycon cephalus (Actinopterygii: Bryconidae); freshwater; surface of intestine and pyloric caeca; metacestode; Amazon River basin; Brazil (Andrade et al. 2001).

Cichla ocellaris (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; fish farms in Northeastern Brazil (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).

Cichlasoma amazonarum (Actinopterygii: Cichlidae); freshwater; intestine; adult; Amazon River basin; Peru (de Chambrier et al. 2015a).

Note: these tapeworms belong to a new species and genus that will be described in a forthcoming paper.

Colossoma macropomum (Actinopterygii: Serrasalmidae); freshwater; body cavity; metacestode; fish farms in Northeastern Brazil (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).

Corydoras atropersonatus (Actinopterygii: Callichthyidae); freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Corydoras reticulatus (Actinopterygii: Callichthyidae); freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Corydoras sychri (Actinopterygii: Callichthyidae); freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Crenicichla lepidota (Actinopterygii: Cichlidae); freshwater; intestine; adult (immature specimens); Paraná River Basin; Paraguay (de Chambrier and Vaucher 1999).

Cynoscion striatus (Actinopterygii: Sciaenidae); marine; intestine; metacestode; WTSA; Brazil (Rego et al. 1974).

Note: certainly not a larva of the Proteocephalidae.

Cyprinus carpio (Actinopterygii: Cyprinidae); freshwater; body cavity; metacestode; fish farms in Northeastern Brazil (Ceará, Pernambuco, Piauí and Rio Grande do Norte states); Brazil (Békési et al. 1992).

Note: introduced fish host (Froese and Pauly 2016).

Galeocharax knerii (Actinopterygii: Characidae); freshwater; intestine; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).

Geophagus brasiliensis (Actinopterygii: Cichlidae); freshwater; body cavity, gallbladder; metacestode; Paraná River basin; Brazil (Bellay et al. 2012).

Geophagus proximus (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; Paraná River basin; Brazil (Zago et al. 2013).

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Hypophthalmichthys nobilis (Actinopterygii: Cyprinidae); freshwater; body cavity; metacestode; fish farms in Northeastern Brazil (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).

Note: introduced fish host (Froese and Pauly 2016).

Hypostomus cf. *ternetzi* (Actinopterygii: Loricariidae); freshwater; intestine; adult (immature specimens); Paraná River Basin; Paraguay (de Chambrier and Vaucher 1999).

Iheringichthys labrosus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Moreira et al. 2005).

Laetacara curviceps (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; Amazon River basin; Brazil (Bittencourt et al. 2014).

Leporellus vittatus (Actinopterygii: Anostomidae); freshwater; site of infection not given; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).

Leporinus aff. *friderici* (Actinopterygii: Anostomidae); freshwater; intestine; adult; Paraná River Basin; Paraguay (de Chambrier and Vaucher 1999).

Loricariichthys platymetopon (Actinopterygii: Loricariidae); freshwater; body cavity, internal organs; metacestode; Paraná River basin; Brazil (Schäeffer et al. 1992).

Megalonema platanum (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens); Paraná River Basin; Paraguay (de Chambrier and Vaucher 1999).

Oxydoras kneri (Actinopterygii: Doradidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Oreochromis sp. (Actinopterygii: Cichlidae); freshwater; body cavity; metacestode; fish farms in Northeast (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).
Note: introduced fish host (Froese and Pauly 2016).

Phractocephalus hemioliopterus (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Amazon River basin; Brazil (Rego 1984b).
Note: the author reported the presence of operculate eggs released from a contracted proglottid, but this unique report needs confirmation.

Piaractus brachypomus (Actinopterygii: Serrasalmidae); freshwater; body cavity; metacestode; fish farms in northeastern Brazil (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).
Note: host reported as *Colossoma brachipomum*.

Pimelodella gracilis (Actinopterygii: Heptapteridae); freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult (immature specimens), metacestode; Paraná and São Francisco River basins; Brazil, Paraguay (de Chambrier and Vaucher 1999; Brasil-Sato 2003).

Pimelodus pohli (Actinopterygii: Pimelodidae); freshwater; intestine; metacestode; São Francisco River basin; Brazil (Sabas and Brasil-Sato 2014).

Pimelodus sp. (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Prochilodus brevis (Actinopterygii: Prochilodontidae); freshwater; body cavity; metacestode; fish farms in Northeast (Ceará, Pernambuco, Piauí and Rio Grande do Norte States); Brazil (Békési et al. 1992).
Note: host reported as *Prochilodus cearensis*.

Prochilodus lineatus (Actinopterygii: Prochilodontidae); freshwater; body cavity; metacestode; Paraná River basin; Brazil (Lizama et al. 2005, 2006).

Psellogrammus kennedyi (Actinopterygii: Characidae); freshwater; site of infection not given; metacestode; Paraná River basin; Brazil (Takemoto et al. 2009).

Pseudoplatystoma corruscans (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Rego and Vicente 1988).

Rhamdia quelen (Actinopterygii: Heptapteridae); freshwater; intestine; adult; Guaíba River estuary (Fortes and Hoffman 1987; Rego and Gibson 1989).
Note: host reported as *Rhamdia sapo*; in both studies, the authors reported hyperparasitism caused by metacestodes of proteocephalids.

Sardinella sp. (Actinopterygii: Clupeidae); marine; intestine; metacestode; WTSA; Brazil (Feijó et al. 1979).
Note: certainly not a larva of the Proteocephalidae.

Satanoperca pappaterra (Actinopterygii: Cichlidae); freshwater; intestine; metacestode; Paraná River basin; Brazil (Yamada et al. 2007).

Trinectes maculatus (Actinopterygii: Achiridae); brackish, freshwater; mesentery; metacestode; Amazon River basin; Peru (de Chambrier et al. 2006a).

Hybrid fish host (Actinopterygii: Serrasalmidae); freshwater; intestine; metacestode; Amazon River basin; Brazil (Silva et al. 2013).
Note: the host is a hybrid of *Colossoma macropomum* and *Piaractus mesopotamicus*.

Species inquirendae

***Acanthobothroides peruensis* López, 1994**

Dasyatis dipterura (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; WTSP; Peru (López 1994).
Notes: type host; it was reported as *D. brevis*. Only *A. thorsoni* and *A. pacificum* Marques, Brooks & Ureña, 1996 are considered valid species in the genus (Marques et al. 1996).

***Monticellia diesingii* (Monticelli, 1891) La Rue, 1911**

[Syns. *Taenia diesingii* Monticelli, 1891; *Tetracotylus diesingii* Monticelli, 1891; *Ichthyotaenia diesingii* (Monticelli, 1891) Riggenbach, 1896]

'*Silurus dorgado*' (unknown fish host); freshwater; intestine; adult; unknown specific locality (Monticelli 1891; Riggenbach 1896b; La Rue 1911, 1914).

***Monticellia macrocotylea* (Monticelli, 1891) La Rue, 1911**

[Syns. *Taenia macrocotylea* Monticelli, 1891; *Tetracotylus macrocotylea* Monticelli, 1891; *Ichthyotaenia macrocotylea* (Monticelli, 1891) Riggenbach, 1896]

'*Silurus megacephalus*' (unknown fish host); freshwater; intestine; adult; unknown locality (Monticelli 1891; Riggenbach 1896b; La Rue 1911, 1914).

***Nomimoscolex arandasregoi* Fortes, 1981**

Genidens barbus (Actinopterygii: Ariidae); anadromous; intestine; adult; Guaíba River estuary, WTSA; Brazil (Fortes 1981; Fortes and Hoffmann 1995; Tavares and Luque 2004; Tavares and Luque 2008).

Note: host reported under four different names, *Tachysurus agassizii*, *T. upsulonophorus*, *T. barbus* and *Netuma barba*.

Genidens genidens (Actinopterygii: Ariidae); anadromous; intestine; adult; Guaíba River estuary; Brazil (Fortes 1981).

Note: Fortes (1981) did not designate the type host.

Genidens sp. (Actinopterygii: Ariidae); anadromous; intestine; adult; Guaíba River estuary; Brazil (Rego and Gibson 1989).

Note: Rego and Gibson (1989) reported hyperparasitism caused by metacestodes of proteocephalids.

***Platybothrium parvum* Linton, 1901**

Sphyrna zygaena (Elasmobranchii: Sphyrnidae); marine; spiral valve; adult; WTSP; Peru (Rivera and Sarmiento 1990).

Note: for details on the taxonomic status of this species, see Healy (2003).

Order Phyllobothriidea Caira, Jensen, Waeschenbach, Olson & Littlewood, 2014
Family Phyllobothriidae Braun, 1900

***Crossobothrium antonioi* Ivanov, 2009**

Notorynchus cepedianus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov 2009).

Note: type host.

***Crossobothrium dohrni* (Örley, 1885) Ruhnke, 1996**

[Syns. *Orygmatobothrium dohrni* Örley, 1885; *Phyllobothrium dohrni* (Örley, 1885) Zschokke, 1888]

Hexanchus griseus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).

Note: tapeworms reported as *Phyllobothrium dorhni*.

Crossobothrium laciniatum* Linton, 1889

[Syn. *Phyllobothrium laciniatum* (Linton, 1889) Southwell, 1925]

Hexanchus griseus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; Magellanic; Chile (Caira et al. 2014).

Notes: position of the species within Phyllobothriidae, based on molecular data, is unclear. Sequences of partial 18S (KF685824) and 28S (KF685883) (Caira et al. 2014).

***Crossobothrium pequeae* Ivanov, 2009**

Notorynchus cepedianus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov 2009).
Note: type host.

Nandocestus guariticus* (Marques, Brooks & Lasso, 2001) Reyda, 2008

[Syn. *Anindobothrium guariticus* Marques, Brooks & Lasso, 2001]

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; Amazon and Orinoco River basins; Peru, Venezuela (Marques et al. 2001; Reyda and Olson 2003; Reyda 2008; Caira et al. 2014).
Notes: type host. Reyda and Olson (2003) and Reyda (2008) reported hyperparasitism by larval stages of proteocephalids. Sequences of partial 28S (KF685888) and 18S (KF685817) (Caira et al. 2014).

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult (immature); Amazon River basin; Peru (Reyda 2008).
Note: host reported as *Potamotrygon* cf. *castexi* and it may represent an undescribed species of *Potamotrygon* (see Reyda 2008).

***Orygmatobothrium juani* Ivanov, 2008**

Mustelus fasciatus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov 2008).
Note: type host.

Orygmatobothrium musteli* (van Beneden, 1849) Diesing, 1863

[Syns. *Anthobothrium musteli* van Beneden, 1850 (*pro parte*); *Orygmatobothrium versatile* (Diesing, 1854) Diesing, 1863; *Tetrabothrium versatile* Diesing, 1854]

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974; Whittaker and Carvajal 1980).

Mustelus whitneyi (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Escalante and Carvajal 1981).

***Orygmatobothrium schmittii* Suriano & Labriola, 2001**

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1973; Suriano and Labriola 2001b; Alarcos et al. 2006; Ivanov 2008).
Notes: type host. Ivanov (2008) redescribed this species and re-assigned the specimens described as *O. velamentum* Yoshida, 1917 by Ostrowski de Núñez (1973) to *O. schmitti*.

***Orygmatobothrium* sp.**

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Tresierra et al. 1986).

***Paraorygmatobothrium angustum* (Linton, 1889) Ruhnke, 2011**

[Syns. *Orygmatobothrium angustum* Linton, 1889; *Crossobothrium angustum* (Linton, 1889) Linton, 1901; *Phyllobothrium angustum* (Linton, 1889) Euzet, 1952; *Scyphophyllidium angustum* (Linton, 1889) Riser, 1955]

Alopias vulpinus (Elasmobranchii: Alopiidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).

Note: tapeworms reported as *C. angustum*.

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal 1974; Escalante 1986).

Note: tapeworms reported as *C. angustum*.

***Paraorygmatobothrium filiforme* (Yamaguti, 1952) Ruhnke, 1996**

[Syns. *Phyllobothrium filiforme* Yamaguti, 1952; *Crossobothrium filiforme* (Yamaguti, 1952) Williams, 1968]

Carcharhinus longimanus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; TSA; Brazil (Rego 1977).

Note: tapeworms reported as *Phyllobothrium filiforme*.

Paraorygmatobothrium prionacis* (Yamaguti, 1934) Ruhnke, 1994

[Syns. *Phyllobothrium prionacis* Yamaguti, 1934; *Crossobothrium prionacis* (Yamaguti, 1934) Williams, 1968; *Anthobothrium minutum* Guiart, 1935]

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; TSA; Brazil (Rego and Mayer 1976).

Notes: type host. Tapeworms reported as *P. prionacis*.

***Paraorygmatobothrium triacis* (Yamaguti, 1952) Ruhnke, 1996**

[Syns. *Phyllobothrium triacis* Yamaguti, 1952; *Crossobothrium triacis* (Yamaguti, 1952) Euzet, 1959]

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).

Note: tapeworms reported as *C. triacis*.

Mustelus whitneyi (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Escalante and Carvajal 1981).

Note: tapeworms reported as *C. triacis*.

Phyllobothrium lactuca* van Beneden, 1850

Dipturus trachyderma (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).

Notes: tapeworms reported as *Phyllobothrium* cf. *lactuca* and the host as *Raja trachyderma*. The identification of this cestode is most likely erroneous, since sharks are the definitive hosts for species of the genus *Phyllobothrium* van Beneden, 1849 (see Ruhnke 2011).

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal 1974; Escalante and Carvajal 1981; Ruhnke and Workman 2013; Caira et al. 2014).

Notes: tapeworms reported as *Phyllobothrium* cf. *lactuca* by Ruhnke and Workman (2013) and Caira et al. (2014). Sequences of partial 18S (KF685770) and 28S (KF685845, KC505628) (Ruhnke and Workman 2013; Caira et al. 2014).

***Phyllobothrium* sp.**

Dipturus flavirostris (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).

Note: host reported as *Raja flavirostris*.

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Tresierra et al. 1986).

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (La Plata River estuary); Uruguay (Brooks et al. 1981a).

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1971).

Notes: host reported as *Psammobatis microps*. The author found only specimens without scolex, which she supposed to belong to *Phyllobothrium* (see Ostrowski de Núñez 1971).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; mesentery; metacestode: WTSA; Argentina (Szidat 1961).

Zapteryx brevirostris (Elasmobranchii: Rhinobatidae); marine; spiral valve; metacestode; WTSA; Argentina (Ostrowski de Núñez 1971).

***Scyphophyllidium uruguayense* Brooks, Marques, Perroni & Sidagis, 1999**

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Uruguay (Brooks et al. 1999).

Note: type host.

***Thysanocephalum thysanocephalum* (Linton, 1889) Braun, 1900**

[Syns. *Phyllobothrium thysanocephalum* Linton, 1889 *nec Thysanocephalum crispum* (Linton, 1889) Linton, 1890 (*nomen nudum*)]

Sphyrna zygaena (Elasmobranchii: Sphyrnidae); marine; spiral valve; adult; WTSP; Peru (López de McDonald and Tantaleán 1985).

Notes: tapeworms reported as *Thysanocephalum crispum*. According to Ruhnke (2011), the genus *Thysanocephalum* should be provisionally retained in the Phyllobothriidae.

Unidentified phyllobothriideans

Merluccius australis (Actinopterygii: Merlucciidae); marine; intestine; metacestode; Magellanic; Chile, Falkland Islands (MacKenzie and Longshaw 1995).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; intestine; metacestode; Magellanic; Argentina, Falkland Islands (MacKenzie and Longshaw 1995).

Mugil liza (Actinopterygii: Mugilidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).
Note: host reported as *Mugil platanus*.

Oncorhynchus mykiss (Actinopterygii: Salmonidae); anadromous; intestine; metacestode; Aisén River basin; Chile (Torres et al. 2000).

Prionotus sp. (Actinopterygii: Triglidae); marine; intestine; metacestode; TSA; Brazil (Vicente and Santos 1974).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2004; Luque and Poulin 2004).

Urophycis mystaceus (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2002c; Luque and Poulin 2004).

Urophycis sp. (Actinopterygii: Phycidae); marine; intestine; metacestode; TSA; Brazil (Vicente and Santos 1974).

Taxa *incertae sedis*

Guidus argentinense* Ivanov, 2006

Bathyraja brachyurops (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov 2006).
Notes: type host. Ruhnke (2011) considers *Guidus* to represent a genus *incertae sedis*.

***Phyllobothrium sinuosiceps* Williams, 1959**

Hexanchus griseus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).
Notes: type host. This species somewhat resembles members of the genus *Crossobothrium*, but it is treated as *incertae sedis* by Ruhnke (2011).

Order Rhinebothriidea Healy, Caira, Jensen, Webster & Littlewood, 2009
Family Anthocephaliidae Ruhnke, Caira & Cox, 2015

Anthocephalum gracile* Linton, 1890

[Syns. *Phyllobothrium centrurum* Southwell, 1925; *Anthocephalum centrurum* (Southwell, 1925) Ruhnke, 1994]

Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: tapeworms reported as *Phyllobothrium centrurum*.

***Anthocephalum hobergi* (Zamparo, Brooks & Barriga, 1999) Marques & Caira, 2016**

[Syn. *Pararhinebothroides hobergi* Zamparo, Brooks & Barriga, 1999]

Urobatis tumbesensis (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TEP; Ecuador (Zamparo et al. 1999; Marques and Caira 2016).
Notes: type host. Sequences of partial 18S (KU295561–KU295564) and 28S (KU295565–KU295568) (Marques and Caira 2016).

***Anthocephalum kingae* (Schmidt, 1978) Ruhnke & Seaman, 2009**
[Syn. *Phyllobothrium kingae* Schmidt, 1978]
Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).
Note: tapeworms reported as *Anthocephalum* cf. *kingae* by Brooks and Mayes (1980).
Urobatis jamaicensis (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).
Note: type host; it was reported as *Urolophus jamaicensis*. Tapeworms reported as *Anthocephalum* cf. *kingae* by Brooks and Mayes (1980).

Family Echeneibothriidae de Beauchamp, 1905

***Echeneibothrium megalosoma* Carvajal & Dailey, 1975**
Dipturus flavirostris (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).
Note: host reported as *Raja flavirostris*.
Zearaja chilensis (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Dailey 1975).
Notes: type host; it was reported as *Raja chilensis*.

***Echeneibothrium multiloculatum* Carvajal & Dailey, 1975**
Dipturus flavirostris (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).
Note: host reported as *Raja flavirostris*.
Zearaja chilensis (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Dailey 1975; Carvajal et al. 1985).
Note: type host; it was reported as *Raja chilensis*.

***Echeneibothrium williamsi* Carvajal & Dailey, 1975**
Dipturus flavirostris (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).
Note: host reported as *Raja flavirostris*.
Zearaja chilensis (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Carvajal and Dailey 1975; Carvajal et al. 1985).
Note: type host; it was reported as *Raja chilensis*.

Notomegarhynchus navonae* Ivanov & Campbell, 2002
Atlantoraja castelnaui (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Ivanov and Campbell 2002).
Note: type host.

Family Rhinebothriidae Euzet, 1953

***Rhinebothrium brooksi* Reyda & Marques, 2011**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).
Notes: type host. Sequences of partial *cox*1 (JF803719–JF803724) under the name *Rhinebothrium* sp. 1 (Reyda and Marques 2011).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).

***Rhinebothrium chilensis* Euzet & Carvajal, 1973**

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Tanzola et al. 1998).

Sympterygia lima (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile (Euzet and Carvajal 1973).
Note: type host; it was reported as *Psammobatis lima*.

***Rhinebothrium copianullum* Reyda, 2008**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Tocantins-Araguaia River basins; Brazil, Peru (Reyda 2008; Healy et al. 2009; Reyda and Marques 2011).
Notes: type host. Reyda and Marques (2011) redescribed this species and considered *Rhinebothrium* sp. 1 of Reyda (2008) to be conspecific; the latter author reported hyperparasitism caused by metacestodes of proteocephalids. Tapeworms reported as *Rhinebothrium* sp. 8 by Healy et al. (2009). Sequences of partial *cox*1 (JF803694–JF803698, JF803700, JF803701, JF803703–JF803710, JF803712–JF803714, JF803726–JF803728), 18S (FJ177090) and 28S (FJ177130) (Healy et al. 2009; Reyda and Marques 2011).

Potamotrygon henlei (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Tocantins-Araguaia River basin; Brazil (Reyda and Marques 2011).

Potamotrygon leopoldi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).
Note: sequence of *cox*1 (JF803711) (Reyda and Marques 2011).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult (immature); Amazon River basin; Brazil (Reyda and Marques 2011).
Note: accidental host (Reyda and Marques 2011).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).

Potamotrygon schroederi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult (immature); Amazon River basin; Brazil (Reyda and Marques 2011).
Note: accidental host, according to Reyda and Marques (2011).

Potamotrygon tatianae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult (immature); Amazon River basin; Peru (Reyda and Marques 2011).

Notes: accidental host. Sequence of *cox*1 (JF803699) (Reyda and Marques 2011).

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Tocantins-Araguaia River basins; Brazil (Reyda and Marques 2011).

Notes: the authors distinguished four morphotypes of the host. Sequence of *cox*1 (JF803702, JF803715–JF803718) (Reyda and Marques 2011).

***Rhinebothrium corbatai* Menoret & Ivanov, 2011**

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina (Menoret and Ivanov 2011).

Note: type host.

***Rhinebothrium corymbum* Campbell, 1975**

Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Note: type host.

***Rhinebothrium fulbrighti* Reyda & Marques, 2011**

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin (estuary); Brazil (Reyda and Marques 2011).

Notes: type host. Sequences of *cox*1 (JF803725, JF803729–JF803734) under the name *Rhinebothrium* sp. 2 (Reyda and Marques 2011).

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin (estuary); Brazil (Reyda and Marques 2011).

***Rhinebothrium jaimei* Marques & Reyda, 2015**

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin (estuary); Brazil (Marques and Reyda 2015).

Note: type host.

Potamotrygon scobina (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin (estuary); Brazil (Marques and Reyda 2015).

***Rhinebothrium leiblei* Euzet & Carvajal, 1973**

Sympterygia lima (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile (Euzet and Carvajal 1973).

Note: type host; it was reported as *Psammobatis lima*.

***Rhinebothrium margaritense* Mayes & Brooks, 1981**

Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Note: type host.

Rhinebothrium mistyae **Menoret & Ivanov, 2011**

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina (Menoret and Ivanov 2011).
Note: type host.

Rhinebothrium paratrygoni **Rego & Dias, 1976**

[Syn. *Rhinebothrium paranaensis* Menoret & Ivanov, 2009]

Potamotrygon brachyura (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Reyda and Marques 2011).

Potamotrygon falkneri (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina, Brazil, Paraguay (Brooks et al. 1981b; Lacerda et al. 2008, 2009; Menoret and Ivanov 2009a; Reyda and Marques 2011).
Note: sequences of *cox*1 (JF803684, JF803685, JF803687–JF803689, JF803691, JF803692) (Reyda and Marques 2011).

Potamotrygon histrix (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Brazil (Reyda and Marques 2011).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil (Brooks and Amato 1992; Reyda and Marques 2011).
Note: sequences of *cox*1 (JF803686, JF803690, JF803693) (Reyda and Marques 2011).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Orinoco River basin; Venezuela (Brooks et al. 1981b).
Note: host reported as *P. hystrix* and *P. reticulatus* (for details, see Brooks and Amato 1992).

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil (Rego and Dias 1976; Reyda and Marques 2011).
Note: type host; it was reported as *Elipesurus* sp.; Reyda and Marques (2011) distinguished two morphotypes of hosts that are most likely new for science.

Rhinebothrium rhinobati **Dailey & Carvajal, 1976**

Rhinobatos planiceps (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Dailey and Carvajal 1976; Tantaleán 1991; Iannacone et al. 2011).
Note: type host.

Rhinebothrium scobinae **Euzet & Carvajal, 1973**

Psammobatis scobina (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSP; Chile (Euzet and Carvajal 1973).
Note: type host.

***Rhinebothrium tetralobatum* Brooks, 1977**

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks 1977).
Note: type host.

***Rhinebothrium* sp.**

Gobionellus oceanicus (Actinopterygii: Gobiidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda and Olson 2003).
Notes: they reported hyperparasitism caused by metacestodes of proteocephalids. Sequences of partial 28S (AY193880–AY193883) from adult tapeworms and partial 28S (AY193877–AY193879) from the encysted larval forms (Reyda and Olson 2003).

Scomber colias (Actinopterygii: Scombridae); marine; intestine, pyloric caeca; metacestode; WTSA; Brazil (Rego and Santos 1983).
Note: host reported as *S. japonicus*, but specimens from the Atlantic were reassigned as *S. colias*, according to Froese and Pauly (2016).

Synodus scituliceps (Actinopterygii: Synodontidae); marine; intestine, pyloric caeca; metacestode; WTSP; Peru (Escalante et al. 1987).

***Rhinebothroides campbelli* Ivanov, 2004**

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina (Ivanov 2004).
Notes: type host. Ivanov (2004) arose doubts concerning the conspecificity between *Rhinebothroides venezuelensis* and *R. circularisi* proposed by Marques and Brooks (2003).

***Rhinebothroides freitasi* (Rego, 1979) Brooks, Mayes & Thorson, 1981**

[Syns. *Rhinebothrium freitasi* Rego, 1979; *Rhinebothroides circularisi* Mayes, Brooks & Thorson, 1981; *Rhinebothroides venezuelensis* Brooks, Mayes & Thorson, 1981]

Potamotrygon constellata (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Mayes et al. 1981).
Note: host reported as *P. circularis*, whereas tapeworms were reported as *R. circularisi*.

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil, Peru (Marques and Brooks 2003; Healy et al. 2009; Caira et al. 2014).
Notes: Healy et al. (2009) and Caira et al. (2014) reported the tapeworms as *Rhinebothroides* cf. *freitasi* and the hosts as *P.* cf. *castexi*. Sequences of partial 18S (FJ177092) and 28S (FJ177132) (Healy et al. 2009; Caira et al. 2014).

Potamotrygon henlei (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Tocantins-Araguaia River basin; Brazil (Marques and Brooks 2003).

Potamotrygon leopoldi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Argentina, Brazil (Brooks and Amato 1992; Marques and Brooks 2003).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Orinoco River basins; Brazil, Venezuela (Rego 1979; Brooks et al. 1981b; Marques and Brooks 2003).

Notes: type host; it was reported as *P. hystrix* by Rego (1979) and Brooks et al. (1981b).

Potamotrygon schroederi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003).

Potamotrygon scobina (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003).

Potamotrygon yepezi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Maracaibo basin; Venezuela (Brooks et al. 1981b).

***Rhinebothroides glandularis* Brooks, Mayes & Thorson, 1981**

Potamotrygon henlei (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Tocantins-Araguaia River basin; Brazil (Marques and Brooks 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Argentina, Brazil (Marques and Brooks 2003; Reyda and Marques 2011).

Note: sequence of partial *cox*1 (JF803682) (Reyda and Marques 2011).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Orinoco River basins; Brazil, Venezuela (Brooks et al. 1981b; Marques and Brooks 2003; Ivanov 2004).

Notes: type host; it was originally reported as *P. hystrix*. Ivanov (2004) studied the type specimens deposited in USNPC and HWML.

Potamotrygon scobina (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003).

Potamotrygon signata (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Parnaíba River basin; Brazil (Marques and Brooks 2003).

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Orinoco River basins; Venezuela (Marques and Brooks 2003; Reyda and Marques 2011).

Notes: Marques and Brooks (2003) reported immature specimens. Sequence of partial *cox*1 (JF803683) (Reyda and Marques 2011).

***Rhinebothroides mclennanae* Brooks & Amato, 1992**

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Argentina, Brazil (Brooks and Amato 1992; Ivanov 2004).

Notes: type host. Marques and Brooks (2003) synonymized this species with *R. glandularis*, but after evaluation of newly collected material, Ivanov (2004) considered this taxon a valid species.

Rhinebothroides moralarai* (Brooks & Thorson, 1976) Mayes, Brooks & Thorson, 1981

[Syn. *Rhinebothrium moralarai* Brooks & Thorson, 1976]

Potamotrygon magdalenae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Magdalena River basin; Colombia (Brooks and Thorson 1976; Brooks et al. 1981b).

Notes: type host. Brooks et al. (1981b) studied the type specimens deposited in USNM.

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003; Reyda and Marques 2011).

Note: sequence of partial *cox*1 (JF803681) (Reyda and Marques 2011).

***Rhinebothroides scorzai* (Lopez-Neyra & Diaz-Ungria, 1958) Mayes, Brooks & Thorson, 1981**

[Syn. *Rhinebothrium scorzai* Lopez-Neyra & Diaz-Ungria, 1958]

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques and Brooks 2003).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil (Rego and Dias 1976; Marques and Brooks 2003).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Orinoco River basin; Venezuela (Lopez-Neyra and Diaz-Ungria 1958).

Note: type host; it was originally reported as *P. hystrix*.

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Reyda and Marques 2011).

Note: sequence of partial *cox*1 (JF803680) (Reyda and Marques 2011).

***Rhinebothroides* sp.**

Paratrygon aiereba (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult (immature); Amazon River basin; Peru (Reyda 2008).

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Brazil, Peru (Reyda and Olson 2003; Reyda 2008; Reyda and Marques 2011).

Notes: Reyda and Olson (2003) and Reyda (2008) reported the host as *Potamotrygon castexi* and the latter reported the cestodes as *Rhinebothroides* sp. 2. Reyda and Olson (2003) found metacestodes of proteocephalids parasitizing *Rhinebothroides* sp. Sequence of partial *cox*1 (JF803678) (Reyda and Marques 2011).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda and Olson 2003, Reyda 2008).
Note: the authors reported hyperparasitism caused by metacestodes of proteocephalids.

Potamotrygon tatianae (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda and Marques 2011).
Note: sequence of partial *cox*1 (JF803679) (Reyda and Marques 2011).

***Rhodobothrium mesodesmatum* (Bahamonde & Lopez, 1962) Campbell & Carvajal, 1979**

[Syns. *Proboscidosaccus mesodesmatis* Bahamonde & Lopez, 1962; *Anthobothrium peruanum* Rego, Vicente & Herrera, 1968]

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Campbell and Carvajal 1979; Rodríguez and Tantaleán 1980; Oliva 1982; Tresierra et al. 1986).
Note: type host. Tapeworm originally described by Bahamonde and Lopez (1962) from a clam, *Mesodesma donacium* (Lamarck), parasitizing the palleal cavity of this intermediate host, which is a common prey for *M. chilensis*.

Myliobatis peruvianus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (López de McDonald and Tantaleán 1985).

Sarda chiliensis (Actinopterygii: Scombridae); marine; intestine; adult (?); WTSP; Peru (Rego et al. 1968).

***Rhodobothrium paucitesticulare* Mayes & Brooks, 1981**

Rhinoptera bonasus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981; Brooks et al. 1981a).
Note: type host.

Rhodobothrium pulvinatum* Linton, 1889

[Syns. *Anthobothrium pulvinatum* Linton, 1890 *nec A. pulvinatum* Linton, 1889 (*nomen nudum*); *Inermiphyllidium pulvinatum* (Linton, 1890) Riser, 1955]

Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

***Scalithrium magniphallum* (Brooks, 1977) Ball, Neifar & Euzet, 2003**

[Syn. *Rhinebothrium magniphallum* Brooks, 1977]

Dasyatis americana (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).

Dasyatis guttata (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia, Venezuela (Brooks 1977; Mayes and Brooks 1981).
Note: type host.

Urobatis jamaicensis (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).
Notes: host reported as *Urolophus jamaicensis*.

Urotrygon venezuelae (Elasmobranchii: Urotrygonidae); marine; spiral valve; adult; TNA; Colombia (Brooks and Mayes 1980).

Taxa *incertae sedis*

Anindobothrium anacolum* (Brooks, 1977) Marques, Brooks & Lasso, 2001
[Syn. *Caulobothrium anacollum* Brooks, 1977]

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Colombia (Brooks 1977).
Notes: type host. The genus *Anindobothrium* is likely a member of the Rhinebothriidea (see Ruhnke 2011) and it is already treated as such in the GCD within Anthocephalidae (Caira et al. 2012).

***Anindobothrium lisae* Marques, Brooks & Lasso, 2001**

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Marques et al. 2001).
Note: type host.

***Phyllobothrium auricula* van Beneden, 1858**

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Tantaleán 1991).
Note: the species is likely a member of the Rhinebothriidea (see Ruhnke 2011) and it is already treated as such in the GCD (Caira et al. 2012).

Myliobatis peruvianus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Peru (Tantaleán 1991).

***Phyllobothrium discopygi* Campbell & Carvajal, 1987**

Discopyge tschudi (Elasmobranchii: Torpedinidae); marine; spiral valve; adult; WTSP; Chile (Campbell and Carvajal 1987).
Notes: type host. This species is likely a member of the Rhinebothriidea (see Ruhnke 2011) and it is already treated as such in the GCD (Caira et al. 2012).

***Phyllobothrium myliobatidis* Brooks, Mayes & Thorson, 1981**

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (La Plata River estuary); Uruguay (Brooks et al. 1981a).
Notes: type host. The tapeworm is likely a member of the Rhinebothriidea (see Ruhnke 2011) and it is already treated as such in the GCD (Caira et al. 2012).

Order 'Tetraphyllidea' Carus, 1863

***Anthobothrium laciniatum* Linton, 1890**

Carcharhinus longimanus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; TSA; Brazil (Rego 1977).

***Anthobothrium galeorhini* Suriano, 2002**

Galeorhinus galeus (Elasmobranchii: Triakidae); marine; spiral valve; adult; Magellanic; Argentina (Suriano 2002).
Note: type host.

***Calliobothrium australis* Ostrowski de Núñez, 1973**

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Argentina, Uruguay (Ostrowski de Núñez 1973; Ivanov and Brooks 2002; Alarcos et al. 2006; Bernot et al. 2015).
Notes: type host. This species was originally described as *C. verticillatum australis*, but Ivanov and Brooks (2002) redescribed this species and they raised this subspecies to the species level as *C. australis*. Sequences of partial 28S (KP128030, KP128031) (Bernot et al. 2015).

Calliobothrium verticillatum* (Rudolphi, 1819) van Beneden, 1850

[Syns. *Bothriocephalus verticillatus* Rudolphi, 1819; *Acanthobothrium verticillatum* (Rudolphi, 1819) van Beneden, 1849]

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal 1974; Escalante and Carvajal 1981).

***Calliobothrium* sp.**

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Tresierra et al. 1986).

***Caulobothrium myliobatidis* Carvajal, 1977**

Myliobatis chilensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Carvajal 1977; Tantaleán 1991).
Note: type host.

***Caulobothrium ostrowskiae* Brooks, Mayes & Thorson, 1981**

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (La Plata River estuary); Uruguay (Brooks et al. 1981a).
Note: type host.

***Caulobothrium uruguayense* Brooks, Mayes & Thorson, 1981**

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA (La Plata River estuary); Uruguay (Brooks et al. 1981a).
Note: type host; it was reported as *Myliobatis uruguayensis*.

***Dioecotaenia campbelli* Mayes & Brooks, 1981**

Rhinoptera bonasus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Brooks et al. 1981a; Mayes and Brooks 1981).
Note: type host.

Reesium paciferum* (Sproston, 1948) Euzet, 1955

[Syn. *Dinobothrium paciferum* Sproston, 1948]

Cetorhinus maximus (Elasmobranchii: Cetorhinidae); marine; spiral valve; adult; WTSP; Peru (Rivera and Sarmiento 1990).
Note: type host.

Serendip deborahae* Brooks & Barriga, 1995

Rhinoptera steindachneri (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TEP; Ecuador (Brooks and Barriga 1995).
Notes: type host. *Serendip* Brooks & Barriga, 1995 is likely a member of the Rhinebothriidea, according to Ruhnke et al. (2015).

***Symcallio barbarae* (Ivanov & Brooks, 2002) Bernot, Caira & Pickering, 2015**

[Syns. *Calliobothrium eschrichti* of Ostrowski de Núñez (1973) *nec* van Beneden, 1850; *Calliobothrium barbarae* Ivanov & Brooks, 2002]

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Argentina, Uruguay (Ostrowski de Núñez 1973; Ivanov and Brooks 2002; Alarcos et al. 2006; Bernot et al. 2015).
Note: type host. Ostrowski de Núñez (1973) reported the tapeworms as *Calliobothrium eschrichti* van Beneden, 1850. Sequence of partial 28S (KP128023) (Bernot et al. 2015).

***Symcallio lintoni* (Euzet, 1954) Bernot, Caira & Pickering, 2015**

[Syn. *Calliobothrium lintoni* Euzet, 1954]

Mustelus whitneyi (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Peru (Escalante and Carvajal 1981).

***Symcallio lunae* (Ivanov & Brooks, 2002) Bernot, Caira & Pickering, 2015**

[Syns. of *Calliobothrium lintoni* of Ostrowski de Núñez (1973) *nec* Euzet, 1974; *Calliobothrium lunae* Ivanov & Brooks, 2002]

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Argentina, Uruguay (Ostrowski de Núñez 1973; Ivanov and Brooks 2002; Alarcos et al. 2006).
Notes: type host. Ostrowski de Núñez (1973) reported this species as *Calliobothrium lintoni* (see Caira et al. 2012).

Collective group name for larval 'tetraphyllideans' and Unidentified taxa

'*Scolex pleuronectis*' Müller, 1788; '*Scolex polymorphus*' Rudolphi, 1819; '*Scolex* sp.'; 'Tetraphyllidea gen. sp.'

Anchoa tricolor (Actinopterygii: Engraulidae); marine; intestine; metacestode; WTSA; Brazil (Tavares et al. 2005).

Aphos porosus (Actinopterygii: Batrachoididae); marine; site of infection not given; metacestode; WTSP; Chile (Torres et al. 1993; Cortés and Muñoz 2008, 2009).

Aspistor luniscutis (Actinopterygii: Ariidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).
Note: host reported as *Sciadeichthys luniscutis*.

Balistes capriscus (Actinopterygii: Balistidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves et al. 2005).

Balistes vetula (Actinopterygii: Balistidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Brachyplatystoma sp. (Actinopterygii: Pimelodidae); freshwater; site of infection not given; metacestode; Amazon River basin (estuary of Amazon River); Brazil (Rego et al. 1999a).

Caranx latus (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Alves 2001).

Cilus gilberti (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSP; Chile (García et al. 2001).

Conger orbignianus (Actinopterygii: Congridae); marine; intestine; metacestode; WTSA; Argentina (Tanzola and Guagliardo 2000; Timi and Lanfranchi 2013).
Note: Tanzola and Guagliardo (2000) distinguished two morphotypes.

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; pyloric caeca, intestine; metacestode; WTSA; Argentina (Timi et al. 2005, 2010b).

Dactylopterus volitans (Actinopterygii: Dactylopteridae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Cordeiro and Luque 2005).

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; stomach, intestine; metacestode; Magellanic; Falkland Islands (Brickle et al. 2006; Brown et al. 2013).

Eleginops maclovinus (Actinopterygii: Eleginopsidae); marine; intestine; metacestode; Magellanic; Falkland Islands (Brickle and MacKenzie 2007).

Engraulis anchoita (Actinopterygii: Engraulidae); marine; pyloric caeca; metacestode; Magellanic, WTSA; Argentina (Timi 2003; Timi and Poulin 2003; Timi et al. 2010b).

Engraulis ringens (Actinopterygii: Engraulidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Moscoso 2013).

Ethmidium maculatum (Actinopterygii: Clupeidae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).

Note: host reported as *Brevoortia maculata*.

Euthynnus alletteratus (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves and Luque 2006).

Genidens barbus (Actinopterygii: Ariidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004; Tavares and Luque 2004, 2008).

Note: Luque and Poulin (2004) and Tavares and Luque (2004) reported the host as *Netuma barba*.

Genypterus blacodes (Actinopterygii: Ophidiidae); marine; intestine; metacestode; WTSA; Argentina (Sardella et al. 1998).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; intestine; metacestode; WTSA; Brazil (Alves et al. 2002a, b; Luque and Poulin 2004).

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; intestine; metacestode; WTSP; Chile, Peru (Escalante et al. 1987; Muñoz and George-Nascimento 2008).

Gobiesox marmoratus (Actinopterygii: Gobiesocidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz 2014).

Gymnothorax moringa (Actinopterygii: Muraenidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Haemulon steindachneri (Actinopterygii: Haemulidae); marine; intestine; metacestode; WTSA; Brazil (Luque et al. 1995, 1996a, b; Luque and Poulin 2004).

Helcogrammoides chilensis (Actinopterygii: Tripterygiidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz and Delorme 2011).

Hippoglossina macrops (Actinopterygii: Paralichthyidae); marine; intestine; metacestode WTSP; Chile (Riffo 1991; González et al. 2001, 2008; Oliva et al. 2004).

Hyporhamphus unifasciatus (Actinopterygii: Hemiramphidae); brackish, marine; body cavity; metacestode; TSA (Itamaracá Island); Brazil (Palm 1997).

Katsuwonus pelamis (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSA, WTSP; Brazil, Peru (Escalante et al. 1987; Alves and Luque 2006)

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; intestine; metacestode; Magellanic; Chile (Oliva 2001).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Merluccius australis (Actinopterygii: Merlucciidae); marine; intestine; metacestode; Magellanic; Chile (Fernández 1985).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; intestine; metacestode; WTSP; Chile (Oliva and Ballón 2002).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; pyloric caeca, intestine; metacestode; Magellanic, WTSA; Argentina, Uruguay (Szidat 1955, 1961; Sardella and Timi 1996, 2004).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; intestine; metacestode; WTSA; Brazil (Alves and Luque 2001a; Luque and Poulin 2004; Luque et al. 2010).

Mugil cephalus (Actinopterygii: Mugilidae); marine; intestine; metacestode; WTSP; Chile (Fernández 1987).

Mugil liza (Actinopterygii: Mugilidae); catadromous; intestine; metacestode; WTSA; Brazil (Knoff et al. 1997; Luque and Poulin 2004).
Notes: host reported as *Mugil platanus*. Knoff et al. (1997) distinguished two morphotypes.

Normanichthys crockeri (Actinopterygii: Normanichthyidae); marine; intestine, ovary; metacestode; Magellanic; Chile (Sepúlveda et al. 2004).

Notothenia cf. *angustata* (Actinopterygii: Nototheniidae); marine; intestine; metacestode; WTSP; Chile (Muñoz et al. 2001).

Odontesthes argentinensis (Actinopterygii: Atherinopsidae); brackish, marine; site of infection not given; metacestode; WTSA (Mar Chiquita coastal lagoon); Argentina (Alarcos and Etchegoin 2010).

Odontesthes regia (Actinopterygii: Atherinopsidae); brackish, marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).
Note: host reported as *O. regia regia*.

Odontesthes smitti (Actinopterygii: Atherinopsidae); marine; body cavity, mesentery, stomach; metacestode; Magellanic; Argentina (Carballo et al. 2011, 2012).

Oligoplites palometa (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSA; Brazil (Takemoto et al. 1996a, b; Luque and Poulin 2004).

Oligoplites saliens (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSA; Brazil (Takemoto et al. 1996a, b; Luque and Poulin 2004).

Oligoplites saurus (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSA; Brazil (Takemoto et al. 1996a, b; Luque and Poulin 2004).

Oncorhynchus kisutch (Actinopterygii: Salmonidae); anadromous; intestine; metacestode; lakes in Chiloé Island; Chile (Torres et al. 1990, 2010).

Orthopristis ruber (Actinopterygii: Haemulidae); marine; intestine; metacestode; WTSA; Brazil (Luque et al. 1995, 1996a, b; Luque and Poulin 2004).

Pagrus pagrus (Actinopterygii: Sparidae); marine; mesentery; metacestode; WTSA; Brazil (Paraguassú et al. 2002; Luque and Poulin 2004; Soares et al. 2014).

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; intestine; metacestode; WTSP; Chile (Riffo 1995; Oliva et al. 1996).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; stomach, intestine; metacestode; WTSA; Argentina, Brazil (Felizardo et al. 2010; Alarcos and Timi 2012; Alarcos et al. 2016).

Paralichthys microps (Actinopterygii: Paralichthyidae); marine; intestine; metacestode; WTSP; Chile (Riffo 1995).

Paralichthys orbignyanus (Actinopterygii: Paralichthyidae); marine; site of infection not given; metacestode; WTSA (Mar Chiquita coastal lagoon); Argentina (Alarcos and Etchegoin 2010).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; stomach, intestine; metacestode; WTSA; Argentina (Alarcos and Timi 2012).

Paralonchurus brasiliensis (Actinopterygii: Sciaenidae); marine; intestine; metacestode; WTSA; Brazil (Ribeiro et al. 2002; Luque et al. 2003; Luque and Poulin 2004).

Parona signata (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSA; Argentina (Szidat 1969).

Percophis brasiliensis (Actinopterygii: Percophidae); marine; intestine; metacestode; WTSA; Argentina, Uruguay (Braicovich and Timi 2008, 2010).

Pinguipes brasilianus (Actinopterygii: Pinguipedidae); marine; intestine; metacestode; WTSA; Argentina, Brazil (Timi et al. 2008, 2009, 2010a).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; intestine; metacestode; WTSA; Brazil (Luque and Chaves 1999; Luque and Poulin 2004).
Note: host reported as *P. saltator*.

Porichthys porosissimus (Actinopterygii: Batrachoididae) marine; intestine; metacestode; WTSA (Bahía Blanca estuary); Argentina (Tanzola et al. 1997; Guagliardo et al. 2009).
Notes: Tanzola et al. (1997) distinguished two morphotypes. Guagliardo et al. (2009) studied lesions caused by the larvae.

Priacanthus arenatus (Actinopterygii: Priacanthidae); marine; intestine; metacestode; WTSA; Brazil (Tavares et al. 2001; Luque and Poulin 2004).

Prolatilus jugularis (Actinopterygii: Pinguipedidae); marine; intestine, ovary; metacestode; Magellanic; Chile (Sepúlveda et al. 2004).

Pseudopercis numida (Actinopterygii: Pinguipedidae); marine; intestine; metacestode; WTSA; Brazil (Luque et al. 2008).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; intestine; metacestode; Magellanic, WTSA; Argentina, Brazil (Luque et al. 2008; Timi and Lanfranchi 2009a).

Raneya brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSA; Argentina (Vales et al. 2011).

Salmo trutta (Actinopterygii: Salmonidae); anadromous; intestine; metacestode; lakes in Chiloé Island; Chile (Torres et al. 1990).

Sarda chiliensis (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).
Note: host reported as *S. chiliensis chiliensis*.

Sarda sarda (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSA; Brazil (Alves and Luque 2006).

Sardinella brasiliensis (Actinopterygii: Clupeidae); marine; pyloric caeca (cysts); metacestode; WTSA; Brazil (Lima et al. 1997).

Sardinops sagax (Actinopterygii: Clupeidae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).
Note: host reported as *S. sagax sagax*.

Scomber colias (Actinopterygii: Scombridae); marine; body cavity, intestine, stomach; metacestode; WTSA; Argentina, Brazil (Rego and Santos 1983; Cremon-

te and Sardella 1997; Abdallah et al. 2002; Alves et al. 2003; Luque and Poulin 2004; Oliva et al. 2008b).

Note: host reported as *S. japonicus*.

Scomber japonicus (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987; Cruces et al. 2014).

Scomberomorus brasiliensis (Actinopterygii: Scombridae); marine; intestine; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves and Luque 2006).

Sebastes capensis (Actinopterygii: Sebastidae); marine; site of infection not given; metacestode; WTSP; Chile (González and Poulin 2005a, b; González et al. 2006).

Sicyases sanguineus (Actinopterygii: Gobiesocidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz and Delorme 2011).

Stellifer minor (Actinopterygii: Sciaenidae); marine; intestine; metacestode; WTSP; Peru (Luque 1991).

Strongylura scapularis (Actinopterygii: Belonidae); marine; intestine; metacestode; WTSP; Peru (Luque 1991).

Note: host reported as *Belone scapularis*.

Synodus scituliceps (Actinopterygii: Synodontidae); marine; intestine; metacestode; WTSP; Peru (Escalante et al. 1987).

Trachurus lathami (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSA; Argentina, Brazil (Luque and Poulin 2004; Braicovich et al. 2012).

Trachurus murphyi (Actinopterygii: Carangidae); marine; intestine; metacestode; WTSP; Chile, Peru (Oliva 1982, 1994, 1999; Luque 1991; Jara 1998).

Note: Luque (1991) and Jara (1998) reported the host as *Trachurus symmetricus murphyi*.

Trichiurus lepturus (Actinopterygii: Trichiuridae); marine; stomach, intestine; metacestode; WTSA; Brazil (Silva et al. 2000a, b; Luque and Poulin 2004; Carvalho and Luque 2011; Bueno et al. 2014).

Tylosurus acus acus (Actinopterygii: Belonidae); marine; intestine; metacestode; WTSA; Brazil (Tavares et al. 2004; Luque and Poulin 2004).

Note: host reported as *Tylosurus acus*.

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; intestine; metacestode; WTSA; Argentina, Brazil (Szidat 1960, 1961; Alves et al. 2004; Luque and Poulin 2004; Pereira et al. 2014).

Note: Pereira et al. (2014) distinguished two morphotypes of larvae.

Urophycis mystaceus (Actinopterygii: Phycidae); marine; intestine; metacestode; WTSA; Brazil (Alves et al. 2002c; Luque and Poulin 2004).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; stomach, intestine; metacestode; WTSA; Argentina (Szidat 1961; Alarcos and Timi 2012, 2013).

Taxon *incertae sedis*

***Anthobothrium pristis* Woodland, 1934**

Pristis pristis (Elasmobranchii: Pristidae); brackish, freshwater, marine; spiral valve; adult; Amazon River basin; Brazil (Woodland 1934c).

Note: host reported as *P. perotteti*. Ruhnke and Caira (2009) did not place this species among six species of *Anthobothrium* (*sensu stricto*); therefore, it is considered *incertae sedis* (see Caira et al. 2012).

Order Trypanorhyncha Diesing, 1863
Suborder Trypanoselachoida Olson, Caira, Jensen, Overstreet, Palm & Beveridge, 2010
Superfamily Gymnorhynchoidea Dollfus, 1935
Family Gilquiniidae Dollfus, 1935

Gilquinia squali* (Fabricius, 1794) Dollfus, 1930

[Syns. *Taenia squali* Fabricius, 1794; *Bothriocephalus paleaceus* Rudolphi, 1810; *Rhynchobothrium tetrabothrium* van Beneden, 1849; *Tetrarhynchobothrium affine* Diesing, 1854; *Tetrarhynchus anteroporus* Hart, 1936]

Etmopterus granulosus (Elasmobranchii: Etmopteridae); marine; spiral valve; adult (immature); WTSP; Chile (Carvajal 1974).

Note: host reported as *Centroscyllium granulosus*.

***Gilquinia* sp.**

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA (La Plata River estuary); Argentina (Suriano 1966).

Note: Pereira (1993) suggested that these specimens might have corresponded to two different species of *Pterobothrium* Diesing, 1850.

Family Gymnorhynchidae Dollfus, 1935

***Gymnorhynchus isuri* Robinson, 1959**

Isurus oxyrinchus (Elasmobranchii: Lamnidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002, 2007).

Molicola horridus* (Goodsir, 1841) Dollfus, 1935

[For synonyms, see Palm (2004)]

Isurus oxyrinchus (Elasmobranchii: Lamnidae); marine; spiral valve; adult; WTSA; Brazil (Palm 2004).

