



Complete Genome Sequence of *Lactobacillus jensenii* Strain SNUV360, a Probiotic for Treatment of Bacterial Vaginosis Isolated from the Vagina of a Healthy Korean Woman

Sunghye Lee,^{a,b} Hyun Ju You,^{c,d,e} Bomi Kwon,^c GwangPyo Ko^{a,b,c,e}

KoBioLabs, Inc., Seoul, Republic of Korea^a; N-Bio, Seoul National University, Seoul, Republic of Korea^b; Department of Environmental Health Sciences, Graduate School of Public Health, Seoul National University, Seoul, Republic of Korea^c; Institute of Health and Environment, Seoul National University, Seoul, Republic of Korea^d; Center for Human and Environmental Microbiome, Seoul National University, Seoul, Republic of Korea^e

ABSTRACT *Lactobacillus jensenii* SNUV360 is a potential probiotic strain that shows antimicrobial activity for the treatment of bacterial vaginosis. Here, we present the complete genomic sequence of *L. jensenii* SNUV360, isolated from a vaginal sample from a healthy Korean woman. Analysis of the sequence may provide insight into its functional activity.

Bacterial vaginosis (BV) is a common condition associated with numerous adverse health outcomes in women of reproductive age. BV can be characterized by a shift in the vaginal flora from *Lactobacillus* spp. dominance to a more diverse microbial environment (1). Some *Lactobacillus* spp. strains have been shown