Mola ramsayi (Actinopterygii: Molidae); marine; liver; metacestode; WTSP; Chile (Villalba and Fernández 1985).

Note: tapeworms reported as *Gymnorhynchus* (*M.*) *horridus*.

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002, 2004c).

***Molicola* sp.**

Mola mola (Actinopterygii: Molidae); marine; site of infection not given; metacestode; WTSA; Brazil (Palm 2004).

Thyrsites atun (Actinopterygii: Gempylidae); marine; muscles; metacestode; WTSP; Chile (Torres et al. 2014).

Superfamily Lacistorhynchoidea Guiart, 1927
Family Lacistorhynchidae Guiart, 1927

Callitetrarhynchus gracilis* (Rudolphi, 1819) Pintner, 1931

[For synonyms, see Palm (2004)]

Balistes capriscus (Actinopterygii: Balistidae); marine; liver, mesentery; metacestode; WTSA; Brazil (Dias et al. 2009).

Balistes vetula (Actinopterygii: Balistidae); marine; body cavity, muscle; metacestode; WTSA; Brazil (São Clemente et al. 1995).

Caranx crysos (Actinopterygii: Carangidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Caranx hippos (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Alves 2001; Luque and Poulin 2004).

Caranx latus (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Alves 2001; Luque and Poulin 2004; Ferreira et al. 2006).

Centropomus undecimalis (Actinopterygii: Centropomidae); amphidromous; peritoneum; metacestode; NBS (Marajó Island); Brazil (Dollfus 1942).
Note: the cestodes were collected by Göldi in 1896.

Chloroscombrus chrysurus (Actinopterygii: Carangidae); marine; body cavity; metacestode; TSA, WTSA; Brazil (Palm 1997, 2004).

Cynoscion acoupa (Actinopterygii: Sciaenidae); marine; muscles; metacestode; NBS; Brazil (Dias et al. 2011b).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity, kidney; metacestode; WTSA; Argentina, Brazil, Uruguay (Pereira and Boeger 2005; Timi et al. 2005, 2010b).

Euthynnus alletteratus (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves and Luque 2006).

Genidens barbus (Actinopterygii: Ariidae); marine; body cavity, viscera; metacestode; WTSA; Brazil (São Clemente et al. 1991a).
Note: host reported as *Netuma barba*.

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; body cavity, mesentery, muscles; metacestode; WTSA; Brazil (São Clemente et al. 2004; Knoff et al. 2008).

Haemulon aurolineatum (Actinopterygii: Haemulidae); marine; body cavity, muscles; metacestode; TSA; Brazil (Palm 1997).

Harengula clupeola (Actinopterygii: Clupeidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Hemilutjanus macrophthalmos (Actinopterygii: Sciaenidae); marine; surface of internal organs, serous membrane; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Hyporthodus niveatus (Actinopterygii: Serranidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).
Note: host reported as *Epinephelus niveatus*.

Larimus breviceps (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Lutjanus synagris (Actinopterygii: Lutjanidae); marine; muscles; metacestode; WTSA; Brazil (Silva and São Clemente 2001).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; body cavity, kidney, mesentery; metacestode; WTSA; Brazil (Sabas and Luque 2003; Luque and Poulin 2004; Pereira and Boeger 2005).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; WTSP; Peru (Durán and Oliva 1980; Chero et al. 2014a).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity, kidney, mesentery; metacestode; TNA, WTSA; Brazil, Venezuela (São Clemente 1986a, b, 1987; Vicente et al. 1989; Pereira 1993; Alves and Luque 1999, 2001a; Luque and Poulin 2004; Palm 2004; Pereira and Boeger 2005; Luque et al. 2010; Timi et al. 2010b).

Mustelus canis (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Brazil (São Clemente and Gomes 1989b; São Clemente et al. 1991b).

Oligoplites palometa (Actinopterygii: Carangidae); marine; body cavity; metacestode; TSA, WTSA; Brazil (Takemoto et al. 1996a, b; Palm 1997; Luque and Poulin 2004).

Oligoplites saurus (Actinopterygii: Carangidae); marine; body cavity; metacestode; WTSA; Brazil (Takemoto et al. 1996a, b; Luque and Poulin 2004).

Opisthonema oglinum (Actinopterygii: Clupeidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Pagrus pagrus (Actinopterygii: Sparidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; muscles; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; body cavity, kidney, mesentery, spleen; metacestode; WTSA; Brazil (Fonseca et al. 2012).

Paralonchurus peruanus (Actinopterygii: Sciaenidae); marine; surface of internal organs, serous membrane; metacestode; WTSP; Peru (Escalante and Carvajal 1984).
Note: host reported as *Polyclemus peruanus*.

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina, Uruguay (Braicovich and Timi 2008, 2010).

Pinguipes brasilianus (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; WTSA; Brazil (Timi et al. 2010a).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; muscles; metacestode; Amazon River basin; Brazil (Silva 2010).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; body cavity, mesentery, peritoneum; metacestode; WTSA; Brazil (Carvajal and Rego 1985; Carvajal et al. 1987; São Clemente et al. 1997; Palm 2004, Ferreira et al. 2006).
Note: host reported as *P. saltator* by some authors.

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002; Pinto et al. 2006).

Pseudopercis numida (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; WTSA; Brazil (Luque et al. 2008).

Sardinella brasiliensis (Actinopterygii: Clupeidae); marine; body cavity; metacestode; WTSA; Brazil (Moreira et al. 2015).

Sciaena deliciosa (Actinopterygii: Sciaenidae); marine; surface of internal organs, serous membrane; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Scomberomorus brasiliensis (Actinopterygii: Scombridae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: host reported as *S. maculatus*.

Scomberomorus cavalla (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Dias et al. 2011a).

Selene setapinnis (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Cordeiro and Luque 2004; Luque and Poulin 2004).

Selene vomer (Actinopterygii: Carangidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Sphyraena guachancho (Actinopterygii: Sphyraenidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Trachurus lathami (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Braicovich et al. 2012).

Trichiurus lepturus (Actinopterygii: Trichiuridae); marine; body cavity, mesentery, stomach; WTSA; Brazil (Silva et al. 2000a, b; Carvalho and Luque 2011; Bueno et al. 2014).

Umbrina canosai (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; site of infection not given; metacestode; WTSA; Argentina (Pereira et al. 2014).

***Callitetrarhynchus speciosus* (Linton, 1897) Carvajal & Rego, 1985**

[Syns. *Rhynchobothrium speciosum* Linton, 1897; *Tentacularia pseudodera* Schuler, 1938]

Aluterus monoceros (Actinopterygii: Monacanthidae); marine; liver, mesentery; metacestode; WTSA; Brazil (Dias et al. 2010).

Balistes capriscus (Actinopterygii: Balistidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Dias et al. 2009).

Balistes vetula (Actinopterygii: Balistidae); marine; body cavity, mesentery, muscles; metacestode; WTSA; Brazil (São Clemente et al. 1995; Luque and Poulin 2004).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Pereira and Boeger 2005; Pinto et al. 2006).

Genidens barbus (Actinopterygii: Ariidae); marine; body cavity, viscera; metacestode; WTSA; Brazil (São Clemente et al. 1991a).

Note: host reported as *Netuma barba*.

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Pereira 1993; Pereira and Boeger 2005; Pinto et al. 2006).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; body cavity, mesentery; metacestode; WTSA; Brazil (Carvajal and Rego 1985; São Clemente et al. 1997; Ferreira et al. 2006).

Note: type host; it was reported as *P. saltator* by some authors.

Priacanthus arenatus (Actinopterygii: Priacanthidae); marine; mesentery, serosa of intestine and ovary; metacestode; WTSA; Brazil (Kuraiem et al. 2016).

Scomberomorus cavalla (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Dias et al. 2011a).

Sphyrna zygaena (Elasmobranchii: Sphyrnidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002; Pinto et al. 2006).

Stephanolepis hispidus (Actinopterygii: Monacanthidae); marine; mesentery; metacestode; WTSA; Brazil (Palm 2004).

***Callitetrarhynchus* sp.**

Balistes capriscus (Actinopterygii: Balistidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2005).

Balistes vetula (Actinopterygii: Balistidae); marine; body cavity, mesentery, muscles; metacestode; WTSA; Brazil (Alves et al. 2005).

Conodon nobilis (Actinopterygii: Haemulidae); marine; body cavity; metacestode; WTSA; Brazil (Paschoal et al. 2015).

***Dasyrhynchus giganteus* (Diesing, 1850) Pintner, 1929**

[Syns. *Anthocephalus giganteus* Diesing, 1850; *Rhynchobothrium insigne* Linton, 1924; *Sbesterium insigne* Dollfus, 1929]

Caranx hippos (Actinopterygii: Carangidae); marine; brain; metacestode; NBS, TSA; Brazil (São Clemente et al. 1993; Palm 1997).

Oligoplites saliens (Actinopterygii: Carangidae); marine; subcutaneous; metacestode; NBS (estuary of Tapajós River); Brazil (Diesing 1850, 1856).
Notes: type host; it was originally reported as *Chorinemus saliens*. Beveridge and Campbell (1993) redescribed this species.

***Dasyrhynchus pacificus* Robinson, 1965**

Carcharhinus brachyurus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (São Clemente and Gomes 1989a; São Clemente et al. 1991b).

Carcharhinus limbatus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (Beveridge and Campbell 1993; Palm 2004).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity, haemal arches, mesentery, kidney; metacestode; WTSA; Argentina, Brazil, Uruguay (Pereira and Boeger 2005; Timi et al. 2005, 2010b).

Cynoscion jamaicensis (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; body cavity, pericardium, kidney; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; body cavity, mesentery, kidney; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Sciaena deliciosa (Actinopterygii: Sciaenidae); marine; peritoneum; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Scyliorhinus haeckelii (Elasmobranchii: Scyliorhinidae); marine; spiral valve; adult; WTSA; Brazil (Beveridge and Campbell 1993; Palm 2004).

Sphyrna sp. (Elasmobranchii: Sphyrnidae); marine; spiral valve; adult; WTSA; Brazil (Beveridge and Campbell 1993; Palm 2004).

Floriceps saccatus* Cuvier, 1817

[Syns. *Anthocephalus elongatus* Rudolphi, 1819; *Rhynchobothrium ingens* Linton, 1921; *R. carangis* MacCallum, 1921; *Floriceps caballeroi* Ceuz-Reyes, 1977]

Aluterus monoceros (Actinopterygii: Monacanthidae); marine; liver, mesentery; metacestode; WTSA; Brazil (Dias et al. 2010).

Centropomus nigrescens (Actinopterygii: Centropomidae); amphidromous; peritoneum; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; muscles; metacestode; WTSA; Brazil (Silva and São Clemente 2001).

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002; Pinto et al. 2006).

Seriola lalandi (Actinopterygii: Carangidae); marine; muscle; metacestode; WTSP; Chile (Soto and Carvajal 1979).
Note: host reported as *S. mazatlana*.

***Grillotia* (*Christianella*) *carvajalregorum* Menoret & Ivanov, 2009**

[Syns. *Progrillotia dollfusi* Carvajal & Rego 1983; *Grillotia* (*Progrillotia*) *dollfusi* (Carvajal & Rego, 1983) Palm, 2004; *G. carvajalregorum* Menoret & Ivanov, 2009]

Acanthistius brasilianus (Actinopterygii: Serranidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2009b).

Carcharhinus signatus (Elasmobranchii: Carcharhinidae); marine; stomach; adult; WTSA; Brazil (Knoff et al. 2002, 2004c).

Conger orbignianus (Actinopterygii: Congridae); marine; intestinal surface, mesentery; metacestode; WTSA; Argentina (Timi and Lanfranchi 2013).

Ctenosciaena gracilicirrhus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity, mesentery; metacestode; WTSA; Argentina, Brazil (Sabas and Luque 2003; Luque and Poulin 2004; Pereira and Boeger 2005; Timi et al. 2005, 2010b; Menoret and Ivanov 2009b).

Cynoscion jamaicensis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Cynoscion striatus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Rego et al. 1974; Carvajal and Rego 1983).

Notes: type host. Carvaval and Rego (1983) re-identified the larvae collected by Rego et al. (1974).

Dules auriga (Actinopterygii: Serranidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2012; Braicovich and Timi 2015).

Note: Menoret and Ivanov (2012) reported the host as *Serranus auriga* (Cuvier).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2002a, b; Luque and Poulin 2004; São Clemente et al. 2004).

Note: after morphological re-evalution of specimens deposited by Alves et al. (2002b), São Clemente et al. (2004) assigned this species to *P. dollfusi* (syn. of *G.* (*C.*) *carvajalregorum*).

Heptranchias perlo (Elasmobranchii: Hexanchidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002, 2004c).

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; body cavity; metacestode; WTSA; Brazil (São Clemente et al. 2007).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; mesentery, body cavity; metacestode; WTSA; Brazil (Sabas and Luque 2003; Luque and Poulin 2004; Pereira and Boeger 2005).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Menticirrhus littoralis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Pereira and Boeger 2005; Menoret and Ivanov 2009b).

Nemadactylus bergi (Actinopterygii: Cheilodactylidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2009b; Rossin and Timi 2010).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; body cavity, liver, mesentery, muscles; metacestode; WTSA; Argentina, Brazil (Felizardo et al. 2010; Alarcos and Timi 2012; Alarcos et al. 2016).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; body cavity, mesentery; metacestode; WTSA; Argentina, Brazil (Alarcos and Timi 2012; Fonseca et al. 2012).

Paralonchurus brasiliensis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Parona signata (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2012).

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2009b; Braicovich and Timi 2010).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2012).

Porichthys porosissimus (Actinopterygii: Batrachoididae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2012).

Prionotus nudigula (Actinopterygii: Triglidae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Prionotus punctatus (Actinopterygii: Triglidae); marine; intestinal surface, mesentery; metacestode; WTSA; Argentina, Brazil (Bicudo et al. 2005; Menoret and Ivanov 2009b).

Pseudopercis numida (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; WTSA; Brazil (Luque et al. 2008).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; WTSA; Brazil (Luque et al. 2008).

Raneya brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; Magellanic, WTSA; Argentina (Vales et al. 2011).

Squalus sp. (Elasmobranchii: Squalidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002, 2004c).

Squatina guggenheim (Elasmobranchii: Squatinidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2009b).

Note: Menoret and Ivanov (2009b) provided a new name for *P. dollfusi* and a detailed description of adult forms.

Trachurus lathami (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Menoret and Ivanov 2009b; Braicovich et al. 2012).

Umbrina canosai (Actinopterygii: Sciaenidae); marine; body cavity, mesentery; metacestode; WTSA; Argentina, Brazil (Pereira and Boeger 2005; Menoret and Ivanov 2009b).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2009b; Pereira et al. 2014).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; body cavity, mesentery; metacestode; WTSA; Argentina, Brazil (Menoret and Ivanov 2009b; Alarcos and Timi 2012, 2013; Fonseca et al. 2012).

Grillotia* (*Christianella*) *minuta* (van Beneden, 1849) Guiart, 1931

[Syns. *Tetrarhynchus minutus* van Beneden, 1849; *Grillotia minuta* (van Beneden, 1849); *T. smarisgora* Wagener, 1854; *T. smarismaenae* Wagener, 1854; *T. smaridum* Pintner, 1893; *G. angeli* Dollfus, 1969; *G. bothridiopunctata* Dollfus, 1969]

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Argentina, Uruguay (Timi et al. 2005).

Notes: tapeworms reported as *G. bothridiopunctata*. *G.* (*C.*) *minuta* is distributed across the Northeastern Atlantic and morphologically similar to *G.* (*C.*) *carvajalregorum*; therefore, its occurrence in the Southwestern Atlantic is doubtful (Menoret and Ivanov 2012).

***Grillotia* (*Grillotia*) *borealis* Keeney & Campbell, 2001**

Macrourus carinatus (Actinopterygii: Macrouridae); marine; mesentery; metacestode; Magellanic; Argentina (Palm 2004).

Note: this species was described from fishes in the North Pacific Ocean and it is morphologically similar to *G. patagonica* Menoret and Ivanov, 2012; thus, its occurrence in the Southwestern Atlantic is doubtful (Menoret and Ivanov 2012).

Salilota australis (Actinopterygii: Moridae); marine; mesentery; metacestode; Magellanic; Argentina (Palm 2004).

***Grillotia* (*Grillotia*) *dollfusi* Carvajal, 1971**

Dipturus flavirostris (Elasmobranchii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Leible et al. 1990).

Note: host reported as *Raja flavirostris*.

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; body cavity; metacestode; WTSP; Chile (Oliva 2001).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; body cavity, intestinal serosa, gonads, liver, stomach wall; metacestode; WTSP; Chile (Carvajal and Cattan 1978; Carvajal et al. 1979; George-Nascimento 1996; Oliva and Ballón 2002).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; WTSP; Peru (Chero et al. 2014a).

Zearaja chilensis (Elasmobrachii: Rajidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1971; Whittaker et al. 1982).

Notes: type host; it was reported as *Raja chilensis*. Beveridge and Campbell (2007) redescribed *D. dollfusi* based on the type-specimens.

***Grillotia* (*Grillotia*) *erinaceus* (van Beneden, 1858) Guiart, 1927**

[Syns. *Tetrarhynchus erinaceus* van Beneden, 1858; *Rhynchobothrium imparispine* Linton, 1897; *Grillotia pseuderinaceus* Dollfus, 1969; *G. recurvispinis* Dollfus, 1969]

Conger orbignianus (Actinopterygii: Congridae); marine; mesentery; metacestode; WTSA; Argentina (Tanzola and Guagliardo 2000).

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; mesentery, stomach, intestine wall; metacestode; Magellanic; Falkland Islands (Brickle et al. 2006; Brown et al. 2013).

Eleginops maclovinus (Actinopterygii: Eleginopsidae); marine; mesentery; metacestode; Magellanic; Falkland Islands (Brickle and MacKenzie 2007).

Porichthys porosissimus (Actinopterygii: Batrachoididae); marine; mesentery; metacestode; WTSA; Argentina (Tanzola et al. 1997).

Note: Menoret and Ivanov (2012) dissected 7 specimens of this fish host, also from the Argentine Sea, and they only found *G.* (*C.*) *carvajalregorum*; therefore, they questioned the reliability of this report.

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Tanzola et al. 1998).

Note: Menoret and Ivanov (2012) dissected 32 specimens of this fish host, also from the Argentine Sea, and they did not find any *G. erinaceus*; therefore, they questioned the reliability of this report.

***Grillotia* (*Grillotia*) *patagonica* Menoret & Ivanov, 2012**

Cottoperca gobio (Actinopterygii: Bovichtidae); marine; body cavity; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Nemadactylus bergi (Actinopterygii: Cheilodactylidae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Patagonotothen brevicauda brevicauda (Actinopterygii: Nototheniidae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Patagonotothen ramsayi (Actinopterygii: Nototheniidae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

Psammobatis rudis (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; Magellanic; Argentina (Menoret and Ivanov 2012).

Note: type host.

Salilota australis (Actinopterygii: Moridae); marine; mesentery; metacestode; Magellanic; Argentina (Menoret and Ivanov 2012).

***Grillotia heptanchi* (Vaullegeard, 1899) Dollfus, 1942** (*Grillotia* sensu lato)

[For synonyms, see Beveridge and Campbell (2013)]

Genypterus chilensis (Actinopterygii: Ophidiidae); marine; muscles; metacestode; Magellanic; Chile (Carvajal and Campbell 1979).

Hexanchus griseus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1971, 1974).

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; body cavity, viscera; metacestode; Magellanic; Chile (Carvajal and Campbell 1979; Torres et al. 1993).

Merluccius australis (Actinopterygii: Merlucciidae); marine; mesentery, muscles; metacestode; Magellanic; Chile (Carvajal and Campbell 1979; Fernández 1985; George-Nascimento and Arancibia 1994; González and Carvajal 1994; Chávez et al. 2012).

Note: Carvajal and Campbell (1979) reported the host as *M. polylepis*.

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; muscles; metacestode; WTSP; Chile (Tagle 1951).

***Grillotia* sp.**

Aphos porosus (Actinopterygii: Batrachoididae); marine; body cavity; metacestode; WTSP; Chile (Cortés and Muñoz 2008, 2009).

Bathyraja magellanica (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; Magellanic; Argentina (Menoret and Ivanov 2012).

Note: the specimens are most likely *G.* (*G.*) *patagonica*, but the internal morphology could not be assessed in frozen material, which prevented the precise identification (Menoret and Ivanov 2012).

Coelorinchus chilensis (Actinopterygii: Macrouridae); marine; stomach; metacestode; JFD; Chile (Pardo-Gandarilhas et al. 2008).

Eleginops maclovinus (Actinopterygii: Eleginopsidae); marine; site of infection not given; metacestode; WTSP; Chile (Henríquez et al. 2011).

Lutjanus analis (Actinopterygii: Lutjanidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Note: Palm (2004) suggested the genus *Pseudolacistorhynchus* Palm, 1995 as the correct generic identification.

Merluccius australis (Actinopterygii: Merlucciidae); marine; body cavity, mesentery; metacestode; Magellanic; Falkland Islands (MacKenzie and Longshaw 1995).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; body cavity; metacestode; Magellanic, WTSA; Argentina, Falkland Islands, Uruguay (MacKenzie and Longshaw 1995; Sardella and Timi 1996, 2004).

Micromesistius australis australis (Actinopterygii: Gadidae); marine; site of infection not given; metacestode; Magellanic; Argentina, Chile, Falkland Islands (Niklitschek et al. 2010; George-Nascimento et al. 2011).

Paralabrax humeralis (Actinopterygii: Serranidae); marine; stomach; metacestode; WTSP; Peru (Armas 1983).

Paralichthys orbignyanus (Actinopterygii: Paralichthyidae); marine; site of infection not given; metacestode; WTSA (Mar Chiquita coastal lagoon); Argentina (Alarcos and Etchegoin 2010).

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil, Uruguay (Luque and Poulin 2004; Braicovich and Timi 2008).

Pinguipes brasilianus (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; Magellanic, WTSA; Argentina, Brazil (Timi et al. 2008, 2009, 2010a).

Prionotus nudigula (Actinopterygii: Triglidae); marine; mesentery; metacestode; WTSA; Argentina (Timi and Lanfranchi 2009b).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; Magellanic, WTSA; Argentina (Timi and Lanfranchi 2009a).

Squatina armata (Elasmobranchii: Squatinidae); marine; spiral valve; adult; WTSP; Peru (Tresierra et al. 1986).

***Lacistorhynchus dollfusi* Beveridge & Sakanari, 1987**

[Syn. *Lacistorhynchus tenuis* (van Beneden, 1858) (*pro parte*)]

Cheilotrema fasciatum (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSP; Peru (Oliva and Luque 1998).
Note: host reported as *Sciaena fasciata*.

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; body cavity, muscles; metacestode; WTSP; Chile (Oliva et al. 1996).

Lacistorhynchus tenuis* (van Beneden, 1858) Pintner, 1913

[Syns. *Tetrarhynchus tenuis* van Beneden, 1858; *Rhynchobothrium heterospine* Linton, 1897; *Rhynchobothrium bulbifer* Linton, 1897]

Anisotremus scapularis (Actinopterygii: Haemulidae); marine; mesentery, muscles; metacestode; WTSP; Peru (Luque 1991).
Note: Beveridge and Sakanari (1987) suggested that this apparently cosmopolitan taxon might represent a composite of two or more species and its distribution in the Pacific waters needs confirmation. Therefore, a taxonomic reassessment of *L. tenuis* is pending.

Cheilodactylus variegatus (Actinopterygii: Cheilodactylidae); marine; mesentery, muscles; metacestode; WTSP; Peru (Luque 1991).

Cheilotrema fasciatum (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSP; Peru (Luque 1991)
Note: host reported as *Sciaena fasciata*.

Cilus gilberti (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSP; Chile (García et al. 2001).

Labrisomus philippii (Actinopterygii: Labrisomidae); marine; mesentery, muscles; metacestode; WTSP; Peru (Rivera and Sarmiento 1990; Oliva and Luque 2002; Cruces et al. 2015).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; WTSP; Peru (Durán and Oliva 1980; Jara 1998).

Mugil cephalus (Actinopterygii: Mugilidae); marine; mesentery, muscles; metacestode; WTSP; Peru (Luque 1991).

Odontesthes regia (Actinopterygii: Atherinopsidae); marine; muscles; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; body cavity; metacestode; WTSA; Argentina (Szidat 1961).

Triakis maculata (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).

***Lacistorhynchus* sp.**

Merluccius australis (Actinopterygii: Merlucciidae); marine; muscles; metacestode; Magellanic; Chile, Falkland Island (MacKenzie and Longshaw 1995).

Scartichthys viridis (Actinopterygii: Bleniidae); marine; site of infection not given; metacestode; WTSP; Chile (Flores and George-Nascimento 2009).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; surface of pyloric caeca; metacestode; WTSA; Argentina (Szidat 1961).

Urophycis mystaceus (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2002c; Luque and Poulin 2004).

***Paragrillotia* sp.**

Dipturus trachyderma (Elasmobranchii: Rajidae); marine; site of infection and stage of development not given; WTSP; Chile (Leible et al. 1990).
Notes: host reported as *Raja trachyderma*. Neither were the three known species of *Paragrillotia* Dollfus, 1969 described from rays nor reported from Southeastern Pacific (see Beveridge and Justine 2007a). Since the vouchers were apparently not deposited, we considered this record doubtful.

***Pseudogrillotia peruviana* Escalante & Carvajal, 1984**

Scomberomorus sierra (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSP; Peru (Escalante and Carvajal 1984).
Note: type host; it was reported as *S. maculatus*.

***Pseudogrillotia* sp.**

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; mesentery; metacestode; WTSA; Brazil (Palm 2004).

Pseudolacistorhynchus noodti* Palm, 1995

[Syn. *Rhynchobothrium* sp. of Linton 1909, 1924]

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: type host.

Scomberomorus brasiliensis (Actinopterygii: Scombridae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: host reported as *S. maculatus*.

Family Pterobothriidae Pintner, 1931

***Pterobothrium acanthotruncatum* Escalante & Carvajal, 1984**

[Syns. *Synbothrium hemuloni* MacCallum, 1921; *Gymnorhynchus gigas* (Cuvier, 1817) sensu Southwell (1929); *Pterobothrium heteracanthum* Diesing, 1850 sensu Palm (1995)]

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; gallbladder, mesentery; metacestode; WTSP; Peru (Escalante and Carvajal 1984).
Note: type host.

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Paralonchurus peruanus (Actinopterygii: Sciaenidae); marine; gallbladder, gonads, mesentery, peritoneum; metacestode; WTSP; Peru (Tantaleán and Huiza 1994).

***Pterobothrium crassicole* Diesing, 1850**

[Syns. *Synbothrium felis* of MacCallum – USNMHC 35978; *Synbothrium* sp. of MacCallum – USNMHC 35744]

Aspistor luniscutis (Actinopterygii: Ariidae); marine; body cavity; metacestode; WTSA; Brazil (Tavares and Luque 2008).

Bagre marinus (Actinopterygii: Ariidae); marine; body cavity; metacestode; NBS (estuary of Amazon River); Brazil (Rego 1987a).
Note: host reported as *Bagrus marinus*.

Brachyplatystoma rousseauxii (Actinopterygii: Pimelodidae); freshwater; body cavity; metacestode; Amazon River basin (estuary); Brazil (Rego 1987a).
Note: host reported as *B. flavicans*.

Brachyplatystoma vaillantii (Actinopterygii: Pimelodidae); freshwater; body cavity; metacestode; Amazon River basin (estuary); Brazil (Rego 1987a).

Citharichthys spilopterus (Actinopterygii: Paralichthyidae); marine; muscles; metacestode; TSA; Brazil (Palm 2004).

Cynoscion acoupa (Actinopterygii: Sciaenidae); marine; muscles; metacestode; NBS; Brazil (Dias et al. 2011b).

Cynoscion leiachus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Rego et al. 1974; Palm 2004).
Note: Rego et al. (1974) reported this species as *Pterobohrium* sp., but Pereira (1993) considered them to be *P. crassicole*.

Cynoscion sp. (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSA; Brazil (Ferreira et al. 2006).

Genidens barbus (Actinopterygii: Ariidae); marine; muscles; metacestode; WTSA; Brazil (São Clemente et al. 1991a).
Note: host reported as *Netuma barba*.

Gobioides broussonnetii (Actinopterygii: Gobiidae); brackish; mesentery; metacestode; Amazon River basin (estuary, Marajó Island); Brazil (Videira et al. 2013).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity, liver, mesentery, peritoneum; metacestode; WTSA; Brazil (São Clemente 1986a, b,

1987; Pereira 1993; Campbell and Beveridge 1996; Pereira and Boeger 2005; Porto et al. 2009).

Micropogonias undulatus (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Palm 2004).
Note: The host is assigned to *Tachysurus* sp. in the CHIOC database.

Oligoplites palometa (Actinopterygii: Carangidae); marine; body cavity; metacestode; WTSA; Brazil (Takemoto et al. 1996a, b; Luque and Poulin 2004).
Note: Palm (2004) re-examined the specimens deposited in CHIOC and considered them as *Pterobothrium* sp.

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; stomach serosa; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; body cavity, kidney, liver, mesentery, muscles, stomach serosa; metacestode; WTSA; Brazil (Fonseca et al. 2012).

Pimelodus sp. (Actinopterygii: Pimelodidae); freshwater; mesentery; metacestode; Amazon River basin; Brazil (Diesing 1850, 1856).
Notes: type host. Diesing (1850) reported the specimens from *Erythrinus unitaeniatus* (syn. of *Hoplerythrinus unitaeniatus*), but it was re-identified by Diesing (1856). The Diesing's types deposited in NHMW consist only of cestode fragments (see the redescription of Campbell and Beveridge 1996).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; muscles; metacestode; Amazon River Basin (estuary); Brazil (Silva 2010).

Pogonias cromis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; mesentery; metacestode; WTSA; Brazil (São Clemente et al. 1997).

Scomberomorus cavalla (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Dias et al. 2011a).

Scorpaena sp. (Actinopterygii: Scorpaenidae); marine; mesentery; metacestode; WTSA; Brazil (Palm 2004).

***Pterobothrium heteracanthum* Diesing, 1850**

[Syns. *Syndesmobothrium filicolle* Linton, 1890; *Synbothrium fillicolle* Linton, 1897; *S. hemuloni* MacCallum, 1921; *Gymnorhynchus cymbiumi* Chincholikar & Shinde, 1977; *Neogymnorhynchus platycephali* Bilqees & Shah, 1982]

Cynoscion acoupa (Actinopterygii: Sciaenidae); marine; muscles; metacestode; NBS; Brazil (Dias et al. 2011b).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity, mesentery, peritoneum; metacestode; TSA, WTSA; Argentina, Brazil (São Clemente 1986a, b, 1987; Pereira 1993; Campbell and Beveridge 1996; Alves and Luque 2000, 2001a, b; Luque and Poulin 2004; Pereira and Boeger 2005;

Alarcos and Etchegoin 2010; Luque et al. 2010; Timi et al. 2010b; Mattos et al. 2015).

Note: Campbell and Beveridge (1996) studied specimens collected by São Clemente in 1980 that are deposited in CHIOC; they suggested that these worms are 'topotypes', i.e. specimens that do not belong to the type series, but were collected in the type locality (this term is not considered by the ICZN), even though there is no evidence that J. Natterer collected the worms in the Rio de Janeiro coast as stated by São Clemente.

Micropogonias undulatus (Actinopterygii: Sciaenidae); marine; gallbladder surface, intestine, mesentery; metacestode; NBS (Amazon River estuary), WTSA; Brazil, Uruguay (Diesing 1850, Palm 2004).

Notes: type host; it was reported as *M. lineatus*. Campbell and Beveridge (1996) also mentioned *M. furnieri* as type host, but Diesing (1850) reported the worms only from *M. lineatus*. The type material is no longer extant in NHMW (Campbell and Beveridge 1996).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; muscles; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; muscles; metacestode; Amazon River basin (estuary); Brazil (Silva 2010).

Pogonias cromis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Pomadasys crocro (Actinopterygii: Haemulidae); marine; muscles; metacestode; NBS (Amazon River estuary); Brazil (Diesing 1855).

Note: host reported as *Pristipoma coro*.

Umbrina canosai (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Clupeidae gen. sp. (Actinopterygii); marine; mesentery; metacestode; WTSA; Brazil (Palm 2004).

***Pterobothrium kingstoni* Campbell & Beveridge, 1996**

[Syn. *Synbothrium lintoni* MacCallum, 1921 (*pro parte*)]

Citharichthys spilopterus (Actinopterygii: Paralichthyidae); marine; body cavity, muscles; metacestode; TSA; Brazil (Palm 1997).

Haemulon aurolineatum (Actinopterygii: Haemulidae); marine; muscles; metacestode; TSA; Brazil (Campbell and Beveridge 1996).

Note: the worms were collected by Palm in Brazil's Northeastern coast.

***Pterobothrium* sp.**

Conodon nobilis (Actinopterygii: Haemulidae); marine; body cavity; metacestode; WTSA; Brazil (Paschoal et al. 2015).

Cynoscion acoupa (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Cynoscion striatus (Actinopterygii: Sciaenidae); marine; muscles; metacestode; WTSA; Brazil (Santos and Zogbi 1971).
Note: metacestode described as *Tetrarhynchus fragilis* (*nomen nudum*) and identified as *Pterobotrium* sp. by Palm (2004).

Epinephelus sp. (Actinopterygii: Serranidae); marine; body cavity; metacestode; TNA; Venezuela (Palm 2004).

Gymnura sp. (Elasmobranchii: Gymnuridae); marine; spiral valve; adult; WTSA; Brazil (Palm 2004).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; muscles; metacestode; WTSA; Brazil (Santos and Zogbi 1971).
Note: Santos and Zogbi (1971) reported the metacestode as *T. fragilis* (see Palm 2004).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).
Note: Palm (2004) studied specimens collected by Travassos in 1921 that are deposited in CHIOC.

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity, muscles; metacestode; WTSA; Brazil (Santos and Zogbi 1971; São Clemente 1986a, b, 1987).
Note: Santos and Zogbi (1971) reported the host as *Micropogon opercularis* and the metacestode as *T. fragilis* (see Palm 2004).

Mycteroperca bonaci (Actinopterygii: Serranidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Pagrus pagrus (Actinopterygii: Sparidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Paralichthys sp. (Actinopterygii: Paralichthyidae); marine; muscles; metacestode; WTSA; Brazil (Santos and Zogbi 1971).
Notes: Santos and Zogbi (1971) reported the metacestode as *T. fragilis* (see Palm 2004).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; intestine; metacestode; WTSA; Brazil (Palm 2004).
Note: host reported as *P. saltator*.

Prionotus sp. (Actinopterygii: Triglidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Umbrina canosai (Actinopterygii: Sciaenidae); marine; muscles; metacestode; WTSA; Brazil (Santos and Zogbi 1971).
Notes: Santos and Zogbi (1971) described the metacestodes as *T. fragilis* (see Palm 2004).

Siluriform fish (Actinopterygii); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Unidentified ray (Elasmobranchii); marine; spiral valve; adult; WTSA; Brazil (Rego et al. 1974).
Note: Rego et al. (1974) named the cestodes as Pterobothriidae gen. sp. and Palm (2004) assigned them to *Pterobothrium* sp.

Unidentified pterobothriids

Bagre bagre (Actinopterygii: Ariidae); marine; site of infection not given; metacestode; NBS; Brazil (Vicente and Fernandes 1978).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; NBS; Brazil (Vicente and Fernandes 1978).

Superfamily Otobothrioidea Dollfus, 1942
Family Otobothriidae Dollfus, 1942

***Otobothrium* sp.**

Balistes vetula (Actinopterygii: Balistidae); marine; body cavity, muscles; metacestode; WTSA; Brazil (São Clemente et al. 1995; Alves et al. 2005).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; body cavity, liver, mesentery, intestine, stomach; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Poecilancistrium caryophyllum* (Diesing, 1850) Dollfus, 1929

[For synonyms, see Palm (2004)]

Carcharhinus leucas (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; NBS (estuarine waters); Brazil (Diesing 1850, 1856).
Note: host reported as *Prionodon leucas*.

Cilus gilberti (Actinopterygii: Sciaenidae); marine; peritoneum; metacestode; WTSP; Peru (Escalante and Carvajal 1984).
Note: host reported as *Sciaena gilberti*.

Cynoscion acoupa (Actinopterygii: Sciaenidae); marine; muscles; metacestode; NBS; Brazil (Dias et al. 2011b).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; muscles; metacestode; NBS; Brazil (Oliveira et al. 2009; Dias et al. 2011b).

Micropogonias altipinnis (Actinopterygii: Sciaenidae); marine; muscles; metacestode; TEP; Ecuador (Palm 2004).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity, muscles; metacestode; TNA, WTSA; Brazil, Venezuela (São Clemente 1986a, b, 1987; Vicente et al. 1989; Pereira 1993; Pereira and Boeger 2005).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; muscles; metacestode; Amazon River basin (estuary of Amazon River); Brazil (Silva 2010).

Rhizoprionodon lalandii (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; NBS (estuarine waters); Brazil (Diesing 1850, 1856).
Note: type host; it was reported as *Scoliodon lalandii*.

Family Pseudotobothriidae Palm, 1995

Pseudotobothrium dipsacum* (Linton, 1897) Dollfus 1942
[Syn. *Otobothrium dipsacum* Linton, 1897]

Haemulon plumierii (Actinopterygii: Haemulidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Hyporthodus niveatus (Actinopterygii: Serranidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).
Note: host reported as *Epinephelus niveatus*.

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Suborder Trypanobatoida Olson, Caira, Jensen, Overstreet, Palm & Beveridge, 2010
Superfamily Eutetrarhynchoidea Guiart, 1927
Family Eutetrarhynchidae Guiart, 1927

***Dollfusiella acuta* Menoret & Ivanov, 2015**

Atlantoraja castelnaui (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2015).

Atlantoraja platana (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; Magellanic; Argentina (Menoret and Ivanov 2015).

Sympterygia acuta (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; Magellanic, WTSA; Argentina (Menoret and Ivanov 2015).
Note: type host.

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2015).

***Dollfusiella musteli* (Carvajal, 1974) Beveridge, Neifar & Euzet, 2004**
[Syns. *Prochristianella musteli* Carvajal, 1974; *Eutetrarhynchus musteli* (Carvajal, 1974) Beveridge, 1990]

Mustelus mento (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSP; Chile (Carvajal 1974).
Note: type host.

***Dollfusiella taminii* Menoret & Ivanov, 2014**

Psammobatis bergi (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2014).
Note: type host.

***Dollfusiella vooremi* (São Clemente & Gomes, 1989) Beveridge, Neifar & Euzet, 2004**
[Syn. *Eutetrarhynchus vooremi* São Clemente & Gomes, 1989]

Mustelus canis (Elasmobranchii: Triakidae); marine; spiral valve; adult; WTSA; Brazil (São Clemente and Gomes 1989b; São Clemente et al. 1991b).
Note: type host.

Mustelus schmitti (Elasmobranchii: Triakidae); marine; spiral valve; adult; Magellanic, WTSA; Argentina, Brazil (São Clemente and Gomes 1989b; São Clemente et al. 1991b; Alarcos et al. 2006; Menoret and Ivanov 2014).
Note: tapeworms reported as *E. vooremi* by Alarcos et al. (2006).

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Tanzola et al. 1998).
Notes: Tanzola et al. (1998) reported the tapeworms as *E. vooremi*. Menoret and Ivanov (2014) examined 42 specimens of *S. bonapartii*, also from the Argentinian coast, and none *D. vooremi* was found in these sharks. Thus, they suggested that this report is most likely a result of misidentification.

***Dollfusiella* sp.**

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA, WTSP; Brazil, Chile (Oliva 1999; Pereira and Boeger 2005).
Note: Oliva (1999) reported tapeworms as *Eutetrarhynchus* sp., but most likely they belonged to the closely-related genus *Dollfusiella*.

***Mecistobothrium oblongum* Menoret & Ivanov, 2015**

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; Magellanic; Argentina (Menoret and Ivanov 2015).
Note: type host.

***Parachristianella damiani* Menoret & Ivanov, 2014**

Myliobatis goodei (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2014).
Note: type host.

***Parachristianella monomegacantha* Kruse, 1959**

Himantura schmardae (Elasmobranchii: Dasyatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: tapeworms reported as *P.* cf. *monomegacantha*.

Rhinobatos planiceps (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSP; Chile (Dailey and Carvajal 1976).

Paroncomegas araya* (Woodland, 1934) Campbell, Marques & Ivanov, 1999
[Syns. *Tentacularia araya* Woodland, 1934; *Eutetrarhynchus araya* (Woodland, 1934) Rego & Dias, 1976]

Potamotrygon falkneri (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Paraná River basin; Paraguay (Brooks et al. 1981b; Lacerda et al. 2008, 2009).

Potamotrygon cf. *falkneri* (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Peru (Reyda 2008).
Note: host reported as *Potamotrygon* cf. *castexi*.

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon and Paraná River basins; Argentina, Brazil (Rego and Dias 1976; Campbell et al. 1999; Reyda 2008).
Notes: Rego and Dias (1976) synonymized *E. baeri* Lopez-Neyra & Diaz-Ungria, 1958 with *E. araya*, but Campbell et al. (1999) recognized their morphological distinctness and named the former species as *Paroncomegas* sp. until further studies confirm its validity. Sequences of partial 18S (DQ642963) and 28S (DQ642801) (Olson et al. 2010).

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Orinoco River basin; Venezuela (Brooks et al. 1981b).
Note: host reported as *P. reticulatus*.

Potamotrygon sp. (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Amazon River basin; Brazil (Woodland 1934c).
Note: type host; it was described as *Trygon* sp.

***Paroncomegas* sp.**

[Syn. *Eutetrarhynchus baeri* Lopez-Neyra & Diaz-Ungria, 1958]

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; spiral valve; adult; Orinoco River basin; Venezuela (Lopez-Neyra and Diaz-Ungria, 1958).
Note: host reported as *P. hystrix*. Campbell et al. (1999) assigned these tapeworms to *Paroncomegas* sp. until new morphological data are available (see the note above). It is treated as *P. baeri* (Lopez-Neyra & Diaz-Ungria, 1958) in GCD (Caira et al. 2012).

***Prochristianella heteracantha* Dailey & Carvajal, 1976**

Rhinobatos planiceps (Elasmobranchii: Rhinobatidae); marine; spiral valve; adult; WTSP; Chile, Peru (Dailey and Carvajal 1976; Tantaleán and Rodríguez 1987; Iannacone et al. 2011).
Note: type host.

Family Rhinoptericolidae Carvajal & Campbell, 1975

Rhinoptericola megacantha* Carvajal & Campbell, 1975

Rhinoptera bonasus (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; TNA; Venezuela (Mayes and Brooks 1981).
Note: type host.

Rhinoptera brasiliensis (Elasmobranchii: Myliobatidae); marine; spiral valve; adult; WTSA; Brazil (Napoleão et al. 2015).

Superfamily Tentacularioidea Poche, 1926
Family Sphyriocephalidae Pintner, 1913

***Hepatoxylon megacephalum* (Rudolphi, 1819) Dollfus, 1942**
[Syns. *Tetrarhynchus megacephalus* Rudolphi, 1819; *Tetrarhynchus scyllium canicula* Wagener, 1854]

Trichomycterus punctulatus (Actinopterygii: Trichomycteridae); freshwater; site of infection not given; metacestode; Lima; Peru (Dollfus 1942).

Hepatoxylon trichiuri* (Holten, 1802) Bosc, 1811
[For synonyms, see Palm (2004)]

Brama australis (Actinopterygii: Bramidae); marine; gonads, mesentery, muscles; metacestode; WTSP; Chile (George-Nascimento and Ortiz 1982; George-Nascimento et al. 2002)
Note: host reported as *Lepidotus australis* by George-Nascimento and Ortiz (1982).

Brama japonica (Actinopterygii: Bramidae); marine; body cavity; metacestode; WTSP; Peru (Iannacone and Alvariño 2013).

Coelorinchus chilensis (Actinopterygii: Macrouridae); marine; intestine; metacestode; JFD; Chile (Pardo-Gandarilhas et al. 2008).

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; liver, intestinal serosa, stomach wall; metacestode; WTSA; Brazil (Rudolphi 1819; São Clemente et al. 2001).

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; body cavity, mesentery, stomach wall; metacestode; Magellanic, WTSP; Chile, Falkland Islands (Rodríguez and George-Nascimento 1996; Brickle et al. 2006; Oliva et al. 2008a).

Genypterus blacodes (Actinopterygii: Ophidiidae); marine; stomach; metacestode; WTSP; Chile (Cattan 1977; Riffo 1994).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery, muscles; metacestode; WTSA; Brazil (São Clemente et al. 2004).

Genypterus chilensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSP; Chile (Vergara and George-Nascimento 1982).

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; gonads, mesentery, muscles; metacestode; WTSP; Chile (George-Nascimento and Ortiz 1982; George-Nascimento and Huet 1984; Muñoz and George-Nascimento 2008).

Isacia conceptionis (Actinopterygii: Haemulidae); marine; intestinal surface; metacestode; WTSP; Chile (Dollfus 1930).

Lampris guttatus (Actinopterygii: Lampridae); marine; gonads, mesentery, muscles; metacestode; WTSP; Chile (George-Nascimento and Ortiz 1982).
Note: host reported as *Lampris regia*.

Macruronus magellanicus (Actinopterygii: Merlucciidae); marine; body cavity, gonads, mesentery, muscles; Magellanic; Chile, Falkland Islands (George-Nascimento and Ortiz 1982; Oliva 2001; Chávez et al. 2012; MacKenzie et al. 2013).

Merluccius australis (Actinopterygii: Merlucciidae); marine; body cavity, muscles; metacestode; Magellanic, WTSP; Argentina, Chile, Falkland Islands (Fernández 1985; George-Nascimento and Arancibia 1994; González and Carvajal 1994; MacKenzie and Longshaw 1995; Chávez et al. 2012; Torres et al. 2014).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; body cavity, mesentery; metacestode; Magellanic, WTSP; Chile (Tagle 1951; George-Nascimento 1996; Oliva and Ballón 2002; Chávez et al. 2012).
Note: host reported as *Merluccius gayi* by some authors.

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; body cavity, mesentery; metacestode; Magellanic, WTSA; Argentina, Falkland Islands, Uruguay (Szidat 1955, 1961; MacKenzie and Longshaw 1995; Sardella and Timi 1996, 2004).

Micromesistius australis australis (Actinopterygii: Gadidae); marine; site of infection not given; metacestode; Magellanic; Chile, Falkland Islands (George-Nascimento et al. 2011; Chávez et al. 2012).

Notacanthus sexspinis (Actinopterygii: Notacanthidae); marine; intestine; metacestode; JFD; Chile (Pardo-Gandarilhas et al. 2008).

Oncorhynchus tshawytscha (Actinopterygii: Salmonidae); anadromous; body cavity; metacestode; Magellanic (Curaco de Vélez); Chile (Reyes-Piraino 1982).

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; body cavity, stomach serosa, liver; metacestode; JFD, WTSA, WTSP; Brazil, Chile, Peru (Yáñez 1950; Carvajal 1974; Cattan et al. 1979; Escalante 1986; São Clemente et al. 2001; Knoff et al. 2002, 2004b; Cousin et al. 2003).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; Magellanic; Argentina (Timi and Lanfranchi 2009a).

Salilota australis (Actinopterygii: Moridae); marine; body cavity; metacestode; Magellanic; Argentina (Szidat 1961).

Scomber japonicus (Actinopterygii: Scombridae); marine; site of infection not given; metacestode; WTSP; Chile (Rodríguez et al. 2000).

Sebastes capensis (Actinopterygii: Sebastidae); marine; site of infection not given; metacestode; WTSP; Chile (González and Poulin 2005a, b; González et al. 2006).

Somniosus pacificus (Elasmobranchii: Somniosidae); marine; intestinal surface; metacestode; WTSP; Chile (Reyes-Piraino 1982).

Trachurus murphyi (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento 2000; Rodríguez et al. 2000; Oliva 1999).

***Hepatoxylon* sp.**

Helicolenus lengerichi (Actinopterygii: Sebastidae); marine; mesentery; metacestode; WTSP; Chile (George-Nascimento and Iriarte 1989; Balboa and George-Nascimento 1998).

Macrourus holotrachys (Actinopterygii: Macrouridae); marine; visceral cavity; metacestode; WTSP: Chile (Ñacari and Oliva 2016).

Micromesistius australis australis (Actinopterygii: Gadidae); marine; site of infection not given; metacestode; Magellanic; Argentina, Chile (Niklitschek et al. 2010).

Trachurus murphyi (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Oliva 2015).

***Heterosphyriocephalus tergestinus* (Pintner, 1913) Dallarés, Carrassón & Schaeffner, 2016**

[Syn. *Sphyriocephalus tergestinus* Pintner, 1913]

Sarda chilensis (Actinopterygii: Scombridae); marine; stomach; metacestode; WTSP; Peru (Chero et al. 2015).

Family Tentaculariidae Poche, 1926

***Heteronybelinia annakohnae* Pereira & Boeger, 2005**

Ctenosciaena gracilicirrhus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Notes: type host. Pereira (1998) described *Nybelinia annakohnae* in his thesis (the type-material was deposited in CHIOC and USNPC), but it does not represent a formal publication (ICZN 1999). Six years later, Pereira and Boeger (2005) properly described the species, but transferred it to *Heteronybelinia*.

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Cynoscion jamaicensis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Heteronybelinia estigmena* (Dollfus, 1960) Palm, 1999

[For synonyms, see Palm (2004)]

Cynoscion jamaicensis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Haemulon plumierii (Actinopterygii: Haemulidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Note: tapeworms reported as *Nybelinia senegalensis* Dollfus, 1960.

Sphyraena guachancho (Actinopterygii: Sphyraenidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

***Heteronybelinia mattisi* Menoret & Ivanov, 2013**

Nemadactylus bergi (Actinopterygii: Cheilodactylidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2013).

Raneya brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSA; Argentina (Menoret and Ivanov 2013).

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Menoret and Ivanov 2013).
Notes: type host. Menoret and Ivanov (2013) also found larval forms in the pyloric caeca of *S. bonapartii*.

***Heteronybelinia nipponica* (Yamaguti, 1952) Palm, 1999**
[Syns. *Nybelinia nipponica* Yamaguti, 1952; *N. rougetcampanae* Dollfus, 1960; *H. rougetcampanae* (Dollfus, 1960) Palm, 1999]

Carcharhinus signatus (Elasmobranchii: Carcharhinidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002, 2004a).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; body cavity; metacestode; WTSA; Brazil (São Clemente et al. 2004).

Menticirrhus americanus (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Mullus argentinae (Actinopterygii: Mullidae); marine; body cavity; metacestode; WTSA; Brazil (Luque et al. 2002).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; body cavity, intestine, kidney, muscles; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; stomach; metacestode; WTSA; Brazil (Fonseca et al. 2012).

Sphyrna lewini (Elasmobranchii: Sphyrnidae); marine; spiral valve; adult; WTSA; Brazil (São Clemente et al. 1991b; São Clemente and Gomes 1992).
Note: tapeworms reported as *N.* (*Syngenes*) *rougetcampanae*.

Sphyrna zygaena (Elasmobranchii: Sphyrnidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002; Gomes et al. 2005).

Umbrina canosai (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; mesentery, stomach; metacestode; WTSA; Brazil (Fonseca et al. 2012).

***Heteronybelinia overstreeti* Palm, 2004**

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: Palm (1997) reported plerocercoids of *Nybelinia* cf. *lingualis* Cuvier, 1817 from this host, but later (Palm 2004) re-assigned these larvae to *H. overstreeti*.

***Heteronybelinia perideraeus* (Shipley & Hornell, 1906) Palm, 1999**
[Syns. *Tetrarhynchus perideraeus* Shipley & Hornell, 1906; *Stenobothrium perideraeum* (Shipley & Hornell, 1906) Pintner, 1913; *Nybelinia dakari* Dollfus, 1960]

Notorynchus cepedianus (Elasmobranchii: Hexanchidae); marine; spiral valve; adult; WTSA; Brazil (São Clemente et al. 1991b; São Clemente and Gomes 1992).

Note: host reported as *N. pectorosus*. São Clemente and Gomes (1992) recorded the tapeworms as *Nybelinia* (*Nybelinia*) *bisulcata* (Linton, 1889), but Palm (2004) re-assigned the material deposited in CHIOC to *H. perideraeus*.

***Heteronybelinia yamagutii* (Dollfus, 1960) Palm, 1999**

[Syn. *Nybelinia yamagutii* Dollfus, 1960]

Carcharhinus signatus (Elasmobranchii: Carcharhinidae); marine; spiral valve; metacestode; WTSA; Brazil (Knoff et al. 2002, 2004a).

***Heteronybelinia* sp.**

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Mixonybelinia beveridgei* (Palm, Walter, Schwerdtfeger & Reimer, 1997) Palm, 1999

[Syn. *Nybelinia beveridgei* Palm, Walter, Schwerdtfeger & Reimer, 1997]

Dipturus trachyderma (Elasmobranchii: Rajidae); marine; stomach; metacestode; WTSA; Brazil (Knoff et al. 2002, 2004a).

Note: host reported as *D. trachydermus*.

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; liver, mesentery, serosa of stomach; metacestode; WTSA; Brazil (São Clemente et al. 2004).

***Mixonybelinia edwinlintoni* (Dollfus, 1960) Palm & Walter, 2000**

[Syn. *Nybelinia edwinlintoni* Dollfus, 1960]

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

***Mixonybelinia* sp.**

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; muscles; metacestode; WTSA; Brazil (São Clemente et al. 2007).

***Nybelinia africana* Dollfus, 1960**

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 2004).

***Nybelinia bisulcata* (Linton, 1889) Dollfus, 1929**

[Syns. *Rhynchobothrium bisulcatum* Linton, 1889; *Tetrarhynchus bisulcatus* (Linton, 1889) Linton, 1890]

Umbrina canosai (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Note: the taxonomic status of this species is problematic since the type material is a mixture of different species under the same name (Palm 2004).

***Nybelinia erythraea* Dollfus, 1960**

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; stomach; metacestode; WTSA; Brazil (Fonseca et al. 2012).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; stomach; metacestode; WTSA; Brazil (Fonseca et al. 2012).

***Nybelinia fayapaulazariahi* Reimer, 1980**

Rhizoprionodon terraenovae (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (Palm 2004).

***Nybelinia indica* Chandra, 1986**

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Nybelinia lingualis* (Cuvier, 1817) Dollfus, 1929

[For synonyms, see Palm (2004)]

Cynoscion sp. (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Ortubay 1944).

Haemulon plumierii (Actinopterygii: Haemulidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: tapeworms reported as *N*. cf. *lingualis*.

Isurus oxyrinchus (Elasmobranchii: Lamnidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002; Gomes et al. 2005).

Mustelus canis (Elasmobranchii: Carcharhinidae); marine; spiral valve; metacestode; WTSA; Brazil (São Clemente and Gomes 1989b; São Clemente et al. 1991b).

Mustelus schmitti (Elasmobranchii: Carcharhinidae); marine; spiral valve; metacestode; WTSA; Brazil (São Clemente and Gomes 1989b; São Clemente et al. 1991b).
Note: tapeworms reported as *N*. (*N*.) *lingualis*.

Oncopterus darwinii (Actinopterygii: Pleuronectidae); marine; intestinal surface; metacestode; WTSA; Argentina (Szidat 1961).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; stomach, intestine, mesentery, spleen serosa, muscles; metacestode; WTSA; Brazil (Felizardo et al. 2010).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; body cavity, mesentery, stomach; metacestode; WTSA; Argentina, Brazil (Szidat 1961; Fonseca et al. 2012).

Porichthys porosissimus (Actinopterygii: Batrachoididae); marine; body cavity; metacestode; WTSA; Argentina (Tanzola et al. 1997).

Pseudupeneus maculatus (Actinopterygii: Mullidae); marine; body cavity; metacestode; TSA; Brazil (Palm 2004).

Selene vomer (Actinopterygii: Carangidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).
Note: tapeworms reported as *N*. cf. *lingualis*.

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; metacestode; WTSA; Argentina (Tanzola et al. 1998).

Trachurus murphyi (Actinopterygii: Carangidae); marine; wall of pharynx; metacestode; WTSP; Chile (Palm 2004).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; body cavity, mesentery, stomach; metacestode; WTSA; Brazil (Fonseca et al. 2012).

***Nybelinia surmenicola* Okada in Dollfus, 1929**

Hippoglossina macrops (Actinopterygii: Paralichthyidae); marine; intestine; metacestode; WTSP; Chile (González et al. 2001, 2008; Oliva et al. 2004).

Merluccius gayi gayi (Actinopterygii: Merlucciidae); marine; body cavity, mesentery; metacestode; WTSP; Chile (Oliva and Ballón 2002).

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; body cavity, gill arches, muscles; metacestode; WTSP; Chile (Oliva et al. 1996).

Trachurus murphyi (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSP; Chile, Peru (Oliva 1999).

***Nybelinia* sp.**

Aphos porosus (Actinopterygii: Batrachoididae); marine; body cavity; metacestode; WTSP; Chile (Cortés and Muñoz 2008, 2009).

Balistes capriscus (Actinopterygii: Balistidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves et al. 2005).

Brama australis (Actinopterygii: Bramidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento et al. 2002).

Brama japonica (Actinopterygii: Bramidae); marine; body cavity; metacestode; WTSP; Peru (Iannacone and Alvariño 2013).

Caranx hippos (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Alves 2001; Luque and Poulin 2004).

Caranx latus (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Alves 2001; Luque and Poulin 2004).

Cilus gilberti (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSP; Chile (García et al. 2001).

Conger orbignianus (Actinopterygii: Congridae); marine; intestinal surface, mesentery; metacestode; WTSA; Argentina (Timi and Lanfranchi 2013).

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; body cavity, stomach; metacestode; WTSP; Peru (López de McDonald and Tantaleán 1985; Vásquez-Ruiz and Jara-Campos 2012).

Cynoscion analis (Actinopterygii: Sciaenidae); marine; intestine, surface of stomach; metacestode; WTSP; Peru (Llerena et al. 2001).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; body cavity, mesentery; metacestode; WTSA; Brazil (Sabas and Luque 2003; Luque and Poulin 2004; Timi et al. 2005).

Dactylopterus volitans (Actinopterygii: Dactylopteridae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Cordeiro and Luque 2005).

Diapterus rhombeus (Actinopterygii: Gerreidae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; body cavity, mesentery, intestinal serosa; metacestode; WTSA; Brazil (Alves et al. 2002a, b; Luque and Poulin 2004; São Clemente et al. 2004).

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Huet 1984; Muñoz and George-Nascimento 2008).

Hippoglossina macrops (Actinopterygii: Paralichthyidae); marine; gill arches; metacestode; WTSP; Chile (Riffo 1991).

Isacia conceptionis (Actinopterygii: Haemulidae); marine; mesentery; metacestode; WTSP; Peru (Iannacone et al. 2015).

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; mesentery, body cavity, muscles; metacestode; WTSA; Brazil (São Clemente et al. 2007).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Sabas and Luque 2003; Luque and Poulin 2004).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; intestine; metacestode; WTSP; Peru (Durán and Oliva 1980).

Merluccius hubbsi (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; Magellanic; Argentina (Sardella and Timi 2004).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; TSA; Brazil (Luque et al. 2010).

Mola ramsayi (Actinopterygii: Molidae); marine; intestine; metacestode; WTSP; Chile (Villalba and Fernández 1985).
Note: the authors distinguished two different morphotypes.

Mullus argentinae (Actinopterygii: Mullidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Luque et al. 2002; Luque and Poulin 2004; Lanfranchi et al. 2009).

Odontesthes regia (Actinopterygii: Atherinopsidae); marine; site of infection not given; metacestode; WTSP; Chile (Torres et al. 1993).
Note: host reported as *Austromenidia laticlava*.

Paralabrax humeralis (Actinopterygii: Serranidae); marine; site of infection not given; metacestode; WTSP; Chile (Henríquez and González 2014).

Paralichthys adspersus (Actinopterygii: Paralichthyidae); marine; intestine; metacestode; WTSP; Chile (Riffo 1995).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; intestine, stomach; metacestode; WTSA; Argentina, Brazil (Luque and Poulin 2004; Alarcos and Timi 2012).

Paralichthys microps (Actinopterygii: Paralichthyidae); marine; intestine; metacestode; WTSP; Chile (Riffo 1995).

Paralichthys patagonicus (Actinopterygii: Paralichthyidae); marine; intestine, stomach; metacestode; WTSA; Argentina (Alarcos and Timi 2012).

Paralonchurus brasiliensis (Actinopterygii: Sciaenidae); marine; mesentery; metacestode; WTSA; Brazil (Ribeiro et al. 2002; Luque et al. 2003; Luque and Poulin 2004).

Percophis brasiliensis (Actinopterygii: Percophidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil, Uruguay (Luque and Poulin 2004; Braicovich and Timi 2008, 2010).

Note: Luque and Poulin (2004) distinguished two morphotypes.

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Chaves 1999; Luque and Poulin 2004).

Notes: host reported as *P. saltator*. Luque and Poulin (2004) distinguished two different morphotypes.

Prionotus punctatus (Actinopterygii: Triglidae); marine; intestine; metacestode; WTSA; Brazil (Luque and Poulin 2004; Bicudo et al. 2005).

Note: Luque and Poulin (2004) distinguished two different morphotypes.

Pseudopercis numida (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; WTSA; Brazil (Luque et al. 2008).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; mesentery; metacestode; Magellanic, WTSA; Argentina (Timi and Lanfranchi 2009a).

Raneya brasiliensis (Actinopterygii: Ophidiidae); marine; site of infection not given; metacestode; Magellanic, WTSA; Argentina (Vales et al. 2011).

Sarda chilensis (Actinopterygii: Scombridae); marine; site of infection not given; metacestode; WTSP; Peru (Pérez et al. 1999).

Sardinella brasiliensis (Actinopterygii: Clupeidae); marine; body cavity; metacestode; WTSA; Brazil (Moreira et al. 2015).

Sciaena deliciosa (Actinopterygii: Scombridae); marine; body cavity, intestinal surface; metacestode; WTSP; Peru (Pérez et al. 1999; Llerena et al. 2001; Chero et al. 2014c).

Scomber japonicus (Actinopterygii: Scombridae); marine; site of infection not given; metacestode; WTSP; Chile (Rodríguez et al. 2000).

Scomberomorus brasiliensis (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Luque and Poulin 2004; Alves and Luque 2006).

Selene setapinnis (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Cordeiro and Luque 2004; Luque and Poulin 2004).

Seriolella porosa (Actinopterygii: Centrolophidae); marine; body cavity; metacestode; Magellanic; Argentina (Guagliardo et al. 2014).

Sympterygia bonapartii (Elasmobranchii: Arhynchobatidae); marine; spiral valve; adult; WTSA; Argentina (Ostrowski de Núñez 1971).

Note: host reported as *Psammobatis microps*.

Trachurus lathami (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSA; Brazil (Braicovich et al. 2012).

Trachurus murphyi (Actinopterygii: Carangidae); marine; body cavity, mesentery; metacestode; WTSP; Chile, Peru (Soto and Carvajal 1979; Oliva 1982, 1994, 1999; George-Nascimento and Arancibia 1992; Jara 1998; Pérez et al. 1999; George-Nascimento 2000; Rodríguez et al. 2000; George-Nascimento and Oliva 2015).

Umbrina canosai (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSA; Brazil (Luque and Poulin 2004).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2004; Luque and Poulin 2004).
Note: Alves et al. (2004) distinguished two different morphotypes.

Urophycis mystaceus (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Brazil (Alves et al. 2002c; Luque and Poulin 2004).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; intestine, stomach; metacestode; WTSA; Argentina (Alarcos and Timi 2012).

Tentacularia coryphaenae* Bosc, 1797

[For synonyms, see Palm (2004)]

Carcharhinus longimanus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; TSA, WTSA; Brazil (Rego 1977; Knoff et al. 2002, 2004b).

Carcharhinus obscurus (Elasmobranchii: Carcharhinidae); marine; spiral valve; adult; WTSA; Brazil (Knoff et al. 2002, 2004b).

Carcharodon carcharias (Elasmobranchii: Lamnidae); marine; duodenum; adult; specific locality not given; Brazil (Dollfus 1942).

Centropomus nigrescens (Actinopterygii: Centropomidae); amphidromous; peritoneum; metacestode; WTSP; Peru (Escalante and Carvajal 1984).

Coryphaena equiselis (Actinopterygii: Coryphaenidae); marine; body cavity; metacestode; specific locality not given; Brazil (Rudolphi 1819; Dollfus 1942).

Coryphaena hippurus (Actinopterygii: Coryphaenidae); marine; body cavity, liver, gonads, muscles; metacestode; WTSA, WTSP; Brazil, Peru (Escalante and Carvajal 1984; López de McDonald and Tantaleán 1985; Silva and São Clemente 2001; Vásquez-Ruiz and Jara-Campos 2012).
Note: type host.

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSA; Brazil (São Clemente et al. 2004).

Katsuwonus pelamis (Actinopterygii: Scombridae); marine; abdominal flaps, body cavity, muscles; metacestode; WTSA, WTSP; Brazil, Peru (Escalante and Carvajal 1984; Amato et al. 1990; Alves and Luque 2006).

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; muscles; metacestode; WTSA; Brazil (São Clemente et al. 2007).

Merluccius gayi peruanus (Actinopterygii: Merlucciidae); marine; mesentery; metacestode; WTSP; Peru (Durán and Oliva 1980; Jara 1998).

Polyprion oxygeneios (Actinopterygii: Polyprionidae); marine; peritoneal cavity; metacestode; JFD; Chile (Cattan et al. 1979).

Prionace glauca (Elasmobranchii: Carcharhinidae); marine; stomach, spiral valve; adult; JFD, WTSA, WTSP; Brazil, Chile, Peru (Cattan et al. 1979; Escalante 1986; Knoff et al. 2002, 2004b).

Scomber japonicus (Actinopterygii: Scombridae); marine; body cavity; metacestode; WTSP; Chile, Peru (Llerena et al. 2001; Oliva et al. 2008b).

Scomberomorus cavalla (Actinopterygii: Scombridae); marine; mesentery; metacestode; WTSA; Brazil (Dias et al. 2011a).

Trachurus murphyi (Actinopterygii: Carangidae); marine; mesentery; metacestode; WTSP; Chile, Peru (Soto and Carvajal 1979; Oliva 1982 ,1994, 1999; Pérez et al. 1999; Jara 1998).

Carangidae gen. sp. (Actinopterygii: Carangidae); marine; kidney, mesentery; metacestode; WTSP; Peru (Palm 2004).
Note: material is deposited in H. Palm's personal collection.

Unidentified tentacullariids

Dules auriga (Actinopterygii: Serranidae); marine; mesentery; metacestode; WTSA; Argentina (Braicovich and Timi 2015).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; site of infection not given; metacestode; WTSA; Brazil (Alarcos et al. 2016).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; mesentery; metacestode; WTSA; Argentina, Brazil (Pereira et al. 2014).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; intestine, mesentery; metacestode; WTSA; Argentina (Alarcos and Timi 2013).

Unidentified trypanorhynchs

Antimora rostrata (Actinopterygii: Moridae); marine; visceral cavity; metacestode; WTSP: Chile (Ñacari and Oliva 2016).

Auxis thazard (Actinopterygii: Scombridae); marine; body cavity; metacestode; WTSA; Brazil (Faria and Silva 1934).

Balistes vetula (Actinopterygii: Balistidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Brama australis (Actinopterygii: Bramidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento et al. 2002).
Note: George-Nascimento et al. (2002) distinguished two morphotypes.

Caranx crysos (Actinopterygii: Carangidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Carcharhinus limbatus (Elasmobranchii: Carcharhinidae); marine; site of infection not given; adult; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Carcharias limbatus*.

Carcharodon carcharias (Elasmobranchii: Lamnidae); marine; site of infection not given; adult; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Carcharias lamia*.

Centropomus undecimalis (Actinopterygii: Centropomidae); amphidromous; intestine; metacestode; Paraíba do Sul River basin (estuary of Guandú River); Brazil (Azevedo et al. 2011).

Coryphaenoides ariommus (Actinopterygii: Macrouridae); marine; visceral cavity; metacestode; WTSP: Chile (Ñacari and Oliva 2016).

Cynoscion acoupa (Actinopterygii: Scianidae); marine; mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Cynoscion jamaicensis (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Cynoscion leiarchus (Actinopterygii: Scianidae); marine; mesentery; metacestode; WTSA; Brazil (Faria and Silva 1934).

Cynoscion striatus (Actinopterygii: Scianidae); marine; body cavity; metacestode; WTSA; Brazil (Faria and Silva 1934; Rego et al. 1974).
Note: Rego et al. (1974) distinguished two morphotypes.

Cynoscion sp. (Actinopterygii: Scianidae); marine; muscles; metacestode; TNA; Venezuela (Vogelsang and Mayaudon 1959).
Note: tapeworms reported as *Tetrarhynchus fragilis*.

Dissostichus eleginoides (Actinopterygii: Nototheniidae); marine; site of infection not given; metacestode; Magellanic; Falkland Islands (Brown et al. 2013).

Epinephelus morio (Actinopterygii: Serranidae); marine; liver, mesentery, muscles, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Cerna morio*.

Epinephelus sp. (Actinopterygii: Serranidae); marine; muscles; metacestode; TNA; Venezuela (Vogelsang and Mayaudon 1959).
Note: tapeworms reported as *Tetrarhynchus fragilis*.

Euthynnus alletteratus (Actinopterygii: Scombridae); marine; liver, mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Gymnosarda alleterada*.

Genidens barbus (Actinopterygii: Ariidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).
Note: host reported as *Netuma barba*.

Genidens sp. (Actinopterygii: Ariidae); marine; mesentery; metacestode; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Thachysurus* sp. (sic!)

Genypterus blacodes (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSP; Chile (Riffo 1994).

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; WTSA; Argentina (Sardella et al. 1998).

Genypterus maculatus (Actinopterygii: Ophidiidae); marine; site of infection not given; metacestode; WTSP; Chile (Muñoz and George-Nascimento 2008).

Helicolenus lengerichi (Actinopterygii: Sebastidae); marine; intestinal serosa, mesentery, peritoneum, stomach serosa; WTSP; Chile (George-Nascimento and Iriarte 1989; Balboa and George-Nascimento 1998).

Hyporthodus niveatus (Actinopterygii: Serranidae); marine; liver, mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).
Note: host reported as *Garrupa niveata*.

Lophius gastrophysus (Actinopterygii: Lophiidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Lutjanus analis (Actinopterygii: Lutjanidae); marine; body cavity; metacestode; TSA; Brazil (Hermida et al. 2014).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; body cavity; metacestode; WTSA; Brazil (Pereira and Boeger 2005).

Macrourus holotrachys (Actinopterygii: Macrouridae); marine; visceral cavity; metacestode; WTSP; Chile (Ñacari and Oliva 2016).

Masturus lanceolatus (Actinopterygii: Molidae); marine; liver; metacestode; TSA (estuary of Una River); Brazil (Araújo et al. 2010).

Micropogonias furnieri (Actinopterygii: Scianidae); marine; body cavity, mesentery, peritoneum; metacestode; WTSA; Brazil, Uruguay (Faria and Silva 1934; Bertullo 1965; São Clemente 1986b; Pereira 1993; Pereira and Boeger 2005; Cardoso et al. 2006).

Notes: Faria and Silva (1934) reported the host as *Micropogon opercularis* and Bertullo (1965) reported the cestode as *Tetrarhynchus fragilis*.

Micropogonias undulatus (Actinopterygii: Scianidae); marine; mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Note: host reported as *Micropogon undulatus*.

Mola mola (Actinopterygii: Molidae); marine; oral cavity, heart, intestine, liver, muscles, stomach; metacestode; TSA; Brazil (Ahid et al. 2009).

Mycteroperca bonaci (Actinopterygii: Serranidae); marine; liver, mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Note: host reported as *Epinephelus bonaci*.

Notothenia cf. *angustata* (Actinopterygii: Nototheniidae); marine; body cavity; metacestode; WTSP; Chile (Muñoz et al. 2001).

Orthopristis ruber (Actinopterygii: Haemulidae); marine; mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Pagrus pagrus (Actinopterygii: Sparidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Paralabrax humeralis (Actinopterygii: Serranidae); marine; site of infection not given; metacestode; WTSP; Peru (Iannacone and Alvariño 2009).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; mesentery, intestine; peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934; Travassos et al. 1967; Gomes et al. 1972; Luque and Chaves 1999).

Note: host reported as *Cheilodipterus saltator* and *P. saltator* by Faria and Silva (1934) and Luque and Chaves (1999), respectively.

Porichthys porosissimus (Actinopterygii: Batrachoididae); marine; liver, mesentery; metacestode; WTSA; Brazil (Faria and Silva 1934).

Pseudopercis numida (Actinopterygii: Pinguipedidae); marine; mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Rhinobatos percellens (Elasmobranchii: Rhinobatidae); marine; site of infection not given; adult; WTSA; Brazil (Faria and Silva 1934).

Note: host reported as 'cação viola', vernacular name of *R. percellens*.

Rhizoprionodon terraenovae (Elasmobranchii: Carcharhinidae); marine; site of infection not given; adult; WTSA; Brazil (Faria and Silva 1934).

Note: host reported as 'cação alecrim' vernacular name of *R. terraenovae*.

Sardinella brasiliensis (Actinopterygii: Clupeidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Sardinella sp. (Actinopterygii: Clupeidae); marine; liver; metacestode; WTSA; Brazil (Feijó et al. 1979; Rodrigues et al. 1990).

Scomber colias (Actinopterygii: Scombridae); marine; body cavity, mesentery, peritoneum, stomach; metacestode; WTSA; Brazil (Faria and Silva 1934; Rego and Santos 1983).

Selene vomer (Actinopterygii: Carangidae); marine; mesentery, peritoneum; metacestode; WTSA; Brazil (Faria and Silva 1934).

Trachurus murphyi (Actinopterygii: Carangidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento and Arancibia 1992; George-Nascimento 2000).

Trichiurus lepturus (Actinopterygii: Trichiuridae); marine; muscle; metacestode; WTSA; Brazil (Bueno et al. 2014).

Urophycis brasiliensis (Actinopterygii: Phycidae); marine; body cavity; metacestode; WTSA; Brazil (Cardoso et al. 2006).

Sciaenidae gen. sp. (Actinopterygii: Sciaenidae); site of infection not given; metacestode; WTSA; Brazil (Travassos et al. 1967).

Note: host reported as 'pescadinha', vernacular name of sciaenid fishes.

Species inquirendae

***Otobothrium cysticum* (Mayer, 1842) Dollfus, 1942**

[Syn. *Tetrarhynchus cysticus* Mayer, 1842]

Genypterus brasiliensis (Actinopterygii: Ophidiidae); marine; body cavity, mesentery; metacestode; WTSA; Brazil (São Clemente et al. 2004).

Note: Palm (2004) treated the type-species of the genus, *O. crenacolle* Linton, 1890, and *O. curtum* (Linton, 1909) as synonyms of *O. cysticum*, but Beverige and Justine (2007b) re-established those species and considered *O. cysticum* as *species inquirenda*.

Pagrus pagrus (Actinopterygii: Sparidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Pomatomus saltatrix (Actinopterygii: Pomatomidae); marine; body cavity; metacestode; WTSA; Brazil (Palm 2004).

Note: host reported as *P. saltator*.

Scomberomorus brasiliensis (Actinopterygii: Scombridae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

Note: Palm (1997) reported the host as *S. maculatus* (Mitchill).

Sphyraena guachancho (Actinopterygii: Sphyraenidae); marine; body cavity; metacestode; TSA; Brazil (Palm 1997).

***Pterobothrium fragile* (Diesing, 1850) Dollfus, 1942**

[Syns. *Synbothrium fragile* Diesing, 1850; *Syndesmobothrium fragile* (Diesing, 1850) Diesing, 1863; *Pterobothrium* (*Synbothrium*) *fragile* (Diesing, 1850) sensu Dollfus 1942]

Pristis pristis (Elasmobranchii: Pristidae); marine; intestine; adult; NBS (estuary of Amazon River); Brazil (Diesing 1850).

Notes: host reported as *P. perotteti*. Type specimens collected by J. Natterer are poorly preserved and the validity of *P. fragile* needs confirmation (see Campbell and Beveridge 1996).

***Pterobothrium interruptum* (Rudolphi, 1819) Diesing, 1850**

[Syn. *Anthocephalus interruptum* Rudolphi, 1819]

Trichiurus lepturus (Actinopterygii: Trichiuridae); marine; body cavity; metacestode; type locality not given; Brazil (Diesing 1850, 1856).

Note: the type material deposited in MNHB was destroyed during the World War II (Campbell and Beveridge 1996).

Nomen nudum

Pterobothrium macrourum* (Rudolphi, 1819) Diesing, 1850

[Syn. *Anthocephalus macrourus* Rudolphi, 1819]

Sparidae gen. sp. (Actinopterygii: Sparidae); marine; liver, mesentery; metacestode; type locality not given; Brazil (Rudolphi 1819, Diesing 1850).

Note: type specimens collected by Olfers are barely recognized as cestodes and basic taxonomic information is missing in the original description; therefore, Cambpell and Beveridge (1996) considered this taxon, type species of *Pterobothrium*, as *nomen nudum*.

Unidentified cestodes

Acestrorhynchus altus (Actinopterygii: Acestrorhynchidae); freshwater; intestine; stage of development not given; Paraná River basin; Brazil (Takemoto et al. 2009).

Ageneiosus inermis (Actinopterygii: Auchenipteridae); freshwater; site of infection and stage of development not given; Amazon and Paraná River basins; Brazil (Travassos et al. 1927; Travassos and Teixeira de Freitas 1964).

Note: hosts reported as *A. valenciennesi* and *Pseudoageneiosus brevifilis* by Travassos and Teixeira de Freitas (1964) and Travassos et al. (1927), respectively.

Astyanax altiparanae (Actinopterygii: Characidae); freshwater; mesentery; metacestode; Paraná River basin; Brazil (Lizama et al. 2008).

Astyanax bimaculatus (Actinopterygii: Characidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos 1945).

Astyanax sp. (Actinopterygii: Characidae); freshwater; site of infection and stage of development not given; Amazon River basin; Brazil (Travassos and Teixeira de Freitas 1964).

Australoheros facetus (Actinopterygii: Cichlidae); freshwater; site of infection and stage of development not given; Doce River basin; Brazil (Travassos and Teixeira de Freitas 1948).

Brachyplatystoma sp. (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos and Teixeira de Freitas 1942, 1943).

Brama australis (Actinopterygii: Bramidae); marine; site of infection not given; metacestode; WTSP; Chile (George-Nascimento et al. 2002).

Brevoortia aurea (Actinopterygii: Clupeidae); marine; site of infection not given; metacestode; WTSA; Argentina (Alarcos and Etchegoin 2010).
Notes: host collected in a coastal lagoon.

Calophysus macropterus (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Amazon River basin; Brazil (Travassos and Teixeira de Freitas 1964).

Caranx hippos (Actinopterygii: Carangidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Centropomus sp. (Actinopterygii: Centropomidae); amphidromous; site of infection and stage of development not given; Doce River basin; Brazil (Travassos and Teixeira de Freitas 1948).

Cetopsis coecutiens (Actinopterygii: Cetopsidae); freshwater; site of infection and stage of development not given; Amazon River basin; Brazil (Travassos and Teixeira de Freitas 1964).

Cichla ocellaris (Actinopterygii: Cichlidae); freshwater; site of infection not given; adult; Pereira de Miranda fishpond, Ceará State; Brazil (Kohn et al. 2004).

Cichlasoma bimaculatum (Actinopterygii: Cichlidae); freshwater; site of infection not given; adult; Pereira de Miranda fishpond, Ceará State, Paraná River basin; Brazil (Travassos 1940; Kohn et al. 2004).

Colossoma macropomum (Actinopterygii: Serrasalmidae); freshwater; site of infection not given; metacestode; Paraná River basin; Brazil (Kohn et al. 1985).

Coryphaena hippurus (Actinopterygii: Coryphaenidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Crenicichla haroldoi (Actinopterygii: Cichlidae); freshwater; intestine; adult (immature); Paraná River basin; Brazil (Kohn et al. 2011).

Cynoscion guatucupa (Actinopterygii: Sciaenidae); marine; site of infection not given; metacestode; WTSA; Argentina (Timi et al. 2010b).

Dules auriga (Actinopterygii: Serranidae); marine; mesentery; metacestode; WTSA; Argentina (Braicovich and Timi 2015).

Galaxias maculatus (Actinopterygii: Galaxiidae); amphidromous; intestine; metacestode; Maullin River basin; Chile (Bravo et al. 2007).

Galeocharax knerii (Actinopterygii: Characidae); freshwater; intestine; adult (immature); Paraná River basin; Brazil (Kohn et al. 2011).

Geophagus brasiliensis (Actinopterygii: Characidae); freshwater; site of infection and stage of development not given; Doce River basin; Brazil (Travassos 1944; Travassos and Teixeira de Freitas 1948).

Gymnocharacinus bergii (Actinopterygii: Characidae); freshwater; site of infection not given; metacestode; Negro River Basin; Argentina (Ortubay et al. 1994).

Gymnotus carapo (Actinopterygii: Gymnotidae); freshwater; site of infection and stage of development not given; Doce River basin; Brazil (Travassos and Teixeira de Freitas 1948).

Note: host reported as *Giton fasciatus*.

Gymnotus inaequilabiatus (Actinopterygii: Gymnotidae); freshwater; intestine; stage of development not given; Paraná River basin; Brazil (Takemoto et al. 2009).

Harengula sp. (Actinopterygii: Clupeidae); marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Hemisorubim platyrhynchos (Actinopterygii: Pimelodidae); freshwater; mesentery, intestine; metacestode, adult; Amazon and Paraná River basins; Brazil, Peru (Travassos and Teixeira de Freitas 1942, 1943; Kohn et al. 2011).

Hoplerythrinus unitaeniatus (Actinopterygii: Erythrinidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos 1940; Travassos and Teixeira de Freitas 1943).

Hoplias malabaricus (Actinopterygii: Erythrinidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos 1940).

Lagocephalus laevigatus (Actinopterygii: Tetraodontidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Leporinus obtusidens (Actinopterygii: Anostomidae); freshwater; intestine; adult; Paraná River basin; Brazil (Kohn et al. 2011).

Loricariichthys sp. (Actinopterygii: Loricariidae); freshwater; intestine; adult (immature); Paraná River basin; Brazil (Kohn et al. 2011).

Lutjanus jocu (Actinopterygii: Lutjanidae); brackish, freshwater; marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Macrodon ancylodon (Actinopterygii: Sciaenidae); marine; mesentery, muscles; metacestode; WTSA; Brazil (Oliveira 1985).

Merluccius australis (Actinopterygii: Merlucciidae); marine; mesentery, muscles; metacestode; WTSP; Chile (Fernández 1985).

Micropogonias furnieri (Actinopterygii: Sciaenidae); marine; mesentery, muscles; metacestode; WTSA; Brazil (Oliveira 1985).

Micropogonias sp. (Actinopterygii: Sciaenidae); marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Oligoplites saurus (Actinopterygii: Carangidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Oreochromis niloticus (Actinopterygii: Cichlidae); freshwater; site of infection not given; metacestode; Pereira de Miranda fishpond, Ceará and Paraná States; Brazil (Kohn et al. 2004; Graça and Machado 2007).

Note: introduced fish host (Froese and Pauly 2016).

Paralichthys isosceles (Actinopterygii: Paralichthyidae); marine; site of infection not given; metacestode; WTSA; Brazil (Alarcos et al. 2016).

Piaractus mesopotamicus (Actinopterygii: Characidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Kohn et al. 2011).

Pimelodella gracilis (Actinopterygii: Heptapteridae); freshwater; intestine, mesentery; adult, metacestode; Amazon and Paraná River basins; Brazil (Kohn et al. 2011).

Pimelodella lateristriga (Actinopterygii: Heptapteridae); freshwater; intestine; adult; Paraná River basin; Brazil (Kohn et al. 2011).

Pimelodus maculatus (Actinopterygii: Pimelodidae); freshwater; intestine; adult, metacestode; Paraná River basin; Brazil (Travassos and Teixeira de Freitas 1940, 1943; Travassos 1947; Kohn et al. 1985; Kohn and Fernandes 1987).
Notes: host reported as *P. clarias* by all authors.

Pimelodus ortmanni (Actinopterygii: Pimelodidae); freshwater; intestine; metacestode; Paraná River basin; Brazil (Kohn et al. 1988).

Pinirampus pirinampu (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos et al. 1927).

Plagioscion squamosissimus (Actinopterygii: Sciaenidae); freshwater; mesentery, intestine; metacestode, adult (immature); Amazon, Paraná and Tocantins-Araguaia River basins; Brazil (Kohn et al. 2011; Lacerda et al. 2012).

Poecilia vivipara (Actinopterygii: Poeciliidae); freshwater; site of infection not given; adult; Pereira de Miranda fishpond, Ceará State; Brazil (Kohn et al. 2004).

Potamotrygon motoro (Elasmobranchii: Potamotrygonidae); freshwater; intestine; adult; Paraná River basin; Brazil (Travassos and Teixeira de Freitas 1940, 1941, 1942, 1943; Kohn et al. 2011).
Notes: Travassos and Teixeira de Freitas (1940, 1941, 1942, 1943) did not specify the site of infection and the stage of development; they also reported the host as *Ellipesurus motoro*.

Potamotrygon orbignyi (Elasmobranchii: Potamotrygonidae); freshwater; site of infection and stage of development not given; Amazon River basin; Brazil (Travassos and Teixeira de Freitas 1964).
Note: host reported as *Paratrygon hystrix*.

Prochilodus argenteus (Actinopterygii: Prochilodontidae); freshwater; intestinal serosa; metacestode; São Francisco River Basin; Brazil (Monteiro et al. 2009).

Pseudopercis semifasciata (Actinopterygii: Pinguipedidae); marine; stomach wall; metacestode; Magellanic, WTSA; Argentina (Timi and Lanfranchi 2009a).

Pseudoplatystoma fasciatum (Actinopterygii: Pimelodidae); freshwater; intestine; adult; Paraná River basin; Brazil (Campos et al. 2008, 2009b).

Pseudoplatystoma sp. (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Amazon and Paraná River basins; Brazil (Travassos et al. 1927; Travassos 1940; Travassos and Teixeira de Freitas 1964).
Note: Travassos et al. (1927) reported the host as *P. tigrinum*, which does not occur in the Paraná River basin (Froese and Pauly 2016).

Raneya brasiliensis (Actinopterygii: Ophidiidae); marine; mesentery; metacestode; Magellanic, WTSA; Argentina (Vales et al. 2011).

Rhaphiodon vulpinus (Actinopterygii: Cynodontidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos and Teixeira de Freitas 1942).

Salilota australis (Actinopterygii: Moridae); marine; body cavity; metacestode; Magellanic; Argentina (Szidat 1961).

Salminus brasiliensis (Actinopterygii: Bryconidae); freshwater; intestine; adult (immature); Paraná River basin; Brazil (Ceccarrelli et al. 2006; Kohn et al. 2011).

Scomber colias (Actinopterygii: Scombridae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Selene vomer (Actinopterygii: Carangidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Sphoeroides testudineus (Actinopterygii: Tetraodontidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Steindachneridion parahybae (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos and Teixeira de Freitas 1943; Kohn and Fernandes 1987).

Note: this fish is found only in the Paraíba do Sul and Jequitinhonha River basins (Froese and Pauly 2016).

Sternopygus macrurus (Actinopterygii: Sternopygidae); freshwater; intestine; adult; Paraná River basin; Brazil (Kohn et al. 2011).

Tilapia rendalli (Actinopterygii: Cichlidae); freshwater; intestine; metacestode; Ingá Lake, Paraná State, Brazil (Graça and Machado 2007).

Trachurus murphyi (Actinopterygii: Carangidae); marine; body cavity; metacestode; WTSP; Chile (George-Nascimento and Oliva 2015).

Umbrina coroides (Actinopterygii: Sciaenidae); brackish, marine; site of infection and stage of development not given; TSA; Brazil (Travassos et al. 1967).

Xystreurys rasile (Actinopterygii: Paralichthyidae); marine; mesentery; metacestode; WTSA; Argentina (Alarcos and Timi 2012, 2013).

Zungaro jahu (Actinopterygii: Pimelodidae); freshwater; site of infection and stage of development not given; Paraná River basin; Brazil (Travassos 1940, 1947; Travassos and Teixeira de Freitas 1942, 1943).

Note: host reported as *P. luetkeni*.

Host-Parasite List

Phylum Chordata
Class Actinopterygii
Order Anguilliformes

Family Congridae
Conger orbignianus Valenciennes: *Grillotia* (*Christianella*) *carvajalregorum* (L), *G.* (*Grillotia*) *erinaceus* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Family Muraenidae
Gymnothorax moringa (Cuvier): '*Scolex* spp.' (L)

Order Atheriniformes
Family Atherinopsidae
Basilichthys australis Eigenmann: *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L).
Odontesthes argentinensis (Valenciennes): '*Scolex* spp.' (L)
Odontesthes bonariensis (Valenciennes): *Cangatiella macdonaghi*
Odontesthes hatcheri (Eigenmann): *Cangatiella macdonaghi*
Odontesthes mauleanum (Steindachner): *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L).
Odontesthes regia (Humboldt): *Diphyllobothrium* sp. (L), *Lacistorhynchus tenuis* (L), *Nybelinia* sp. (L), unidentified 'pseudophyllidean' (L), '*Scolex* spp.' (L)
Odontesthes smitti (Lahille): unidentified bothriocephalidean (L), '*Scolex* spp.' (L)

Order Aulopiformes
Family Synodontidae
Synodus scituliceps Jordan & Gilbert: *Rhinebothrium* sp. (L), '*Scolex* spp.' (L)

Order Batrachoidiformes
Family Batrachoididae
Aphos porosus (Valenciennes): *Clestobothrium crassiceps*, *Grillotia* sp. (L), *Nybelinia* sp. (L), unidentified 'pseudophyllidean' (L), '*Scolex* spp.' (L)
Porichthys porosissimus (Cuvier): *Grillotia* (*Christianella*) *carvajalregorum* (L), *G.* (*Grillotia*) *erinaceus* (?) (L), *Nybelinia lingualis* (L), unidentified trypanorhynch (L), '*Scolex* spp.' (L)

Order Beloniformes
Family Belonidae
Strongylura scapularis (Jordan & Gilbert): '*Scolex* spp.' (L)
Tylosurus acus acus (Lacépède): '*Scolex*' spp. (L)

Family Hemiramphidae
Hyporhamphus unifasciatus (Ranzani): '*Scolex* spp.' (L)

Order Characiformes
Family Acestrorhynchidae
Acestrorhynchus altus Menezes: *Monticellia dlouhyi*, unidentified cestode

Family Anostomidae
Leporellus vittatus (Valenciennes): unidentified proteocephalid
Leporinus friderici (Bloch): *Proteocephalus vazzolerae*, unidentified proteocephalid
Leporinus lacustris Amaral Campos: *Proteocephalus vazzolerae*
Leporinus obtusidens (Valenciennes): unidentified cestode

Family Bryconidae
Brycon cephalus (Günther): unidentified proteocephalid
Brycon orbignyanus (Valenciennes): *Monticellia* sp.
Salminus brasiliensis (Cuvier): *Monticellia coryphicephala*, unidentified cestode
Salminus franciscanus Lima & Britski: *Monticellia coryphicephala*

Family Characidae
Aphyocharax anisitsi Eigenmann & Kennedy: unidentified proteocephalid
Astyanax altiparanae Garutti & Britski: *Senga* sp., unidentified proteocephalid, unidentified cestode
Astyanax bimaculatus (Linnaeus): unidentified cestode
Astyanax scabripinnis (Jenyns): *Senga* sp.
***Astyanax* sp.**: unidentified cestode
Galeocharax knerii (Steindachner): unidentified cestode
Gymnocharacinus bergii Steindachner: unidentified cestode
Psellogrammus kennedyi (Eigenmann): unidentified proteocephalid

Family Cynodontidae
Rhaphiodon vulpinus Spix & Agassiz: *Choanoscolex abscisus*, unidentified cestode

Family Erythrinidae
Hoplerythrinus unitaeniatus (Spix & Agassiz): *Nomimoscolex matogrossensis* (?), *Proteocephalus mahnerti*, unidentified cestode
Hoplias malabaricus (Bloch): *Nomimoscolex matogrossensis*, *Proteocephalus regoi*, unidentified cestode

Family Prochilodontidae
Prochilodus argenteus Spix & Agassiz: *Valipora* sp. (L), unidentified cestode
Prochilodus brevis Steindachner: unidentified proteocephalid

Prochilodus lineatus (Valenciennes): unidentified proteocephalid, *Valipora campylancristrota* (L)

Family Serrasalmidae
Colossoma macropomum (Cuvier): unidentified proteocephalid, unidentified cestode
Colossoma macropomum* × *Piaractus mesopotamicus: unidentified proteocephalid
Piaractus brachypomus (Cuvier): unidentified proteocephalid
Piaractus mesopotamicus (Holmberg): *Proteocephalus vazzolerae*, unidentified cestode
Pygocentrus nattereri Kner: *Proteocephalus serrasalmus*
Serrasalmus maculatus Kner: *Proteocephalus serrasalmus*

Order Clupeiformes
Family Clupeidae
Brevoortia aurea (Spix & Agassiz): unidentified cestode
Ethmidium maculatum (Valenciennes): '*Scolex* spp.' (L)
Harengula clupeola (Cuvier): *Callitetrarhynchus gracilis* (L)
***Harengula* sp.**: unidentified cestode
Opisthonema oglinum (Lesueur): *Callitetrarhynchus gracilis* (L)
Sardinella brasiliensis (Steindachner): *Callitetrarhynchus gracilis* (L), *Nybelinia* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L)
***Sardinella* sp.**: unidentified proteocephalid, unidentified trypanorhynch
Sardinops sagax (Jenyns): '*Scolex* spp.' (L)
Clupeidae gen. sp.: *Pterobothrium heteracanthum* (L)

Family Engraulidae
Anchoa tricolor (Spix & Agassiz): '*Scolex* spp.' (L)
Engraulis anchoita Hubbs & Marini: *Bothriocephalus* sp., unidentified 'pseudophyllidean', '*Scolex* spp.' (L)
Engraulis ringens Jenyns: *Bothriocephalus* sp., *Diphyllobothrium* sp. (L), '*Scolex* spp.' (L)

Order Cypriniformes
Family Cyprinidae
Cyprinus carpio Linnaeus: *Schyzocotyle acheilognathi*, unidentified caryophyllidean, unidentified proteocephalid
Hypophthalmichthys nobilis (Richardson): unidentified proteocephalid
Pethia conchonius (Hamilton): *Schyzocotyle acheilognathi*

Order Cyprinodontiformes
Family Poeciliidae
Poecilia reticulata Peters: *Glossocercus auritus* (L), *Schyzocotyle acheilognathi*
Poecilia vivipara Bloch & Schneider: unidentified cestode
Xiphophorus hellerii Heckel: *Schyzocotyle acheilognathi*
Xiphophorus maculatus (Günther): *Schyzocotyle acheilognathi*

Order Gadiformes

Family Gadidae

Micromesistius australis australis Norman: *Clestobothrium crassiceps*, *Diphyllobothrium* sp. (L), *Grillotia* sp. (L), *Hepatoxylon trichiuri* (L), *Hepatoxylon* sp. (L), unidentified 'pseudophyllidean'

Family Macrouridae

Coelorinchus chilensis Gilbert & Thompson: *Grillotia* sp. (L), *Hepatoxylon trichiuri* (L)

Coryphaenoides ariommus Gilbert & Thompson: unidentified trypanorhynch

Macrourus carinatus (Günther): *Grillotia* (*Grillotia*) *borealis* (?) (L)

Macrourus holotrachys Günther: *Hepatoxylon* sp. (L), *Parabothriocephalus* sp. (L), unidentified trypanorhynch

Family Merlucciidae

Macruronus magellanicus Lönnberg: *Clestobothrium crassiceps*, *Grillotia* (*Grillotia*) *dollfusi* (L), *G. heptanchi* (L), *Hepatoxylon trichiuri* (L), unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Merluccius australis (Hutton): *Clestobothrium splendidum*, *Diphyllobothrium* sp. (L), *Grillotia heptanchi* (L), *Grillotia* sp. (L), *Hepatoxylon trichiuri* (L), *Lacistorhynchus* sp. (L), unidentified phyllobothriidean, '*Scolex* spp.' (L), unidentified cestode

Merluccius gayi gayi (Guichenot): *Clestobothrium crassiceps*, *Grillotia* (*Grillotia*) *dollfusi* (L), *G. heptanchi* (L), *Hepatoxylon trichiuri* (L), *Nybelinia surmenicola* (L), unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Merluccius gayi peruanus (Ginsburg): *Adenocephalus pacificus* (L), *Callitetrarhynchus gracilis* (L), *Clestobothrium crassiceps*, *Diphyllobothrium* sp. (L), *Grillotia* (*Grillotia*) *dollfusi* (L), *Lacistorhynchus tenuis* (L), *Nybelinia* sp. (L), *Tentacularia coryphaenae* (L)

Merluccius hubbsi Marini: *Clestobothrium cristinae*, *Diphyllobothrium* sp. (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Grillotia* sp. (L), *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L), unidentified phyllobothriidean, unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

***Merluccius* sp.**: *Clestobothrium crassiceps*

Family Moridae

Antimora rostrata (Günther): unidentified trypanorhynch

Salilota australis (Günther): *Grillotia* (*Grillotia*) *borealis* (?) (L), *G.* (*G.*) *patagonica* (L), *Hepatoxylon trichiuri* (L), unidentified cestode

Family Phycidae

Urophycis brasiliensis (Kaup): *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia* sp. (L), *Lacistorhynchus* sp. (L), *Nybelinia* sp. (L), *Phyllobothrium* sp. (L), unidentified phyllobothriidean, unidentified trypanorhynch, '*Scolex* spp.' (L)

Urophycis mystaceus Ribeiro: *Lacistorhynchus* sp. (L), *Nybelinia* sp. (L), unidentified phyllobothriidean, '*Scolex* spp.' (L)
***Urophycis* sp.**: unidentified phyllobothriidean

Order Gobiesociformes
Family Gobiesocidae
Gobiesox marmoratus Jenyns: unidentified 'pseudophyllidean', '*Scolex* spp.' (L)
Sicyases sanguineus Müller & Troschel: unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Order Gymnotiformes
Family Gymnotidae
Gymnotus carapo Linnaeus: *Nomimoscolex chubbi*, *N. dechambrieri*, *N. guillermoi*, *Proteocephalus* sp., unidentified cestode
Gymnotus inaequilabiatus (Valenciennes): unidentified cestode
***Gymnotus* sp.**: *Nomimoscolex chubbi*

Family Sternopygidae
Sternopygus macrurus (Bloch & Schneider): unidentified cestode

Order Lampriformes
Family Lampridae
Lampris guttatus (Brünnich): *Hepatoxylon trichiuri* (L)

Order Lophiiformes
Family Lophiidae
Lophius gastrophysus Miranda Ribeiro: *Diphyllobothrium* sp. (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Mixonybelinia* sp. (L), *Nybelinia* sp. (L), *Tentacularia coryphaenae* (L), unidentified trypanorhynch

Ordem Mugiliformes
Family Mugilidae
Mugil cephalus Linnaeus: *Lacistorhynchus tenuis* (L), '*Scolex* spp.' (L)
Mugil liza Valenciennes: unidentified phyllobothriidean, '*Scolex* spp.' (L)

Order Notacanthiformes
Family Notacanthidae
Notacanthus sexspinis Richardson: *Hepatoxylon trichiuri* (L)

Order Ophidiiformes
Family Ophidiidae
Genypterus blacodes (Forster): *Anonchocephalus chilensis*, *Hepatoxylon trichiuri* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Genypterus brasiliensis Regan: *Anonchocephalus chilensis*, *Callitetrarhynchus gracilis* (L), *Diphyllobothrium* sp. (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Hepatoxylon trichiuri* (L), *Heteronybelinia nipponica* (L), *Mixonybelinia beveridgei* (L), *Nybelinia* sp. (L), *Otobothrium cysticum* (L), *Tentacularia coryphaenae* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Genypterus chilensis (Guichenot): *Anonchocephalus chilensis*, *Grillotia heptanchi* (L), *Hepatoxylon trichiuri* (L)

Genypterus maculatus (Tschudi): *Adenocephalus pacificus* (L), *Anonchocephalus chilensis*, *Diphyllobothrium* sp. (L), *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L), unidentified trypanorhynch, '*Scolex* spp.'

Raneya brasiliensis (Kaup): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia mattisi* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L), unidentified cestode

Order Osmeriformes

Family Galaxiidae

Aplochiton taeniatus Jenyns: unidentified bothriocephalidean

Aplochiton zebra Jenyns: *Ailinella mirabilis*

Galaxias maculatus (Jenyns): *Ailinella mirabilis*, *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L), unidentified cestode

Galaxias platei Steindachner: *Diphyllobothrium latum* (L), *Diphyllobothrium* sp. (L), *Galaxitaenia toloi*

Order Osteoglossiformes

Family Arapaimidae

Arapaima gigas (Schinz): *Nesolecithus janickii*, *Schizochoerus liguloideus*

Order Perciformes

Family Blenniidae

Hypsoblennius sordidus (Bennett): unidentified 'pseudophyllidean'

Scartichthys viridis (Valenciennes): *Lacistorhynchus* sp. (L), unidentified 'pseudophyllidean'

Family Bovichtidae

Cottoperca gobio (Günther): *Bothriocephalus timii*, *Grillotia* (*Grillotia*) *patagonica* (L)

Family Bramidae

Brama australis Valenciennes: *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L), unidentified trypanorhynch, unidentified cestode

Brama japonica Hilgendorf: *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L)

Family Carangidae

Caranx crysos (Mitchill): *Callitetrarhynchus gracilis* (L), unidentified trypanorhynch

Caranx hippos (Linnaeus): *Callitetrarhynchus gracilis* (L), *Dasyrhynchus giganteus* (L), *Nybelinia* sp. (L), unidentified cestode
Caranx latus Agassiz: *Callitetrarhynchus gracilis* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L)
Chloroscombrus chrysurus (Linnaeus): *Callitetrarhynchus gracilis* (L)
Oligoplites palometa (Cuvier): *Callitetrarhynchus gracilis* (L), *Pterobothrium crassicole* (L), '*Scolex* spp.' (L)
Oligoplites saliens (Bloch): *Dasyrhynchus giganteus* (L), '*Scolex* spp.' (L)
Oligoplites saurus (Bloch & Schneider): *Callitetrarhynchus gracilis* (L), '*Scolex* spp.' (L), unidentified cestode
Parona signata (Jenyns): *Grillotia* (*Christianella*) *carvajalregorum* (L), '*Scolex* spp.' (L)
Selene setapinnis (Mitchill): *Callitetrarhynchus gracilis* (L), *Nybelinia* sp. (L)
Selene vomer (Linnaeus): *Callitetrarhynchus gracilis* (L), *Nybelinia lingualis* (L), unidentified trypanorhynch, unidentified cestode
Seriola lalandi Valenciennes: *Floriceps saccatus* (L)
Trachinotus paitensis Cuvier: *Adenocephalus pacificus* (L)
Trachurus lathami Nichols: *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), unidentified 'pseudophyllidean', '*Scolex* spp.' (L)
Trachurus murphyi Nichols: *Adenocephalus pacificus* (L), *Diphyllobothrium* sp. (L), *Hepatoxylon trichiuri* (L), *Hepatoxylon* sp. (L), *Nybelinia lingualis* (L), *N. surmenicola* (L), *Nybelinia* sp. (L), *Tentacularia coryphaenae* (L), unidentified 'pseudophyllidean', unidentified trypanorhynch, '*Scolex* spp.' (L), unidentified cestode
Carangidae gen. sp.: *Tentacularia coryphaenae* (L)

Family Centrolophidae
Seriolella porosa Guichenot: *Nybelinia* sp. (L).
Seriolella violacea Guichenot: *Adenocephalus pacificus* (L), *Neobothriocephalus aspinosus*

Family Centropomidae
Centropomus nigrescens Günther: *Floriceps saccatus* (L), *Tentacularia coryphaenae* (L)
Centropomus undecimalis (Bloch): *Callitetrarhynchus gracilis* (L), unidentified trypanorhynch
***Centropomus* sp.**: unidentified cestode

Family Cheilodactylidae
Cheilodactylus variegatus Valenciennes: *Lacistorhynchus tenuis* (L)
Nemadactylus bergi (Norman): *Grillotia* (*Christianella*) *carvajalregorum* (L), *G.* (*Grillotia*) *patagonica* (L), *Heteronybelinia mattisi* (L)

Family Cichlidae
Aequidens tetramerus (Heckel): unidentified proteocephalid
Astronotus ocellatus (Agassiz): *Proteocephalus gibsoni*, unidentified proteocephalid
***Astronotus* sp.**: *Proteocephalus gibsoni*
Australoheros facetus (Jenyns): *Parvitaenia macropeos* (L), unidentified cestode

Cichla kelberi Kullander & Ferreira: *Proteocephalus macrophallus*, *P. microscopicus*, *Sciadocephalus megalodiscus*
Cichla monoculus Spix: *Proteocephalus macrophallus*, *P. microscopicus*, *Sciadocephalus megalodiscus*, unidentified bothriocephalidean
Cichla ocellaris Schneider: *Proteocephalus macrophallus*, *P. microscopicus*, unidentified proteocephalid, unidentified cestode
Cichla piquiti Kullander & Ferreira: *Proteocephalus macrophallus*, *P. microscopicus*, *Sciadocephalus megalodiscus*
***Cichla* sp.**: *Proteocephalus macrophallus*, *P. microscopicus*
Cichlasoma amazonarum Kullander: unidentified proteocephalid (new species and genus, see the parasite-host list)
Cichlasoma bimaculatum (Linnaeus): unidentified cestode
Crenicichla britskii Kullander: *Valipora* sp. (L)
Crenicichla haroldoi Luengo & Britski: unidentified cestode
Crenicichla lepidota Heckel: unidentified proteocephalid
Geophagus brasiliensis (Quoy & Gaimard): unidentified caryophyllidean, *Proteocephalus gibsoni*, *Valipora campylancristrota* (L), unidentified proteocephalid, unidentified cestode
Geophagus proximus (Castelnau): unidentified proteocephalid
Laetacara curviceps (Ahl): unidentified proteocephalid
Oreochromis niloticus (Linnaeus): unidentified cestode
***Oreochromis* sp.**: unidentified proteocephalid
Satanoperca pappaterra (Heckel): unidentified cyclophyllidean, unidentified proteocephalid
Tilapia rendalli (Boulenger): unidentified cestode

Family Coryphaenidae
Coryphaena equiselis Linnaeus: *Tentacularia coryphaenae* (L)
Coryphaena hippurus Linnaeus: *Floriceps saccatus* (L), *Hepatoxylon trichiuri* (L), *Pterobothrium acanthotruncatum*, *Nybelinia* sp., *Tentacularia coryphaenae*, '*Scolex* spp.', unidentified cestode

Family Eleginopsidae
Eleginops maclovinus (Cuvier): *Bothriocephalus* sp., *Grillotia* (*Grillotia*) *erinaceus* (L), *Grillotia* sp. (L), unidentified bothriocephalidean, '*Scolex* spp.' (L)

Family Eleotridae
Dormitator maculatus (Bloch): unidentified cyclophyllidean (L)

Family Gempylidae
Thyrsites atun (Euphrasen): *Molicola* sp. (L)

Family Gerreidae
Diapterus rhombeus (Cuvier): *Nybelinia* sp. (L)

Family Gobiidae
Gobioides broussonnetii Lacépède: *Pterobothrium crassicole* (L)
Gobionellus oceanicus (Pallas): *Rhinebothrium* sp. (L)

Family Haemulidae
Conodon nobilis (Linnaeus): *Callitetrarhynchus* sp. (L), *Pterobothrium* sp. (L)
Haemulon aurolineatum Cuvier: *Callitetrarhynchus gracilis* (L), *Pterobothrium kingstoni* (L)
Haemulon plumierii (Lacépède): *Heteronybelinia estigmena* (L), *Nybelinia lingualis* (L), *Pseudotobothrium dipsacum* (L)
Haemulon steindachneri (Jordan & Gilbert): '*Scolex* spp.' (L)
Isacia conceptionis (Cuvier): *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L)
Orthopristis ruber (Cuvier): unidentified trypanorhynch, '*Scolex* spp.' (L)
Pomadasys crocro (Cuvier): *Pterobothrium heteracanthum* (L)

Family Kyphosidae
Girella laevifrons (Tschudi): unidentified bothriocephalidean

Family Labrisomidae
Labrisomus philippii (Steindachner): *Lacistorhynchus tenuis* (L)

Family Lutjanidae
Lutjanus analis (Cuvier): *Grillotia* sp. (L), unidentified trypanorhynch
Lutjanus jocu (Bloch & Schneider): unidentified cestode
Lutjanus synagris (Linnaeus): *Callitetrarhynchus gracilis* (L)

Family Mullidae
Mullus argentinae Hubbs & Marini: *Heteronybelinia nipponica* (L), *Nybelinia* sp. (L)
Pseudupeneus maculatus (Bloch): *Heteronybelinia overstreeti* (L), *Mixonybelinia edwinlintoni* (L), *Nybelinia africana* (L), *N. indica* (L), *N. lingualis* (L), *Pseudolacistorhynchus noodti* (L), *Pseudotobothrium dipsacum* (L)

Family Nototheniidae
Dissostichus eleginoides Smitt: *Clestobothrium crassiceps* (?), *Grillotia* (*Grillotia*) *erinaceus* (L), *Hepatoxylon trichiuri* (L), unidentified 'pseudophyllidean', unidentified trypanorhynch, '*Scolex* spp.' (L)
Notothenia* cf. *angustata Hutton: unidentified diphyllidean, unidentified trypanorhynch, '*Scolex* spp.' (L)
Patagonotothen brevicauda brevicauda (Lönnberg): *Grillotia* (*Grillotia*) *patagonica* (L)
Patagonotothen ramsayi (Regan): *Grillotia* (*Grillotia*) *patagonica* (L)

Family Percichthyidae

Percichthys trucha (Valenciennes): *Diphyllobothrium dendriticum* (L), *D. latum* (L), unidentified bothriocephalidean, unidentified cyclophyllidean (L)

***Percichthys* sp.**: *Diphyllobothrium latum* (L)

Family Perciliidae

Percilia gillissi Girard: *Diphyllobothrium dendriticum* (L)

Family Percophidae

Percophis brasiliensis Quoy & Gaimard: *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Grillotia* sp. (L), *Nybelinia* sp. (L), unidentified bothriocephalidean, unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Family Pinguipedidae

Pinguipes brasilianus Cuvier: *Anonchocephalus* sp., *Callitetrarhynchus gracilis* (L), *Grillotia* sp. (L), '*Scolex* spp.' (L)

Prolatilus jugularis (Valenciennes): '*Scolex* spp. (L)', unidentified bothriocephalidean unidentified 'pseudophyllidean'

Pseudopercis numida Miranda Ribeiro: *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Pseudopercis semifasciata (Cuvier): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Grillotia* sp. (L), *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L), unidentified cestode

Family Polyprionidae

Polyprion oxygeneios (Schneider & Forster): *Tentacularia coryphaenae* (L)

Family Pomatomidae

Pomatomus saltatrix (Linnaeus): *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), *Otobothrium cysticum* (L), *Pseudogrillotia* sp. (L), *Pterobothrium crassicole* (L), *Pterobothrium* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Family Priacanthidae

Priacanthus arenatus Cuvier: *Callitetrarhynchus speciosus* (L), '*Scolex* spp.' (L)

Family Sciaenidae

Cheilotrema fasciatum Tschudi: *Lacistorhynchus dollfusi* (L), *L. tenuis* (L)

Cilus gilberti (Abbott): *Adenocephalus pacificus* (L), *Diphyllobothrium* sp. (L), *Lacistorhynchus tenuis* (L), *Poecilancistrium caryophyllum* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Ctenosciaena gracilicirrhus (Metzelaar): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia annakohnae* (L)

Cynoscion acoupa (Lacépède): *Callitetrarhynchus gracilis* (L), *Poecilancistrium caryophyllum* (L), *Pterobothrium crassicole* (L), *P. heteracanthum* (L), *Pterobothrium* sp. (L), unidentified trypanorhynch

Cynoscion analis (Jenyns): *Adenocephalus pacificus* (L), *Diphyllobothrium* sp. (L), *Nybelinia* sp. (L)

Cynoscion guatucupa (Cuvier): *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Dasyrhynchus pacificus* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *G.* (*C.*) *minuta* (?) (L), *Heteronybelinia annakohnae* (L), *Nybelinia* sp. (L), *Pterobothrium heteracanthum* (L),'*Scolex* spp.' (L), unidentified cestode

Cynoscion jamaicensis (Vaillant & Bocourt): *Dasyrhynchus pacificus* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia annakohnae* (L), *H. estigmena* (L), unidentified trypanorhynch

Cynoscion leiarchus (Cuvier): *Pterobothrium crassicole* (L), unidentified trypanorhynch

Cynoscion striatus (Cuvier): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Pterobothrium* sp. (L), unidentified proteocephalid, unidentified trypanorhynch

***Cynoscion* sp.**: *Nybelinia lingualis* (L), *Pterobothrium crassicole* (L), unidentified trypanorhynch

Larimus breviceps Cuvier: *Callitetrarhynchus gracilis* (L)

Macrodon ancylodon (Bloch & Schneider): *Callitetrarhynchus gracilis* (L), *Dasyrhynchus pacificus* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), *Poecilancistrium caryophyllum* (L), *Pterobothrium* sp. (L), unidentified trypanorhynch, unidentified cestode

Menticirrhus americanus (Linnaeus): *Dasyrhynchus pacificus* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia annakohnae* (L), *H. nipponica* (L), *Pterobothrium* sp. (L), '*Scolex* spp.' (L)

Menticirrhus littoralis (Holbrook): *Grillotia* (*Christianella*) *carvajalregorum* (L)

Menticirrhus ophicephalus (Jenyns): *Adenocephalus pacificus* (L)

Micropogonias altipinnis (Günther): *Poecilancistrium caryophyllum* (L)

Micropogonias furnieri (Desmarest): *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Dollfusiella* sp. (L), *Gilquinia* sp. (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), *Poecilancistrium caryophyllum* (L), *Pterobothrium acanthotruncatum* (L), *P. crassicole* (L), *P. heteracanthum* (L), *Pterobothrium* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L), unidentified cestode

Micropogonias undulatus (Linnaeus): *Pterobothrium crassicole* (L), *P. heteracanthum* (L), unidentified trypanorhynch

***Micropogonias* sp.**: unidentified cestode

Paralabrax humeralis (Valenciennes): *Adenocephalus pacificus* (L), *Grillotia* sp. (L), *Nybelinia* sp. (L), unidentified bothriocephalidean, unidentified trypanorhynch

Paralonchurus brasiliensis (Steindachner): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Paralonchurus peruanus (Steindachner): *Adenocephalus pacificus* (L), *Callitetrarhynchus gracilis* (L), *Diphyllobothrium* sp. (L), *Pterobothrium acanthotruncatum* (L)

Plagioscion squamosissimus (Heckel): *Callitetrarhynchus gracilis* (L), *Pterobothrium crassicole* (L), *P. heteracanthum* (L), *Poecilancistrium caryophyllum* (L), unidentified bothriocephalidean, unidentified cestode

Pogonias cromis (Linnaeus): *Pterobothrium crassicole* (L), *P. heteracanthum* (L)

Sciaena callaensis Hildebrand: *Adenocephalus pacificus* (L), *Diphyllobothrium* sp. (L).

Sciaena deliciosa (Tschudi): *Adenocephalus pacificus* (L), *Callitetrarhynchus gracilis* (L), *Dasyrhynchus pacificus* (L), *Diphyllobothrium* sp. (L), *Nybelinia* sp. (L)

Umbrina canosai Berg: *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia nipponica* (L), *Nybelinia bisulcata* (?) (L), *Nybelinia* sp. (L), *Pterobothrium heteracanthum* (L), *Pterobothrium* sp. (L)

Umbrina coroides Cuvier: unidentified cestode

Sciaenidae gen. sp.: unidentified trypanorhynch

Family Scombridae

Auxis thazard (Lacépède): unidentified trypanorhynch

Euthynnus alletteratus (Rafinesque): *Callitetrarhynchus gracilis* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Katsuwonus pelamis (Linnaeus): *Tentacularia coryphaenae* (L), '*Scolex* spp.' (L)

Sarda chiliensis (Cuvier): *Adenocephalus pacificus* (L), *Acanthobothrium chilense*, *Nybelinia* sp. (L), *Rhodobothrium mesodesmatum* (L), *Sphyriocephalus tergestinus* (L), '*Scolex* spp.' (L)

Scomber colias Gmelin: *Rhinebothrium* sp. (L), unidentified trypanorhynch, unidentified cestode, '*Scolex* spp.' (L)

Scomber japonicus Houttuyn: *Diphyllobothrium* sp. (L), *Hepatoxylon trichiuri* (L), *Nybelinia* sp. (L), *Tentacularia coryphaenae* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Scomberomorus brasiliensis Collette, Russo & Zavala-Camin: *Callitetrarhynchus gracilis* (L), *Nybelinia* sp. (L), *Otobothrium cysticum* (L), *Pseudolacistorhynchus noodti* (L), '*Scolex* spp.' (L)

Scomberomorus cavalla (Cuvier): *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Pterobothrium crassicole* (L), *Tentacularia coryphaenae* (L)

Scomberomorus sierra Jordan & Starks: *Adenocephalus pacificus* (L), *Pseudogrillotia peruviana* (L)

Stellifer minor (Tschudi): '*Scolex* spp.' (L)

Family Serranidae

Acanthistius brasilianus (Cuvier): *Grillotia* (*Christianella*) *carvajalregorum* (L)

Dules auriga Cuvier: *Grillotia* (*Christianella*) *carvajalregorum* (L), unidentified trypanorhynch, unidentified cestode

Epinephelus morio (Valenciennes): unidentified trypanorhynch

***Epinephelus* sp.**: *Pterobothrium* sp. (L), unidentified trypanorhynch

Hemilutjanus macrophthalmos (Tschudi): *Callitetrarhynchus gracilis* (L)

Hyporthodus niveatus (Valenciennes): *Callitetrarhynchus gracilis* (L), *Pseudotobothrium dipsacum* (L), unidentified trypanorhynch

Mycteroperca bonaci (Poey): *Pterobothrium* sp. (L), unidentified trypanorhynch

Family Sparidae

Pagrus pagrus (Linnaeus): *Callitetrarhynchus gracilis* (L), *Pterobothrium* sp. (L), *Otobothrium cysticum* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Sparidae gen. sp.: *Pterobothrium macrourum* (L)

Family Sphyraenidae

Sphyraena guachancho Cuvier: *Callitetrarhynchus gracilis* (L), *Heteronybelinia estigmena* (L), *Otobothrium cysticum* (L)

Family Trichiuridae

Trichiurus lepturus Linnaeus: *Callitetrarhynchus gracilis* (L), *Pterobothrium interruptum* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Family Tripterygiidae

Helcogrammoides chilensis (Cancino): unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Order Pleuronectiformes

Family Achiridae

Trinectes maculatus (Bloch & Schneider): unidentified cestode

Family Paralichthyidae

Citharichthys spilopterus Günther: *Pterobothrium crassicole* (L), *P. kingstoni* (L)

Hippoglossina macrops Steindachner: *Neobothriocephalus* sp., *Nybelinia surmenicola* (L), *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Paralichthys adspersus (Steindachner): *Adenocephalus pacificus* (L), *Lacistorhynchus dollfusi* (L), *Neobothriocephalus* sp., *Nybelinia surmenicola* (L), *Nybelinia* sp. (L), unidentified bothriocephalidean, '*Scolex* spp.' (L)

Paralichthys isosceles Jordan: *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Diphyllobothrium* sp. (L), *Heteronybelinia nipponica* (L), *Nybelinia lingualis* (L), *Nybelinia* sp. (L), *Otobothrium* sp. (L), *Pterobothrium crassicole* (L), *P. heteracanthum* (L), *Pterobothrium* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L), unidentified cestode

Paralichthys microps (Günther): *Neobothriocephalus* sp., *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Paralichthys orbignyanus (Valenciennes): *Grillotia* sp. (L), '*Scolex* spp.' (L)

Paralichthys patagonicus Jordan: *Anonchocephalus patagonicus*, *Callitetrarhynchus gracilis* (L), *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia nipponica* (L), *Lacistorhynchus tenuis* (L), *Nybelinia erythraea* (L), *N. lingualis* (L), *Nybelinia* sp. (L), *Pterobothrium crassicole* (L), '*Scolex* spp.' (L)

***Paralichthys* sp.**: *Pterobothrium* sp. (L)

Xystreurys rasile (Jordan): *Anonchocephalus argentinensis*, *Grillotia* (*Christianella*) *carvajalregorum* (L), *Heteronybelinia nipponica* (L), *Nybelinia erythraea* (L), *N. lingualis* (L), *Nybelinia* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L), unidentified cestode

Family Pleuronectidae
Oncopterus darwinii Steindachner: *Nybelinia lingualis* (L)

Order Salmoniformes
Family Salmonidae
Oncorhynchus kisutch (Walbaum): *Diphyllobothrium dendriticum* (L), *Diphyllobothrium* sp. (L), '*Scolex* spp.' (L)
Oncorhynchus mykiss (Walbaum): *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L), unidentified bothriocephalidean, unidentified phyllobothriidean
Oncorhynchus tshawytscha (Walbaum): *Hepatoxylon trichiuri* (L)
Salmo salar Linnaeus: *Diphyllobothrium dendriticum* (L)
Salmo trutta Linnaeus: *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L), '*Scolex* spp.' (L)
Salvelinus fontinalis (Mitchill): *Diphyllobothrium dendriticum* (L), *D. latum* (L), *Diphyllobothrium* sp. (L), unidentified bothriocephalidean

Order Scorpaeniformes
Family Dactylopteridae
Dactylopterus volitans (Linnaeus): *Nybelinia* sp. (L), '*Scolex* spp.' (L)

Family Normanichthyidae
Normanichthys crockeri Clark: '*Scolex* spp.' (L)

Family Scorpaenidae
***Scorpaena* sp.**: *Pterobothrium crassicole* (L)

Family Sebastidae
Helicolenus lengerichi Norman: *Bothriocephalus* sp., *Hepatoxylon* sp. (L), unidentified trypanorhynch
Sebastes capensis (Gmelin): *Diphyllobothrium* sp. (L), *Hepatoxylon trichiuri* (L), unidentified diphyllidean, '*Scolex* spp.' (L)

Family Triglidae
Prionotus nudigula Ginsburg: *Grillotia* (*Christianella*) *carvajalregorum* (L), *Grillotia* sp. (L)
Prionotus punctatus (Bloch): *Grillotia* (*Christianella*) *carvajalregorum* (L), *Nybelinia* sp. (L)
***Prionotus* sp.**: *Pterobothrium* sp. (L), unidentified phyllobothriidean

Order Siluriformes
Family Ariidae
Ariopsis seemanni (Günther): *Adenocephalus pacificus* (L)

Aspistor luniscutis (Valenciennes): *Pterobothrium crassicole* (L), '*Scolex* spp.' (L)
Bagre bagre (Linnaeus): unidentified trypanorhynch
Bagre marinus (Mitchill): *Pterobothrium crassicole* (L)
Galeichthys peruvianus Lütken: *Adenocephalus pacificus* (L)
Genidens barbus (Lacépède): *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Nomimoscolex arandasregoi*, *Pterobothrium crassicole* (L), unidentified trypanorhynch, '*Scolex* spp.' (L)
Genidens genidens (Cuvier): *Nomimoscolex arandasregoi*
***Genidens* sp.**: *Nomimoscolex arandasregoi*, unidentified trypanorhynch

Family Auchenipteridae
Ageneiosus inermis (Linnaeus): *Ageneiella brevifilis*, *Ageneiella* sp., *Gibsoniela mandube*, *G. meursaulti*, *Luciaella ivanovae*, unidentified cestode
Ageneiosus militaris Valenciennes: *Ageneiella brevifilis*, *Gibsoniela meursaulti*
Ageneiosus pardalis Lütken: *Corallotaenia* sp. (?), *Goezeella danbrooksi*
***Ageneiosus* sp.**: *Gibsoniela mandube*
Auchenipterus nigripinnis (Boulenger): unidentified proteocephalid
Auchenipterus osteomystax (Miranda Ribeiro): *Endorchis auchenipteri*
Tocantinsia piresi (Miranda Ribeiro): *Frezella vaucheri*
Trachelyopterus galeatus (Linnaeus): *Cangatiella arandasi*, *Nupelia tomasi*
Trachelyopterus striatulus (Steindachner): *Endorchis* sp.
Trachelyopterus* cf. *striatulus: *Nupelia tomasi*

Family Callichthyidae
Callichthys callichthys (Linnaeus): *Margaritaella gracilis*
Corydoras atropersonatus Weitzman & Nijssen: unidentified proteocephalid
Corydoras reticulatus Fraser-Brunner: unidentified proteocephalid
Corydoras sychri Weitzman: unidentified proteocephalid
Hoplosternum littorale (Hancock): *Valipora campylancristrota* (L)

Family Cetopsidae
Cetopsis coecutiens (Lichtenstein): *Brooksiella praeputialis*, *Goezeella siluri*, unidentified cestode
Cetopsis othonops (Eigenmann): *Brooksiella praeputialis*, *Goezeella siluri*

Family Diplomystimidae
Diplomystes camposensis Arratia: *Diphyllobothrium latum* (L)
Olivaichthys viedmensis (MacDonagh): *Nomimoscolex semenasae*

Family Doradidae
Franciscodoras marmoratus (Lütken): *Proteocephalus renauldi*, *Proteocephalus* sp.
Megalodoras uranoscopus (Eigenmann & Eigenmann): *Proteocephalus kuyukuyu*
Oxydoras kneri Bleeker: *Proteocephalus hobergi*, unidentified proteocephalid

Oxydoras niger (Valenciennes): *Proteocephalus hobergi*, *P. kuyukuyu*
Platydoras costatus (Linnaeus): *Proteocephalus renaudi* (?), *P. soniae* (?), *Proteocephalus* sp. (?)
Pterodoras granulosus (Valenciennes): *Monticellia belavistensis*, *Proteocephalus kuyukuyu*, *Proteocephalus* sp.
***Pterodoras* sp.**: *Proteocephalus kuyukuyu*

Family Heptapteridae
Goeldiella eques (Müller & Troschel): *Nupelia* sp.
Pimelodella gracilis (Valenciennes): unidentified proteocephalid, unidentified cestode
Pimelodella lateristriga (Lichtenstein): unidentified cestode
Rhamdia quelen (Quoy & Gaimard): *Lenhataenia megacephala*, *Proteocephalus bagri*, *P. rhamdiae*, *Travassiella jandia*, unidentified proteocephalid

Family Loricariidae
Hypostomus* cf. *ternetzi (Boulenger): unidentified proteocephalid
Loricariichthys platymetopon Isbrücker & Nijssen: unidentified proteocephalid
***Loricariichthys* sp.**: unidentified cestode
***Paraloricaria* sp.**: *Proteocephalus pilarensis*

Family Pimelodidae
Brachyplatystoma capapretum Lundberg & Akama: *Amazotaenia yvettae, Endorchis piraeeba*
Brachyplatystoma filamentosum (Lichtenstein): *Amphoteromorphus ninoi*, *A. piraeeba*, *Nomimoscolex piraeeba*, *N. suspectus*, unidentified proteocephalid
Brachyplatystoma* cf. *filamentosum: *Amphoteromorphus ovalis*, *Endorchis piraeeba*, *Nomimoscolex suspectus*
Brachyplatystoma rousseauxii (Castelnau): *Amphoteromorphus peniculus*, *A. piriformis*, *Nomimoscolex dorad*, *N. piraeeba*, *N. suspectus*, *Nomimoscolex* sp., *Pterobothrium crassicole* (L)
Brachyplatystoma vaillantii (Valenciennes): *Amazotaenia yvettae*, *Amphoteromorphus ninoi*, *Chambriella* sp., *Goezeella siluri*, *Harriscolex piramutab*, *Nomimoscolex suspectus*, *Pterobothrium crassicole* (L)
***Brachyplatystoma* sp.**: *Amphoteromorphus ovalis*, '*Scolex* spp.' (L), unidentified cestode
Calophysus macropterus (Lichtenstein): *Monticellia amazonica*, *Rudolphiella piracatinga*, unidentified cestode
Hemisorubim platyrhynchos (Valenciennes): *Chambriella paranaensis*, *Manaosia bracodemoca*, *Mariauxiella piscatorum*, *Spatulifer maringaensis*, unidentified cestode
Iheringichthys labrosus (Lütken): unidentified proteocephalid
Luciopimelodus pati (Valenciennes): *Monticellia ventrei*, *Nomimoscolex* sp., *Proteocephalus fossatus*, *Rudolphiella lobosa* (?), *R. szidati*, *Rudolphiella* sp.
Megalonema platanum (Günther): *Monticellia santafesina*, *Rudolphiella lobosa*, unidentified proteocephalid

Megalonema platycephalum Eigenmann: *Monticellia santafesina*

Phractocephalus hemioliopterus (Schneider): *Chambriella* sp., *Ephedrocephalus microcephalus*, *Proteocephalus hemioliopteri*, *Proteocephalus* sp., *Pseudocrepidobothrium eirasi*, *P. ludovici*, *Pseudocrepidobothrium* sp., *Scholzia emarginata*, *Zygobothrium megacephalum*, Monticelliinae gen. sp., unidentified proteocephalid

Pimelodus albicans (Valenciennes): *Monticellia magna*, *Nomimoscolex microacetabula*

Pimelodus altissimus Eigenmann & Pearson: *Endorchis* sp.

Pimelodus argenteus Perugia: *Monticellia magna*

Pimelodus blochii Valenciennes: *Nomimoscolex alovarius*, *Proteocephalus* sp.

Pimelodus* cf. *blochii: *Monticellia magna* (?)

Pimelodus maculatus Lacépède: *Chambriella agostinhoi*, *Monticellia magna*, *Nomimoscolex microacetabula*, *Nomimoscolex* sp., *Proteocephalus pimelodi*, *Proteocephalus* sp., *Valipora* sp. (L), unidentified proteocephalid, unidentified cestode

Pimelodus* cf. *maculatus: *Endorchis* sp., *Monticellia magna*

Pimelodus ornatus Kner: *Mariauxiella pimelodi*, *Nomimoscolex microacetabula*, *Nomimoscolex* sp., *Spasskyellina mandi*, *Spasskyellina* sp.

Pimelodus ortmanni Haseman: unidentified cestode

Pimelodus pohli Ribeiro & Lucena: unidentified proteocephalid

***Pimelodus* sp.**: *Mariauxiella pimelodi*, *Pterobothrium crassicole* (L), unidentified proteocephalid

Pinirampus pirinampu (Spix & Agassiz): *Goezeella siluri*, *Monticellia ventrei*, *Monticellia* sp., *Nomimoscolex admonticellia*, *Proteocephalus vladimirae*, *Rudolphiella myoides*, *R. piranabu*, *Rudolphiella* sp., unidentified proteocephalid, unidentified cestode

Platynematichthys notatus (Jardine): *Brayela karuatayi*

Pseudoplatystoma corruscans (Agassiz): *Choanoscolex abscisus*, *Harriscolex kaparari* (?), *H. nathaliae*, *Megathylacus travassosi*, *Megathylacus* sp., *Monticellia* sp., *Nomimoscolex pertierrae*, *Nomimoscolex sudobim* (?), *Nomimoscolex* sp., *Peltidocotyle rugosa*, *Spasskyellina spinulifera*, unidentified proteocephalid

Pseudoplatystoma fasciatum (Linnaeus) (*sensu lato*): *Chambriella* sp., *Choanoscolex abscisus*, *Choanoscolex* sp., *Euzetiella tetraphylliformis*, *Harriscolex kaparari*, *Houssayela sudobim*, *Megathylacus travassosi*, *Megathylacus* sp., *Monticellia* sp., *Nomimoscolex lopesi*, *N. sudobim*, *Nomimoscolex* sp., *Peltidocotyle rugosa*, *Pseudocrepidobothrium chanaorum*, *Regoella brevis*, *Spasskyellina spinulifera*, *Spasskyellina* sp., *Spatulifer rugosa*, unidentified proteocephalid, unidentified cestode

Pseudoplatystoma tigrinum (Valenciennes): *Choanoscolex* sp., *Harriscolex kaparari*, *Nomimoscolex sudobim*, *Spasskyellina spinulifera*, *Spatulifer surubim*, *Spatulifer* sp.

***Pseudoplatystoma* sp.**: *Proteocephalus platystomi*, unidentified cestode

Sorubim lima (Bloch & Schneider): *Manaosia bracodemoca*, *Nupelia portoriquensis*, *Spasskyellina spinulifera*, *Spatulifer maringaensis*

Sorubimichthys planiceps (Spix & Agassiz): *Chambriella* sp., *Choanoscolex* sp., *Lenhataenia megacephala*, *Nomimoscolex lenha*, *Peltidocotyle lenha*, *Spasskyellina lenha*

Steindachneridion parahybae (Steindachner): unidentified cestode

Zungaro jahu (Ihering): *Chambriella agostinhoi*, *Choanoscolex abscisus*, *Euzetiella tetraphylliformis*, *Jauella glandicephalus*, *Megathylacus jandia*, *Peltidocotyle lenha*, *P. rugosa*, *Peltidocotyle* sp., *Travassiella jandia*, unidentified cestode

Zungaro zungaro (Humboldt): *Amphoteromorphus parkamoo*, *Chambriella agostinhoi*, *Euzetiella tetraphylliformis*, *Jauella glandicephalus*, *Megathylacus jandia*, *Peltidocotyle lenha*, *Proteocephalus sophiae*, *Travassiella jandia*

Family Pseudopimelodidae

Pseudopimelodus mangurus (Valenciennes): *Peltidocotyle rugosa*

Family Trichomycteridae

Trichomycterus punctulatus Valenciennes: *Hepatoxylon megacephalum* (L)

Unidentified siluriform fish

'*Silurus dorgado*': *Monticellia diesingii*

'*Silurus megacephalus*': *Monticellia macrocotylea*

Siluriform fish: *Pterobothrium* sp. (L)

Order Tetraodontiformes

Family Balistidae

Balistes capriscus Gmelin: *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Callitetrarhynchus* sp. (L), *Nybelinia* sp. (L), unidentified 'pseudophyllidean', '*Scolex* spp.' (L)

Balistes vetula Linnaeus: *Callitetrarhynchus gracilis* (L), *C. speciosus* (L), *Callitetrarhynchus* sp. (L), *Otobothrium* sp. (L), unidentified trypanorhynch, '*Scolex* spp.' (L)

Family Molidae

Masturus lanceolatus Liénard: unidentified trypanorhynch

Mola mola (Linnaeus): *Anchistrocephalus microcephalus*, *Molicola* sp. (L), unidentified trypanorhynch

Mola ramsayi (Giglioli): *Anchistrocephalus microcephalus*, *Molicola horridus* (L), *Nybelinia* sp. (L)

Family Monacanthidae

Aluterus monoceros (Linnaeus): *Callitetrarhynchus speciosus* (L), *Floriceps saccatus* (L)

Stephanolepis hispidus (Linnaeus): *Callitetrarhynchus speciosus* (L)

Family Tetraodontidae

Lagocephalus laevigatus (Linnaeus): unidentified cestode

Sphoeroides testudineus (Linnaeus): unidentified cestode

Class Chondrichthyes
Subclass Elasmobranchii
Order Carcharhiniformes
Family Carcharhinidae
Carcharhinus brachyurus (Günther): *Cathetocephalus australis*, *Dasyrhynchus pacificus*
Carcharhinus leucas (Müller & Henle): *Cathetocephalus thatcheri*, *Poecilancistrium caryophyllum*
Carcharhinus limbatus (Müller & Henle): *Dasyrhynchus pacificus*, unidentified trypanorhynch
Carcharhinus longimanus (Poey): *Anthobothrium laciniatum*, *Disculiceps galapagoensis*, *Paraorygmatobothrium filiforme*, *Tentacularia coryphaenae*
Carcharhinus obscurus (Lesueur): *Tentacularia coryphaenae*
Carcharhinus porosus (Ranzani): *Disculiceps pileatus*
Carcharhinus signatus (Poey): *Grillotia* (*Christianella*) *carvajalregorum*, *Heteronybelinia nipponica* (L), *H. yamagutii* (L)
Prionace glauca (Linnaeus): *Callitetrarhynchus gracilis* (L), *Floriceps saccatus*, *Hepatoxylon trichiuri* (L), *Molicola horridus*, *Paraorygmatobothrium angustum*, *P. prionacis*, *Platybothrium auriculatum*, *Prosobothrium armigerum*, *Tentacularia coryphaenae*
Rhizoprionodon lalandii (Müller & Henle): *Poecilancistrium caryophyllum*
Rhizoprionodon terraenovae (Richardson): *Nybelinia fayapaulazariahi*, unidentified trypanorhynch

Family Scyliorhinidae
Scyliorhinus besnardi Springer & Sadowsky: *Ahamulina catarina*
Scyliorhinus haeckelii (Miranda Ribeiro): *Dasyrhynchus pacificus*

Family Sphyrnidae
Sphyrna lewini (Griffith & Smith): *Heteronybelinia nipponica*
Sphyrna zygaena (Linnaeus): *Callitetrarhynchus speciosus* (L), *Heteronybelinia nipponica* (L), *Platybothrium parvum*, *Platybothrium* sp., *Thysanocephalum thysanocephalum*
***Sphyrna* sp.**: *Dasyrhynchus pacificus*

Family Triakidae
Galeorhinus galeus (Linnaeus): *Anthobothrium galeorhini*
Mustelus canis (Mitchill): *Callitetrarhynchus gracilis*, *Dollfusiella vooremi*, *Nybelinia lingualis* (L)
Mustelus fasciatus (Garman): *Orygmatobothrium juani*
Mustelus mento Cope: *Calliobothrium verticillatum*, *Calliobothrium* sp., *Dollfusiella musteli*, *Orygmatobothrium musteli*, *Orygmatobothrium* sp., *Paraorygmatobothrium triacis*, *Phyllobothrium lactuca*, *Phyllobothrium* sp., *Scyphophyllidium uruguayense*
Mustelus schmitti Springer: *Calliobothrium australis*, *Coronocestus notoguidoi*, *Dollfusiella vooremi*, *Gyrocotyle maxima* (?), *Nybelinia lingualis*, *Orygmatobothrium schmittii*, *Symcallio barbarae*, *S. lunae*

Mustelus whitneyi Chirichigno: *Orygmatobothrium musteli*, *Paraorygmatobothrium triacis*, *Symcallio lintoni*
Triakis maculata Kner & Steindachner: *Lacistorhynchus tenuis*

Order Hexanchiformes
Family Hexanchidae
Heptranchias perlo (Bonnaterre): *Grillotia* (*Christianella*) *carvajalregorum* (L)
Hexanchus griseus (Bonnaterre): *Crossobothrium dohrni*, *C. laciniatum*, *Grillotia heptanchi*, *Phyllobothrium sinuosiceps*
Notorynchus cepedianus (Péron): *Crossobothrium antonioi*, *C. pequeae*, *Heteronybelinia perideraeus*

Order Lamniformes
Family Alopiidae
Alopias vulpinus (Bonnaterre): *Paraorygmatobothrium angustum*

Family Cetorhinidae
Cetorhinus maximus (Gunnerus): *Reesium paciferum*

Family Lamnidae
Carcharodon carcharias (Linnaeus): *Tentacularia coryphaenae*, unidentified trypanorhynch
Isurus oxyrinchus Rafinesque: *Gymnorhynchus isuri*, *Molicola horridus*, *Nybelinia lingualis*

Order Myliobatiformes
Family Dasyatidae
Dasyatis americana Hildebrand & Schroeder: *Acanthobothrium americanum*, *Anthocephalum gracile*, *A. kingae*, *Lecanicephalum peltatum*, *Polypocephalus medusia*, *Rhinebothrium corymbum*, *R. margaritense*, *Rhodobothrium pulvinatum*, *Scalithrium magniphallum*
Dasyatis dipterura (Jordan & Gilbert): *Acanthobothroides peruensis*, *A. thorsoni*
Dasyatis guttata (Bloch & Schneider): *Acanthobothrium tasajerasi*, *A. urotrygoni*, *Acanthobothroides thorsoni*, *Rhinebothrium margaritense*, *Rhodobothrium pulvinatum*, *Scalithrium magniphallum*
Dasyatis longa (Garman): *Acanthobothrium campbelli*, *A. costarricense*, *A. obuncum*
Himantura schmardae (Werner): *Acanthobothrium himanturi*, *A. tasajerasi*, *Acanthobothroides thorsoni*, *Anindobothrium anacolum*, *Parachristianella monomegacantha*, *Rhinebothrium tetralobatum*, *Scalithrium magniphallum*

Family Gymnuridae
Gymnura afuerae (Hildebrand): *Acanthobothrium atahualpai*
Gymnura micrura (Bloch & Schneider): *Acanthobothrium fogeli*
***Gymnura* sp.**: *Pterobothrium* sp.

Family Myliobatidae

Aetobatus narinari (Euphrasen): *Acanthobothrium colombianum*, *A. monksi*, *A. tortum* *Disculiceps* sp. (?)

Myliobatis chilensis Philippi: *Acanthobothrium batailloni*, *A. coquimbense*, *A. holorhini*, *Acanthobothrium* sp., *Caulobothrium myliobatidis*, *Phyllobothrium auricula*, *Rhodobothrium mesodesmatum*

Myliobatis goodei Garman: *Aberrapex arrhynchum*, *Acanthobothrium* sp., *Caulobothrium ostrowskiae*, *C. uruguayense*, *Halysioncum megacanthum*, *Mecistobothrium oblongum*, *Parachristianella damiani*, *Phyllobothrium myliobatidis*, *Phyllobothrium* sp.

Myliobatis peruvianus Garman: *Acanthobothrium brevissime*, *A. gonzalesmugaburoi*, *Phyllobothrium auricula*, *Rhodobothrium mesodesmatum*

Rhinoptera bonasus (Mitchill): *Dioecotaenia campbelli*, *Rhinoptericola megacantha*, *Rhodobothrium paucitesticulare*, *Tylocephalum brooksi*, *Tylocephalum* sp.

Rhinoptera brasiliensis Müller: *Rhinoptericola megacantha*

Rhinoptera steindachneri Evermann & Jenkins: *Serendip deborahae*

Family Potamotrygonidae

Paratrygon aiereba (Müller & Henle): *Acanthobothrium terezae*, *Nandocestus guariticus*, *Potamotrygonocestus fitzgeraldae*, *P. travassosi*, *Potamotrygonocestus* sp., *Rhinebothrium brooksi*, *R. copianullum*, *Rhinebothrium* sp., *Rhinebothroides scorzai*, *Rhinebothroides* sp.

Plesiotrygon iwamae Rosa, Castello & Thorson: *Potamotrygonocestus chaoi*, *P. marajoara*

Potamotrygon brachyura (Günther): *Rhinebothrium paratrygoni*

Potamotrygon constellata (Vaillant): *Acanthobothrium amazonense*, *Potamotrygonocestus amazonensis*, *P. travassosi*, *Rhinebothroides freitasi*

Potamotrygon falkneri Castex & Maciel: *Acanthobothrium regoi*, *Paroncomegas araya*, *Potamotrygonocestus amazonensis*, *P. travassosi*, *Rhinebothrium paratrygoni*

Potamotrygon* cf. *falkneri: *Acanthobothrium peruviense*, *Nandocestus guariticus*, *Paroncomegas araya*, *Potamotrygonocestus fitzgeraldae*, *Potamotrygonocestus* sp., *Rhinebothroides freitasi*, *Rhinebothroides* sp.

Potamotrygon henlei (Castelnau): *Potamotrygonocestus* sp., *Rhinebothrium copianullum*, *Rhinebothroides freitasi*, *R. glandularis*

Potamotrygon histrix (Müller & Henle): *Rhinebothrium paratrygoni*

Potamotrygon leopoldi Castex & Castello: *Potamotrygonocestus fitzgeraldae*, *Rhinebothrium copianullum*, *Rhinebothroides freitasi*

Potamotrygon magdalenae (Duméril): *Acanthobothrium quinonese*, *Potamotrygonocestus magdalenensis*, *Rhinebothroides moralarai*

Potamotrygon motoro (Müller & Henle): *Acanthobothrium peruviense*, *A. ramiroi*, *A. regoi*, *A. terezae*, *Paroncomegas araya*, *Potamotrygonocestus amazonensis*, *P. fitzgeraldae*, *P. travassosi*, *Potamotrygonocestus* sp., *Rhinebothrium copianullum*, *R. corbatai*, *R. mistyae*, *R. paratrygoni*, *Rhinebothroides campbelli*, *R. freitasi*, *R. glandularis*, *R. mclennanae*, *R. scorzai*, unidentified cestode

Potamotrygon orbignyi (Castelnau): *Acanthobothrium regoi*, *Anindobothrium lisae*, *Paroncomegas araya*, *Paroncomegas* sp., *Potamotrygonocestus amazonensis*, *P. fitzgeraldae*, *P. maurae*, *P. travassosi*, *Rhinebothrium brooksi*, *R. copianullum*, *R. fulbrighti*, *R. jaimei*, *R. paratrygoni*, *Rhinebothroides freitasi*, *R. glandularis*, *R. scorzai*, unidentified cestode

Potamotrygon schroederi Fernández-Yépez: *Potamotrygonocestus* sp., *Rhinebothrium copianullum*, *Rhinebothroides freitasi*

Potamotrygon scobina Garman: *Potamotrygonocestus amazonensis*, *Rhinebothrium jaimei*, *Rhinebothroides freitasi*, *R. glandularis*

Potamotrygon signata Garman: *Rhinebothroides glandularis*

Potamotrygon tatianae Silva & Carvalho: *Rhinebothrium copianullum*, *Rhinebothroides* sp.

Potamotrygon yepezi Castex & Castello: *Acanthobothrium quinonese*, *Potamotrygonocestus amazonensis*, *Rhinebothroides freitasi*

***Potamotrygon* sp.**: *Paroncomegas araya*, *Potamotrygonocestus amazonensis*, *Rhinebothrium copianullum*, *R. fulbrighti*, *R. paratrygoni*, *Rhinebothroides glandularis*, *R. moralarai*, *R. scorzai*

Family Urotrygonidae

Urobatis jamaicensis (Cuvier): *Acanthobothrium cartaginense*, *Anthocephalum kingae*, *Scalithrium magniphallum*

Urobatis tumbesensis (Chirichigno & McEachran): *Acanthobothrium minusculum*, *Anthocephalum hobergi*

Urotrygon venezuelae Schultz: *Acanthobothrium urotrygoni*, *Scalithrium magniphallum*

Order Pristiformes

Family Pristidae

Pristis pristis (Linnaeus): *Anthobothrium pristis*, *Pterobothrium fragile*

Order Rajiformes

Family Arhynchobatidae

Atlantoraja castelnaui (Miranda Ribeiro): *Acanthobothrium marplatense*, *Dollfusiella acuta*, *Notomegarhynchus navonae*

Atlantoraja platana (Günther): *Dollfusiella acuta*

Bathyraja brachyurops (Fowler): *Guidus argentinense*

Bathyraja magellanica (Philippi): *Grillotia* sp.

Psammobatis bergi Marini: *Dollfusiella taminii*

Psammobatis rudis Günther: *Grillotia* (*Grillotia*) *patagonica*

Psammobatis scobina (Philippi): *Acanthobothrium psammobati*, *Rhinebothrium scobinae*

Sympterygia acuta Garman: *Dollfusiella acuta*

Sympterygia bonapartii Müller & Henle: *Dollfusiella acuta*, *D. vooremi* (?), *Grillotia* (*Grillotia*) *erinaceus* (?), *Heteronybelinia mattisi*, *Nybelinia lingualis*, *Nybelinia* sp., *Phyllobothrium* sp., *Rhinebothrium chilensis*

Sympterygia brevicaudata (Cope): *Acanthobothrium lusarmientoi*, *A. psammobati*, *Acanthobothrium* sp.
Sympterygia lima (Poeppig): *Halysioncum euzeti*, *Rhinebothrium chilensis*, *R. leiblei*

Family Rajidae
Dipturus flavirostris (Philippi): *Echeneibothrium megalosoma*, *E. multiloculatum*, *E. williamsi*, *Grillotia* (*Grillotia*) *dollfusi*, *Phyllobothrium* sp.
Dipturus trachyderma (Krefft & Stehmann): *Mixonybelinia beveridgei* (L), *Paragrillotia* sp. (?), *Phyllobothrium lactuca* (?)
Zearaja chilensis (Guichenot): *Acanthobothrium annapinkiense*, *Echeneibothrium megalosoma*, *E. multiloculatum*, *E. williamsi*, *Grillotia* (*Grillotia*) *dollfusi*

Family Rhinobatidae
Rhinobatos percellens (Walbaum): unidentified trypanorhynch
Rhinobatos planiceps Garman: *Acanthobothrium olseni*, *A. robustum*, *Parachristianella monomegacantha*, *Prochristianella heteracantha, Rhinebothrium rhinobati*
Zapteryx brevirostris (Müller & Henle): *Acanthobothrium zapterycum*, *Acanthobothrium* sp., *Halysioncum pigmentatum*, *Phyllobothrium* sp. (L)

Order Squaliformes
Family Etmopteridae
Etmopterus granulosus (Günther): *Gilquinia squali*

Family Somniosidae
Somniosus pacificus Bigelow & Schroeder: *Hepatoxylon trichiuri* (L)

Family Squalidae
***Squalus* sp.**: *Grillotia* (*Christianella*) *carvajalregorum*

Order Squatiniformes
Family Squatinidae
Squatina armata (Philippi): *Grillotia* sp.
Squatina guggenheim Marini: *Grillotia* (*Christianella*) *carvajalregorum*, *Paraberrapex atlanticus*

Order Torpediniformes
Family Narcinidae
Discopyge tschudii Heckel: *Phyllobothrium discopygi*
Narcine brasiliensis (Olfers): *Acanthobothrium electricolum*, *A. lintoni*

Unidentified ray order
Unidentified ray: *Acanthobothrium dasybati* (?), *Pterobothrium* sp.

Subclass Holocephali
Order Chimaeriformes
Family Callorhinchidae
Callorhinchus callorynchus (Linnaeus): *Gyrocotyle maxima*, *G. rugosa*

Results and discussion

The database compiled from the available literature on fish cestodes in South America comprises records of 297 species recognized as valid as well as unidentified ones included in 120 genera and 32 families, associated with 401 cartilaginous and bony fish hosts (Tables 1, 2). Among the recognized 19 orders of tapeworms, 13 have been found in marine and freshwater systems in South America (excluding the doubtful reports of the Caryophyllidea). The recently erected order Onchoproteocephalidea, which accommodates several taxa previously placed in the tetraphyllidean family Onchobothriidae and the entire former order Proteocephalidea, is the most diverse group, being represented by 148 species in 43 genera.

The tapeworm with the widest spectrum of definitive hosts is *Rhinebothroides freitasi* (Rhinebothriidea) that parasitizes nine species of stingrays of the genus *Potamotrygon,* even though it exhibits only a mesostenoxenous specificity, i.e. occurrence limited to a single host genus. Conversely, members of five orders, namely Amphilinidea, Cathetocephalidea, Diphyllidea, Lecanicephalidea, 'Tetraphyllidea' and most likely Gyrocotylidea (see the checklist records for details), showed only a single fish host (oioxenous specificity). It is also worth noting the usually broad spectrum of intermediate teleost hosts for metacestodes, mainly diphyllobothriideans, 'tetraphyllideans' and trypanorhynchs, which is reflected in the higher number of actinopterygian (315) than chondrichthyean (86) hosts. However, the stingray *Potamotrygon motoro* harbours the highest number of cestodes (17) belonging to the species-rich genera *Acanthobothrium* Blanchard, 1848, *Potamotrygonocestus* Brooks & Thorson, 1976, *Rhinebothrium* Linton, 1890 and *Rhinebothroides* Mayes, Brooks & Thorson, 1981, in addition to *Paroncomegas araya*.

A total of 208 species of tapeworms are found across seven major ecoregions of South American coast (one additional species is found in Galapagos), being the highest species richness reported from WTSP (66) and WTSA (60), whereas 209 species are found throughout six major river basins of South America (Fig. 1). The major number of species comes from the Amazon and Paraná River basins, with 95 and 80 species, respectively. At least four species were reported from particular lakes, mostly parasitizing osmeriforms and salmoniforms in Argentina and Chile.

The number of taxonomic studies has been steadily growing since 1940, but only 16 papers were based on an integrated taxonomy approach, using molecular data as an important tool. The number of general parasitological surveys has also increased since the beginning of the last century, whereas ecological studies have launched the first publications only in the mid-sixties, with a peak in the last sixteen years, noticeably higher than the previous period (Fig. 2).

Table 1. Survey of fish cestodes from South America according to their high taxonomic level classification.

Order	Family	No. of genera	No. of species	Identified to generic level	No. of sequences*
Amphilinidea	Amphilinidae	2	2	0	4
Bothriocephalidea	Bothriocephalidae	4	5	2	12
	Echinophallidae	2	1	2	4
	Triaenophoridae	4	6	1	1
Cathetocephalidea	Cathetocephalidae	1	2	0	0
	Disculicepitidae	1	2	1	0
Cyclophyllidea	Gryporhynchidae	3	2	1	0
Diphyllidea	Echinobothriidae	3	5	0	12
Diphyllobothriidea	Diphyllobothriidae	2	3	1	26
Gyrocotylidea	Gyrocotylidae	1	2	0	0
Lecanicephalidea	Aberrapecidae	1	1	0	0
	Cephalobothriidae	1	1	1	0
	Lecanicephalidae	1	1	0	0
	Paraberrapecidae	1	1	0	0
	Polypocephalidae	1	1	0	0
Onchoproteocephalidea	Onchobothriidae	4	45	3	3
	Prosobothriidae	1	1	0	0
	Proteocephalidae	38	102	15	164
Phyllobothriidea	Phyllobothriidae	7	15	2	7
Rhinebothriidea	Anthocephaliidae	1	3	0	6
	Echeneibothriidae	2	4	0	0
	Rhinebothriidae	4	24	2	57
'Tetraphyllidea'	*incertae sedis*	7	13	1	3
Trypanorhyncha	Eutetrarhynchidae	5	9	2	2
	Gilquiniidae	1	1	1	0
	Gymnorhynchidae	2	2	1	0
	Lacistorhynchidae	9	16	5	0
	Otobothriidae	2	1	1	0
	Pseudotobothriidae	1	1	0	0
	Pterobothriidae	1	4	1	0
	Rhinoptericolidae	1	1	0	0
	Sphyriocephalidae	2	3	1	0
	Tentaculariidae	4	17	3	0
	Total	**120**	**297**	**61**	**301**

*Only sequences of cestodes collected in South America were considered.

Taxonomic resolution

Among the genera of fish cestodes reported from South America, one half was either identified only at generic level or they were specifically identified in some reports and at generic level in others. The numerous papers published in the last 30 years, mostly those ecological ones (see Fig. 2) focused on marine teleost hosts as models, include a high number of records of unidentified larvae. Most of them corresponded to the

Table 2. Survey of fish hosts that harbour cestodes in South America.

Class	Subclass	Order	No. of genera	No. of species	No. of cestodes reported*
ACTINOPTERYGII	Neopterygii	Anguilliformes	2	2	2
		Atheriniformes	2	7	4
		Aulopiformes	1	1	0
		Batrachoidiformes	2	2	4
		Beloniformes	3	3	0
		Characiformes	18	27	8
		Clupeiformes	8	9	2
		Cypriniformes	3	3	1
		Cyprinodontiformes	2	4	2
		Gadiformes	9	14	14
		Gobiesociformes	2	2	0
		Gymnotiformes	2	3	3
		Lampriformes	1	1	1
		Lophiiformes	1	1	2
		Mugiliformes	1	2	1
		Notacanthiformes	1	1	1
		Ophidiiformes	2	5	11
		Osmeriformes	2	4	4
		Osteoglossiformes	1	1	2
		Perciformes	87	125	46
		Pleuronectiformes	6	9	14
		Salmoniformes	3	6	3
		Scorpaeniformes	6	6	3
		Siluriformes	45	68	95
		Tetraodontiformes	7	9	5
CHONDRICHTHYES	Elasmobranchii	Carcharhiniformes**	8	21	44
		Hexanchiformes**	3	3	8
		Lamniformes**	4	4	6
		Myliobatiformes***	11	33	86
		Pristiformes***	1	1	2
		Rajiformes***	8	17	30
		Squaliformes**	3	2	3
		Squatiniformes**	1	2	2
		Torpediniformes***	2	2	3
	Holocephali	Chimaeriformes	1	1	2
		TOTAL	**259**	**401**	**414**

*Cestodes with no specific identification were not counted; **Selachii (sharks); ***Batoidea (rays)

'Tetraphyllidea' named as '*Scolex* spp.', even though these individuals can belong to other orders that were previously included within this catch-all group (Chambers et al. 2000; Jensen and Bullard 2010). Poulin and Leung (2010) stated that the presence of larval stages in a community is inversely proportional to the taxonomic resolution

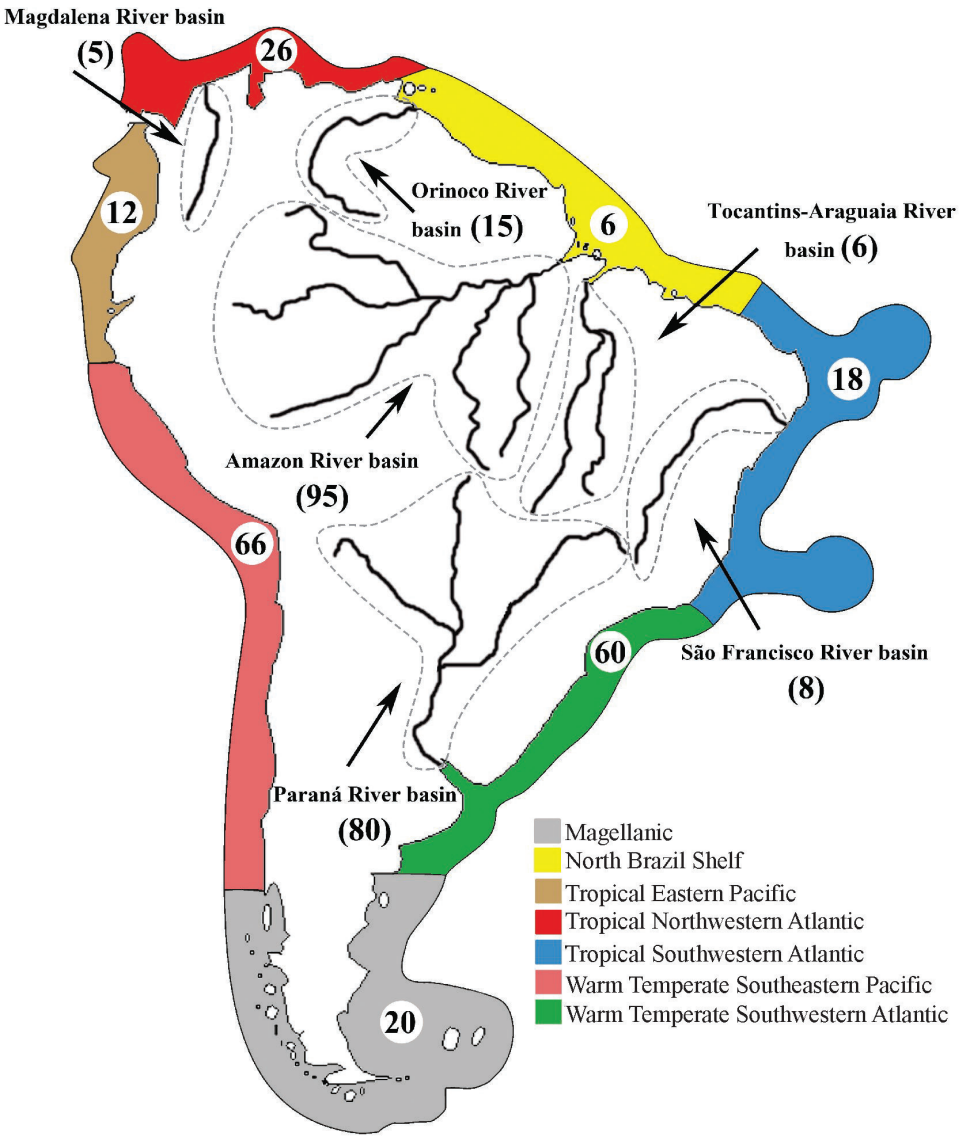

Figure 1. The geographical distribution of tapeworms in South America associated with their fish hosts from the major marine ecoregions of Spalding et al. (2007) and river basins in the continent. Each species may occur in more than one basin or ecoregion.

achieved and those parasites in fish hosts exhibit lower taxonomic resolution than endohelminths parasitizing birds and mammals.

The accurate identification of larval stages of cestodes is usually challenging, because they lack key morphological traits that are present in their adult forms, and studies dealing with their genetic characterization are rare in South America (Rozas et al. 2012). An even more important concern is the high number of records of uni-

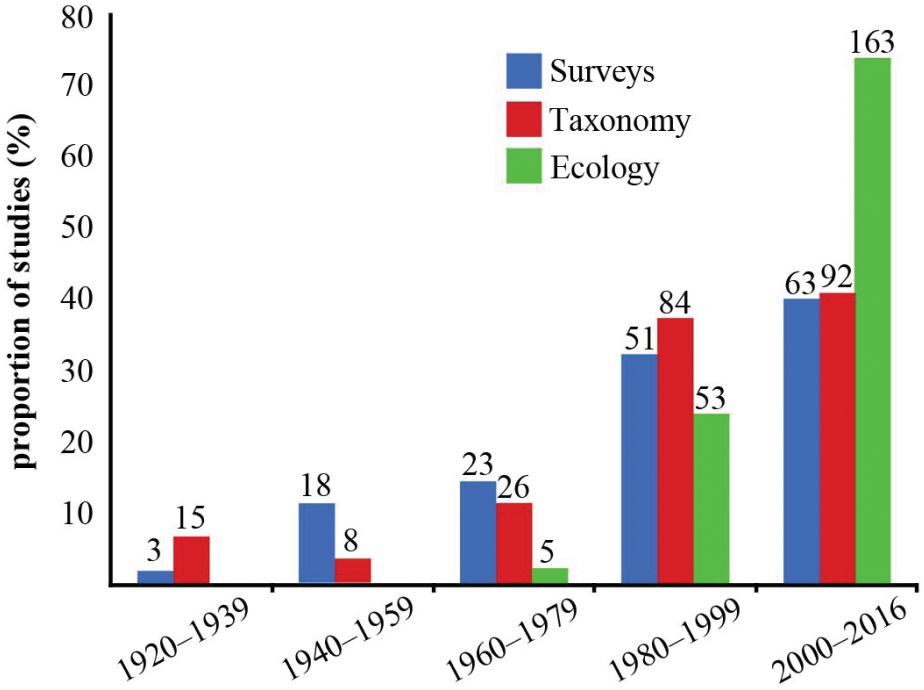

Figure 2. Proportion of articles on the fish cestodes from South America published per intervals of years sorted by categories. The numerals above individual bars indicate the absolute number of articles.

dentified diphyllobothrid plerocercoids in teleosts (see the Parasite-Host list), because these metacestodes can infect humans who consume raw or undercooked fish and may cause a disease known as diphyllobothriosis (Scholz et al. 2009; Kuchta et al. 2015). Larval trypanorhynchs are the only exception, because they may be precisely identified based on their tentacular armature (Palm 2004; Caira and Jensen 2014). For instance, all three valid species of *Pterobothrium* Diesing, 1850 originally described from South America have teleost fishes as type hosts.

One of the main obstacles that hampers our understanding of the diversity of fish cestodes in South America is the deficient knowledge of their life cycles and failure to match the morphologically amorphous or divergent larval forms to their adult stages; to date, no life cycle studies have been undertaken in this continent. Jensen and Bullard (2010) performed the most comprehensive study combining molecular and morphological approaches to elucidate life cycles of marine cestodes from four metazoan phyla in the Gulf of Mexico. They found as many as eight larval types which could be associated with their adult forms and provided a useful morphological key for the 15 recognized types, including larvae of the currently recognized Onchoproteocephalidea, Phyllobothriidea, Rhinebothriidea and 'Tetraphyllidea'.

Unlike the poor taxonomic resolution of marine larvae from teleosts, adult forms, typically those infecting freshwater catfishes (Siluriformes) and potamotrygonid sting-

rays (Potamotrygonidae), have been fairly well-documented (Reyda and Marques 2011; de Chambrier et al. 2015b). Their characterisation using modern descriptive tools, e.g. scanning electron micrographs and molecular data, associated with the traditional morphological approach, deeply contributed to the improvement of their taxonomic resolution and to elucidating the high cestode diversity associated with these groups of hosts.

Elasmobranch and teleost fish hosts

Miloslavich et al. (2011) estimated the fish diversity in five subregions along the South American coast and suggested the occurrence of more than 5000 species in these marine systems. Reis (2013) estimated a value slightly higher for fishes from freshwater drainage systems in South America, *c*. 5400 species. Considering that predictions for estimating the global species richness of parasites suggest that they exceed twice the number of their hosts (Dobson 2008) and that only 4% of the potential fish hosts have been scrutinized for cestodes in South America, it is straightforward to conclude that our knowledge of the diversity of these parasites is far from adequate. Similar results were also found for trematodes infecting freshwater fishes in the same continent (Choudhury et al. 2016) and it may be valid also for others groups of helminths.

Contrasting the generally poorly-known diversity of fish cestodes in South America, some groups of hosts have been extensively studied compared to others. Among the elasmobranch hosts, the stingrays (Myliobatiformes) have been steadily examined for tapeworms, exhibiting the highest proportion of records (39%), which were mainly reported from marine and freshwater systems (e.g. Brooks et al. 1981a, b; Reyda and Marques 2011). Regarding teleosts, members of the order Perciformes are the most representative hosts, representing *c*. 40% of all records among this group. The majority of these studies have been conducted by ecological research teams interested in unravelling the structure of fish parasite communities and, more recently, their use as biological tags for stock discrimination (e.g. Luque et al. 2010; Timi et al. 2010a).

According to Luque and Poulin (2007), the study effort and local priorities of research teams play an important role on the uneven knowledge of parasite species richness in Neotropical fishes. Since cestodes are ubiquitously distributed in fishes from South America, it is likely that the higher the number of elasmobranchs and teleosts examined in parasitological surveys, the higher the number of parasite-host associations that will be identified.

Accurate identification of fish hosts

During the development of this checklist, we have faced several examples of problematic identification and controversial taxonomy of hosts, which may compromise the reliability of any parasitological survey and limit our understanding of host specificity, the rela-

tionship between parasite and host phylogenies, as well as the establishment of trophic links elucidated by life-cycle studies (Naylor et al. 2012). Some genera, such as *Cichla*, *Pimelodus*, *Potamotrygon*, *Pseudoplatystoma* and *Zungaro*, have a convoluted taxonomic history and their species boundaries can diverge depending upon the approach used. Kullander and Ferreira (2006) for instance, recognized 15 species of *Cichla* distributed across South American rivers, based on morphological characters. However, Willis et al. (2012) recognized only eight species using multi-locus genetic data, suggesting that the number of *Cichla* species in South America may have been overestimated.

Therefore, we recommend that parasitologists keep a piece of host tissue in a molecular-grade ethanol for sequencing and to work in synergy with fish taxonomists to be as accurate as possible in fish identification, as already advocated by Naylor et al. (2012) for elasmobranch hosts. Caira and Jensen (2014) provided a field-sampling protocol that may be useful not only for parasite taxonomists, but also for those who are interested in general host-parasite associations.

Conclusions

Poulin et al. (2016) tested the completeness of 25 checklists of metazoan parasites in vertebrate hosts from several geographic regions based on three approaches. None of the studies analyzed performed well and only three of them passed two of the tests. Several obstacles contribute to a lack of completeness of checklists, including: (1) the reliability of information depends on the accuracy of the description or report; (2) geographically biased studies may not reflect the real distribution of diversity; (3) cryptic species, i.e. genetically distinct species that look similar morphologically, may contribute to an underestimate of the true number of species; and (4) only a small fraction of the potential fish hosts in South America have been examined for parasites. To mitigate these issues, we have attempted to critically gather as much information as possible and have obtained expert opinions. Therefore, we hope that we provide here the most robust database up to date that may help in a reliable estimation of the true diversity of fish cestodes in South America.

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References

Abdallah VD, Luque JL, Alves DR, Paraguassú AR (2002) Aspectos quantitativos das infrapopulações de metazoários parasitos da cavalinha, *Scomber japonicus* (Osteichthyes: Scombridae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Universidade Rural, Série Ciências da Vida 22: 103–107.

Ahid SMM, Filgueira KD, Fonseca ZAAS, Soto-Blanco B, Oliveira MF (2009) Ocorrência de parasitismo em *Mola mola* (Linnaeus, 1758) por metazoários no litoral do Rio Grande do Norte, Brasil. Acta Veterinaria Brasilica 3: 43–47. https://doi.org/10.21708/avb.2009.3.1.1118

Alarcos AJ, Etchegoin JA (2010) Parasite assemblages of estuarine-dependent marine fishes from Mar Chiquita coastal lagoon (Buenos Aires Province, Argentina). Parasitology Research 107: 1083–1091. https://doi.org/10.1007/s00436-010-1974-z

Alarcos AJ, Ivanov VA, Sardella NH (2006) Distribution patterns and interactions of cestodes in the spiral intestine of the narrownose smooth-hound shark, *Mustelus schmitti* Springer, 1939 (Chondrichthyes, Carcharhiniformes). Acta Parasitologica 51: 100–106. https://doi.org/10.2478/s11686-006-0015-7

Alarcos AJ, Pereira AN, Taborda NL, Luque JL, Timi JT (2016) Parasitological evidence of stocks of *Paralichthys isosceles* (Pleuronectiformes: Paralichthyidae) at small and large geographical scales in South American Atlantic coasts. Fisheries Research 173: 221–228. https://doi.org/10.1016/j.fishres.2015.07.018

Alarcos AJ, Timi JT (2012) Parasite communities in three sympatric flounder species (Pleuronectiformes: Paralichthyidae). Parasitology Research 110: 2155–2166. https://doi.org/10.1007/s00436-011-2741-5

Alarcos AJ, Timi JT (2013) Stocks and seasonal migrations of the flounder *Xystreurys rasile* as indicated by its parasites. Journal of Fish Biology 83: 531–541. https://doi.org/10.1111/jfb.12190

Albuquerque MC, Santos MD, Monteiro CM, Martins AN, Ederli NB, Brasil-Sato MC (2008) Helmintos endoparasitos de *Pimelodus maculatus* Lacépède, 1803 (Siluriformes, Pimelodidae) de duas localidades (Lagoa e Calha) do Rio Guandu, Estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 17: 113–119.

Alcântara NM, Tavares-Dias M (2015) Structure of the parasites communities in two Erythrinidae fish from Amazon river system (Brazil). Revista Brasileira de Parasitologia Veterinária 24: 183–190. https://doi.org/10.1590/S1984-29612015039

Alves DR, Luque JL (1999) Aspectos quantitativos das infrapopulações de metazoários parasitos de indivíduos jovens da corvina, *Micropogonias furnieri* (Osteichthyes: Sciaenidae) do litoral do Estado do Rio de Janeiro, Brasil. Contribuições Avulsas sobre a História Natural do Brasil, Série Zoologia 10: 1–4.

Alves DR, Luque JL (2000) Metazoários parasitos de *Micropogonias furnieri* (Osteichthyes: Sciaenidae) do litoral do Estado do Rio de Janeiro, Brasil. Parasitología al Día 24: 40–45. https://doi.org/10.4067/S0716-07202000000100006

Alves DR, Luque JL (2001a) Community ecology of the metazoan parasites of white croaker, *Micropogonias furnieri* (Osteichthyes: Sciaenidae), from the coastal zone of the State of Rio de Janeiro, Brazil. Memórias do Instituto Oswaldo Cruz 96: 145–153. https://doi.org/10.1590/S0074-02762001000200002

Alves DR, Luque JL (2001b) Aspectos quantitativos das infrapopulações de metazoários parasitos de *Micropogonias furnieri* (Osteichthyes: Sciaenidae) do litoral do Estado do Rio de Janeiro, Brasil. Parasitología al Día 25: 30–35. https://doi.org/10.4067/S0716-07202001000100006

Alves DR, Luque JL (2006) Ecologia das comunidades de metazoários parasitos de cinco espécies de escombrídeos (Perciformes: Scombridae) do litoral do estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 15: 167–181.

Alves DR, Luque JL, Abdallah VD (2003) Metazoan parasites of chub mackerel, *Scomber japonicus* Houttuyn (Osteichthyes: Scombridae), from the coastal zone of the State of Rio de Janeiro, Brazil. Revista Brasileira de Parasitologia Veterinária 12: 164–170.

Alves DR, Luque JL, Paraguassú AR (2002a) Metazoários parasitos do "congro-rosa" *Genypterus brasiliensis* Regan, 1903 (Osteichthyes, Ophidiidae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Brasileira de Zoociências 4: 133–142.

Alves DR, Luque JL, Paraguassú AR (2002b) Community ecology of the metazoan parasites of pink cusk-eel, *Genypterus brasiliensis* (Osteichthyes: Ophidiidae), from the coastal zone of the State of Rio de Janeiro, Brazil. Memórias do Instituto Oswaldo Cruz 97: 683–689. https://doi.org/10.1590/S0074-02762002000500018

Alves DR, Luque JL, Paraguassú AR, Jorge DS, Viñas RA (2002c) Ecologia da comunidade de metazoários parasitos da abrótea, *Urophycis mystaceus* Ribeiro, 1903 (Osteichthyes, Phycidae), do litoral do Estado do Rio de Janeiro. Revista Brasileira de Zoociências 4: 19–30.

Alves DR, Paraguassú AR, Luque JL (2004) Metazoários parasitos da abrótea, *Urophycis brasiliensis* (Kaup, 1858),(Osteichthyes: Phycidae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 13: 49–55.

Alves DR, Paraguassú AR, Luque JL (2005) Community ecology of the metazoan parasites of the grey triggerfish, *Balistes capriscus* Gmelin, 1789 and queen triggerfish *B. vetula* Linnaeus, 1758 (Osteichthyes: Balistidae) from the State of Rio de Janeiro, Brazil. Revista Brasileira de Parasitologia Veterinária 14: 71–77.

Alves PV, de Chambrier A, Scholz T, Luque JL (2015) A new genus and species of proteocephalidean tapeworm (Cestoda), first parasite found in the driftwood catfish *Tocantinsia piresi* (Siluriformes: Auchenipteridae) from Brazil. Folia Parasitologica 62: 006. https://doi.org/10.14411/fp.2015.006

Amato JFR, São Clemente SC, Oliveira GA (1990) *Tentacularia coryphaenae* Bosc, 1801 (Eucestoda: Trypanorhyncha) in the inspection and technology of the skipjack tuna, *Katsuwonus pelamis* (L.) (Pisces: Scombridae). Atlantica 12: 73–77.

Andrade SMS, Malta JCO, Ferraz E (2001) Parasitological fauna of matrinchã fries *Brycon cephalus* (Günther, 1869) collected in Negro and Solimões Rivers, in Central Amazônia. Acta Amazonica 31: 263–263. https://doi.org/10.1590/1809.4392200-1312273

Araújo CSO, Tavares-Dias M, Gomes ALS, Andrade SMS, Lemos JRG, Oliveira AT, Cruz WR, Affonso EG (2009) Infecções parasitárias e parâmetros sanguíneos em *Arapaima gigas* Schinz, 1822 (Arapaimidae) cultivados no estado do Amazonas, Brasil. In: Tavares-Dias M (Ed.) Manejo e sanidade de peixes em cultivo. Embrapa Amapá, Macapá, 389–424.

Araújo ME, Silva-Falcão EC, Falcão PD, Marques VM, Joca IR (2010) Stranding of *Masturus lanceolatus* (Actinopterygii: Molidae) in the estuary of the Una River, Pernambuco, Brazil: natural and anthropogenic causes. Marine Biodiversity Records 3: e69. https://doi.org/10.1017/S1755267210000588

Armas G (1983) Estudio de los endoparásitos (cestodos y nematodos) de *Paralabrax humeralis* Cuvier & Valenciennes, 1828, pez serránido de consumo humano em el Perú. Revista Ciencias Veterinárias Colegio Médico Eduardo Aragua (Venezuela) 12: 56–66.

Arredondo NJ, de Chambrier A, Gil de Pertierra AA (2013) A new genus and species of the Monticelliinae (Eucestoda: Proteocephalidea), a parasite of *Pseudoplatystoma fasciatum* (Pisces: Siluriformes) from the Paraná River basin (Argentina), with comments on microtriches of proteocephalideans. Folia Parasitologica 60: 248–256. https://doi.org/10.14411/fp.2013.028

Arredondo NJ, Gil de Pertierra AA (2008) The taxonomic status of *Spatulifer* cf. *maringaensis* Pavanelli & Rego, 1989 (Eucestoda: Proteocephalidea) from *Sorubim lima* (Bloch & Schneider) (Pisces: Siluriformes), and the use of the microthrix pattern in the discrimination of *Spatulifer* spp. Systematic Parasitology 70: 223–236. https://doi.org/10.1007/s11230-008-9142-x

Arredondo NJ, Gil de Pertierra AA (2010) *Monticellia santafesina* n. sp. (Cestoda: Proteocephalidea), a parasite of *Megalonema platanum* (Günther) (Siluriformes: Pimelodidae) in the Paraná River basin, Argentina. Systematic Parasitology 76: 103–110. https://doi.org/10.1007/s11230-010-9238-y

Arredondo NJ, Gil de Pertierra AA (2012) *Margaritaella gracilis* gen. n. et sp. n. (Eucestoda: Proteocephalidea), a parasite of *Callichthys callichthys* (Pisces: Siluriformes) from the Paraná River basin, Argentina. Folia Parasitologica 59: 99–106. https://doi.org/10.14411/fp.2012.015

Arredondo NJ, Gil de Pertierra AA, de Chambrier A (2014) A new species of *Pseudocrepidobothrium* (Cestoda: Proteocephalidea) from *Pseudoplatystoma reticulatum* (Pisces: Siluriformes) in the Paraná River basin (Argentina). Folia Parasitologica 61: 462–472. https://doi.org/10.14411/fp.2014.051

Azevedo GB, Madi RR, Ueta MT (2007) Metazoários parasitas de *Astyanax altiparanae* (Pisces: Characidae) na Fazenda Rio das Pedras, Campinas, SP, Brasil. Bioikos 21: 89–96.

Azevedo RK, Abdallah VD, Luque JL (2010) Acanthocephala, Annelida, Arthropoda, Myxozoa, Nematoda and Platyhelminthes parasites of fishes from the Guandu river, Rio de Janeiro, Brazil. Check List 6: 659–667.

Azevedo RK, Abdallah VD, Luque JL (2011) Biodiversity of fish parasites from Guandu River, southeastern Brazil: an ecological approach. Neotropical Helminthology 5: 185–199.

Bachmann F, Greinert JA, Bertelli PW, Silva Filho HH, Lara NOT, Ghiraldelli L, Martins ML (2007) Parasitofauna de *Pimelodus maculatus* (Osteichthyes: Pimelodidae) do rio Itajaí-Açu em Blumenau, Estado de Santa Catarina, Brasil. Acta Scientiarum: Biological Sciences 29: 109–114. https://doi.org/10.4025/actascibiolsci.v29i1.159

Baer JG (1969) *Diphyllobothrium pacificum*, a tapeworm from sea lions endemic in man along the coastal area of Peru. Journal of the Fisheries Board of Canada 26: 717–723. https://doi.org/10.1139/f69-071

Bahamonde N, Lopez MT (1962) *Proboscidosaccus mesodesmatis* n. sp. parásito de *Mesodesma donacium* Lamarck. Investigaciones Zoológicas Chilenas 8: 43–56.

Balboa L, George-Nascimento M (1998) Variaciones ontogenéticas y entre años en las infracomunidades de parásitos metazoos de dos especies de peces marinos de Chile. Revista Chilena de Historia Natural 71: 27–37.

Bandoni SM, Brooks DR (1987) Revision and phylogenetic analysis of the Amphilinidea Poche, 1922 (Platyhelminthes: Cercomeria: Cercomeromorpha). Canadian Journal of Zoology 65: 1110–1128. https://doi.org/10.1139/z87-175

Békési L, Feitosa VA, Cabral FAB (1992) Metacestodosis caused by plerocercoids of Proteocephalidea (Cestoda) in fish fry cultured in large scale in the Brazilian Northeast. Parasitologia Hungarica 25: 9–13.

Bellay S, Ueda BH, Takemoto RM, Lizama MAP, Pavanelli GC (2012) Fauna parasitária de *Geophagus brasiliensis* (Perciformes: Cichlidae) em reservatórios do estado do Paraná, Brasil. Revista Brasileira de Biociências 10: 74–78.

Bernot JP, Caira JN, Pickering M (2015) The dismantling of *Calliobothrium* (Cestoda: Tetraphyllidea) with erection of *Symcallio* n. gen. and description of two new species. Journal of Parasitology 101: 167–181. https://doi.org/10.1645/14-571.1

Bertullo V (1965) Infestación masiva de músculos de corvina (*Micropogom opercularis* L.) por *Tretrarhynchus fragilis* (Diesing). Revista del Instituto de Investigaciones Pesqueras 1: 344–348.

Bethular A, Mancini M, Salinas V, Echaniz S, Vignatti A, Larriestra A (2014) Alimentación, condición corporal y principales parásitos del pejerrey (*Odontotesthes bonaeriensis*) del embalse San Roque (Argentina). Biología Acuática 30: 59–68.

Beveridge I, Campbell RA (1993) A revision of *Dasyrhynchus* Pintner (Cestoda: Trypanorhyncha), parasitic in elasmobranch and teleost fishes. Systematic Parasitology 24: 129–157. https://doi.org/10.1007/BF00009597

Beveridge I, Campbell RA (2007) Revision of the *Grillotia erinaceus* (van Beneden, 1858) species complex (Cestoda: Trypanorhyncha), with the description of *G. brayi* n. sp. Systematic Parasitology 68: 1–31. https://doi.org/10.1007/s11230-006-9082-2

Beveridge I, Justine J (2007a) *Paragrillotia apecteta* n. sp. and redescription of *P. spratti* (Campbell & Beveridge, 1993) n. comb. (Cestoda, Trypanorhyncha) from hexanchid and carcharhinid sharks off New Caledonia. Zoosystema 29: 381–391.

Beveridge I, Justine J (2007b) Redescriptions of four species of *Otobothrium* Linton, 1890 (Cestoda: Trypanorhyncha), including new records from Australia, New Caledonia and Malaysia, with the description of *O. parvum* n. sp. Zootaxa 1587: 1–25.

Beveridge I, Sakanari JA (1987) *Lacistorhynchus dollfusi* sp. nov. (Cestoda: Trypanorhyncha) in elasmobranch fishes from Australian and North American coastal waters. Transactions of the Royal Society of South Australia 111: 147–154.

Bicudo AJ, Tavares LE, Luque JL (2004) Metazoários parasitos da cabrinha *Prionotus punctatus* (Bloch, 1793) (Osteichthyes: Triglidae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 14: 27–33.

Bittencourt LS, Pinheiro DA, Cárdenas MQ, Fernandes BM, Tavares-Dias M (2014) Parasites of native Cichlidae populations and invasive *Oreochromis niloticus* (Linnaeus, 1758) in tributary of Amazonas River (Brazil). Revista Brasileira de Parasitologia Veterinária 23: 44–54. https://doi.org/10.1590/S1984-29612014006

Boni TA, Padial AA, Prioli SMAP, Lucio LC, Maniglia TC, Bignotto TS, Panarari-Antunes RS, Prioli RA, Prioli AJ (2011) Molecular differentiation of species of the genus *Zungaro* (Siluriformes, Pimelodidae) from the Amazon and Paraná-Paraguay River basins in Brazil. Genetics and Molecular Research 10: 2795– 2805. https://doi.org/10.4238/2011.November.10.2

Brabec J, Waeschenbach A, Scholz T, Littlewood DTJ, Kuchta R (2015) Molecular phylogeny of the Bothriocephalidea (Cestoda): molecular data challenge morphological classification. International Journal for Parasitology 45: 761–771. https://doi.org/10.1016/j.ijpara.2015.05.006

Braicovich PE, Luque JL, Timi JT (2012) Geographical patterns of parasite infracommunities in the rough scad, *Trachurus lathami* Nichols, in the southwestern Atlantic Ocean. Journal of Parasitology 98: 768–777. https://doi.org/10.1645/GE-2950.1

Braicovich PE, Timi JT (2008) Parasites as biological tags for stock discrimination of the Brazilian flathead *Percophis brasiliensis* in the south-west Atlantic. Journal of Fish Biology 73: 557–571. https://doi.org/10.1111/j.1095-8649.2008.01948.x

Braicovich PE, Timi JT (2010) Seasonal stability in parasite assemblages of the Brazilian flathead, *Percophis brasiliensis* (Perciformes: Percophidae): predictable tools for stock identification. Folia Parasitologica 57: 206–212. https://doi.org/10.14411/fp.2010.027

Braicovich PE, Timi JT (2015) Homogeneity of parasite assemblages of *Dules auriga* (Serranidae) in hydrographically heterogeneous sites. Journal of Fish Biology 86: 1363–1376. https://doi.org/10.1111/jfb.12648

Brasil-Sato MC (2003) Parasitos de peixes da bacia do São Francisco. In: Godinho HP, Godinho AL (Eds) Águas, peixes e pescadores do São Francisco das Minas Gerais. PUC Minas, Belo Horizonte, 149–165.

Bravo S, Almonacid C, Oyarzo C, Silva MT (2007) The parasite fauna of *Galaxias maculatus* in the estuary of Maullin River, Chile. Bulletin of the European Association of Fish Pathologists 27: 10–17.

Bray RA (1994) Order Nippotaeniidea. In: Khalil LF, Jones A, Bray RA (Eds) Keys to the Cestode Parasites of Vertebrates. CAB International, Wallingford, 253–255.

Bray RA, Jones A, Andersen KI (1994) Order Pseudophyllidea Carus, 1863. In: Khalil LF, Jones A, Bray RA (Eds) Keys to the cestode parasites of vertebrates. CAB International, Wallingford, 205–247.

Brickle P, MacKenzie K (2007) Parasites as biological tags for *Eleginops maclovinus* (Teleostei: Eleginopidae) around the Falkland Islands. Journal of Helminthology 81: 147–153. https://doi.org/10.1017/S0022149X07750514

Brickle P, MacKenzie K, Pike A (2006) Variations in the parasite fauna of the Patagonian toothfish (*Dissostichus eleginoides* Smitt, 1898), with length, season, and depth of habitat around the Falkland Islands. Journal of Parasitology 92: 282–291. https://doi.org/10.1645/GE-539R.1

Brooks DR (1977) Six new species of tetraphyllidean cestodes, including a new genus, from a marine stingray *Himantura schmardae* (Werner, 1904) from Colombia. Proceedings of the Helminthological Society of Washington 44: 51–59.

Brooks DR, Amato JFR (1992) Cestode parasites in *Potamotrygon motoro* (Natterer) (Chondrichthyes: Potamotrygonidae) from southwestern Brazil, including *Rhinebothroides mclennanae* n. sp. (Tetraphyllidea: Phyllobothriidae), and a revised host-parasite checklist for helminths inhabiting Neotropical freshwater stingrays. Journal of Parasitology 78: 393–398. https://doi.org/10.2307/3283633

Brooks DR, Barriga R (1995) *Serendip deborahae* n. gen. and n. sp. (Eucestoda: Tetraphyllidea: Serendipidae n. fam.) *in Rhinoptera steindachneri* Evermann and Jenkins, 1891 (Chondrichthyes: Myliobatiformes: Myliobatidae) from Southeastern Ecuador. Journal of Parasitology 81: 80–84.

Brooks DR, Deardorff TL (1980) Three proteocephalid cestodes from Colombian siluriform fishes, including *Nomimoscolex alovarius* sp. n. (Monticelliidae: Zygobothriinae). Proceedings of the Biological Society of Washington 47: 15–21.

Brooks DR, Marques F, Perroni C, Sidagis C (1999) *Scyphophyllidium uruguayense* n. sp. (Eucestoda: Tetraphyllidea) in *Mustelus mento* (Cope, 1877) (Chondrichthyes: Carcharhiniformes: Triakidae) from La Paloma, Uruguay. Journal of Parasitology 85: 490–494. https://doi.org/10.2307/3285784

Brooks DR, Mayes MA (1978) *Acanthobothrium electricolum* sp. n. and *A. lintoni* Goldstein, Henson, and Schlicht 1969 (Cestoda: Tetraphyllidea) from *Narcine brasiliensis* (Olfers) (Chondrichthyes: Torpedinidae) in Colombia. Journal of Parasitology: 617–619. https://doi.org/10.2307/3279945

Brooks DR, Mayers MA (1980) Cestodes in four species of euryhaline stingrays from Colombia. Proceedings of the Helminthological Society of Washington 47: 22–29.

Brooks DR, Mayes MA, Thorson TB (1981a) Cestode parasites in *Myliobatis goodei* Garman (Myliobatiformes: Myliobatidae) from Rio de la Plata, Uruguay, with a summary of cestodes collected from South American elasmobranchs during 1975-1979. Proceedings of the Biological Society of Washington 93: 1239–1252.

Brooks DR, Mayes MA, Thorson TB (1981b) Systematic review of cestodes infecting freshwater stingrays (Chondrichthyes: Potamotrygonidae) including four new species from Venezuela. Proceedings of the Biological Society of Washington 48: 43–64.

Brooks DR, Rasmussen G (1984) Proteocephalidean cestodes from Venezuelan siluriform fishes, with a revised classification of the Monticelliidae. Proceedings of the Biological Society of Washington 97: 748–760.

Brooks DR, Thorson TB (1976) Two tetraphyllidean cestodes from the freshwater stingray *Potamotrygon magdalenae* Dumeril 1852 (Chondrichthyes: Potamotrygonidae) from Colombia. Journal of Parasitology 62: 943–947. https://doi.org/10.230-7/3279188

Brown J, Brickle P, Scott BE (2013) The parasite fauna of the Patagonian toothfish *Dissostichus eleginoides* off the Falkland Islands. Journal of Helminthology 87: 501–509. https://doi.org/10.1017/S0022149X12000636

Bueno GBF, Aguiar JCC, Santos SMC (2014) Community structure of metazoan parasites of *Trichiurus lepturus* (Perciformes, Trichiuridae) from Ubatuba, Southwestern Atlantic Ocean, Brazil. Acta Scientiarum: Biological Sciences 36: 357–364. https://doi.org/10.4025/actascibiolsci.v36i3.21908

Caira JN, Jensen K (2014) A digest of elasmobranch tapeworms. Journal of Parasitology 100: 373–391. https://doi.org/10.1645/14-516.1

Caira JN, Jensen K (2015) Insights on the identities of sharks of the *Rhizoprionodon acutus* (Elasmobranchii: Carcharhiniformes) species complex based on three new species of *Phoreiobothrium* (Cestoda: Onchoproteocephalidea). Zootaxa 4059: 335–350. https://doi.org/10.11646/zootaxa.4059.2.5

Caira JN, Jensen K, Barbeau E (Eds) (2012) Global Cestode Database. World Wide Web electronic publication. http://www.tapewormdb.uconn.edu/ [accessed 1.V.2016]

Caira JN, Jensen K, Waeschenbach A, Olson PD, Littlewood DTJ (2014) Orders out of chaos – molecular phylogenetics reveals the complexity of shark and stingray tapeworm relationships. International Journal for Parasitology 44: 55–73. https://doi.org/10.1016/j.ijpara.2013.10.004

Caira JN, Littlewood DTJ (2013) Worms, Platyhelminthes. In: Levin SA (Ed.) Encyclopedia of Biodiversity. Academic Press, San Diego, 437–469. https://doi.org/10.1016/B978-0-12-384719-5.00166-0

Caira JN, Marques FPL, Jensen K, Kuchta R, Ivanov VA (2013) Phylogenetic analysis and reconfiguration of genera in the cestode order Diphyllidea. International Journal for Parasitology 43: 621–639. https://doi.org/10.1016/j.ijpara.2013.03.001

Caira JN, Orringer DJ (1995) Additional information on the morphology of *Potamotrygonocestus magdalenensis* (Tetraphyllidea: Onchobothriidae) from the freshwater stingray *Potamotrygon magdalenae* in Colombia. Journal of the Helminthological Society of Washington 62: 22–26.

Campbell RA, Beveridge I (1996) Revision of the family Pterobothriidae Pintner, 1931 (Cestoda: Trypanorhyncha). Invertebrate Systematics 10: 617–662. https://doi.org/10.1071/IT9960617

Campbell RA, Beveridge I (2002) The genus *Acanthobothrium* (Cestoda: Tetraphyllidea: Onchobothriidae) parasitic in Australian elasmobranch fishes. Invertebrate Systematics 16: 237–344. https://doi.org/10.1071/IT01004

Campbell RA, Carvajal JG (1979) Synonymy of the phyllobothriid genera *Rhodobothrium* Linton, 1889, *Inermiphyllidium* Riser, 1955, and *Sphaerobothrium* Euzet, 1959 (Cestoda: Tetraphyllidea). Proceedings of the Helminthological Society of Washington 46: 88–97.

Campbell RA, Carvajal JG (1980) *Echinobothrium euzeti*, a new cestode from the spiral valve of a Chilean elasmobranch. Proceedings of the Helminthological Society of Washington 47: 165–167.

Campbell RA, Carvajal JG (1987) *Phyllobothrium discopygi* n. sp. (Cestoda: Tetraphyllidea) from Chile, with a critical comparison of the affinities of *P. auricula* van Beneden, 1858 and *P. foliatum* Linton, 1890. Systematic Parasitology 10: 159–164. https://doi.org/10.1007/BF00009621

Campbell RA, Marques FPL, Ivanov VA (1999) *Paroncomegas araya* (Woodland, 1934) n. gen. et comb. (Cestoda: Trypanorhyncha: Eutetrarhynchidae) from the freshwater stingray *Potamotrygon motoro* in South America. Journal of Parasitology 85: 313–320. https://doi.org/10.2307/3285640

Campos CM, Fonseca VE, Takemoto RM, Moraes FR (2008) Fauna parasitária de cachara *Pseudoplatystoma fasciatum* (Siluriforme: Pimelodidae) do rio Aquidauana, Pantanal Sul Matogrossense, Brasil. Acta Scientiarum: Biological Sciences: 91–96. https://doi.org/10.4025/actascibiolsci.v30i1.1469

Campos CM, Fonseca VE, Takemoto RM, Moraes FR (2009a) Ecology of the parasitic endohelminth community of *Pseudoplatystoma fasciatum* (Linnaeus, 1776) (Siluriformes: Pimelodidae) from the Aquidauana River, Pantanal, State of Mato Grosso do Sul, Brazil. Brazilian Journal of Biology 69: 93–99. https://doi.org/10.1590/S1519-69842009000100011

Campos CM, Moraes JRE, Rondini J, Moraes FR (2009b) Histopatologia do intestino de *Pseudoplatystoma fasciatum* (Osteichthyes, Pimelodidae) parasitados com cestodas proteocefalídeos e nematodas. Boletim do Instituto de Pesca: 153–158.

Carballo MC, Cremonte F, Navone GT, Timi JT (2012) Similarity in parasite community structure may be used to trace latitudinal migrations of *Odontesthes smitti* along Argentinean coasts. Journal of Fish Biology 80: 15–28. https://doi.org/10.1111/j.1095-8649.2011.03125.x

Carballo MC, Navone GT, Cremonte F (2011) Parasites of the silversides *Odontesthes smitti* and *Odontesthes nigricans* (Pisces: Atherinopsidae) from Argentinean Patagonia. Comparative Parasitology 78: 95–103. https://doi.org/10.1654/4445.1

Cardoso TP, Salgado RL, Andrade PF, São Clemente SC, Lima FC (2006) Nematóides da família Anisakidae e cestóides da ordem Trypanorhyncha em peixes teleósteos comercializados no estado do Rio de Janeiro. Revista Brasileira de Ciência Veterinária 13: 98–101.

Carfora M, de Chambrier A, Vaucher C (2003) Le genre *Amphoteromorphus* (Cestoda: Proteocephalidea), parasite de poissons-chats d'Amérique tropicale: étude morphologique et approche biosystématique par électrophorèse des protéines. Revue Suisse de Zoologie 110: 381–409. https://doi.org/10.5962/bhl.part.80190

Carvajal JG (1971) *Grillotia dollfusi* sp. n. (Cestoda: Trypanorhyncha) from the skate, *Raja chilensis*, from Chile, and a note on *G. heptanchi*. Journal of Parasitology 57: 1269–1271. https://doi.org/10.2307/3277978

Carvajal JG (1974) Records of cestodes from Chilean sharks. Journal of Parasitology 60: 29–34. https://doi.org/10.2307/3278674

Carvajal JG (1977) Description of the adult and larva of *Caulobothrium myliobatidis* sp. n. (Cestoda: Tetraphyllidea) from Chile. Journal of Parasitology 63: 99–103. https://doi.org/10.2307/3280111

Carvajal JG, Barros C, Whittaker FH (1985) Scanning electron microscopy of scolices of some tetraphyllidean cestodes in Chilean skates. Microscopía Electrónica y Biología Celular 9: 23–33.

Carvajal JG, Barros C, Whittaker FH (1987) Scanning electron microscopy of the scolex of the plerocercus *Callitetrarhynchus gracilis* (Rudolphi, 1819) (Cestoda: Trypanorhyncha). Journal of Parasitology 73: 1265–1267. https://doi.org/10.2307/3282323

Carvajal JG, Campbell RA (1979) Identificación de las larvas de cestodos tetrarínquidos presentes en las merluzas y congrios de Puerto Montt, Chile. Boletín Chileno de Parasitología 34: 65–67.

Carvajal JG, Cattan PE (1978) Occurrence of the plerocercus of *Grillotia dollfusi* Carvajal 1971 (Cestoda: Trypanorhyncha) in the Chilean hake *Merluccius gayi* (Guichenot 1848). Journal of Parasitology 64: 695–695. https://doi.org/10.2307/3279962

Carvajal JG, Cattan PE, Castillo C, Schatte P (1979) Larval anisakids and other helminths in the hake, *Merluccius gayi* (Guichenot) from Chile. Journal of Fish Biology 15: 671–677. https://doi.org/10.1111/j.1095-8649.1979.tb03676.x

Carvajal JG, Dailey MD (1975) Three new species of *Echeneibothrium* (Cestoda: Tetraphyllidea) from the skate, *Raja chilensis* Guichenot, 1848, with comments on mode of attachment and host specificity. Journal of Parasitology 61: 89–94. https://doi.org/10.2307/3279115

Carvajal JG, Goldstein RJ (1969) *Acanthobothrium psammobati* sp. n. (Cestoda: Tetraphyllidea: Onchobothriidae) from the skate, *Psammobatis scobina* (Chondrichthyes: Rajidae) from Chile. Zoologischer Anzeiger 182: 432–435.

Carvajal JG, Goldstein RJ (1971) *Acanthobothrium annapinkiensis* n. sp. (Cestoda: Tetraphyllidea: Onchobothriidae) from the skate, *Raja chilensis* (Chondrichthyes: Rajidae) from Chile. Zoologischer Anzeiger 186: 158–162.

Carvajal JG, Jeges JG (1980) Cestode parasites of *Myliobatis chilensis* with a description of a new species of *Acanthobothrium*. Anales del Centro de Ciencias del Mar y Limnologia 7: 51–56.

Carvajal JG, Rego AA (1983) *Progrillotia dollfusi* sp. n. (Cestoda: Trypanorhyncha) parasito de pescada do litoral brasileiro. Memórias do Instituto Oswaldo Cruz 78: 231–234. https://doi.org/10.1590/S0074-02761983000200012

Carvajal JG, Rego AA (1985) Critical studies on the genus *Callitetrarhynchus* (Cestoda: Trypanorhyncha) with recognition of *Rhynchobothrium speciosum* Linton, 1897 as a valid species of the genus *Callitetrarhynchus*. Systematic Parasitology 7: 161–167. https://doi.org/10.1007/BF00011449

Carvajal JG, Ruíz G (1987) Fijación de dos especies de *Acanthobothrium* (Cestoda: Tetraphyllidea) a la válvula espiral de rayas del género *Sympterygia*: estudio de la interfase parásito--huésped. Parasitología al Día 11: 49–55.

Carvalho AR, Luque JL (2011) Seasonal variation in metazoan parasites of *Trichiurus lepturus* (Perciformes: Trichiuridae) of Rio de Janeiro, Brazil. Brazilian Journal of Biology 71: 771–782. https://doi.org/10.1590/S1519-69842011000400024

Carvalho-Costa LF, Piorski NM, Willis SC, Galetti PM, Ortí G (2011) Molecular systematics of the neotropical shovelnose catfish genus *Pseudoplatystoma* Bleeker 1862 based on nuclear and mtDNA markers. Molecular Phylogenetics and Evolution 59: 177–194. https://doi.org/10.1016/j.ympev.2011.02.005

Cattan PE (1977) El congrio dorado *Genypterus blacodes*, Schneider, nuevo registro en Chile, para *Hepatoxylon trichiuri* Holten, 1802 (Cestoda, Trypanorhyncha). Boletín Chileno de Parasitología 32: 92–93.

Cattan PE, Carvajal JG, Torres D, Yáñez JL (1979) Primera comunicación sobre cestodos Trypanorhyncha en peces del archipiélago de Juan Fernández. Boletín Chileno de Parasitología 34: 44–46.

Ceccarelli PS, Adriano EA, Santos SMC, Rego RF, Silva LOL (2006) Levantamento qualiquantitativo da fauna parasitológica de peixes do Pantanal Mato-Grossense. In: Centro Nacional de Pesquisa e Gestão de Recursos Pesqueiros Continentais-Cepta (Ed.) Pesquisas patológicas e genéticas em recursos pesqueiros da Bacia do Alto Paraguai. IBAMA, Pirassununga, 16–116.

Chambers CB, Cribb TH, Jones MK (2000) Tetraphyllidean metacestodes of teleosts of the Great Barrier Reef, and the use of *in vitro* cultivation to identify them. Folia Parasitologica 47: 285–292. https://doi.org/10.14411/fp.2000.050

Chávez RA, González MT, Oliva ME, Valdivia IM (2012) Endoparasite fauna of five Gadiformes fish species from the coast of Chile: host ecology versus phylogeny. Journal of Helminthology 86: 10–15. https://doi.org/10.1017/S0022149X10000921

Chávez RA, Valdivia IM, Oliva ME (2007) Local variability in metazoan parasites of the pelagic fish species, *Engraulis ringens*: implications for fish stock assessment using parasites as biological tags. Journal of Helminthology 81: 113–116. https://doi.org/10.1017/S0022149X07726573

Chemes SB, Takemoto RM (2011) Diversity of parasites from middle Paraná system freshwater fishes, Argentina. International Journal of Biodiversity and Conservation 3: 249–266.

Chero J, Cruces C, Iannacone J, Sáez G, Alvariño L, Rodríguez C, Rodríguez H, Tuesta E, Pacheco A, Huamani N (2014a) Índices parasitológicos de la merluza peruana *Merluccius gayi peruanus* (Ginsburg, 1954) (Perciformes: Merlucciidae) adquiridos del terminal pesquero de Ventanilla, Callao, Perú. Neotropical Helminthology 8: 141–162.

Chero J, Cruces C, Iannacone J, Sáez G, Sanchez L, Minaya D, Alvariño L, Mendoza-Vidaurre C, Luque JL (2015) First record of *Unitubulotestis pelamydis* (Trematoda: Didymozoidae) and *Sphyriocephalus tergestinus* (Cestoda: Sphyriocephalidae) in eastern Pacific bonito, *Sarda chiliensis* (Perciformes: Scombridae) in Peru. Neotropical Helminthology 9: 313–323.

Chero J, Iannacone J, Cruces C, Sáez G, Alvariño L (2014b) Comunidad de metazoos parásitos de la corvina *Cilus gilberti* (Abbott, 1899) (Perciformes: Sciaenidae) en la zona costera de Chorrillos, Lima, Perú. Neotropical Helminthology 8: 163–182.

Chero J, Sáez G, Iannacone J, Aquino W (2014c) Aspectos ecológicos de los helmintos parásitos de lorna *Sciaena deliciosa* (tschudi, 1846) (Perciformes: Sciaenidae) adquiridos del terminal pesquero de Ventanilla, Callao, Perú. Neotropical Helminthology 8: 59–76.

Choudhury A, Aguirre-Macedo ML, Curran SS, Ostrowski de Núñez M, Overstreet RM, Pérez-Ponce de León G, Santos CP (2016) Trematode diversity in freshwater fishes of the Globe II: 'New World'. Systematic Parasitology 93: 271–282. https://doi.org/10.1007/s11230-016-9632-1

Cordeiro AS, Luque JL (2004) Community ecology of the metazoan parasites of Atlantic moonfish, *Selene setapinnis* (Osteichthyes: Carangidae) from the coastal zone of the State of Rio de Janeiro, Brazil. Brazilian Journal of Biology 64: 399–406. https://doi.org/10.1590/S1519-69842004000300004

Cordeiro AS, Luque JL (2005) Metazoários parasitos do coió *Dactylopterus volitans* (Linnaeus, 1758) (Osteichthyes: Dactylopteridae) do litoral do Estado do Rio de Janeiro, Brasil. Acta Scientiarum: Biological Sciences 27: 119–123. https://doi.org/10.4025/actascibiolsci.v27i2.1320

Cortés Y, Muñoz G (2008) Infracomunidades de parásitos eumetazoos del bagre de mar *Aphos porosus* (Valenciennes, 1837) (Actinopterygii: Batrachoidiformes) en Chile central. Revista de Biología Marina y Oceanografía 43: 255–263. https://doi.org/10.4067/S0718-19572008000200004

Cortés Y, Muñoz G (2009) Metazoan parasite infracommunities of the toadfish *Aphos porosus* (Pisces: Batrachoidiformes) in central Chile: how variable are they over time? Journal of Parasitology 95: 753–756. https://doi.org/10.1645/GE-1651.1

Cousin JCB, Pereira J Jr., Gonzales JF (2006) Histopatologia no fígado de *Prionace glauca* (Chondrichthyes, Squaliformes, Carcharhinidae) causada por *Hepatoxylon trichiuri* (Eucestoda, Trypanorhyncha). Biociências 11: 167–172.

Cremonte F, Sardella NH (1997) The parasita Fauna of *Scomber japonicus* Houttuyn, 1782 (Pisces: Scombridae) in two zones of the Argentine Sea. Fisheries Research 31: 1–9. https://doi.org/10.1016/S0165-7836(97)00024-6

Cruces C, Chero J, Iannacone J, Diestro A, Sáez G, Alvariño L (2014) Metazoos parásitos de "caballa" *Scomber japonicus* Houttuyn, 1782 (Perciformes: Scombridae), del puerto de Chicama, La Libertad, Perú. Neotropical Helminthology 8: 357–381.

Cruces C, Chero J, Iannacone J, Sáez G, Alvariño L (2015) Comunidad de endohelmintos parásitos del trambollo de boca amarilla *Labrisomus philippii* (Steindachner, 1866) (Perciformes: Labrisomidae) de la costa central del Perú. The Biologist 13: 91–109.

Dailey MD, Carvajal JG (1976) Helminth parasites of *Rhinobatos planiceps* Garman 1880, including two new species of cestodes, with comments on host specificity of the genus *Rhinebothrium* Linton 1890. Journal of Parasitology 62: 939–942. https://doi.org/10.2307/3279187

de Chambrier A (1990) Redescription de *Proteocephalus paraguayensis* (Rudin, 1917) (Cestoda: Proteocephalidae) parasite de *Hydrodynastes gigas* (Dum., Bibr. & Dum., 1854) du Paraguay. Systematic Parasitology 16: 85–87. https://doi.org/10.1007/BF00009608

de Chambrier A (2001) A new tapeworm from the Amazon, *Amazotaenia yvettae* gen. n., sp. n., (Eucestoda: Proteocephalidea) from the siluriform fishes *Brachyplatystoma filamentosum* and *B. vaillanti* (Pimelodidae). Revue Suisse de Zoologie 108: 303–316. https://doi.org/10.5962/bhl.part.79632

de Chambrier A (2003) Systematic status of *Manaosia bracodemoca* Woodland, 1935 and *Paramonticellia itaipuensis* Pavanclli et Rego, 1991 (Eucestoda: Proteocephalidea), parasites of *Sorubim lima* (Siluriformes: Pimelodidae) from South America. Folia Parasitologica 50: 121–127. https://doi.org/10.14411/fp.2003.021

de Chambrier A, Gil de Pertierra AA (2002) Redescription of *Travassiella avitellina* Rego & Pavanelli, 1987 (Proteocephalidea: Monticelliidae, Zygobothriinae), a parasite of *Paulicea luetkeni* (Siluriformes) from South America. Memórias do Instituto Oswaldo Cruz 97: 657–661. https://doi.org/10.1590/S0074-02762002000500013

de Chambrier A, Kuchta R, Scholz T (2015a) Tapeworms (Cestoda: Proteocephalidea) of teleost fishes from the Amazon River in Peru: additional records as an evidence of unexplored species diversity. Revue Suisse de Zoologie 122: 149–163. https://doi.org/10.5281/zenodo.14580

de Chambrier A, Rego AA (1994) *Proteocephalus sophiae* n. sp. (Cestoda: Proteocephalidae), a parasite of the siluroid fish *Paulicea luetkeni* (Pisces: Pimelodidae) from the Brazilian Amazon. Revue Suisse de Zoologie 101: 361–368. https://doi.org/10.5962/bhl.part.79911

de Chambrier A, Rego AA (1995) *Mariauxiella pimelodi* n. g., n. sp. (Cestoda: Monticelliidae): A parasite of pimelodid siluroid fishes from South America. Systematic Parasitology 30: 57–65. https://doi.org/10.1007/BF00009245

de Chambrier A, Rego AA, Gil de Pertierra AA (2005) Redescription of two cestodes (Eucestoda: Proteocephalidea) parasitic in *Phractocephalus hemioliopterus* (Siluriformes) from the Amazon and erection of *Scholzia* gen. n. Revue Suisse de Zoologie 112: 735–752. https://doi.org/10.5962/bhl.part.80323

de Chambrier A, Rego AA, Mariaux J (2004a) Redescription of *Brooksiella praeputialis* and *Goezeella siluri* (Eucestoda: proteocephalidea), parasites of *Cetopsis coecutiens* (Siluriformes) from the Amazon and proposition of *Goezeella dankbrooksi* sp. n. Revue Suisse de Zoologie 111: 111–120. https://doi.org/10.5962/bhl.part.80230

de Chambrier A, Rego AA, Vaucher C (1999) *Euzetiella tetraphylliformis* n. gen., n. sp., (Eucestoda: Proteocephalidae), parasite du poisson d'eau douce néotropical *Paulicea luetkeni* (Siluriforme, Pimelodidae). Parasite 6: 43–47. https://doi.org/10.1051/parasite/1999061043

de Chambrier A, Scholz T (2005) Redescription of *Houssayela sudobim* (Woodland, 1935) (Cestoda: Proteocephalidea), a parasite of *Pseudoplatystoma fasciatum* (Pisces: Siluriformes) from the River Amazon. Systematic Parasitology 62: 161–169. https://doi.org/10.1007/s11230-005-5487-6

de Chambrier A, Scholz T (2008) Tapeworms (Cestoda: Proteocephalidea) of firewood catfish *Sorubimichthys planiceps* (Siluriformes: Pimelodidae) from the Amazon River. Folia Parasitologica 55: 17–28. https://doi.org/10.14411/fp.2008.004

de Chambrier A, Scholz T, Kuchta R (2014) Taxonomic status of Woodland's enigmatic tapeworms (Cestoda: Proteocephalidea) from Amazonian catfishes: back to museum collections. Systematic Parasitology 87: 1–19. https://doi.org/10.1007/s11230-013-9457-0

de Chambrier A, Scholz T, Kuchta R, Posel P, Mortenthaler M, Guardia CC (2006a) Tapeworms (Cestoda: Proteocephalidea) of fishes from the Amazon River in Peru. Comparative Parasitology 73: 111–120. https://doi.org/10.1654/4182.1

de Chambrier A, Scholz T, Vaucher C (1996) Tapeworms (Cestoda: Proteocephalidea) of *Hoplias malabaricus* (Pisces: Characiformes, Erythrinidae) in Paraguay: description of *Proteocephalus regoi* sp. n., and redescription of *Nomimoscolex matogrossensis*. Folia Parasitologica 43: 133–140.

de Chambrier A, Takemoto RM, Pavanelli GC (2006b) *Nomimoscolex pertierrae* n. sp. (Eucestoda: Proteocephalidea), a parasite of *Pseudoplatystoma corruscans* (Siluriformes: Pimelodidae) in Brazil and redescription of *N. sudobim* Woodland, 1935, a parasite of *P. fasciatum*. Systematic Parasitology 64: 191–202. https://doi.org/10.1007/s11230-006-9031-0

de Chambrier A, Vaucher C (1994) Etude morpho-anatomique et génétique de deux nouveaux *Proteocephalus* Weinland, 1858 (Cestoda: Proteocephalidae) parasites de *Platydoras costatus* (L.), poisson siluriforme du Paraguay. Systematic Parasitology 27: 173–185. https://doi.org/10.1007/BF00008479

de Chambrier A, Vaucher C (1997) Révision des cestodes (Monticelliidae) décrits par Woodland (1934) chez *Brachyplatystoma filamentosum* avec redéfinition des genres *Endorchis* Woodland, 1934 et *Nomimoscolex* Woodland, 1934. Systematic Parasitology 37: 219–233. https://doi.org/10.1023/A:1005863808627

de Chambrier A, Vaucher C (1999) Proteocephalidae et Monticelliidae (Eucestoda: Proteocephalidea) parasites de poissons d'eau douce au Paraguay, avec descriptions d'un genre nouveau et de dix espèces nouvelles. Revue Suisse de Zoologie 106: 165–240. https://doi.org/10.5962/bhl.part.80074

de Chambrier A, Waeschenbach A, Fisseha M, Scholz T, Mariaux J (2015b) A large 28S rDNA-based phylogeny confirms the limitations of established morphological characters for classification of proteocephalidean tapeworms (Platyhelminthes, Cestoda). ZooKeys 500: 25–59. https://doi.org/10.3897/zookeys.500.9360

de Chambrier A, Zehnder M, Vaucher C, Mariaux J (2004b) The evolution of the Proteocephalidea (Platyhelminthes, Eucestoda) based on an enlarged molecular phylogeny, with comments on their uterine development. Systematic Parasitology 57: 159–171. https://doi.org/10.1023/B:SYPA.0000019083.26876.34

Dias FJE, São Clemente SC, Knoff M (2009) Cestóides Trypanorhyncha parasitos de peroá, *Balistes capriscus* Gmelin, 1789 comercializados no estado do Rio de Janeiro, Brasil. Revista Brasileira de Ciência Veterinária 16: 19–21. https://doi.org/10.4322/rbcv.2014.163

Dias FJE, São Clemente SC, Knoff M (2010) Nematoides anisaquídeos e cestoides Trypanorhyncha de importância em saúde pública em *Aluterus monoceros* (Linnaeus, 1758) no Estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 19: 94–97. https://doi.org/10.1590/S1984-29612010000200005

Dias FJE, São Clemente SC, Pinto RM, Knoff M (2011a) Anisakidae nematodes and Trypanorhyncha cestodes of hygienic importance infecting the king mackerel *Scomberomorus cavalla* (Osteichthyes: Scombridae) in Brazil. Veterinary Parasitology 175: 351–355. https://doi.org/10.1016/j.vetpar.2010.10.014

Dias LNS, Paiva RS, São Clemente SC, Rodrigues AE, Peralta ASL, Matos ER (2011b) Cestóides Trypanorhyncha parasitos de scianideos de importância comercial, capturados no litoral amazônico, Brasil. Revista Brasileira de Ciência Veterinária 18: 3–5. https://doi.org/10.4322/rbcv.2014.111

Diesing KM (1850) Systema Helminthum, vol I. Braumüller, Vienna, 679 pp.

Diesing KM (1855) Sechzehn Gattungen von Binnenwürmern und ihre Arten. Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Classe 9: 171–185.

Diesing KM (1856) Zwanzig Arten von Cephalocotyllen. Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Classe 12: 23–38.

Dobson A, Lafferty KD, Kuris AM, Hechinger RF, Jetz W (2008) Homage to Linnaeus: how many parasites? How many hosts? Proceedings of the National Academy of Sciences 105: 11482–11489. https://doi.org/10.1073/pnas.0803232105

Dollfus RP (1930) Sur les Tétrarhynches. I. Définition des genres (suite). Mémoires de la Société Zoologique de France 29: 139–216.

Dollfus RP (1942) Études critiques sur les Tétrarhynques du Muséum de Paris. Archives du Muséum National d'Histoire Naturelle, Paris, 466 pp.

Drago FB (2012) Community structure of metazoan parasites of silverside, *Odontesthes bonariensis* (Pisces, Atherinopsidae) from Argentina. Iheringia: Série Zoologia 102: 26–32. https://doi.org/10.1590/S0073-47212012000100004

Durán LE, Oliva ME (1980) Estudio parasitológico en *Merluccius gayi peruanus* Gingsburg, 1954. Boletín Chileno de Parasitología 35: 18–21.

Eiras JC, Rego AA, Pavanelll GC (1986) Histopathology in *Paulicea lutkeni* (Pisces: Pimelodidae) resulting from infections with *Megathylacus brooksi* and *Jauela glandicephalus* (Cestoda: Proteocephalidae). Journal of Fish Biology 28: 359–365. https://doi.org/10.1111/j.1095-8649.1986.tb05172.x

Escalante H (1983) Larvas plerocercoides de Diphyllobothridae Lühe, 1910: hallazgo en peces marinos de consumo humano en la costa peruana. Boletín Chileno de Parasitología 38: 50–52.

Escalante H (1986) Cestodes de elasmobranquios de la costa peruana. Revista de Ciencias (Lima) 74: 70–74.

Escalante H, Arigaza P, Moreno B (1987) Metacéstodos de tetrafilídeos em teleosteos em la zona norte del mar peruano. Rebiol 6: 15–23.

Escalante H, Carvajal JG (1981) Cestodos tetrafilididos de peces del genero *Mustelus* en la costa peruana. Boletín Chileno de Parasitología 36: 76–78.

Escalante H, Carvajal JG (1984) Larval trypanorhynch cestodes from Peruvian teleost fishes, with descriptions of two new species. Studies on Neotropical Fauna and Environment 19: 185–194. https://doi.org/10.1080/01650528409360658

Escalante H, Miranda H (1986) *Diphyllobothrium pacificum*: hallazgo de larvas plerocercoides en peces marinos del Perú y desarrollo de formas adultas del parásito en *Canis familiaris*. Boletín Chileno de Parasitología 41: 7–13.

Euzet L, Carvajal JG (1973) *Rhinebothrium* (Cestoda, Tetraphyllidea) parasites de raies du genre *Psammobatis* au Chili. Bulletin du Museum National d'Histoire Naturelle, Paris 137: 779–787.

Faria A, Silva D (1934) Garoupa vermelha de Abrolhos e São Tomé "Garoupa Bichada" *Tetrarhynchus*. Primeiro Congresso Nacional de Pesca, Rio de Janeiro 1: 237–250.

Feijó LMF, Rodrigues HO, Rodrigues SS (1979) Contribuição ao estudo da fauna helmintológica de sardinhas (*Sardinella* sp.) do litoral do Estado do rio de Janeiro. Atas da Sociedade de Biologia do Rio de Janeiro 20: 23–27.

Felizardo NN, Torres EJL, Fonsesca MCG, Pinto RM, Gomes DC, Knoff M (2010) Cestodes of the flounder *Paralichthys isosceles* Jordan, 1890 (Osteichthyes – Paralichthyidae) from the state of Rio de Janeiro, Brazil. Neotropical Helminthology 4: 113–126.

Fernández J (1985) Estudio parasitológico de *Merluccius australis* (Hutton, 1872) (Pisces: Merluccidae): aspectos sistemáticos, estadísticos y zoogeográficos. Boletín de la Sociedad de Biología de Concepción 56: 31–41.

Fernández J (1987) Los parásitos de la lisa *Mugil cephalus* L., en Chile: sistemática y aspectos poblacionales (Perciformes: Mugilidae). Gayana Zoología 51: 3–58.

Fernández J, Villalba C, Albiña A (1986) Parásitos del pejegallo, *Callorhynchus callorhynchus* (L.), en Chile: aspectos biológicos y sistemáticos. Biología Pesquera 15: 63–73.

Fernández MV, Brugni NL, Viozzi GP, Semenas L (2010) The relationship between fish assemblages and the helminth communities of a prey fish, in a group of small shallow lakes. Journal of Parasitology 96: 1066–1071. https://doi.org/10.1645/GE-2380.1

Fernández MV, Semenas L, Viozzi GP (2012) Parasites of the "Peladilla," *Aplochiton zebra* (Osmeriformes: Galaxiidae), from Patagonia (Argentina and Chile). Comparative Parasitology 79: 231–237. https://doi.org/10.1654/4561.1

Ferraris CJ Jr (2007) Checklist of catfishes, recent and fossil (Osteichthyes: Siluriformes), and catalogue of siluriform primary types. Zootaxa 1418: 1–628.

Ferreira MF, São Clemente SC, Tortelly R, Lima FC, Nascimento ER, Oliveira GA, Lima AR (2006) Parasitas da ordem Trypanorhyncha: sua importância na inspeção sanitária do pescado. Revista Brasileira de Ciência Veterinária 13: 190–193. https://doi.org/10.4322/rbcv.2014.297

Flores K, George-Nascimento M (2009) Las infracomunidades de parásitos de dos especies de *Scartichthys* (Pisces: Blenniidae) en localidades cercanas del norte de Chile. Revista Chilena de Historia Natural 82: 63–71. https://doi.org/10.4067/S0716-078X2009000100004

Fonseca MCG, São Clemente SC, Felizardo NN, Gomes DC, Knoff M (2012) Trypanorhyncha cestodes of hygienic-sanitary importance infecting flounders *Paralichthys patagonicus* Jordan, 1889 and *Xystreurys rasile* (Jordan, 1891) of the Neotropical region, Brazil. Parasitology Research 111: 865–874. https://doi.org/10.1007/s00436-012-2912-z

Fortes E (1981) Descrição de quatro espécies novas de helmintos de bagres (Pisces, Bagridae) do estuário do Guaíba, Porto Alegre, RS, Brasil. Arquivos da Faculdade de Veterinária UFRGS 9: 69–78.

Fortes E, Hoffmann RP (1987) Hiperparasitismo de cestódeos de *Rhamdia sapo* Valenciennes, 1840 do estuário do Guaíba, Rio Grande do Sul, Brasil. Revista Brasileira de Medicina Veterinária 9: 114.

Fortes E, Hoffmann RP (1995) Levantamento da fauna parasitária de peixes do lago Guaíba, Porto Alegre, Rio Grande do Sul, Brasil. Revista Brasileira de Medicina Veterinária 17: 107–111.

Franceschini L, Zago AC, Zocoller-Seno MC, Veríssimo-Silveira R, Ninhaus-Silveira A, Silva RJ (2013) Endohelminths in *Cichla piquiti* (Perciformes, Cichlidae) from the Paraná River, São Paulo State, Brazil. Revista Brasileira de Parasitologia Veterinária 22: 475–484. https://doi.org/10.1590/S1984-29612013000400006

Freze VI (1965) [Proteocephalata. Tapeworms of Fish, Amphibians and Reptiles]. In: Skryabin KI (Ed.) [Principles of Cestodology. Vol. 5]. Izdatel'stvo «Nauka«, Moscow, 538 pp. [In Russian] [English translation, Israel Program of Scientific Translation, 1969, Cat. No. 1853. v + 597 pp.]

Froese R, Pauly D (Eds) 2016. FishBase. World Wide Web electronic publication. http://www.fishbase.org, version (06/2016) [accessed 1.V.2016]

Fuhrmann O (1916) Eigentümliche Fischcestoden. Zoologischer Anzeiger 46: 385–398.

Fuhrmann O (1934) Vier Diesing'sche Typen (Cestoda). Revue Suisse de Zoologie 41: 545–564.

Fuster de Plaza ML, Boschi EE (1957) Desnutrición y deformaciones vertebrales en pejerreyes de los embalses de Córdoba. Ministerio de Agricultura y Ganadería. Departamento de Investigaciones Pesqueras, Buenos Aires, 26 pp.

Garcías F, Mendoza R, George-Nascimento M (2001) Variación entre años de las infracomunidades de parásitos metazoos de la corvina *Cilus gilberti* (Pisces: Sciaenidae) en Chile. Revista Chilena de Historia Natural 74: 833–840. https://doi.org/10.4067/S0716-078X2001000400010

George-Nascimento M (1996) Populations and assemblages of parasites in hake, *Merluccius gayi*, from the southeastern Pacific Ocean: stock implications. Journal of Fish Biology 48: 557–568. https://doi.org/10.1111/j.1095-8649.1996.tb01452.x

George-Nascimento M (2000) Geographical variations in the jack mackerel *Trachurus symmetricus murphyi* populations in the southeastern Pacific Ocean as evidenced from the associated parasite communities. Journal of Parasitology 86: 929–932. https://doi.org/10.1645/0022-3395(2000)086[0929:GVITJM]2.0.CO;2

George-Nascimento M, Arancibia H (1992) Stocks ecológicos del jurel (*Trachurus symmetricus murphyi* Nichols) en tres zonas de pesca frente a Chile, detectados mediante comparación de su fauna parasitaria y morfometría. Revista Chilena de Historia Natural 65: 453–470.

George-Nascimento M, Arancibia H (1994) La fauna parasitaria y la morfometría de la merluza austral *Merluccius australis* (Hutton) como indicadoras de unidades de stock. Biología Pesquera 23: 31–47.

George-Nascimento M, Garcías F, Muñoz G (2002) Parasite body volume and infracommunity patterns in the southern pomfret *Brama australis* (Pisces: Bramidae). Revista Chilena de Historia Natural 75: 835–839. https://doi.org/10.4067/S0716-078X2002000400016

George-Nascimento M, Huet B (1984) Una aproximación ecológica al estudio del parasitismo en el congrio negro *Genypterus maculatus* (Tschudi) (Pisces: Ophidiidae). Biología Pesquera 13: 23–30.

George-Nascimento M, Iriarte JL (1989) Las infracomunidades de parásitos metazoos del chancharro *Helicolenus lengerichi* Norman, 1937 (Pisces, Scorpaenidae): un ensamble no interactivo de especies. Revista Chilena de Historia Natural 62: 217–227.

George-Nascimento M, Mellado A, Saavedra S, Carvajal JG (2009) Variabilidad de las comunidades de parásitos metazoos del róbalo *Eleginops maclovinus* (Cuvier & Valenciennes, 1830) (Pisces: Eleginopidae) en Chile. Revista Chilena de Historia Natural 82: 199–207. https://doi.org/10.4067/S0716-078X2009000200003

George-Nascimento M, Moscoso D (2013) Variación local y geográfica de las infracomunidades de parásitos de la anchoveta *Engraulis ringens* en Chile. Revista de Biología Marina y Oceanografía 48: 207–212. https://doi.org/10.4067/S0718-19572013000100020

George-Nascimento M, Moscoso D, Niklitschek E, González K (2011) Variación geográfica de las comunidades de parásitos de la merluza de tres aletas *Micromesistius australis* al sur de Sudamérica. Revista de Biología Marina y Oceanografía 46: 53–58. https://doi.org/10.4067/S0718-19572011000100007

George-Nascimento M, Oliva M (2015) Fish population studies using parasites from the Southeastern Pacific Ocean: considering host population changes and species body size as sources of variability of parasite communities. Parasitology 142: 25–35. https://doi.org/10.1017/S0031182014001127

George-Nascimento M, Ortiz E (1982) Nuevos registros de huésped para el plerocercoide de *Hepatoxylon trichiuri* (Holten, 1802) (Cestoda: Trypanorhyncha) en peces marinos chilenos. Parasitología al Día 6: 39.

Gibson DI (1994) Order Amphilinidea Poche, 1922. In: Khalil LF, Jones A, Bray RA (Eds) Keys to the Cestode Parasites of Vertebrates. CAB International, Wallingford, 3–10.

Gibson DI, Bray RA, Harris EA (Compilers) (2005) Host-Parasite Database of the Natural History Museum, London. World Wide Web electronic publication. http://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-parasites/ [accessed 1.IV.2016]

Gil de Pertierra AA (1995) *Nomimoscolex microacetabula* sp. n. y *N. pimelodi* sp. n. (Cestoda: Proteocephalidea) parasitos de Siluriformes del Rio de la Plata. Neotrópica 41: 19–26.

Gil de Pertierra AA (2002a) *Nomimoscolex semenasae* n. sp. (Proteocephalidea: Monticelliidae), a cestode parasite of *Diplomystes viedmensis* (Pisces: Siluriformes) from the Patagonian region of Argentina. Systematic Parasitology 53: 183–190. https://doi.org/10.1023/A:1021151225726

Gil de Pertierra AA (2002b) Redescription of *Proteocephalus bagri* and *P. rhamdiae* (Cestoda: Proteocephalidac), parasites of *Rhamdia quelen* (Siluriformes: Pimelodidae) from South America, with comments on morphological variation. Folia Parasitologica 49: 55–66. https://doi.org/10.14411/fp.2002.012

Gil de Pertierra AA (2003) Two new species of *Nomimoscolex* (cestoda: Proteocephalidea, Monticelliidae) from *Gymnotus carapo* (Pisces: gymnotiformes) in Argentina. Memórias do Instituto Oswaldo Cruz 98: 345–352. https://doi.org/10.1590/S0074-02762003000300009

Gil de Pertierra AA (2004) Redescription of *Monticellia magna* (Rego, dos Santos & Silva, 1974) (Eucestoda: Monticelliidae) parasite of *Pimelodus* spp. (Pisces: Siluriformes) from Argentina, and morphological study of microtriches. Revue Suisse de Zoologie 111: 11–20. https://doi.org/10.5962/bhl.part.80222

Gil de Pertierra AA (2005) Comparative study of the microtriches of adult cestodes (Proteocephalidea: Monticelliidae), and some comments on their systematic value. Zoologischer Anzeiger 243: 295–304. https://doi.org/10.1016/j.jcz.2005.01.002

Gil de Pertierra AA (2009) *Luciaella ivanovae* n. g., n. sp. (Eucestoda: Proteocephalidea: Peltidocotylinae), a parasite of *Ageneiosus inermis* (L.) (Siluriformes: Auchenipteridae) in Argentina. Systematic Parasitology 73: 71–80. https://doi.org/10.1007/s11230-009-9174-x

Gil de Pertierra AA, Arredondo NJ, Kuchta R, Incorvaia IS (2015) A new species of *Bothriocephalus* Rudolphi, 1808 (Eucestoda: Bothriocephalidea) from the channel bull blenny *Cottoperca gobio* (Günther) (Perciformes: Bovichtidae) on the Patagonian shelf off Argentina. Systematic Parasitology 90: 247–256. https://doi.org/10.1007/s11230-015-9549-0

Gil de Pertierra AA, de Chambrier A (2000) *Rudolphiella szidati* sp. n. (Proteocephalidea: Monticelliidae, Rudolphiellinae) parasite of *Luciopimelodus pati* (Valenciennes, 1840) (Pisces: Pimelodidae) from Argentina with new observations on *Rudolphiella lobosa* (Riggenbach, 1895). Revue Suisse de Zoologie 107: 81–95. https://doi.org/10.5962/bhl.part.80119

Gil de Pertierra AA, de Chambrier A (2013) *Harriscolex nathaliae* n. sp. (Cestoda: Proteocephalidea) from *Pseudoplatystoma corruscans* (Siluriformes: Pimelodidae) in the Paraná River Basin, Argentina. Journal of Parasitology 99: 480–486. https://doi.org/10.1645/12-11.1

Gil de Pertierra AA, Incorvaia IS, Arredondo NJ (2011) Two new species of *Clestobothrium* (Cestoda: Bothriocephalidea), parasites of *Merluccius australis* and *M. hubbsi* (Gadiformes: Merlucciidae) from the Patagonian shelf of Argentina, with comments on *Clestobothrium crassiceps* (Rudolphi, 1819). Folia Parasitologica 58: 121–134. https://doi.org/10.14411/fp.2011.012

Gil de Pertierra AA, Ostrowski de Núñez M (1990) Seasonal dynamics and maturation of the cestode *Proteocephalus jandia* (Woodland, 1933) in the catfish (*Rhamdia sapo*). Acta Parasitologica Polonica 35: 305–313.

Gil de Pertierra AA, Semenas LG (2005) *Galaxitaenia toloi* n. gen., n. sp. (Eucestoda: Pseudophyllidea) from *Galaxias platei* (Pisces: Osmeriformes, Galaxiidae), in the Patagonian region of Argentina. Journal of Parasitology 91: 900–908. https://doi.org/10.1645/GE-437R.1

Gil de Pertierra AA, Semenas LG (2006) *Ailinella mirabilis* gen. n., sp. n. (Eucestoda: Pseudophyllidea) from *Galaxias maculatus* (Pisces: Galaxiidae) in the Andean-Patagonian region of Argentina. Folia Parasitologica 53: 276–286. https://doi.org/10.14411/fp.2006.034

Gil de Pertierra AA, Viozzi GP (1999) Redescription of *Cangatiella macdonaghi* (Szidat y Nani, 1951) comb. nov. (Cestoda: Proteocephalidae) a parasite of the atheriniform fish *Odontesthes hatcheri* (Eigenmann, 1909) from the Patagonian region of Argentina. Neotrópica 45: 13–20.

Gomes DC, Fabio SP, Rolas FJT (1972) Contribution to the knowledge of the parasites of fishes in Guanabara State. Memórias do Instituto Oswaldo Cruz 70: 541–553. https://doi.org/10.1590/S0074-02761972000400005

Gomes DC, Knoff M, São Clemente SC, Lanfredi RM, Pinto RM (2005) Taxonomic reports of Homeacanthoidea (Eucestoda: Trypanorhyncha) in lamnid and sphyrnid elasmobranchs collected off the coast of Santa Catarina, Brazil. Parasite 12: 15–22. https://doi.org/10.1051/parasite/2005121015

Gonçalves RA, Oliveira MSB, Neves LR, Tavares-Dias M (2016) Seasonal pattern in parasite infracommunities of *Hoplerythrinus unitaeniatus* and *Hoplias malabaricus* (Actinopterygii: Erythrinidae) from the Brazilian Amazon. Acta Parasitologica 61: 119–129. https://doi.org/10.1515/ap-2016-0016

González H, Garrido V, Landeta M, Martens P (1980) New data for the identification of *Diphyllobothrium* sp. in Lake Rupanco, Chile. Boletín Chileno de Parasitología 35: 10–14.

González H, Garrido V, Martens P, Aguirrebeña R (1978) Identificación de *Diphyllobothrium* sp. en especies salmonideas del lago Rupanco, Chile. Boletín Chileno de Parasitología 33: 25–34.

González L, Carvajal JG (1994) Estudio parasitológico de *Merluccius australis* (Hutton, 1972) del mar interior de Aysén. Investigación Pesquera 38: 75–85.

González MT, Acuña E, Oliva ME (2001) Metazoan parasite fauna of the bigeye flounder, *Hippoglossina macrops*, from northern Chile. Influence of host age and sex. Memórias do Instituto Oswaldo Cruz 96: 1049–1054. https://doi.org/10.1590/S0074-02762001000800004

González MT, Barrientos C, Moreno C (2006) Biogeographical patterns in endoparasite communities of a marine fish (*Sebastes capensis* Gmelin) with extended range in the Southern Hemisphere. Journal of Biogeography 33: 1086–1095. https://doi.org/10.1111/j.1365-2699.2006.01488.x

González MT, Poulin R (2005a) Nested patterns in parasite component communities of a marine fish along its latitudinal range on the Pacific coast of South America. Parasitology 131: 569–577. https://doi.org/10.1017/S0031182005007900

González MT, Poulin R (2005b) Spatial and temporal predictability of the parasite community structure of a benthic marine fish along its distributional range. International Journal for Parasitology 35: 1369–1377. https://doi.org/10.1016/j.ijpara.2005.07.016

González MT, Vásquez R, Acuna E (2008) Biogeographic patterns of metazoan parasites of the bigeye flounder, *Hippoglossina macrops*, in the Southeastern Pacific coast. Journal of Parasitology 94: 429–435. https://doi.org/10.1645/GE-1265.1

Graça RJ, Machado MH (2007) Ocorrência e aspectos ecológicos de metazoários parasitos de peixes do Lago do Parque do Ingá, Maringá, Estado do Paraná. Acta Scientiarum: Biological Sciences 29: 321–326. https://doi.org/10.4025/actascibiolsci.v29i3.507

Guagliardo S, Schwerdt C, Galeano N, González R, Tanzola RD (2014) Helminthic assemblages of *Seriolella porosa* Guichenot 1848 (Pisces: Centrolophidae) from San Matías Gulf (Argentina). Neotropical Helminthology 8: 84–96.

Guagliardo S, Tanzola D, Schwerdt C, Galeano N (2009) Host-parasite relationships between *Porichthys porosissimus* (Pisces) and cestode larvae of the *Scolex* group. Helminthologia 46: 162–167. https://doi.org/10.2478/s11687-009-0031-x

Guidelli G, Tavechio WLG, Takemoto RM, Pavanelli GC (2006) Fauna parasitária de *Leporinus lacustris* e *Leporinus friderici* (Characiformes, Anostomidae) da planície de inundação do alto rio Paraná, Brasil. Acta Scientiarum: Biological Sciences 28: 281–290. https://doi.org/10.4025/actascibiolsci.v28i3.228

Guidelli G, Tavechio WLG, Takemoto RM, Pavanelli GC (2011) Relative condition factor and parasitism in anostomid fishes from the floodplain of the Upper Paraná River, Brazil. Veterinary Parasitology 177: 145–151. https://doi.org/10.1016/j.vetpar.2010.11.035

Guidelli GM, Isaac A, Takemoto RM, Pavanelli GC (2003) Endoparasite infracommunities of *Hemisorubim platyrhynchos* (Valenciennes, 1840) (Pisces: Pimelodidae) of the Baía River, upper Paraná River floodplain, Brazil: specific composition and ecological aspects. Brazilian Journal of Biology 63: 261–268. https://doi.org/10.1590/S1519-69842003000200011

Healy CJ (2003) A revision of *Platybothrium* Linton, 1890 (Tetraphyllidea: Onchobothriidae), with a phylogenetic analysis and comments on host-parasite associations. Systematic Parasitology 56: 85–139. https://doi.org/10.1023/A:1026135528505

Healy CJ, Caira JN, Jensen K, Webster BL, Littlewood DTJ (2009) Proposal for a new tapeworm order, Rhinebothriidea. International Journal for Parasitology 39: 497–511. https://doi.org/10.1016/j.ijpara.2008.09.002

Henríquez VP, González MT (2014) Patterns of variation in parasite component communities and infracommunities of a littoral fish species from the northern coast of Chile. Journal of Helminthology 88: 89–96. https://doi.org/10.1017/S0022149X12000788

Henríquez VP, González MT, Licandeo R, Carvajal JG (2011) Metazoan parasite communities of rock cod *Eleginops maclovinus* along southern Chilean coast and their use as biological tags at a local spatial scale. Journal of Fish Biology 79: 1851–1865. https://doi.org/10.1111/j.1095-8649.2011.03126.x

Hermida M, Carvalho BFL, Cruz C, Saraiva A (2014) Parasites of the mutton snapper *Lutjanus analis* (Perciformes: Lutjanidae) in Alagoas, Brazil. Revista Brasileira de Parasitologia Veterinária 23: 241–243. https://doi.org/10.1590/S1984-29612014023

Hernández-Orts JS, Scholz T, Brabec J, Kuzmina T, Kuchta R (2015) High morphological plasticity and global geographical distribution of the Pacific broad tapeworm *Adenocephalus pacificus* (syn. *Diphyllobothrium pacificum*): molecular and morphological survey. Acta Tropica 149: 168–178. https://doi.org/10.1016/j.actatropica.2015.05.017

Holcman-Spector B, Mañé-Garzón F (1988) Two new species of the genus *Proteocephalus* Weinland, 1858 (Proteocephalidea, Eucestoda) of freshwater fishes *Rhamdia sapo* (Valenciennes, 1840), from Uruguay. Parasitología al Día 12: 148–154.

Hypša V, Škeříková A, Scholz T (2005) Phylogeny, evolution and host–parasite relationships of the order Proteocephalidea (Eucestoda) as revealed by combined analysis and secondary structure characters. Parasitology 130: 359–371. https://doi.org/10.1017/S0031182004006638

Iannacone J (2003) Three metazoan parasites of palm ruff *Seriolella violacea* Guichenot (Pisces, Centrolophidae), Callao, Peru. Revista Brasileira de Zoologia 20: 257–260. https://doi.org/10.1590/S0101-81752003000200014

Iannacone J, Alvariño L (2009) Dinámica poblacional de la diversidad parasitaria de la "Cabrilla" *Paralabrax humeralis* (Teleostei: Serranidae) en Chorrillos, Lima, Perú. Neotropical Helminthology 3: 73–88.

Iannacone J, Alvariño L (2013) Parasitological indices of Pacific pomfret *Brama japonica* Hilgendorf, 1878 (Osteichthyes, Bramidae) acquired at the fishing terminal of Chorrillos Lima, Peru. Neotropical Helminthology 7: 117–132.

Iannacone J, Alvariño L, Chero J, Sáez G (2015) Comunidad parasitaria de cabinza *Isacia conceptionis* (Cuvier & Valenciennes, 1830) (Perciformes: Haemulidae) en la Zona de Chorrillos, Lima, Perú. Revista de Investigaciones Veterinarias del Perú 26: 96–110. https://doi.org/10.15381/rivep.v26i1.10943

Iannacone J, Avila-Peltroche J, Rojas-Perea S, Salas-Sierralta M, Neira-Cruzado K, Palomares-Torres R, Valdivia-Alarcón S, Pacheco-Silva A, Benvenutto-Vargas V, Ferrario-Bazalar V (2011) Dinámica poblacional de los parásitos metazoos del pez guitarra del pacífico *Rhinobatos planiceps* (Batoidea: Rajiformes) de la zona costera marina de Lima, Perú. Neotropical Helminthology 5: 265–278.

Isaac A, Guidelli GM, França JG, Pavanelli GC (2004) Composição e estrutura das infracomunidades endoparasitárias de *Gymnotus* spp. (Pisces: Gymnotidae) do rio Baía, Mato Grosso do Sul, Brasil. Acta Scientiarum: Biological Sciences 26: 453–462. https://doi.org/10.4025/actascibiolsci.v26i4.1527

Ivanov VA (1997) *Echinobothrium notoguidoi* n. sp. (Cestoda: Diphyllidea) from *Mustelus schmitti* (Chondrichthyes: Carcharhiniformes) in the Argentine Sea. Journal of Parasitology 83: 913–916. https://doi.org/10.2307/3284288

Ivanov VA (2004) A new species of *Rhinebothroides* Mayes, Brooks & Thorson, 1981 (Cestoda: Tetraphyllidea) from the ocellate river stingray in Argentina, with amended descriptions of two other species of the genus. Systematic Parasitology 58: 159–174. https://doi.org/10.1023/B:SYPA.0000032933.71645.1e

Ivanov VA (2005) A new species of *Acanthobothrium* (Cestoda: Tetraphyllidea: Onchobothriidae) from the ocellate river stingray, *Potamotrygon motoro* (Chondrichthyes: Potamotrygonidae), in Argentina. Journal of Parasitology 91: 390–396. https://doi.org/10.1645/GE-354R1

Ivanov VA (2006) *Guidus* n. gen. (Cestoda: Tetraphyllidea), with description of a new species and emendation of the generic diagnosis of *Marsupiobothrium*. Journal of Parasitology 92: 832–840. https://doi.org/10.1645/GE-767R.1

Ivanov VA (2008) *Orygmatobothrium* spp. (Cestoda: Tetraphyllidea) from triakid sharks in Argentina: redescription of *Orygmatobothrium schmittii* and description of a new species. Journal of Parasitology 94: 1087–1097. https://doi.org/10.1645/GE-1482.1

Ivanov VA (2009) New species of *Crossobothrium* (Cestoda: Tetraphyllidea) from the broadnose sevengill shark, *Notorynchus cepedianus*, in Argentina. Journal of Parasitology 95: 1479–1488. https://doi.org/10.1645/GE-2096.1

Ivanov VA, Brooks DR (2002) *Calliobothrium* spp. (Eucestoda: Tetraphyllidea: Onchobothriidae) in *Mustelus schmitti* (Chondrichthyes: Carcharhiniformes) from Argentina and Uruguay. Journal of Parasitology 88: 1200–1213. https://doi.org/10.2307/3285494

Ivanov VA, Campbell RA (1998a) A new species of *Acanthobothrium* van Beneden, 1849 (Cestoda: Tetraphyllidea) from *Rioraja castelnaui* (Chondrichthyes: Rajoidei) in coastal waters of Argentina. Systematic Parasitology 40: 203–212. https://doi.org/10.1023/A:1006049404646

Ivanov VA, Campbell RA (1998b) *Echinobothrium megacanthum* sp. n. (Cestoda: Diphyllidea) from the eagle ray *Myliobatis goodei* (Chondrichthyes: Rajoidei) from the Patagonian shelf of Argentina. Folia Parasitologica 45: 225–229.

Ivanov VA, Campbell RA (2000) Emendation of the generic diagnosis of *Tylocephalum* (Cestoda: Lecanicephalidea: Tetragonocephalidae), and description of *Tylocephalum brooksi* n. sp. Journal of Parasitology 86: 1085–1092. https://doi.org/10.2307/3284827

Ivanov VA, Campbell RA (2002) *Notomegarhynchus navonae* n. gen. and n. sp. (Eucestoda: Tetraphyllidea), from skates (Rajidae: Arhynchobatinae) in the Southern Hemisphere. Journal of Parasitology 88: 340–349. https://doi.org/10.1645/0022-3395(2002)088[0340:NNNGAN]2.0.CO;2

Janicki C (1808) Über den Bau von *Amphilina liguloidea* Diesing. Zeitschrift für Wissenschaftliche Zoologie 89: 568–597. http://www.biodiversitylibrary.org/part/26850

Jara CA (1998) Prevalencia e intensidad de parasitismo por helmintos en cuatro especies de peces de la zona norte del mar peruano. Revista Peruana de Parasitología 13: 76–83.

Jensen K (2001) Four new genera and five new species of lecanicephalideans (Cestoda: Lecanicephalidea) from elasmobranchs in the Gulf of California, Mexico. Journal of Parasitology 87: 845–861. https://doi.org/10.1645/0022-3395(2001)087[0845:FNGAFN]2.0.CO;2

Jensen K, Bullard SA (2010) Characterization of a diversity of tetraphyllidean and rhinebothriidean cestode larval types, with comments on host associations and life-cycles. International Journal for Parasitology 40: 889–910. https://doi.org/10.1016/j.ijpara.2009.11.015

Jensen K, Caira JN, Cielocha JJ, Littlewood DTJ, Waeschenbach A (2016) When proglottids and scoleces conflict: phylogenetic relationships and a family-level classification of the Lecanicephalidea (Platyhelminthes: Cestoda). International Journal for Parasitology 46: 291–310. https://doi.org/10.1016/j.ijpara.2016.02.002

Jerônimo GT, Ventura AS, Pádua SBD, Satake F, Ishikawa MM, Martins ML (2013) Parasitofauna de cachara cultivado em tanque-rede no Rio Paraguai. Pesquisa Agropecuária Brasileira: 1163–1166. https://doi.org/10.1590/S0100-204X2013000800054

Karling LC, Isaac A, Affonso IP, Takemoto RM, Pavanelli GC (2013a) The impact of a dam on the helminth fauna and health of a neotropical fish species *Salminus brasiliensis* (Cuvier 1816) from the upper Paraná River, Brazil. Journal of Helminthology 87: 245–251. https://doi.org/10.1017/S0022149X1200034X

Karling LC, Lacerda ACF, Takemoto RM, Pavanelli GC (2013b) Ecological relationships between endoparasites and the fish *Salminus brasiliensis* (Characidae) in a Neotropical floodplain. Neotropical Helminthology 7: 219–230.

Kennedy CR, Andersen KI (1982) Redescription of *Anchistrocephalus microcephalus* (Rudolphi) (Cestoda, Pseudophyllidea) from the sunfish *Mola mola*. Zoologica Scripta 11: 101–105. https://doi.org/10.1111/j.1463-6409.1982.tb00522.x

Khalil LF, Jones A, Bray RA (1994) Keys to the Cestode Parasites of Vertebrates. CAB international, Wallingford, 768 pp.

Knoff M, Luque JL, Amato JF (1997) Community ecology of the metazoan parasites of grey mullets, *Mugil platanus* (Osteichthyes: Mugilidae) from the littoral of the state of Rio de Janeiro, Brazil. Revista Brasileira de Biologia 57: 441–454.

Knoff M, São Clemente SC, Andrada CDG, Lima FC, Padovani RES, Fonseca MCG, Neves RCF, Gomes DC (2008) Cestóides Pseudophyllidea parasitos de congro-rosa, *Genypterus brasiliensis* Regan, 1903 comercializados no estado do Rio de Janeiro, Brasil. Revista Brasileira de Ciência Veterinária 15: 28–32. https://doi.org/10.4322/rbcv.2014.192

Knoff M, São Clemente SC, Fonseca MCG, Felizardo NN, Pinto RM, Gomes DC (2011) Cestodes Diphyllobothriidea parasitizing blackfin goosefish, *Lophius gastrophysus* Miranda-Ribeiro, 1915. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 63: 1033–1038. https://doi.org/10.1590/S0102-09352011000400035

Knoff M, São Clemente SC, Pinto RM, Gomes DC (2002) Prevalência e intensidade de infecção de cestóides Trypanorhyncha de elasmobrânquios nos estados do Paraná e Santa Catarina, Brasil. Parasitología Latinoamericana 57: 149–157. https://doi.org/10.4067/S071777122002000300012

Knoff M, São Clemente SC, Pinto RM, Gomes DC (2004a) Registros taxonômicos de cestóides Trypanorhyncha/Homeacanthoidea em elasmobrânquios coletados na costa do Estado do Paraná, Brasil. Parasitología Latinoamericana 59: 31–36. https://doi.org/10.4067/S0717-77122004000100006

Knoff M, São Clemente SC, Pinto RM, Lanfredi RM, Gomes DC (2004b) New records and expanded descriptions of *Tentacularia coryphaenae* and *Hepatoxylon trichiuri* homeacanth trypanorhynchs (Eucestoda) from carcharhinid sharks from the State of Santa Catarina offshore, Brazil. Revista Brasileira de Parasitologia Veterinária 13: 73–80.

Knoff M, São Clemente SC, Pinto RM, Lanfredi RM, Gomes DC (2004c) Taxonomic reports of Otobothrioidea (Eucestoda, Trypanorhyncha) from elasmobranch fishes of the southern coast off Brazil. Memórias do Instituto Oswaldo Cruz 99: 31–36. https://doi.org/10.1590/S0074-02762004000100006

Knoff M, São Clemente SC, Pinto RM, Lanfredi RM, Gomes DC (2007) Redescription of *Gymnorhynchus isuri* (Cestoda: Trypanorhyncha) from *Isurus oxyrinchus* (Elasmobranchii: Lamnidae). Folia Parasitologica 54: 208–214. https://doi.org/10.14411/fp.2007.028

Koch KR, Jensen K, Caira JN (2012) Three new genera and six new species of lecanicephalideans (Cestoda) from eagle rays of the genus *Aetomylaeus* (Myliobatiformes: Myliobatidae) from northern Australia and Borneo. Journal of Parasitology 98: 175–198. https://doi.org/10.1645/GE-2798.1

Kohn A, Fernandes BMM (1987) Estudo comparativo dos helmintos parasitos de peixes do Rio Mogi Guassu, coletados nas excursões realizadas entre 1927 e 1985. Memórias do Instituto Oswaldo Cruz 82: 483–500. https://doi.org/10.1590/S0074-02761987000400006

Kohn A, Fernandes BMM, Baptista-Farias DMF, Cohen SC, Santos AL, Pamplona-Basilio MC, Vieira MJAF, Feitosa VA (2004) Prevalência de helmintos parasitos dos peixes do açude Pereira de Miranda e dos viveiros do DNOCS (Pentecoste, Ceará, Brasil). Revista Brasileira de Ciência Veterinária 11: 55–57. https://doi.org/10.4322/rbcv.2014.344

Kohn A, Fernandes BMM, Macedo B, Abramson B (1985) Helminths parasites of freshwater fishes from Pirassununga, SP, Brazil. Memórias do Instituto Oswaldo Cruz 80: 327–336. https://doi.org/10.1590/S0074-02761985000300009

Kohn A, Fernandes BMM, Pipolo HV, Godoy MP (1988) Helmintos parasitos de peixes das usinas hidreléticas da Eletrosul (Brasil). II: Reservatórios de Salto Osório e de Salto Santiago, Bacia do Rio Iguaçu. Memórias do Instituto Oswaldo Cruz 83: 299–303. https://doi.org/10.1590/S0074-02761988000300006

Kohn A, Moravec F, Cohen SC, Canzi C, Takemoto RM, Fernandes BMM (2011) Helminths of freshwater fishes in the reservoir of the Hydroelectric Power Station of Itaipu, Paraná, Brazil. Check List 7: 681–690. https://doi.org/10.15560/7.5.681

Kuchta R, Scholz T (2007) Diversity and distribution of fish tapeworms of the "Bothriocephalidea" (Eucestoda). Parassitologia 49: 129–146.

Kuchta R, Scholz T, Bray RA (2008) Revision of the order Bothriocephalidea Kuchta, Scholz, Brabec & Bray, 2008 (Eucestoda) with amended generic diagnoses and keys to families and genera. Systematic Parasitology 71: 81–136. https://doi.org/10.1007/s11230-008-9153-7

Kuchta R, Serrano-Martínez ME, Scholz T (2015) Pacific broad tapeworm *Adenocephalus pacificus* as a causative agent of globally reemerging diphyllobothriosis. Emerging Infectous Diseases 21: 1697–1703. https://doi.org/10.3201/eid2110.150516

Kullander SO, Ferreira EJG (2006) A review of the South American cichlid genus *Cichla*, with descriptions of nine new species (Teleostei: Cichlidae). Ichthyological Exploration of Freshwaters 17: 289–398.

Kuraiem BP, Knoff M, Felizardo NN, Gomes DC, São Clemente SC (2016) *Callitetrarhynchus speciosus* (Linton, 1897) Carvajal & Rego, 1985 Trypanorhyncha (Cestoda) parasitizing *Priacanthus arenatus* (Cuvier, 1829) (Osteichthyes, Priacanthidae) from Rio de Janeiro coast, Brazil. Neotropical Helminthology 10: 33–40.

La Rue GR (1911) A revision of the cestode family Proteocephalidae. Zoologischer Anzeiger 38: 473–482. http://www.biodiversitylibrary.org/item/95304#page/485/mode/1up

La Rue GR (1914) A revision of the cestode family Proteocephalidae. Illinois Biological Monographs 1: 1–349. https://doi.org/10.5962/bhl.title.16792

Lacerda ACF, Takemoto RM, Pavanelli GC (2008) Digenea, Nematoda, Cestoda, and Acanthocephala, parasites in Potamotrygonidae (Chondrichthyes) from the upper Paraná River floodplain, states of Paraná and Mato Grosso do Sul, Brazil. Check List 4: 115–122. https://doi.org/10.15560/4.2.115

Lacerda ACF, Takemoto RM, Pavanelli GC (2009) Ecology of endoparasites of the fluvial stingray *Potamotrygon falkneri* (Chondrichthyes: Potamotrygonidae) from the upper Paraná River floodplain, Brazil. Brazilian Journal of Biology 69: 297–303. https://doi.org/10.1590/S1519-69842009000200009

Lacerda ACF, Takemoto RM, Poulin R, Pavanelli GC (2013) Parasites of the fish *Cichla piquiti* (Cichlidae) in native and invaded Brazilians basins: release not from the enemy, but from its effects. Parasitology Research 112: 279–288. https://doi.org/10.1007/s00436-012-3135-z

Lacerda ACF, Takemoto RM, Tavares-Dias M, Poulin R, Pavanelli GC (2012) Comparative parasitism of the fish *Plagioscion squamosissimus* in native and invaded river basins. Journal of Parasitology 98: 713–717. https://doi.org/10.1645/GE-2882.1

Lanfranchi AL, Rossin MA, Timi JT (2009) Parasite infracommunities of a specialized marine fish species in a compound community dominated by generalist parasites. Journal of Helminthology 83: 373–378. https://doi.org/10.1017/S0022149X09390069

Leible MD, Carvajal JG, Fuentealba MC (1990) Polimorfismo en *Raja* (*Dipturus*) *flavirostris* Philippi, 1892: análisis morfológico y parasitario. Boletín de la Sociedad de Biología de Concepción 61: 93–102.

Lima FC, São Clemente SC, Mesquita EF (1997) Pseudocistos cecais associados a parasitose por *Scolex pleuronectis* em sardinha-verdadeira (*Sardinella brasiliensis*). Parasitología al Día 21: 58–60.

Littlewood DTJ, Bray RA, Waeschenbach A (2015) Phylogenetic patterns of diversity in the cestodes and trematodes. In: Morand S, Krasnov BR, Littlewood DTJ (Eds) Parasite Diver-

sity and Diversification: Evolutionary Ecology Meets Phylogenetics. Cambridge University Press, Cambridge, 304–319. https://doi.org/10.1017/CBO9781139794749.020

Lizama MAP, Takemoto RM, Pavanelli GC (2005) Influence of host sex and age on infracommunities of metazoan parasites of *Prochilodus lineatus* (Valenciennes, 1836) (Prochilodontidae) of the upper Paraná River floodplain, Brazil. Parasite 12: 299–304. https://doi.org/10.1051/parasite/2005124299

Lizama MAP, Takemoto RM, Pavanelli GC (2006) Parasitism influence on the hepato, splenosomatic and weight/length relation and relative condition factor of *Prochilodus lineatus* (Valenciennes, 1836) (Prochilodontidae) of the Upper Paraná River floodplain, Brazil. Revista Brasileira de Parasitologia Veterinária 15: 116–122.

Lizama MAP, Takemoto RM, Pavanelli GC (2008) Ecological aspects of metazoan parasites of *Astyanax altiparanae* Garutti & Britski, 2000 (Characidae) of the upper Paraná River floodplain, Brazil. Boletim do Instituto de Pesca 34: 527–533.

Llerena CZ, Chávez AV, Casas EA (2001) Frecuencia de larvas Diphyllobothriidae y larvas Anisakidae en peces marinos comerciales del terminal pesquero de Ventanilla-Callao. Revista de Investigaciones Veterinarias del Perú 12: 58–62.

Lopes LPC, Varella AMB, Malta JCO (2009) Metazoan parasites of *Pseudoplatystoma punctifer* (Linnaeus, 1766) and *Pseudoplatystoma tigrinum* (Spix & Agassiz, 1829) Siluriformes: Pimelodidae) of the Central Amazon Basin, Brazil. Biologia Geral e Experimental 9: 3–15.

López E (1994) *Acanthobothroides peruensis* n. sp. (Tetraphyllidea: Onchobothriidae) parásito de *Dasyatis brevis* G. (Rajiformes: Dasyatidae) de la costa del Perú. Biotempo 1: 19–20.

López de McDonald E, Tantaleán MV (1985) Contribución al conocimiento de la helmintofauna de peces marinos de la costa peruana. Parasitología al Día 9: 40–43.

López-Neyra CR, Diaz-Ungría C (1958) Cestodes de Venezuela. V. Cestodes de vertebrados Venezolanos. Novedades Cientificas: Contribuciones Ocasionales del Museo de Historia Natural La Salle 23: 1–42.

Luchetti NM, Marques FPL, Charvet-Almeida P (2008) A new species of *Potamotrygonocestus* Brooks & Thorson, 1976 (Eucestoda: Tetraphyllidea) from *Plesiotrygon iwamae* Rosa, Castello & Thorson (Mylliobatoidea: Potamotrygonidae) and a redescription of *Potamotrygonocestus chaoi* Marques, Brooks & Araujo, 2003. Systematic Parasitology 70: 131–145. https://doi.org/10.1007/s11230-008-9135-9

Luque JL (1991) Formas larvarias de helmintos parásitos en especies marinas del Perú. Parasitología al Día 15: 43–48.

Luque JL, Alves DR (2001) Ecologia das comunidades de metazoários parasitos do xaréu, *Caranx hippos* (Linnaeus) e do xerelete, *Caranx latus* Agassiz (Osteichthyes, Carangidae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Brasileira de Zoologia 18: 399–410. https://doi.org/10.1590/S0101-81752001000200011

Luque JL, Alves DR, Ribeiro RS (2003) Community ecology of the metazoan parasites of banded croaker, *Paralonchurus brasiliensis* (Osteichthyes: Sciaenidae), from the coastal zone of the State of Rio de Janeiro, Brazil. Acta Scientiarum: Biological Sciences 25: 273–278. https://doi.org/10.4025/actascibiolsci.v25i2.2009

Luque JL, Amato JFR, Takemoto RM (1995) Helminth larval stages in *Orthopristis ruber* and *Haemulon steindachneri* (Osteichthyes: Haemulidae) from the coast of the state of Rio de Janeiro, Brazil. Revista Brasileira de Biologia 55: 33–38.

Luque JL, Amato JFR, Takemoto RM (1996a) Comparative analysis of the communities of metazoan parasites of *Orthopristis ruber* and *Haemulon steindachneri* (Osteichthyes: Haemulidae) from the southeastern Brazilian litoral: I. structure and influence of the size and sex of hosts. Revista Brasileira de Biologia 56: 279–292.

Luque JL, Amato JFR, Takemoto RM (1996b) Comparative analysis of the communities of metazoan parasites of *Orthopristis ruber* and *Haemulon steindachneri* (Osteichthyes: Haemulidae) from the southeastern Brazilian litoral: II diversity, interspecifc associations, and distribution of the gastrointestinal parasites. Revista Brasileira de Biologia 56: 292–302.

Luque JL, Chaves ND (1999) Ecologia da comunidade de metazoários parasitos da anchova *Pomatomus saltator* (Linnaeus) (Osteichthyes, Pomatomidae) do litoral do estado do Rio de Janeiro, Brasil. Revista Brasileira de Zoologia 16: 711–723. https://doi.org/10.1590/S0101-81751999000300010

Luque JL, Cordeiro AS, Oliva ME (2010) Metazoan parasites as biological tags for stock discrimination of whitemouth croaker *Micropogonias furnieri*. Journal of Fish Biology 76: 591–600. https://doi.org/10.1111/j.1095-8649.2009.02515.x

Luque JL, Felizardo NN, Tavares LER (2008) Community ecology of the metazoan parasites of namorado sandperches, *Pseudopercis numida* Miranda-Ribeiro, 1903 and *P. semifasciata* Cuvier, 1829 (Perciformes: Pinguipedidae), from the coastal zone of the State of Rio de Janeiro, Brazil. Brazilian Journal of Biology 68: 269–278. https://doi.org/10.1590/S1519-69842008000200007

Luque JL, Porrozzi F, Alves DR (2002) Community ecology of the metazoan parasites of Argentine goatfish, *Mullus argentinae* (Osteichthyes: Mullidae), from the coastal zone of the state of Rio de Janeiro, Brazil. Revista Brasileira de Parasitologia Veterinária 11: 33–38.

Luque JL, Poulin R (2004) Use of fish as intermediate hosts by helminth parasites: a comparative analysis. Acta Parasitologica 49: 353–361.

Luque JL, Poulin R (2007) Metazoan parasite species richness in Neotropical fishes: hotspots and the geography of biodiversity. Parasitology 134: 865–878. https://doi.org/10.1017/S0031182007002272

Lynsdale JA (1959) On a new species of *Proteocephalus* from Brazil. Journal of Helminthology 33: 145–150. https://doi.org/10.1017/S0022149X00033381

MacDonagh E (1927) Parásitos de peces comestibles – III. Dos cestodarios: *Gyrocotyle rugosa* del "Pez gallo" y *Gyrocotyle maxima* n. sp. del "Gatuso". La Semana Médica 34: 1232–1235.

MacDonagh E (1932) Sobre una *Ichthyotaenia* y oncosfera del pejerrey. La Semana Médica 25: 1−16.

Machado MH, Pavanelli GC, Takemoto RM (1994) Influence of host's sex and size on endoparasitic infrapopulations of *Pseudoplatystoma corruscans* and *Schizodon borelli* (Osteichthyes) of the high Paraná River, Brazil. Revista Brasileira de Parasitologia Veterinária 3: 143–148.

Machado MH, Pavanelli GC, Takemoto RM (1995) Influence of the type of environment and of the hydrological level variation in endoparasitic infrapopulations of *Pseudoplatystoma corruscans* (Agassiz) and *Schizodon borelli* (Boulenger) (Osteichthyes) of the high Paraná River, Brazil. Revista Brasileira de Zoologia 12: 961–976. https://doi.org/10.1590/S0101-81751995000400023

Machado MH, Pavanelli GC, Takemoto RM (1996) Structure and diversity of endoparasitic infracommunities and the trophic level of *Pseudoplatystoma corruscans* and *Schizodon borelli*

(Osteichthyes) of the high Paraná River. Memórias do Instituto Oswaldo Cruz 91: 441–448. https://doi.org/10.1590/S0074-02761996000400010

Machado PM, Almeida SC, Pavanelli GC, Takemoto RM (2000) Ecological aspects of endohelminths parasitizing *Cichla monoculus* Spix, 1831 (Perciformes: Cichlidae) in the Paraná River near Porto Rico, State of Paraná, Brazil. Comparative Parasitology 67: 210–217.

MacKenzie K, Brickle P, Hemmingsen W, George-Nascimento M (2013) Parasites of hoki, *Macruronus magellanicus*, in the Southwest Atlantic and Southeast Pacific Oceans, with an assessment of their potential value as biological tags. Fisheries Research 145: 1–5. https://doi.org/10.1016/j.fishres.2013.03.008

MacKenzie K, Longshaw M (1995) Parasites of the hakes *Merluccius australis* and *M. hubbsi* in the waters around the Falkland Islands, southern Chile, and Argentina, with an assessment of their potential value as biological tags. Canadian Journal of Fisheries and Aquatic Sciences 52: 213–224. https://doi.org/10.1139/f95-529

Mancini M, Bucco C, Salinas V, Larriestra A, Tanzola R, Guagliardo S (2008) Seasonal variation of parasitism in pejerrey *Odontesthes bonariensis* (Atheriniformes, Atherinopsidae) from La Viña reservoir (Córdoba, Argentina). Revista Brasileira de Parasitologia Veterinária 17: 28–32. https://doi.org/10.1590/S1984-29612008000100006

Mancini M, Grosman F (1998) Aspectos poblacionales del pejerrey *Odontesthes bonariensis* en el embalse Río Tercero, Córdoba. Natura Neotropicalis 2: 137–143.

Mariaux J (1998) A molecular phylogeny of the Eucestoda. Journal of Parasitology 84: 114–124. https://doi.org/10.2307/3284540

Marques FPL, Brooks DR (2003) Taxonomic revision of *Rhinebothroides* (Eucestoda: Tetraphyllidea: Phyllobothriidae), parasites of Neotropical freshwater stingrays (Rajiformes: Myliobatoidei: Potamotrygonidae). Journal of Parasitology 89: 994–1017. https://doi.org/10.1645/GE-3059

Marques F, Brooks DR, Ureña HM (1996) Two new species of tetraphyllidean cestodes in *Himantura pacifica* (Chondrichthyes: Myliobatiformes: Dasyatididae) from the northwest coast of Costa Rica. Journal of Parasitology 82: 302–306. https://doi.org/10.2307/3284165

Marques FPL, Brooks DR, Araújo MLG (2003) Systematics and phylogeny of *Potamotrygonocestus* (Platyhelminthes, Tetraphyllidea, Onchobothriidae) with descriptions of three new species from freshwater potamotrygonids (Myliobatoidei, Potamotrygonidae). Zoologica Scripta 32: 367–396. https://doi.org/10.1046/j.1463-6409.2003.00111.x

Marques FPL, Brooks DR, Barriga R (1997) Six species of *Acanthobothrium* (Eucestoda: Tetraphyllidea) in stingrays (Chondrichthyes: Rajiformes: Myliobatoidei) from Ecuador. Journal of Parasitology 83: 475–484. https://doi.org/10.2307/3284414

Marques FPL, Brooks DR, Lasso CA (2001) *Anindobothrium* n. gen. (Eucestoda: Tetraphyllidea) inhabiting marine and freshwater potamotrygonid stingrays. Journal of Parasitology 87: 666–672. https://doi.org/10.2307/3285110

Marques FPL, Caira JN (2016) *Pararhinebothroides* – neither the sister-taxon of *Rhinebothroides* nor a valid genus. Journal of Parasitology 102: 249–259. https://doi.org/10.1645/15-894

Marques FPL, Jensen K, Caira JN (2012) *Ahamulina* n. gen. (Cestoda: Diphyllidea) from the polkadot catshark, *Scyliorhinus besnardi* (Carcharhiniformes: Scyliorhinidae), off Brazil. Zootaxa: 51–59.

Marques FPL, Reyda FB (2015) *Rhinebothrium jaimei* sp. n. (Eucestoda: Rhinebothriidea: Rhinebothriidae): a new species from Neotropical freshwater stingrays (Potamotrygonidae). Folia Parasitologica 62: 057. https://doi.org/10.14411/fp.2015.057

Martins ML, Pereira J Jr., de Chambrier A, Mouriño JLP (2011) Infection levels of proteocephalidean cestodes in *Cichla piquiti* (Osteichthyes: Cichlidae) of the Volta Grande Reservoir, Minas Gerais, Brazil, relative to host body weight and gender. Journal of Helminthology 85: 462–467. https://doi.org/10.1017/S0022149X10000830

Martins ML, Pereira J Jr., de Chambrier A, Yamashita MM (2009) Proteocephalid cestode infection in alien fish, *Cichla piquiti* Kullander and Ferreira, 2006 (Osteichthyes: Cichlidae), from Volta Grande reservoir, Minas Gerais, Brazil. Brazilian Journal of Biology 69: 189–195. https://doi.org/10.1590/S1519-69842009000100025

Mateo E, Bullock WL (1966) *Neobothriocephalus aspinosus* gen. et sp. n. (Pseudophyllidea: Parabothriocephalidae), from the Peruvian marine fish, *Neptomenus crassus*. Journal of Parasitology 52: 1070–1073. https://doi.org/10.2307/3276350

Mattos DPBG, Verícimo MA, Lopes LMS, São Clemente SC (2015) Immunogenic activity of the fish tapeworm *Pterobothrium heteracanthum* (Trypanorhyncha: Pterobothriidae) in BALB/c mice. Journal of Helminthology 89: 203–207. https://doi.org/10.1017/S0022149X13000795

Mayes MA, Brooks DR (1981) Cestode parasites of some Venezuelan stingrays. Proceedings of the Biological Society of Washington 93: 1230–1238.

Mayes MA, Brooks DR, Thorson TB (1978) Two new species of *Acanthobothrium* van Beneden 1849 (Cestoidea: Tetraphyllidea) from freshwater stingrays in South America. Journal of Parasitology 64: 838–841. https://doi.org/10.2307/3279513

Mayes MA, Brooks DR, Thorson TB (1981) Two new tetraphyllidean cestodes from *Potamotrygon circularis* Garman (Chondrichthyes: Potamotrygonidae) in the Itacuaí River, Brazil. Proceedings of the Biological Society of Washington 48: 38–42.

Mendes MV (1944) Sobre Cestoda de teleósteos marinhos. Boletim da Faculdade de Filosofia, Ciências e Letras da Universidade de São Paulo, Série Zoologia 43: 173–187.

Mendívil-Herrera J (1946) *Gyrocotyle meandrica* n. sp., del intestino espiral de pez gallo, *Callorhynchus callorhynchus* (L.). Comunicaciones Zoologicas del Museo de Historia Natural de Montevideo 36: 1–12.

Menoret A, Ivanov VA (2009a) A new species of tetraphyllidean (Cestoda) from the largespot river stingray, *Potamotrygon falkneri* (Potamotrygonidae: Chondrichthyes), from the Paraná Basin. Journal of Parasitology 95: 994–999. https://doi.org/10.1645/GE-1850.1

Menoret A, Ivanov VA (2009b) New name for *Progrillotia dollfusi* Carvajal et Rego, 1983 (Cestoda: Trypanorhyncha): description of adults from *Squatina guggenheim* (Chondrichthyes: Squatiniformes) off the coast of Argentina. Folia Parasitologica 56: 284–294. https://doi.org/10.14411/fp.2009.033

Menoret A, Ivanov VA (2011) Descriptions of two new freshwater Neotropical species of *Rhinebothrium* (Cestoda: Rhinebothriidea) from *Potamotrygon motoro* (Chondrichthyes: Potamotrygonidae). Folia Parasitologica 58: 178–186. https://doi.org/10.14411/fp.2011.018

Menoret A, Ivanov VA (2012) Description of plerocerci and adults of a new species of *Grillotia* (Cestoda: Trypanorhyncha) in teleosts and elasmobranchs from the Patagonian shelf off Argentina. Journal of Parasitology 98: 1185–1199. https://doi.org/10.1645/GE-3107.1

Menoret A, Ivanov VA (2013) A new species of *Heteronybelinia* (Cestoda: Trypanorhyncha) from *Sympterygia bonapartii* (Rajidae), *Nemadactylus bergi* (Cheilodactylidae) and *Raneya brasiliensis* (Ophidiidae) in the south-western Atlantic, with comments on host specificity of the genus. Journal of Helminthology 87: 467–482. https://doi.org/10.1017/S0022149X12000545

Menoret A, Ivanov VA (2014) Eutetrarhynchid trypanorhynchs (Cestoda) from elasmobranchs off Argentina, including the description of *Dollfusiella taminii* sp. n. and *Parachristianella damiani* sp. n., and amended description of *Dollfusiella vooremi* (Sao Clemente et Gomes, 1989). Folia Parasitologica 61: 411–431. https://doi.org/10.14411/fp.2014.056

Menoret A, Ivanov VA (2015) Trypanorhynch cestodes (Eutetrarhynchidae) from batoids along the coast of Argentina, including the description of new species in *Dollfusiella* Campbell et Beveridge, 1994 and *Mecistobothrium* Heinz et Dailey, 1974. Folia Parasitologica 62: 058. https://doi.org/10.14411/fp.2015.058

Mesquita RLB, Santos SMC, Ceccarelli PS, Luque JL (2012) Metazoários endoparasitos de *Salminus brasiliensis* (Cuvier, 1816) (Characiformes: Characidae) do rio Mogi Guaçu, SP, Brasil. Revista Brasileira de Zoociências 14: 95–102.

Miloslavich P, Klein E, Díaz JM, Hernández CE, Bigatti G, Campos L, Artigas F, Castillo J, Penchaszadeh PE, Neill PE, Carranza A, Retana MV, Díaz de Astarloa JM, Lewis M, Yorio P, Piriz ML, Rodríguez D, Yoneshigue-Valentin Y, Gamboa L, Martín A (2011) Marine biodiversity in the Atlantic and Pacific coasts of South America: knowledge and gaps. PLoS ONE 6: e14631. https://doi.org/10.1371/journal.pone.0014631

Mola P (1906) Di alcune specie poco studiate o mal note di Cestodi. Annuario del Museo Zoologico della R Università di Napoli 2: 1–12.

Monteiro CM, Santos MD, Zuchi NA, Brasil-Sato MC (2009) Ecological parameters of the endohelminths in relation to size and sex of *Prochilodus argenteus* (Actinopterygii: Prochilodontidae) from the Upper São Francisco River, Minas Gerais, Brazil. Zoologia (Curitiba) 26: 753–757. https://doi.org/10.1590/S1984-46702009000400021

Monticelli FS (1891) Noticie su di alcune specie di *Taenia*. Bollettino della Società dei Naturalisti in Napoli 5: 151–174.

Monticelli FS (1892) Appunti sui Cestodaria. Atti della Reale Accademia delle Scienze Fisiche e Matematiche, Napoli 5: 67–78.

Moreira J, Paschoal F, Cezar AD, Luque JL (2015) Community ecology of the metazoan parasites of Brazilian sardinella, *Sardinella brasiliensis* (Steindachner, 1879) (Actinopterygii: Clupeidae) from the coastal zone of the State of Rio de Janeiro, Brazil. Brazilian Journal of Biology 75: 736–741. https://doi.org/10.1590/1519-6984.00114

Moreira LHA, Takemoto RM, Pagotto JPA, Pavanelli GC (2014) Endoparasite community structure of three fish species in tributary streams of the river Pirapó, Paraná state, Brazil. Neotropical Helminthology 8: 97–109.

Moreira ST, Ito KF, Takemoto RM, Pavanelli GC (2005) Ecological aspects of the parasites of *Iheringichthys labrosus* (Lütken, 1874) (Siluriformes: Pimelodidae) in reservoirs of Paraná basin and upper Paraná floodplain, Brazil. Acta Scientiarum: Biological Sciences 27: 317–322. https://doi.org/10.4025/actascibiolsci.v27i4.1265

Moreno AZ, Fuentes JL, Mago Y, Chinchilla O (2008) Descripción, taxonomía e índices ecológicos de parásitos en peces de la laguna de los Mártires, Isla de Margarita, Venezuela. SABER, Revista Multidisciplinaria del Consejo de Investigación de la Universidad de Oriente 20: 3–11.

Müller MI, Madi RR, Ueta MT (2008) Primeiro registro de ocorrência de cestódeos da família Bothriocephalidae Blanchard, 1849 (Pseudophyllidea), parasitando *Cichla monoculus* (Cichlidae) nas lagoas da Fazenda Rio das Pedras, Campinas (SP). Bioikos 22: 45–49.

Muñoz G (2014) Parasites communities in the clingfish *Gobiesox marmoratus* from central Chile. Acta Parasitologica 59: 108–114. https://doi.org/10.2478/s11686-014-0215-5

Muñoz G, Delorme N (2011) Variaciones temporales de las comunidades de parásitos en peces intermareales de Chile central: hospedadores residentes vs temporales. Revista de Biología Marina y Oceanografía 46: 313–327. https://doi.org/10.4067/S0718-19572011000300003

Muñoz G, Garcías F, Valdebenito V, George-Nascimento M (2001) Parasitofauna y alimentación de *Notothenia* cf. *angustata* Hutton, 1875 (Pisces: Nototheniidae) en el intermareal de dos localidades del Golfo de Arauco, Chile. Boletín Chileno de Parasitología 56: 29–33. https://doi.org/10.4067/S0365-94022001000100008

Muñoz G, Olmos V (2008) Revisión bibliográfica de especies endoparásitas y hospedadoras de sistemas acuáticos de Chile. Revista de Biología Marina y Oceanografía 43: 173–245. https://doi.org/10.4067/S0718-19572008000200002

Muñoz G, Randhawa HS (2011) Monthly variation in the parasite communities of the intertidal fish *Scartichthys viridis* (Blenniidae) from central Chile: are there seasonal patterns? Parasitology Research 109: 53–62. https://doi.org/10.1007/s00436-010-2220-4

Muñoz G, Zamora L (2011) Ontogenetic variation in parasite infracommunities of the clingfish *Sicyases sanguineus* (Pisces: Gobiesocidae). Journal of Parasitology 97: 14–19. https://doi.org/10.1645/GE-2445.1

Muñoz SA, George-Nascimento M (2008) The effect of *Anonchocephalus chilensis* Riggenbach (Eucestoda: Bothriocephalidea) on infracommunity patterns in *Genypterus maculatus* Tschudi (Osteichthyes: Ophidiidae). Journal of Helminthology 82: 221–226. https://doi.org/10.1017/S0022149X08960788

Mutti LD, Ivanov VA (2016) A new species of *Paraberrapex* Jensen, 2001 (Cestoda: Lecanicephalidea) from *Squatina guggenheim* Marini (Squatiniformes: Squatinidae) off Argentina. Folia Parasitologica 63: 007. https://doi.org/10.14411/fp.2016.007

Myers N, Mittermeier RA, Mittermeier CG, Fonseca GAB, Kent J (2000) Biodiversity hotspots for conservation priorities. Nature 403: 853–858. https://doi.org/10.1038/35002501

Ñacari LA, Oliva ME (2016) Metazoan parasites of deep-sea fishes from the South Eastern Pacific: exploring the role of ecology and host phylogeny. Deep Sea Research Part I: Oceanographic Research Papers 115: 123–130. https://doi.org/10.1016/j.dsr.2016.06.002

Napoleão SR, Antonucci AM, Amorim AF, Takemoto RM (2015) Occurrence of *Rhinoptericola megacantha* (Cestoda, Trypanorhyncha) in new host and new location. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 67: 1175–1177. https://doi.org/10.1590/1678-4162-7015

Naylor GJP, Caira JN, Jensen K, Rosana KAM, White WT, Last PR (2012) A DNA sequence-based approach to the identification of shark and ray species and its implications for global elasmobranch diversity and parasitology. Bulletin of the American Museum of Natural History 367: 1–262. https://doi.org/10.1206/754.1

Neghme M, Bertín V (1951) *Diphyllobothrium latum* en Chile. IV Estado actual de las investigaciones epidemiológicas. Revista Chilena de Higiene y Medicina Preventiva 13: 8–11.

Neghme M, Bertín V, Tagle I, Silva R, Artigas J (1950) *Diphyllobothrium latum* en Chile. II Primera encuesta en el lago Colico. Boletín de Infecciones Parasitarias Chilenas 5: 16–17.

Niklitschek EJ, Secor DH, Toledo P, Lafon A, George-Nascimento M (2010) Segregation of SE Pacific and SW Atlantic southern blue whiting stocks: integrating evidence from complementary otolith microchemistry and parasite assemblage approaches. Environmental Biology of Fishes 89: 399–413. https://doi.org/10.1007/s10641-010-9695-9

Nock AM, Caira JN (1988) *Disculiceps galapagoensis* n. sp. (Lecanicephalidea: Disculicepitidae) from the shark, *Carcharhinus longimanus*, with comments on *D. pileatus*. Journal of Parasitology 74: 153–158. https://doi.org/10.2307/3282492

Oliva ME (1982) Parásitos en peces marinos de la zona de Antofagasta. Ciencia y Tecnología del Mar 6: 45–51.

Oliva ME (1994) Parasites of the Chilean jack mackerel *Trachurus symmetricus murphyi* (Pisces: Carangidae). Memórias do Instituto Oswaldo Cruz 89: 363–364. https://doi.org/10.1590/S0074-02761994000300011

Oliva ME (1999) Metazoan parasites of the jack mackerel *Trachurus murphyi* (Teleostei, Carangidae) in a latitudinal gradient from South America (Chile and Peru). Parasite 6: 223–230. https://doi.org/10.1051/parasite/1999063223

Oliva ME (2001) Metazoan parasites of *Macruronus magellanicus* from southern Chile as biological tags. Journal of Fish Biology 58: 1617–1622. https://doi.org/10.1006/jfbi.2001.1569

Oliva ME, Ballón I (2002) Metazoan parasites of the Chilean hake *Merluccius gayi gayi* as a tool for stock discrimination. Fisheries Research 56: 313–320. https://doi.org/10.1016/S0165-7836(01)00329-0

Oliva ME, Castro RE, Burgos R (1996) Parasites of the flatfish *Paralichthys adspersus* (Steindachner, 1867) (Pleuronectiformes) from northern Chile. Memórias do Instituto Oswaldo Cruz 91: 301–306. https://doi.org/10.1590/S0074-02761996000300009

Oliva ME, Fernández I, Oyarzun C, Murillo C (2008a) Metazoan parasites of the stomach of *Dissostichus eleginoides* Smitt 1898 (Pisces: Notothenidae) from southern Chile: a tool for stock discrimination? Fisheries Research 91: 119–122. https://doi.org/10.1016/j.fishres.2007.11.012

Oliva ME, González MT, Acuña E (2004) Metazoan parasite fauna as a biological tag for the habitat of the flounder *Hippoglossina macrops* from northern Chile, in a depth gradient. Journal of Parasitology 90: 1374–1377. https://doi.org/10.1645/GE-317R

Oliva ME, Luque JL (1998) Metazoan parasite infracommunities in five sciaenids from the central Peruvian coast. Memórias do Instituto Oswaldo Cruz 93: 175–180. https://doi.org/10.1590/S0074-02761998000200007

Oliva ME, Luque JL (2002) Endohelminth parasites of the trambollo *Labrisomus philippii* (Steindachner) (Osteichthyes: Labrisomidae) from the central Peruvian coast. Comparative Parasitology 69: 100–104. https://doi.org/10.1654/1525-2647(2002)069[0100:EPOTTL]2.0.CO;2

Oliva ME, Valdivia IM, Costa G, Freitas N, Pinheiro de Carvalho MA, Sánchez L, Luque JL (2008b) What can metazoan parasites reveal about the taxonomy of *Scomber japonicus* Houttuyn in the coast of South America and Madeira Islands? Journal of Fish Biology 72: 545–554. https://doi.org/10.1111/j.1095-8649.2007.01725.x

Oliveira MT (1985) Infestação da pescada-foguete, *Macrodon ancylodon* (Bloch, 1801) Jordan, Evermann e Clark, 1830, e da corvina *Micropogon furnieri* (Desmarest, 1822) Jordan, 1884 família Sciaenidae do litoral Sul do Brasil, por larvas de Cestoda. Higiene Alimentar 4: 191–201.

Oliveira SAL, São Clemente SC, Benigno RNM, Knoff M (2009) *Poecilancistrium caryophyllum* (Diesing, 1850) (Cestoda, Trypanorhyncha), parasite of *Macrodon ancylodon* (Bloch & Schneider, 1801) from the Northern littoral of Brazil. Revista Brasileira de Parasitologia Veterinária 18: 71–73. https://doi.org/10.4322/rbpv.01804014

Olson PD, Caira JN (1999) Evolution of the major lineages of tapeworms (Platyhelminthes: Cestoidea) inferred from 18S ribosomal DNA and elongation factor-1α. Journal of Parasitology: 1134–1159. https://doi.org/10.2307/3285679

Olson PD, Caira JN, Jensen K, Overstreet RM, Palm HW, Beveridge I (2010) Evolution of the trypanorhynch tapeworms: parasite phylogeny supports independent lineages of sharks and rays. International Journal for Parasitology 40: 223–242. https://doi.org/10.1016/j.ijpara.2009.07.012

Olson PD, Littlewood DTJ, Bray RA, Mariaux J (2001) Interrelationships and evolution of the tapeworms (Platyhelminthes: Cestoda). Molecular Phylogenetics and Evolution 19: 443–467. https://doi.org/10.1006/mpev.2001.0930

Ortubay SG, Semenas LG, Ubeda CA, Quaggiotto AE, Viozzi GP (1994) Catálogo de peces dulceacuícolas de la Patagonia Argentina y sus parásitos metazoos. Direccion de Pesca, Subsecretaria de Recursos Naturales, San Carlos de Bariloche, 110 pp.

Ostrowski de Núñez M (1971) Estudios preliminares sobre la fauna parasitaria de algunos elasmobranquios del litoral bonaerense, Mar del Plata, Argentina. I. Cestodes y trematodes de *Psammobatis microps* (Günther) y *Zapteryx brevirostris* (Müller y Henle). Physis 30: 425–446.

Ostrowski de Núñez M (1973) Estudios preliminares sobre la fauna parasitaria de algunos elasmobranquios del litoral bonaerense, Mar del Plata, Argentina. II. Cestodes de *Mustelus schmitti* Springer 1939. Physis 32: 1–14.

Palm HW (1997) Trypanorhynch cestodes of commercial fishes from northeast Brazilian coastal waters. Memórias do Instituto Oswaldo Cruz 92: 69–79. https://doi.org/10.1590/S0074-02761997000100014

Palm HW (2004) The Trypanorhyncha Diesing, 1863. PKSPL-IPB Press, Bogor, 710 pp.

Palm HW, Walter T (2000) Tentaculariid cestodes (Trypanorhyncha) from the Muséum d'Histoire Naturelle, Paris. Zoosystema 22: 641–666.

Paraguassú AR, Luque JL, Alves DR (2002) Community ecology of the metazoan parasites of red porgy, *Pagrus pagrus* (L., 1758) (Osteichthyes, Sparidae), from the coastal zone, state of Rio de Janeiro, Brazil. Acta Scientiarum: Biological Sciences 24: 461–467.

Pardo-Gandarillas MC, González K, Ibáñez CM, George-Nascimento M (2008) Parasites of two deep-sea fish *Coelorynchus chilensis* (Pisces: Macrouridae) and *Notacanthus sexspinis* (Pisces: Notacanthidae) from Juan Fernández Archipelago, Chile. Marine Biodiversity Records 1: e76. https://doi.org/10.1017/S1755267207007944

Paschoal F, Cezar AD, Luque JL (2015) Checklist of metazoan associated with grunts (Perciformes, Haemulidae) from the Nearctic and Neotropical regions. Check List 11: 1501. https://doi.org/10.15560/11.1.1501

Pavanelli GC, Machado MH (1990) *Cangatiella arandasi* gen. n. sp. n. (Cestoda-Proteocephalidae), parasito de *Parauchenipterus galeatus* (Siluriformes-Auchenipteridae) do rio Paraná, PR. Revista Brasileira de Zoologia 7: 531–534. https://doi.org/10.1590/S0101-81751990000400012

Pavanelli GC, Machado MH (1991) Proteocefalídeos parasitos de peixes, em especial pimelodídeos do rio Paraná, Paraná. Revista Unimar 13: 163–175.

Pavanelli GC, Machado MH (1992) *Goezeella agostinhoi* sp. n. e *Monticellia loyolai* sp. n., cestóides proteocefalídeos parasitas de peixes pimelodídeos do rio Paraná, Paraná, Brasil. Revista Brasileira de Parasitologia Veterinária 1: 45–50.

Pavanelli GC, Machado MH, Takemoto RM, Santos LC (1994) A new species of proteocephalidean cestode, *Monticellia belavistensis* sp. n., parasite of *Pterodoras granulosus* (Valenciennes) (Pisces, Doradidae) from Itaipu reservoir and Paraná River, Paraná, Brazil. Revista Brasileira de Zoologia 11: 587–595. https://doi.org/10.1590/S0101-81751994000400001

Pavanelli GC, Rego AA (1989) Novas espécies de proteocefalídeos (Cestoda) de *Hemisorubim platyrhynchos* (Pisces: Pimelodidae) do Estado do Paraná. Revista Brasileira de Biologia 49: 381–386.

Pavanelli GC, Rego AA (1991) Cestóides proteocefalídeos de *Sorubim lima* (Schneider, 1801) (Pisces: Pimelodidae) do rio Paraná e reservatório de Itaipu. Revista Brasileira de Biologia 51: 7–12.

Pavanelli GC, Rego AA (1992) *Megathylacus travassosi* sp. n. and *Nomimoscolex sudobim* Woodland, 1935 (Cestoda-Proteocephalidea) parasites of *Pseudoplatystoma corruscans* (Agassiz, 1829) (Siluriformes-Pimelodidae) from the Itaipu reservoir and Paraná River, Paraná state, Brazil. Memórias do Instituto Oswaldo Cruz 87: 191–195. https://doi.org/10.1590/S0074-02761992000500036

Pavanelli GC, Takemoto RM (1995) New species of *Proteocephalus* (Cestoda-Proteocephalidae) parasitic in fishes from the Paraná River, Paraná, Brazil. Memórias do Instituto Oswaldo Cruz 90: 593–596. https://doi.org/10.1590/S0074-02761995000500009

Pavanelli GC, Takemoto RM (1996) *Spasskyellina mandi* n. sp. (Proteocephalidea: Monticelliidae), parasite of *Pimelodus ornatus* Kner, 1857 (Pisces: Pimelodidae) of the Paraná River, Paraná, Brazil. Memórias do Instituto Oswaldo Cruz 91: 723–726. https://doi.org/10.1590/S0074-02761996000600013

Pavanelli GC, Takemoto RM (2000) Aspects of the ecology of proteocephalid cestodes, parasites of *Sorubim lima* (Pimelodidae), of the upper Paraná River, Brazil: II. Interspecific associations and distribution of gastrintestinal parasites. Revista Brasileira de Biologia 60: 585–590. https://doi.org/10.1590/S0034-71082000000400007

Pereira AN, Pantoja C, Luque JL, Timi JT (2014) Parasites of *Urophycis brasiliensis* (Gadiformes: Phycidae) as indicators of marine ecoregions in coastal areas of the South American Atlantic. Parasitology Research 113: 4281–4292. https://doi.org/10.1007/s00436-014-4106-3

Pereira J Jr. (1993) O complexo de espécies de Trypanorhyncha (Cestoda), em corvinas *Micropogonias furnieri* do Rio Grande do Sul. Arquivos da Faculdade de Veterinária UFRGS 21: 58–70.

Pereira J Jr. (1998) Trypanorhyncha (Cercomeromorpha, Eucestoda) nos Sciaenidae (Neopterygii, Perciformes) do Litoral do Rio Grande do Sul: sistemática, estrutura das comunidades componentes e sua utilização como indicadores da estrutura trófica da assembléia hospedeira. PhD thesis, Curitiba, Brazil: Universidade Federal do Paraná.

Pereira J Jr. (2000) New morphologic data on *Anonchocephalus chilensis* (Riggenbach, 1896) (Triaenophoridae: Pseudophyllidea: Cestoda) and emendation of genus diagnosis. Comunicaçoes do Museu de Ciencias e Tecnologia da PUCRS, Serie Zoologia 13: 99–104.

Pereira J Jr., Boeger WA (2005) Larval tapeworms (Platyhelminthes, Cestoda) from sciaenid fishes of the southern coast of Brazil. Zoosystema 27: 5–25.

Pérez I, Chávez A, Casas E (1999) Presencia de formas parasitarias en peces comerciales del mar peruano. Revista de Investigaciones Veterinarias del Perú 10: 34–38.

Piazza RS, Martins ML, Guiraldelli L, Yamashita MM (2006) Parasitic diseases of freshwater ornamental fishes commercialized in Florianópolis, Santa Catarina, Brazil. Boletim do Instituto de Pesca 32: 51–57.

Pinto HA, Melo AL (2011a) Metacestodes of *Parvitaenia macropeos* (Cyclophyllidea, Gryporhynchidae) in *Australoheros facetus* (Pisces, Cichlidae) in Brazil. Neotropical Helminthology 5: 279–283.

Pinto HA, Melo AL (2011b) Metacestodes of *Glossocercus auritus* (Cyclophyllidea, Gryporhynchidae) in *Poecilia reticulata* (Pisces, Poeciliidae) from Brazil. Revista Brasileira de Parasitologia Veterinária 20: 161–164. https://doi.org/10.1590/S1984-29612011000200012

Pinto RM, Knoff M, São Clemente SC, Lanfredi RM, Gomes DC (2006) The taxonomy of some Poecilacanthoidea (Eucestoda: Trypanorhyncha) from elasmobranchs off the southern coast of Brazil. Journal of Helminthology 80: 291–298. https://doi.org/10.1079/JOH2006339

Piorski NM, Garavello JC, Arce HM, Sabaj-Pérez MH (2008) *Platydoras brachylecis*, a new species of thorny catfish (Siluriformes: Doradidae) from northeastern Brazil. Neotropical Ichthyology 6: 481–494. https://doi.org/10.1590/S1679-62252008000300021

Poche F (1922) Zur Kenntnis der Amphilinidea. Zoologischer Anzeiger 54: 276–287. http://www.biodiversitylibrary.org/part/68459

Porto CJS, São Clemente SC, Freitas MQ, São Clemente RRB, Knoff M, Matos E (2009) *Pterobothrium crassicolle* (Eucestoda: Trypanorhyncha) em corvinas, *Micropogonias furnieri*, comercializadas no município de Niterói, Rio de Janeiro, Brasil. Revista Brasileira de Ciência Veterinária 16: 133–135.

Poulin R, Besson AA, Morin MB, Randhawa HS (2016) Missing links: testing the completeness of host-parasite checklists. Parasitology 143: 114–122. https://doi.org/10.1017/S0031182015001559

Poulin R, Leung TLF (2010) Taxonomic resolution in parasite community studies: are things getting worse? Parasitology 137: 1967–1973. https://doi.org/10.1017/S0031182010000910

Rabey JS (1973) Un cestode parasito del tubo digestivo del bagre sapo (*Rhamdia sapo*) de la laguna de Chascomús, Provincia de Buenos Aires. Physis 32: 115–119.

Rauque CA, Viozzi GP, Semenas LG (2003) Component population study of *Acanthocephalus tumescens* (Acanthocephala) in fishes from Lake Moreno, Argentina. Folia Parasitologica 50: 72–78. https://doi.org/10.14411/fp.2003.013

Rego AA (1975) Estudos de cestóides de peixes do Brasil. 2ª Nota: revisão do gênero Monticellia La Rue, 1911 (Cestoda, Proteocephalidae). Revista Brasileira de Biologia 35: 567–586.

Rego AA (1977) Cestóides parasitas de *Carcharinus longimanus* (Poey, 1861). Revista Brasileira de Biologia 37: 847–852.

Rego AA (1979) Contribuição ao conhecimento dos helmintos de raias fluviais Paratrygonidae. Revista Brasileira de Biologia 39: 879–890.

Rego AA (1984a) Proteocephalidea from Amazonian freshwater fishes: new systematic arrangement for the species described by Woodland as *Anthobothrium* (Tetraphyllidea). Acta Amazonica 14: 86–94. https://doi.org/10.1590/1809-43921984142094

Rego AA (1984b) Proteocefalídeos (Cestoda) de *Phractocephalus hemioliopterus*, peixe da Amazônia. Memórias do Instituto Oswaldo Cruz 79: 257–261. https://doi.org/10.1590/S0074-02761984000200013

Rego AA (1987a) Redescription of *Pterobothrium crassicole* Diesing, 1850 (Cestoda: Trypanorhyncha) and revalidation of the species. Memórias do Instituto Oswaldo Cruz 82: 51–53. https://doi.org/10.1590/S0074-02761987000100008

Rego AA (1987b) Cestóides proteocefalídeos do Brasil. Reorganização taxonômica. Revista Brasileira de Biologia 47: 203–212

Rego AA (1989) Cestóides proteocefalídeos de "cachara", *Pseudoplatystoma fasciatus* (L.) (Pisces, Pimelodidae) de Mato Grosso. Memórias do Instituto Oswaldo Cruz 84: 455–461. https://doi.org/10.1590/S0074-02761989000800080

Rego AA (1990) Cestóides proteocefalídeos parasitas de pintado, *Pseudoplatystoma corruscans* (Agassiz) (Pisces, Pimelodidae). Ciência e Cultura 42: 997–1002.

Rego AA (1991) Redescription of *Nomimoscolex piraeeba* Woodland, 1934 (Cestoda, Proteocephalidea), from the Amazon catfishes, *Brachyplatystoma* spp. with proposal of synonyms and invalidation of Endorchiinae and *Endorchis*. Memórias do Instituto Oswaldo Cruz 86: 229–232. https://doi.org/10.1590/S0074-02761991000200013

Rego AA (1992) Redescription of *Gibsoniela mandube* (Woodland, 1935) (Cestoda: Proteocephalidea), a parasite of *Ageneiosus brevifilis* (Pisces: Siluriformes), and reappraisal of the classification of the Proteocephalideans. Memórias do Instituto Oswaldo Cruz 87: 417–422. https://doi.org/10.1590/S0074-02761992000300012

Rego AA (1997) *Senga* sp., occurrence of a pseudophyllid cestode in a Brazilian freshwater fish. Memórias do Instituto Oswaldo Cruz 92: 607. https://doi.org/10.1590/s0074-02761997000500008

Rego AA (1999) Scolex morphology of proteocephalid cestodes parasites of Neotropical freshwater fishes. Memórias do Instituto Oswaldo Cruz 94: 37–52. https://doi.org/10.1590/S0074-02761999000100011

Rego AA (2002) Cestóides proteocefalídeos parasitas de *Pseudoplatystoma* (Pisces, Pimelodidae) da América do Sul. Revista Brasileira de Zoociências 4: 269–282.

Rego AA (2004) Current state of knowledge of cestodes from Neotropical freshwater fishes and rays. Revista Brasileira de Zoociências 6: 61–79.

Rego AA, Chubb JC, Pavanelli GC (1999a) Cestodes in South American freshwater teleost fishes: keys to genera and brief description of species. Revista Brasileira de Zoologia 16: 299–367. https://doi.org/10.1590/S0101-81751999000200003

Rego AA, de Chambrier A (1995) *Crepidobothrium eirasi* n. sp. (Cestoda, Proteocephalidae), a parasites of the siluroid fish *Phractocephalus hemioliopterus* (Schneider, 1801) (Pisces, Pimelodidae) from the Brazilian Amazon. Revue Suisse de Zoologie 102: 3–11. https://doi.org/10.5962/bhl.part.80457

Rego AA, Dias PL (1976) Estudos de cestóides de peixes do Brasil. 3ª Nota: cestóides de raias fluviais Paratrygonidae. Revista Brasileira de Biologia 36: 941–956.

Rego AA, Gibson DI (1989) Hyperparasitism by helminths: new records of cestodes and nematodes in proteocephalid cestodes from South American siluriform fishes. Memórias do Instituto Oswaldo Cruz 84: 371–376. https://doi.org/10.1590/S0074-02761989000300012

Rego AA, Ivanov VA (2001) *Pseudocrepidobothrium eirasi* (Rego and de Chambrier, 1995) gen. n. and comb. nov. (Cestoda, Proteocephalidea), parasite of a South American freshwater fish, and comparative cladistic analysis with *Crepidobothrium* spp. Acta Scientiarum: Biological Sciences 23: 363–367.

Rego AA, Machado PM, Pavanelli G (1999b) *Sciadocephalus megalodiscus* Diesing, 1850 (Cestoda: Corallobothriinae), a parasite of *Cichla monoculus* Spix, 1831 (Cichlidae), in the Paraná river, State of Paraná, Brazil. Journal of the Helminthlogical Society of Washington 66: 133–137.

Rego AA, Mayer MT (1976) Ocorrência de duas espécies de tetrafilídeos em tubarão da costa brasileira e considerações sobre os gêneros *Cylindrophorus*, *Platybothrium* e *Phoreiobothrium* (Cestoda, Tetraphyllidea). Revista Brasileira de Biologia 36: 321–328.

Rego AA, Pavanelli G (1985) *Jauella glandicephalus* gen. n., sp. n. e *Megathylacus brooksi* sp. n., cestóides patogênicos para o jaú, *Paulicea luetkeni*, peixe pimelodídeo. Revista Brasileira de Biologia 45: 643–652.

Rego AA, Pavanelli G (1987) Cestóides proteocefalídeos do jaú, *Paulicea luetkeni*, peixe pimelodídeo do Brasil. Revista Brasileira de Biologia 47: 357–361.

Rego AA, Pavanelli G (1990) Novas espécies de cestóides proteocefálideos: parasitas de peixes näo Siluriformes. Revista Brasileira de Biologia 50: 91–101.

Rego AA, Pavanelli G (1991) *Proteocephalus gibsoni* nom. nov. for *Proteocephalus ocellatus* Rego and Pavanelli, 1990 preoccupied by *Proteocephalus ocellatus* (Rudolphi, 1802). Revista Brasileira de Biologia 51: 701.

Rego AA, Pavanelli GC (1992) Redescription of *Nomimoscolex admonticellia* (Woodland), comb. n (Cestoda: Proteocephalidea) parasite of *Pinirampus pirinampu* (Spix), a freshwater siluriform fish. Revista Brasileira de Zoologia 9: 283–289. https://doi.org/10.1590/S0101-81751992000200015

Rego AA, Santos CP (1983) Helmintofauna de cavalas, *Scomber japonicus* Houtt, do Rio de Janeiro. Memórias do Instituto Oswaldo Cruz 78: 443–448. https://doi.org/10.1590/S0074-02761983000400008

Rego AA, dos Santos JC, Silva PP (1974) Estudos de cestóides de peixes do Brasil. Memórias do Instituto Oswaldo Cruz 72: 187–204. https://doi.org/10.1590/S0074-02761974000200004

Rego AA, Schaeffer GV (1999) Histopatologia das lesões de *Brachyplatystoma* spp. (Pisces: Siluriformes) por *Nomimoscolex piraeeba* Woodland, 1934 (Cestoda: Proteocephalidea). Revista Brasileira de Ciência Veterinária 6: 98–100. https://doi.org/10.4322/rbcv.2015.144

Rego AA, Vicente JJ (1988) Excursão científica à zona do Pantanal, Estado de Mato Grosso, para coleta de helmintos. Ciência e Cultura 40: 65–68.

Rego AA, Vicente JJ, Herrera NI (1968) Sôbre dois novos parasitos de peixe da costa do Peru (Cestoda, Tetraphyllidea). Memórias do Instituto Oswaldo Cruz 66: 145–149. https://doi.org/10.1590/S0074-02761968000200002

Reis RE (2013) Conserving the freshwater fishes of South America. International Zoo Yearbook 47: 65–70. https://doi.org/10.1111/izy.12000

Revenga JE (1993) *Diphyllobothrium dendriticum* and *Diphyllobothrium latum* in fishes from southern Argentina: association, abundance, distribution, pathological effects, and risk of human infection. Journal of Parasitology 79: 379–383. https://doi.org/10.2307/3283573

Revenga JE, Perfumo CJ, Ubeda CA, Semenas LG (1995) Difilobotriasis en salmónidos introducidos en el Parque y Reserva Nacional Nahuel Huapi, Argentina: patología de las lesiones producidas por *Diphyllobothrium* spp. Archivos de Medicina Veterinaria 23: 115–122.

Revenga JE, Semenas LG (1991) Difilobotriasis en salmónidos introducidos en el Parque y Reserva Nacional Nahuel Huapi, Argentina: morfología de plerocercoides. Archivos de Medicina Veterinaria 23: 157–164.

Revenga JE, Torres PF, Baiz M (2005) Impact of a caged-trout farm on parasites of *Galaxias maculatus* in Lake Moreno, southern Argentina. Journal of Parasitology 91: 707–709. https://doi.org/10.1645/ge-441r

Reyda FB (2008) Intestinal helminths of freshwater stingrays in southeastern Peru, and a new genus and two new species of cestode. Journal of Parasitology 94: 684–699. https://doi.org/10.1645/GE-1230.1

Reyda FB, Marques FPL (2011) Diversification and species boundaries of *Rhinebothrium* (Cestoda; Rhinebothriidea) in South American freshwater stingrays (Batoidea; Potamotrygonidae). PLoS ONE 6: e22604. https://doi.org/10.1371/journal.pone.0022604

Reyda FB, Olson PD (2003) Cestodes of cestodes of Peruvian freshwater stingrays. Journal of Parasitology 89: 1018–1024. https://doi.org/10.1645/ge-3143

Reyes-Piraino X (1982) Presencia de *Hepatoxylon trichiuri* (holten, 1802) (Cestoda: Trypanorhyncha) en *Oncorhynchus tshawytscha* y *Somniosus pacificus* capturados en Chile. Investigaciones Marinas (Valparaíso) 10: 41–43.

Ribeiro RS, Luque JL, Alves DR (2002) Aspectos quantitativos dos parasitos da Maria-luiza, *Paralonchurus brasiliensis* (Osteichthyes: Sciaenidae), do litoral do Estado do Rio de Janeiro. Revista Universidade Rural, Série Ciências da Vida 22: 151–154.

Ribeiro TS, Lizama MAP, Takemoto RM (2014) Metazoan endoparasites diversity of *Pseudoplatystoma corruscans* (Siluriformes: Pimelodidae) as an indicator of environmental alterations on a tropical aquatic system. Acta Parasitologica 59: 398–404. https://doi.org/10.2478/s11686-014-0260-0

Ribeiro TS, Takemoto RM (2014) Resposta inflamatória do pintado à infecção por *Nomimoscolex pertierrae* (Eucestoda: Proteocephalidea). Boletim do Instituto de Pesca 40: 111–120.

Riffo R (1991) La fauna de parásitos metazoos del lenguado de ojos grandes *Hippoglossina macrops* Steindachner, 1876 (Pisces: Bothidae): una aproximación ecológica. Medio Ambiente 11: 54–60.

Riffo R (1994) Composición taxonómica y características cuantitativas de la fauna de parásitos metazoos del congrio dorado *Genypterus blacodes* Schneider 1801. Medio Ambiente 12: 27–31.

Riffo R (1995) Análisis comparativo de la fauna de parásitos metazoos de dos especies de lenguados congenéricos y sintópicos: *Paralichthys microps* Gunther 1881 y *Paralichthys adspersus* Steindachner 1867 (Pleuronectiformes: Bothidae) en la Bahía Concepción, Chile. Medio ambiente 12: 51–59.

Riggenbach E (1895) Beiträge zur Kenntnis der Taenien der Süsswasserfische. Centralblatt für Bakteriologie und Parasitenkunde 18: 609–613.

Riggenbach E (1896a) Bemerkungen über das Genus *Bothriotaenia* Railliet. Centralblatt für Bakteriologie, Parasitenkunde und Infektionskrankheiten 20: 222–231.

Riggenbach E (1896b) Das Genus *Ichthyotaenia*. Revue Suisse de Zoologie 4: 165–277. https://doi.org/10.5962/bhl.part.70783

Ringuelet R (1943) Piscicultura del pejerrey o Atherinicultura. Suelo Argentino, Buenos Aires, 162 pp.

Rivera G, Sarmiento L (1990) Cestodes en peces elasmobranquios en Pucusana. Boletin de Lima 12: 87–88.

Rocha CA, Rocha Jr. CAM, da Silva IHF, Alcântara ME, Drago Bisneto MQ, Baker PKB (2016) Ecological aspects of helminth fauna of *Brachyplatystoma rousseauxii* (Siluriformes: Pimelodidae) from Bay of Marajó, Para State, Brazil. Veterinária e Zootecnia 23: 243–250.

Rodrigues HO, Rodrigues SS, Faria Z (1990) Contribution to the knowledge of the helminthological fauna of vertebrates of Maricá, Rio de Janeiro State, Brazil. Memórias do Instituto Oswaldo Cruz 85: 115-116. https://doi.org/10.1590/S0074-02761990000100020

Rodríguez L, Balboa L, George-Nascimento M (2000) Parasitismo en la caballa *Scomber japonicus* Houttuyn, 1782 y el jurel *Trachurus symmetricus* murphyi Nichols, 1920 frente a Chile central. Biología Pesquera 28: 15–21.

Rodríguez L, George-Nascimento M (1996) La fauna de parásitos metazoos del bacalao de profundidad *Dissostichus eleginoides* Smitt, 1898 (Pisces: Nototheniidae) en Chile central: aspectos taxonómicos, ecológicos y zoogeográficos. Revista Chilena de Historia Natural 69: 21–33.

Rodríguez T, Tantaleán MV (1980) A study of the helminths of elasmobranch fishes of the Peruvian coast. 1. New reports of tetraphyllideans. Boletin Peruano de Parasitologia 2: 71–75.

Rossin MA, Timi JT (2010) Parasite assemblages of *Nemadactylus bergi* (Pisces: Latridae): the role of larval stages in the short-scale predictability. Parasitology Research 107: 1373–1379. https://doi.org/10.1007/s00436-010-2011-y

Rozas M, Bohle H, Sandoval A, Ildefonso R, Navarrete A, Bustos P (2012) First molecular identification of *Diphyllobothrium dendriticum* plerocercoids from feral rainbow trout (*Oncorhynchus mykiss*) in Chile. Journal of Parasitology 98: 1220–1226. https://doi.org/10.1645/jp-ge-3097.1

Rudolphi CA (1819) Entozoorum synopsis cui accedunt mantesia duplex et indices locupletissimi. Rücker. S. A, Berolini, 811 pp. https://doi.org/10.5962/bhl.title.9157

Ruedi V, de Chambrier A (2012) *Pseudocrepidobothrium ludovici* sp n. (Eucestoda: Proteocephalidea), a parasite of *Phractocephalus hemioliopterus* (Pisces: Pimelodidae) from Brazilian Amazon. Revue Suisse de Zoologie 119: 137–147. https://doi.org/10.5962/bhl.part.150326

Ruhnke TR (2011) Tapeworms of Elasmobranchs (part III). A monograph on the Phyllobothriidae. Bulletin of the University of Nebraska State Museum 25: 1–208.

Ruhnke TR, Caira JN (2009) Two new species of *Anthobothrium* van Beneden, 1850 (Tetraphyllidea: Phyllobothriidae) from carcharhinid sharks, with a redescription of *Anthobothrium laciniatum* Linton, 1890. Systematic Parasitology 72: 217–227. https://doi.org/10.1007/s11230-008-9168-0

Ruhnke TR, Caira JN, Cox A (2015) The cestode order Rhinebothriidea no longer family-less: a molecular phylogenetic investigation with erection of two new families and description

of eight new species of *Anthocephalum*. Zootaxa 3904: 51–81. https://doi.org/10.11646/zootaxa.3904.1.3

Ruhnke TR, Workman RE (2013) Two new species and a new phyllobothriid cestode genus from sharks of the genus *Negaprion* Whitley (Carcharhiniformes). Systematic Parasitology 85: 37–48. https://doi.org/10.1007/s11230-013-9411-1

Sabas CSS, Brasil-Sato MC (2014) Helminth fauna parasitizing *Pimelodus pohli* (Actinopterygii: Pimelodidae) from the upper São Francisco River, Brazil. Revista Brasileira de Parasitologia Veterinária 23: 375–382. https://doi.org/10.1590/S1984-29612014067

Sabas CSS, Luque JL (2003) Metazoan parasites of weakfish, *Cynoscion guatucupa* and *Macrodon ancylodon* (Osteichthyes: Sciaenidae), from the coastal zone of the State of Rio de Janeiro, Brazil. Revista Brasileira de Parasitologia Veterinária 12: 171–178.

Santos CAML, Zogbi EPV (1971) Infestation of fish in Brazil with *Tetrarhynchus fragilis* larvae. In: Kreuzer R (Ed.) Fish Inspection and Quality Control. Fishing News (Books) Ltd. London, 262–264.

Santos-Clapp MD, Brasil-Sato MC (2014) Parasite community of *Cichla kelberi* (Perciformes, Cichlidae) in the Tres Marias Reservoir, Minas Gerais, Brazil. Revista Brasileira de Parasitologia Veterinária 23: 367–374. https://doi.org/10.1590/s1984-29612014059

Santos MD, Brasil-Sato MC (2004) Parasitos metazoários de *Franciscodoras marmoratus* (Reinhardt, 1874), "serrudo" (Siluriformes: Doradidae) do rio São Francisco, Brasil. Revista Brasileira de Parasitologia Veterinária 13: 18–22.

Santos MD, Brasil-Sato MC (2006) Parasitic community of *Fransciscodoras marmoratus* (Reinhardt, 1874) (Pisces: Siluriformes, Doradidae) from the upper Sao Francisco River, Brazil. Brazilian Journal of Biology 66: 931–938. https://doi.org/10.1590/S1519-69842006000500019

Santos MD, Lemos-Pita SRLC, Brasil-Sato MC (2007) Metazoan parasite fauna of *Pimelodus maculatus* La Cépède, 1803 (Siluriformes, Pimelodidae) from the Guandu river, Rio de Janeiro State, Brazil. Acta Scientiarum: Biological Sciences 29: 101–107. https://doi.org/10.4025/actascibiolsci.v29i1.130

Santos RBS, Tavares-Dias M (2010) Células sanguíneas e resposta hematológica de *Oxydoras niger* (Pisces, Doradidae) oriundos da bacia do médio rio Solimões, estado do Amazonas (Brasil), naturalmente parasitados. Boletim do Instituto de Pesca 36: 283–292.

Santos RS, Roumbedakis K, Marengoni NG, Takahashi HK, Pimenta FDA, Melo CMR, Martins ML (2011) Proteocephalid cestode infection in tucunaré *Cichla* sp. (Osteichthyes: Cichlidae) from Paraná River, São Paulo. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 63: 584–590. https://doi.org/10.1590/S0102-09352011000300008

Santos SMC, Ceccarelli PS, Rêgo RF (2003) Helmintos em peixes do Pantanal sul-mato-grossense: primeira expedição do Programa Pantanal. Boletim Técnico do CEPTA 16: 15–26.

São Clemente SC (1986a) Plerocercos da ordem Trypanorhyncha, parasitos de corvina *Micropogonias furnieri* (Desmarest) no litoral do Estado do Rio de Janeiro. Atas da Sociedade de Biologia do Rio de Janeiro 26: 29–36.

São Clemente SC (1986b) Prevalência e intensidade média de infecção de plerocercos de Trypanorhyncha, parasitando corvina *Micropogonias furnieri* (Desmarest) no litoral do Rio de Janeiro. Atas da Sociedade de Biologia do Rio de Janeiro 26: 37–44.

São Clemente SC (1987) Plerocercos de cestóides da ordem Trypanorhyncha em corvina *Micropogonias furnieri* (Desmarest) e sua importância na inspeção sanitária do pescado. Arquivos Fluminense de Medicina Veterinária 2: 82–83.

São Clemente SC, Coelho MRT, Serra-Freire NM (1991a) Cestóides parasitos de bagre *Netuma barba* (Lacépède, 1803) pescados no litoral do Rio de Janeiro e comercializados para consumo humano. Arquivos da Universidade Federal Rural do Rio de Janeiro 14: 27–34.

São Clemente SC, Gomes DC (1989a) *Dasyrhynchus pacificus* Robinson, 1965 (Trypanorhyncha: Dasyrhynchidae) description of the adult form. Memórias do Instituto Oswaldo Cruz 84: 113–116. https://doi.org/10.1590/S0074-02761989000100020

São Clemente SC, Gomes DC (1989b) Trypanorhyncha from sharks of southern Brazilian coast: *Eutetrarhynchus vooremi* sp. n. and two other species parasites of *Mustelus* (Pisces, Triakidae). Memórias do Instituto Oswaldo Cruz 84: 475–481. https://doi.org/10.1590/S0074-02761989000800083

São Clemente SC, Gomes DC (1992) Description of the adult form of *Nybelinia* (*Syngenes*) *rougetcampanae* Dollfus, 1960 and some new data on *N*. (*N*.) *bisulcata* (Linton, 1889) (Trypanorhyncha: Tentaculariidae). Memórias do Instituto Oswaldo Cruz 87: 251–255. https://doi.org/10.1590/S0074-02761992000500047

São Clemente SC, Gomes DC, Serra-Freire NM (1991b) Prevalência e intensidade de infecção de helmintos da ordem Trypanorhyncha em elasmobrânquios no litoral sul do Brasil. Parasitología al Día 15: 9–14.

São Clemente SC, Knoff M, Lima FC, Andrada CDG, Felizardo NN, Padovani RES, Gomes DC (2007) Cestóides Trypanorhyncha parasitos de peixe sapo-pescador, *Lophius gastrophysus* Miranda-Ribeiro, 1915 comercializados no estado do Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 16: 37–42.

São Clemente SC, Knoff M, Padovani RES, Lima FC, Gomes DC (2004) Cestóides Trypanorhyncha parasitos de Congro-rosa, *Genypterus brasiliensis* Regan, 1903 comercializados nos municípios de Niterói e Rio de Janeiro, Brasil. Revista Brasileira de Parasitologia Veterinária 13: 97–102.

São Clemente SC, Lima FC, Uchoa CMA (1995) Parasitos de *Balistes vetula* (L.) e sua importância na inspeção do pescado. Revista Brasileira de Ciência Veterinária 2: 39–41. https://doi.org/10.4322/rbcv.2015.020

São Clemente SC, Matos E, Uchoa CMA, Matos P (1993) Trypanorhynch plerocerci in fish of commercial importance in Brazil. Parasitología al Día 17: 52–53.

São Clemente SC, Pereira J Jr., Knoff M, Silva CM, Fernandez JG, Cousin JC (2001) *Hepatoxylon trichiuri* (Holten, 1802) Dollfus, 1942, Hepatoxylidae Dollfus, 1940 (Eucestoda: Trypanorhyncha) em *Prionace glauca* (Linnaeus, 1758), do litoral do estado do Rio Grande do Sul e em *Coryphaena hippurus* Linnaeus, 1758, do litoral do estado do Rio de Janeiro, Brasil. Parasitología al Día 25: 135–137. https://doi.org/10.4067/S0716-07202001000300013

São Clemente SC, Silva CM, Gottschalk S (1997) Prevalência e intensidade de infecção de cestóides Trypanorhyncha em anchovas, *Pomatomus saltatrix* (L.) do litoral do Rio de Janeiro, Brasil. Parasitología al Día 21: 54–57.

Sardella NH, Avendaño MF, Timi JT (1998) Parasite communities of *Genypterus blacodes* and *G. brasiliensis* (Pisces: Ophidiidae) from Argentina. Helminthologia 35: 209–218.

Sardella NH, Timi JT (1996) Parasite communities of *Merluccius hubbsi* from the Argentinian-Uruguayan common fishing zone. Fisheries Research 27: 81–88. https://doi.org/10.1016/0165-7836(95)00460-2

Sardella NH, Timi JT (2004) Parasites of Argentine hake in the Argentine Sea: population and infracommunity structure as evidence for host stock discrimination. Journal of Fish Biology 65: 1472–1488. https://doi.org/10.1111/j.0022-1112.2004.00572.x

Schäffer GV, Rego AA, Pavanelli GC (1992) Peritoneal and visceral cestode larvae in Brazilian freshwater fishes. Memórias do Instituto Oswaldo Cruz 87: 257–258. https://doi.org/10.1590/S0074-02761992000500048

Scholz T, Bray RA, Kuchta R, Řepová R (2004) Larvae of gryporhynchid cestodes (Cyclophyllidea) from fish: a review. Folia Parasitologica 51: 131–152. https://doi.org/10.14411/fp.2004.018

Scholz T, de Chambrier A, Kuchta R (2008) Redescription of the tapeworm *Monticellia amazonica* de Chambrier et Vaucher, 1997 (Cestoda, Proteocephalidea), a parasite of *Calophysus macropterus* (Siluriformes, Pimelodidae) from the Amazon River. Acta Parasitologica 53: 30–35. https://doi.org/10.2478/s11686-008-0004-0

Scholz T, de Chambrier A, Kuchta R, Littlewood DT, Waeschenbach A (2013) *Macrobothriotaenia ficta* (Cestoda: Proteocephalidea), a parasite of sunbeam snake (*Xenopeltis unicolor*): example of convergent evolution. Zootaxa 3640: 485–499. https://doi.org/10.11646/zootaxa.3640.3.12

Scholz T, de Chambrier A, Prouza A, Royero R (1996) Redescription of *Proteocephalus macrophallus*, a parasite of *Cichla ocellaris* (Pisces: Cichlidae) from South America. Folia Parasitologica 43: 287–291.

Scholz T, Garcia HH, Kuchta R, Wicht B (2009) Update on the human broad tapeworm (genus *Diphyllobothrium*), including clinical relevance. Clinical Microbiology Reviews 22: 146–160. https://doi.org/10.1128/CMR.00033-08

Scholz T, Kuchta R, Williams C (2011) *Bothriocephalus acheilognathi*. In: P. T. K., Woo, Buchmann K (Eds) Fish Parasites: Pathobiology and Protection. CAB International, Oxfordshire, UK, 282–297.

Scholz T, Priemer J (1989) Notes to the synonymy of cestodes of the genus *Proteocephalus* Weinland, 1858 from perches *P. percae* (Müller, 1780) or *P. ocellatus* (Rudolphi, 1802)? Folia Parasitologica 36: 69–70.

Semenas L, Kreiter A (1995) Epidemiología de la difilobotriasis en la región Andino Patagónica. Revista de la Asociación Bioquímica Argentina 59: 203–206.

Sepúlveda F, Marín SL, Carvajal JG (2004) Metazoan parasites in wild fish and farmed salmon from aquaculture sites in southern Chile. Aquaculture 235: 89–100. https://doi.org/10.1016/j.aquaculture.2003.09.015

Serrano-Martínez E, Tantaleán MV, Leguía GP, Quispe MH, Casas GCV (2015) Parásitos en *Arapaima gigas* de la Amazonía Peruana según grupo etario. Revista de Investigaciones Veterinarias del Perú 26: 303–309. https://doi.org/10.15381/rivep.v26i2.11014

Severino RL, Sarmiento LB (1979) Nueva especie del género *Acanthobothrium* van Beneden 1849; Cestode: Tetraphyllidea de *Myliobatis peruvianus* Garman 1913. Revista de Ciencias (Lima) 71: 38–43.

Severino RL, Verano RM (1980) *Acanthobothrium lusarmientoi* n. sp. (Cestoda: Tetraphyllidea: Onchobothriidae). *Psammobatis caudispina* Hildebrand, 1941 (Chondrichtyes: Rajiidae) de Perú. Revista de Ciencias (Lima) 72: 21–27.

Silva A, Tavares-Dias M, Jerônimo G, Martins M (2011) Parasite diversity in *Oxydoras niger* (Osteichthyes: Doradidae) from the basin of Solimões River, Amazonas state, Brazil, and the relationship between monogenoidean and condition factor. Brazilian Journal of Biology 71: 791–796. https://doi.org/10.1590/S1519-69842011000400026

Silva ACS Jr. (2010) Parasitismo por cestoides da ordem Trypanorhyncha na musculatura de *Plagioscion squamosissimus* pescada branca (Perciforme: Sciaenidae), comercializados em Macapá, AP. Ciência Animal Brasileira 11: 737–742. https://doi.org/10.5216/cab.v11i3.8495

Silva CM, São Clemente SC (2001) Nematóides da família Anisakidae e cestóides da ordem Trypanoryncha em filés de dourado (*Coryphaena hippurus*) e ariocó (*Lutjanus synagris*) e sua importância na inspeção de pescado. Higiene Alimentar 15: 75–79.

Silva LO, Luque JL, Alves DR (2000a) Metazoários parasitos do peixe espada, *Trichiurus lepturus* (Osteichthyes: Trichiuridae) do litoral do Estado do Rio de Janeiro, Brasil. Parasitología al Día 24: 97–101. https://doi.org/10.4067/S0716-07202000000300005

Silva LO, Luque JL, Alves DR, Paraguassú AR (2000b) Ecologia da comunidade de metazoários parasitos do peixe-espada *Trichiurus lepturus* Linnaeus (Osteichthyes, Trichiuridae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Brasileira de Zoociências 2: 115–133.

Silva RM, Tavares-Dias M, Dias MWR, Dias MKR, Marinho RGB (2013) Parasitic fauna in hybrid tambacu from fish farms. Pesquisa Agropecuária Brasileira 48: 1049–1057. https://doi.org/10.1590/S0100-204X2013000800034

Soares IA, Vieira FM, Luque JL (2014) Parasite community of *Pagrus pagrus* (Sparidae) from Rio de Janeiro, Brazil: evidence of temporal stability. Revista Brasileira de Parasitologia Veterinária 23: 216–223. https://doi.org/10.1590/S1984-29612014047

Soto J, Carvajal JG (1979) Parásitos cestodos de algunos peces comerciales de Antofagasta, Chile. Boletín Chileno de Parasitología 34: 67–71.

Spalding MD, Fox HE, Allen GR, Davidson N, Ferdaña ZA, Finlayson M, Halpern BS, Jorge MA, Lombana AL, Lourie SA, Martin KD, McManus E, Molnar J, Recchia CA, Robertson J (2007) Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. BioScience 57: 573–583. https://doi.org/10.1641/B570707

Suriano DM (1966) Estudio de la fauna parasitaria de "*Micropogon opercularis*" en relación con problemas zoogeográficos del Atlantico Sur. Comunicaciones del Museo Argentino de Ciencias Naturales "Bernardo Rivadavia" 1: 31–47.

Suriano DM (2002) *Anthobothrium galeorhini* n. sp. (Eucestoda: Tetraphyllidea) a parasite of *Galeorhinus galeus* (Triakidae) from the Argentine coast. Parasite 9: 121–125. https://doi.org/10.1051/parasite/2002092121

Suriano DM, Labriola JB (1998) Redescription of *Anonchocephalus chilensis* (Riggenbach, 1896) (Pseudophyllidea: Triaenophoridae) and description of *A. patagonicus* n. sp. Boletín Chileno de Parasitología 53: 73–77.

Suriano DM, Labriola JB (2001a) Redescription of *Cathetocephalus australis* Schmidt et Beveridge, 1990 (Cestoda, Cathetocephalidae) parasite of *Carcharinus brachyurus* (Gunther) (Pisces, Carcharhiniformes) from the southwestern Atlantic Ocean. Acta Parasitologica 46: 276–279.

Suriano DM, Labriola JB (2001b) A new *Orygmatobothrium* Diesing, 1863 (Eucestoda, Tetraphyllidea) parasite of *Mustelus schmitti* Springer, 1939 (Carcharhiniformes, Triakidae) from the southwestern Atlantic Ocean. Zoosystema 23: 669–674.

Szidat L (1955) La fauna parásitos de *Merluccius hubbsi* como carácter auxiliar para la solución de los problemas sistemáticos y zoogeográficos del género *Merluccius*. Comunicaciones del Instituto Nacional de Investigaciones de Ciencias Naturales, Buenos Aires 3: 1–54

Szidat L (1960) La parasitología como ciencia auxiliar para develar problemas hidrobiológicos, zoogeográficos y geofísicos del Atlántico Sudamericano. Libro Homenaje al Dr. Eduardo Caballero. Instituto de Biología, UNAM, 577–594.

Szidat L (1961) Versuch einer Zoogeographie des Süd-Atlantik mit Hilfe von Leitparasiten der Meeresfische. Parasitologische Schriftenreihe 13: 1–98.

Szidat L (1964) Vergleichende Helminthologische Untersuchungen an den argentinischen grossmöwen *Larus marinus dominicanus* Lichtenstein und *Larus ridibundus maculipennis* Lichtenstein nebst neuen Beobachtungen über die Artbildung bei Parasiten. Zeitschrift für Parasitenkunde 24: 351–414. https://doi.org/10.1007/BF00260454

Szidat L (1969) Los parasitos de la "Palometa" *Parona signata* (Jenyns, 1842) Berg, 1895 y su aplicacion a problemas zoogeograficos del Atlantico sur. Neotrópica 15: 125–131.

Szidat L, Nani A (1951) Diplostomiasis cerebralis del pejerrey: una grave epizootia que afecta a la economía nacional producida por larvas de trematodes que destruyen el cerebro de los pejerreyes. Revista del Instituto Nacional de Investigación de las Ciencias Naturales 1: 324–384

Szidat L, Soria MF (1957) Difilobotriasis en nuestro país. Sobre una nueva especie de *Sparganum*, parásita de salmónidos y de *Diphyllobothrium*, parásita de gaviotas del lago Nahuel Huapi. Boletín del Museo Argentino de Ciencias Naturales 9: 1–22.

Tagle I (1951) Parásitos en la merluza. Boletin Informativo de Parasitologia del Chile 6: 8–9.

Takemoto RM, Amato JFR, Luque JL (1996a) Comparative analysis of the metazoan parasite communities of leatherjackets, *Oligoplites palometa*, *O. saurus*, and *O. saliens* (Osteichthyes: Carangidae) from Sepetiba Bay, Rio de Janeiro, Brazil. Revista Brasileira de Biologia 56: 639–650.

Takemoto RM, Amato JFR, Luque JL (1996b) Larvas de Eucestoda parasitas de *Oligoplites* (Osteichthyes, Carangidae) do litoral do Estado do Rio de Janeiro, Brasil. Revista Unimar 18: 283–291.

Takemoto RM, Pavanelli GC (1994) Ecological aspects of proteocephalidean cestodes parasites of *Paulicea luetkeni* (Steindachner) (Osteichthyes: Pimelodidae) from the Paraná River, Paraná, Brazil. Revista Unimar 16: 17–26.

Takemoto RM, Pavanelli GC (1996) Proteocephalidean cestodes in the freshwater fish *Cichla monoculus* from the Paraná River, Brazil. Studies on Neotropical Fauna and Environment 31: 123–127. https://doi.org/10.1076/snfe.31.2.123.13326

Takemoto RM, Pavanelli GC (2000) Aspects of the ecology of proteocephalid cestodes parasites of *Sorubim lima* (Pimelodidae) of the upper Paraná River, Brazil: I. Structure and influence of host's size and sex. Revista Brasileira de Biologia 60: 577–584. https://doi.org/10.1590/S0034-71082000000400006

Takemoto RM, Pavanelli GC, Lizama MAP, Lacerda ACF, Yamada FH, Moreira LHA, Ceschini TL, Bellay S (2009) Diversity of parasites of fish from the Upper Paraná River floodplain, Brazil. Brazilian Journal of Biology 69: 691–705. https://doi.org/10.1590/S1519-69842009000300023

Tantaleán MV (1975) Hallazgo de larvas plerocercoides de Diphyllobothriidae Lühe (Cestoda) en peces del mar peruano. Boletín Chileno de Parasitología 30: 18–20.

Tantaleán MV (1991) Nuevos helmintos parasitos en peces elasmobranquio de la costa peruana. Boletin de Lima 13: 25–28.

Tantaleán MV, Huiza A (1994) Sinopsis de los parásitos de peces marinos de la costa peruana. Biotempo 1: 53–101.

Tantaleán MV, Rodríguez J (1987) Nuevos registros de helmintos parásitos de peces elasmobranquios de las costas del Perú. Revista de Biologia Tropical 35: 167–168.

Tanzola R, Guagliardo S (2000) Helminth fauna of the Argentine conger, *Conger orbignyanus* (Pisces: Anguilliformes). Helminthologia 37: 229–232.

Tanzola RD, Guagliardo SE, Brizzola SM, Arias MV (1997) Helminth fauna of *Porichthys porosissimus* (Pisces: Batrachoidiformes) in the estuary of Bahia Blanca Argentina. Helminthologia 34: 221–227.

Tanzola RD, Guagliardo SE, Brizzola SM, Arias MV, Botte SE (1998) Parasite assemblage of *Sympterygia bonapartei* (Pisces: Rajidae), an endemic skate of the Southwest Atlantic. Helminthologia 35: 123–129.

Tavares LER, Bicudo AJA, Luque JL (2004) Metazoan parasites of the needlefish *Tylosurus acus* (Lacépède, 1803) (Osteichthyes: Belonidae) from the coastal zone of the State of Rio de Janeiro, Brazil Revista Brasileira de Parasitologia Veterinária 13: 36–40.

Tavares LER, Luque JL (2004) Community ecology of the metazoan parasites of white sea catfish, *Netuma barba* (Osteichthyes: Ariidae), from the coastal zone of the state of Rio de Janeiro, Brazil. Brazilian Journal of Biology 64: 169–176. https://doi.org/10.1590/S1519-69842004000100019

Tavares LER, Luque JL (2008) Similarity between metazoan parasite communities of two sympatric brackish fish species from Brazil. Journal of Parasitology 94: 985–989. https://doi.org/10.1645/GE-1460.1

Tavares LER, Luque JL, Bicudo AJA (2005) Community ecology of metazoan parasites of the anchovy *Anchoa tricolor* (Osteichthyes: Engraulidae) from the coastal zone of the State of Rio de Janeiro, Brazil. Brazilian Journal of Biology 65: 533–540. https://doi.org/10.1590/S1519-69842005000300019

Tavares LER, Luque JL, Neto SLB (2001) Ecologia da comunidade de metazoários parasitos do olho-de-cão *Priacanthus arenatus* (Cuvier, 1829) (Osteichthyes, Priacanthidae) do litoral do estado do Rio de Janeiro, Brasil. Revista Brasileira de Zoociências 3: 45–59.

Thatcher VE (2006) Amazon Fish Parasites. Pensoft Publishers, Sofia-Moscow, 508 pp.

Timi JT (2003) Parasites of Argentine anchovy in the south-west Atlantic: latitudinal patterns and their use for discrimination of host populations. Journal of Fish Biology 63: 90–107. https://doi.org/10.1046/j.1095-8649.2003.00131.x

Timi JT, Lanfranchi AL (2009a) The metazoan parasite communities of the Argentinean sandperch *Pseudopercis semifasciata* (Pisces: Perciformes) and their use to elucidate the stock structure of the host. Parasitology 136: 1209–1219. https://doi.org/10.1017/S0031182009990503

Timi JT, Lanfranchi AL (2009b) The importance of the compound community on the parasite infracommunity structure in a small benthic fish. Parasitology Research 104: 295–302. https://doi.org/10.1007/s00436-008-1191-1

Timi JT, Lanfranchi AL (2013) Ontogenetic changes in heterogeneity of parasite communities of fish: disentangling the relative role of compositional versus abundance variability. Parasitology 140: 309–317. https://doi.org/10.1017/S0031182012001606

Timi JT, Lanfranchi AL, Etchegoin JA (2009) Seasonal stability and spatial variability of parasites in Brazilian sandperch *Pinguipes brasilianus* from the Northern Argentine Sea: evidence for stock discrimination. Journal of Fish Biology 74: 1206–1225. https://doi.org/10.1111/j.1095-8649.2009.02190.x

Timi JT, Lanfranchi AL, Etchegoin JA, Cremonte F (2008) Parasites of the Brazilian sandperch *Pinguipes brasilianus* Cuvier: a tool for stock discrimination in the Argentine Sea. Journal of Fish Biology 72: 1332–1342. https://doi.org/10.1111/j.1095-8649.2008.01800.x

Timi JT, Lanfranchi AL, Luque JL (2010a) Similarity in parasite communities of the teleost fish *Pinguipes brasilianus* in the southwestern Atlantic: infracommunities as a tool to detect geographical patterns. International Journal for Parasitology 40: 243–254. https://doi.org/10.1016/j.ijpara.2009.07.006

Timi JT, Luque JL, Poulin R (2010b) Host ontogeny and the temporal decay of similarity in parasite communities of marine fish. International Journal for Parasitology 40: 963–968. https://doi.org/10.1016/j.ijpara.2010.02.005

Timi JT, Luque JL, Sardella NH (2005) Parasites of *Cynoscion guatucupa* along South American Atlantic coasts: evidence for stock discrimination. Journal of Fish Biology 67: 1603–1618. https://doi.org/10.1111/j.1095-8649.2005.00867.x

Timi JT, Poulin R (2003) Parasite community structure within and across host populations of a marine pelagic fish: how repeatable is it? International Journal for Parasitology 33: 1353–1362. https://doi.org/10.1016/S0020-7519(03)00203-0

Torres P (1990) Primeros registros de endohelmintos parásitos en el salmón coho *Oncorhynchus kisutch* (Walbaum), introducido en Chile. Archivos de Medicina Veterinaria 22: 105–107.

Torres P, Aedo E, Figueroa L, Siegmund I, Silva R, Navarrete N, Puga S, Marín F, Aedo E (2000) Infección por helmintos parásitos en salmón coho, *Oncorhynchus kisutch*, durante su retorno al río Simpson, Chile. Boletín Chileno de Parasitología 55: 31–35. https://doi.org/10.4067/S0365-94022000000100009

Torres P, Contreras A, Figueroa L, Franjola R, González H, Martín R (1977) Investigaciones sobre Pseudophyllidea (Carus, 1813) en el sur de Chile. I. Estudio preliminar sobre infección por plerocercoides de *Diphyllobothrium* sp. em *Salmo gairdneri* Richardson, 1836 del lago Calafquén (39°32"S, 72°09"O), Chile. Boletín Chileno de Parasitología 32: 73–80.

Torres P, Contreras A, Revenga J, Fritz N (1993) Helminth parasites in fishes from Valdivia and Tornagaleones river estuaries in the south of Chile. Memórias do Instituto Oswaldo Cruz 88: 491–492. https://doi.org/10.1590/S0074-02761993000300021

Torres P, Cubillos V, Aedo E, Silva R, Garrido O, Aedo JE (1995) Prevalencia y aspectos patológicos de la difilobotriasis en salmones de retorno, *Oncorhynchus kisutch* de Coyhaique, XI Región de Chile. Archivos de Medicina Veterinaria 27: 107–114.

Torres P, Cubillos V, Gesche W, Rebolledo C, Montefusco A, Miranda JC, Arenas J, Mira A, Nilo M, Abello C (1991) Difilobotriasis en salmónidos introducidos en lagos del sur de Chile. Aspectos patológicos, relación con infección humana, animales domésticos y aves piscívoras. Archivos de Medicina Veterinaria 23: 165–183.

Torres P, Cuevas C, Tang M, Barra M, Franjola R, Navarrete N, Montefusco A, Otth L, Wilson G, Puga S, Figueroa L, Cerda O (2004) Introduced and native fishes as infection foci of *Diphyllobothrium* spp. in humans and dogs from two localities at Lake Panguipulli in Southern Chile. Comparative Parasitology 71: 111–117. https://doi.org/10.1654/4119

Torres P, Figueroa L, Franjola R (1983) Pseudophyllidea in the South of Chile. IX. Types of plerocercoids in trouts from five lakes and new cases of *Diphyllobothrium latum* in man and *D. pacificum* in a dog. International Journal of Zoonoses 10: 15–21.

Torres P, Franjola R, Contreras LF (1982) Pseudophyllidea (Carus, 1813) en el sur de Chile: VII. Distribución estacional de la infección por plerocercoides en *Salmo gairdneri* (Richardson) del lago Calafquén. Zentralblatt für Veterinärmedizin Reihe B 29: 67–75. https://doi.org/10.1111/j.1439-0450.1982.tb01190.x

Torres P, Franjola R, Figueroa L, Schlatter R, González H, Contreras B, Martin R (1981) Researches on Pseudophyllidea (Carus, 1813) in the south of Chile. IV Occurrence of *Diphyllobothrium dendriticum* (Nitzch). Journal of Helminthology 55: 173–188. https://doi.org/10.1017/S0022149X00026833

Torres P, Franjola R, Pérez J, Auad S, Uherek F, Miranda JC, Flores L, Riquelme J, Salazar S, Hermosilla C, Rojo R (1989a) Epidemiología de la difilobotriasis en la cuenca del río Valdivia, Chile. Revista de Saúde Pública 23: 45–57. https://doi.org/10.1590/S0034-89101989000100007

Torres P, Gesche W, Montefusco A, Miranda J, Dietz P, Huijse R (1998) Diphyllobothriosis humana y en peces del lago Riñihue, Chile: efecto de la actividad educativa, distribución estacional y relación con sexo, talla y dieta de los peces. Archivos de Medicina Veterinaria 30: 31–45. https://doi.org/10.4067/S0301-732X1998000100004

Torres P, Leyán V, Puga S (2012) Prevalence, intensity, and abundance of infection and pathogenesis caused by diphyllobothriosis in vulnerable, native fish and introduced trout in Lake Panguipulli, Chile. Journal of Wildlife Diseases 48: 937–950. https://doi.org/10.7589/2011-08-235

Torres P, Lopez JC, Cubillos V, Lobos C, Silva R (2002) Visceral diphyllobothriosis in a cultured rainbow trout, *Oncorhynchus mykiss* (Walbaum), in Chile. Journal of Fish Diseases 25: 375–379. https://doi.org/10.1046/j.1365-2761.2002.00381.x

Torres P, Puga S (2011) Comparative efficacy of candling and glass plate compression for detection of diphyllobothriosis in rainbow trout (*Oncorhynchus mykiss*) musculature. Revue Scientifique et Technique (International Office of Epizootics) 30: 831–837.

Torres P, Puga S, Castillo L, Lamilla J, Miranda J (2014) Helmintos, myxozoos y microsporidios en músculos de peces comercializados frescos y su importancia como riesgo potencial para la salud humana en la ciudad de Valdivia, Chile. Archivos de Medicina Veterinaria 46: 83–92. https://doi.org/10.4067/S0301-732X2014000100012

Torres P, Quintanilla J, Rozas M, Miranda P, Ibarra R, San Martín M, Raddatz B, Wolter M, Villegas A, Canobra C, Hausdorf M, Silva R (2010) Endohelminth parasites from salmonids in intensive culture from southern Chile. Journal of Parasitology 96: 669–670. https://doi.org/10.1645/GE-2211.1

Torres P, Roman C, Figueroa L, Franjola R (1980) Plerocercoids of *Diphyllobothrium* (Cobbold) in fishes and identification of copepods in plankton from Calafquen Lake, Chile. Indian Journal of Parasitology 4: 207–208.

Torres P, Ruíz E, Rebolledo C, Mira A, Cubillos V, Navarrete N, Gesche W, Montefusco A, Valdés L, Alberdi A (1990) Parasitism in fishes and human riverside communities of the Huillinco and Natri lakes (Great Island of Chiloe), Chile. Boletín Chileno de Parasitología 45: 47–55.

Torres P, Torres J, Garrido O, Thibaut I (1989b) Investigaciones sobre Pseudophyllidea (Carus, 1813) en el sur de Chile. X. Observaciones experimentales sobre la coexistencia de plero-

cercoides de *Diphyllobothrium latum* (L.) y *D. dendriticum* (Nitzch) en salmonidos de la cuenca del rio Valdívia. Archivos de Medicina Veterinaria 21: 51–57.

Travassos L (1940) Relatório da terceira excursão a zona da Estrada de Ferro Noroeste do Brasil realizada em Fevereiro e Março de 1940: I-Introdução. Memórias do Instituto Oswaldo Cruz 35: 607–696. https://doi.org/10.1590/S0074-02761940000300013

Travassos L (1944) Relatório da excursão do Instituto Oswaldo Cruz ao Município de Santa Teresa, no Estado do Espírito Santo, em agôsto e setembro de 1943. Memórias do Instituto Oswaldo Cruz 40: 121–128. https://doi.org/10.1590/S0074-02761944000200002

Travassos L (1945) Relatório da excursão do Instituto Oswaldo Cruz ao Rio Paraná (Porto Cabral), em março e abril de 1944. Memórias do Instituto Oswaldo Cruz 42: 151–165. https://doi.org/10.1590/S0074-02761945000100010

Travassos L (1947) Relatório da excursão do Instituto Oswaldo Cruz realizada no Estado de S. Paulo em novembro e dezembro de 1946. Memórias do Instituto Oswaldo Cruz 45: 619–627. https://doi.org/10.1590/S0074-02761947000300009

Travassos L, Pinto C, Muniz J (1927) Excursão scientifica ao Estado de Matto Grosso na zona do Pantanal (Margens dos Rios S. Lourenço e Cuyabá) realisada em 1922. Memórias do Instituto Oswaldo Cruz 20: 249–269. https://doi.org/10.1590/S0074-02761927000200004

Travassos L, Teixeira de Freitas JF (1940) Relatório da excursão científica realisada na zona da Estrada de Ferro Noroeste do Brasil em Julho de 1939. Memórias do Instituto Oswaldo Cruz 35: 525–556. https://doi.org/10.1590/S0074-02761940000300004

Travassos L, Teixeira de Freitas JF (1941) Relatório da quinta excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em janeiro de 1941: II Pesquisas Parasitológicas. Memórias do Instituto Oswaldo Cruz 36: 272–300. https://doi.org/10.1590/S0074-02761941000300003

Travassos L, Teixeira de Freitas JF (1942) Relatório da sexta excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em novembro de 1941. Memórias do Instituto Oswaldo Cruz 37: 259–286. https://doi.org/10.1590/S0074-02761942000300004

Travassos L, Teixeira de Freitas JF (1943) Relatório da sétima excursão científica do Instituto Oswaldo Cruz, realizada a zona da Estrada de Ferro Noroeste do Brasil, em maio de 1942. Memórias do Instituto Oswaldo Cruz 38: 385–412. https://doi.org/10.1590/S0074-02761943000300007

Travassos L, Teixeira de Freitas JF (1948) Relatório da excursão do Instituto Oswaldo Cruz ao norte do Estado do Espírito Santo, junto ao Parque de Reserva e Refúgio Soóretama, em fevereiro e março de 1948. Memórias do Instituto Oswaldo Cruz 46: 605–631. https://doi.org/10.1590/S0074-02761948000300006

Travassos L, Teixeira de Freitas JF (1964) Pesquisas helmintológicas, realizadas em Maicuru, Estado do Pará. Museu Paraense Emílio Goeldi, Belém, 16 pp.

Travassos L, Teixeira de Freitas JF, Bührnheim PF (1967) Relatório da excursão do Instituto Oswaldo Cruz ao estado do Espírito Santo em novembro de 1964. Boletím do Museu de Biologia Prof Mello Leitâo (Zoologia) 31: 1−54

Tresierra A, Escalante H, Benitez J (1986) Hábitos alimenticios y fauna helmintológica de tres especies de elasmobranquios del mar peruano. Rebiol 6: 35–46.

Tyler GA (2006) Tapeworms of elasmobranchs (Part II). A monograph on the Diphyllidea (Platyhelminthes: Cestoda). Bulletin of the University of Nebraska State Museum 20: 1–142.

Valdivia IM, Chávez RA, Oliva ME (2007) Metazoan parasites of *Engraulis ringens* as tools for stock discrimination along the Chilean coast. Journal of Fish Biology 70: 1504–1511. https://doi.org/10.1111/j.1095-8649.2007.01429.x

Vales DG, García NA, Crespo EA, Timi JT (2011) Parasites of a marine benthic fish in the Southwestern Atlantic: searching for geographical recurrent patterns of community structure. Parasitology Research 108: 261–272. https://doi.org/10.1007/s00436-010-2052-2

Vásquez-Ruiz CE, Jara-Campos CA (2012) Prevalencia e intensidad parasitaria en *Coryphaena hippurus* y *Mugil cephalus* (Teleostei) desembarcados en los puertos Salaverry y Paita (Perú). Sciéndo 15: 22–32.

Vergara LA, George-Nascimento M (1982) Contribucion al estudio del parasitismo en el congrio colorado *Genypterus chilensis* (Guichenot, 1848). Boletín Chileno de Parasitología 37: 9–14.

Vicente JJ, Fernandes GL (1978) Contribuição ao conhecimento dos helmintos de *Bagre bagre* (linnaeus, 1766) Fowler, 1841 e de *Macrodon ancylodon* (Bloch, 1801) Jordan, Evermann e Clark, 1930, no litoral da ilha de São Luís, Estado do Maranhão, Brasil. Boletim do Laboratório de Hidrobiologia 2: 91–95.

Vicente JJ, Pinto RM, Aguilera O (1989) On *Dichelyne* (*Cucullanellus*) *elongatus* (Tornquist, 1931) Petter, 1974: South American correlated species (Nematoda, Cucullanidae) and some other helminths of *Micropogonias furnieri* (Desmarest, 1823) (Pisces, Sciaenidae). Memórias do Instituto Oswaldo Cruz 84: 357–361. https://doi.org/10.1590/S0074-02761989000300010

Vicente JJ, Santos E (1974) Alguns helmintos de peixes do litoral norte fluminense-II. Memórias do Instituto Oswaldo Cruz 72: 173–180. https://doi.org/10.1590/S0074-02761974000200002

Videira M, Velasco M, Dias L, Matos P, Almeida HDF, Costa ML, São Clemente SC, Matos E (2013) *Gobioides broussonnetii* (Gobiidae): a new host for *Pterobothrium crassicolle* (Trypanorhyncha) on Marajó Island, northern Brazil. Revista Brasileira de Parasitologia Veterinária 22: 398–401. https://doi.org/10.1590/S1984-29612013000300013

Villalba CS, Fernández JB (1985) Parásitos de *Mola ramsayi* (Giglioli, 1883) (Pisces: Molidae) en Chile. Boletín de la Sociedad de Biología de Concepción 56: 71–78.

Viozzi G, Semenas L, Brugni N, Flores VR (2009) Metazoan parasites of *Galaxias maculatus* (Osmeriformes: Galaxiidae) from Argentinean Patagonia. Comparative Parasitology 76: 229–239. https://doi.org/10.1654/4328.1

Vogelsang EG, Mayaudon TH (1959) Contribucion al estudio de la parasitología animal en Venezuela (XXIII). Revista de Medicina Veterinaria y Parasitología 18: 5–9.

Waeschenbach A, Webster BL, Littlewood DT (2012) Adding resolution to ordinal level relationships of tapeworms (Platyhelminthes: Cestoda) with large fragments of mtDNA. Molecular Phylogenetics and Evolution 63: 834–847. https://doi.org/10.1016/j.ympev.2012.02.020

Waicheim A, Blasetti G, Cordero P, Rauque C, Viozzi G (2014) Macroparasites of the invasive fish, *Cyprinus carpio*, in Patagonia, Argentina. Comparative Parasitology 81: 270–275. https://doi.org/10.1654/1525-2647-81.2.270

Whittaker FH, Carvajal JG (1980) Scanning electron microscopy of scolices of some cestodes from elasmobranchs. Proceedings of the Helminthological Society of Washington 47: 256–259.

Whittaker FH, Carvajal JG, Apkarian R (1982) Scanning electron microscopy of the scolex of *Grillotia dollfusi* Carvajal 1971 (Cestoda: Trypanorhyncha). Journal of Parasitology 68: 1173–1175. https://doi.org/10.2307/3281120

Wicht B, Yanagida T, Scholz T, Ito A, Jiménez JA, Brabec J (2010) Multiplex PCR for differential identification of broad tapeworms (Cestoda: *Diphyllobothrium*) infecting humans. Journal of Clinical Microbiology 48: 3111–3116. https://doi.org/10.1128/JCM.00445-10

Willis SC, Macrander J, Farias IP, Ortí G (2012) Simultaneous delimitation of species and quantification of interspecific hybridization in Amazonian peacock cichlids (genus *Cichla*) using multi-locus data. BMC Evolutionary Biology 12: 96. https://doi.org/10.1186/1471-2148-12-96

Wolffügel K (1949) ¿Es autóctono el *Diphyllobothrium* en Chile? Boletín de la Sociedad de Biología de Concepción 24: 85–89.

Woodland WNF (1933a) On a new subfamily of proteocephalid cestodes–the Othinoscolecinae–from the Amazon siluroid fish *Platystomatichthys sturio* (Kner). Parasitology 25: 491–500. https://doi.org/10.1017/S0031182000019739

Woodland WNF (1933b) On the anatomy of some fish cestodes described by Diesing from the Amazon. Quarterly Journal of Microscopical Science 76: 175–208.

Woodland WNF (1933c) On two new cestodes from the Amazon siluroid fish *Brachyplatystoma vaillanti* Cuv. and Val. Parasitology 25: 485–490. https://doi.org/10.1017/S0031182000019727

Woodland WNF (1934a) On the Amphilaphorchidinae, a new subfamily of proteocephalid cestodes, and *Myzophorus admonticellia*, gen. et sp. n., parasitic in *Pirinampus* spp. from the Amazon. Parasitology 26: 141–149. https://doi.org/10.1017/S0031182000023441

Woodland WNF (1934b) On some remarkable new cestodes from the Amazon siluroid fish, *Brachyplatystoma filamentosum* (Lichtenstein). Parasitology 26: 268–277. https://doi.org/10.1017/S0031182000023556

Woodland WNF (1934c) On six new cestodes from Amazon fishes. Proceedings of the Zoological Society of London 104: 33–44. https://doi.org/10.1111/j.1469-7998.1934.tb06218.x

Woodland WNF (1935a) Additional cestodes from the Amazon siluroids pirarará, dorad, and sudobim. Proceedings of the Zoological Society of London 104: 851–862. https://doi.org/10.1111/j.1096-3642.1934.tb01669.x

Woodland WNF (1935b) Some more remarkable cestodes from Amazon siluroid fish. Parasitology 27: 207–225. https://doi.org/10.1017/S0031182000023556

Woodland WNF (1935c) Some new proteocephalids and a ptychobothriid (Cestoda) from the Amazon. Proceedings of the Zoological Society of London 105: 619–623. https://doi.org/10.1111/j.1096-3642.1935.tb01685.x

Yamada FH, Takemoto RM (2013) Metazoan parasite fauna of two peacock-bass cichlid fish in Brazil. Check List 9: 1371–1377. https://doi.org/10.15560/9.6.1371

Yamada FH, Takemoto RM, Pavanelli GC (2007) Ecological aspects of ectoparasites from the gills of *Satanoperca pappaterra* (Heckel, 1840) (Cichlidae) from the upper Paraná

river floodplain, Brazil. Acta Scientiarum: Biological Sciences 29: 331–336. https://doi.org/10.4025/actascibiolsci.v29i3.555

Yáñez P (1950) Observación de un *Dibothriorhynchus* parásito del azulejo. Revista de Biología Marina 2: 165–166.

Zago AC, Franceschini L, Zocoller-Seno MC, Veríssimo-Silveira R, Maia AAD, Ikefuti CV, Silva RJ (2013) The helminth community of *Geophagus proximus* (Perciformes: Cichlidae) from a tributary of the Paraná River, Ilha Solteira Reservoir, São Paulo State, Brazil. Journal of Helminthology 87: 203–211. https://doi.org/10.1017/S0022149X12000223

Zamparo D, Brooks DR, Barriga R (1999) *Pararhinebothroides hobergi* n. gen. n. sp. (Eucestoda: Tetraphyllidea) in *Urobatis tumbesensis* (Chondrichthyes: Myliobatiformes) from coastal Ecuador. Journal of Parasitology 85: 534–539. https://doi.org/10.2307/3285791

Zehnder MP, de Chambrier A (2000) Morphological and molecular analyses of the genera *Peltidocotyle* Diesing 1850 and *Othinoscolex* Woodland 1933, and a morphological study of *Woodlandiella* Freze, 1965 (Eucestoda, Proteocephalidea), parasites of South American siluriform fishes (Pimelodidae). Systematic Parasitology 46: 33–43. https://doi.org/10.1023/A:1006252601201

Zehnder MP, de Chambrier A, Vaucher C, Mariaux J (2000) *Nomimoscolex suspectus* n. sp. (Eucestoda: Proteocephalidea: Zygobothriinae) with morphological and molecular phylogenetic analyses of the genus. Systematic Parasitology 47: 157–172. https://doi.org/10.1023/A:1006465026316

Zehnder MP, Mariaux J (1999) Molecular systematic analysis of the order Proteocephalidea (Eucestoda) based on mitochondrial and nuclear rDNA sequences. International Journal for Parasitology 29: 1841–1852. https://doi.org/10.1016/S0020-7519(99)00122-8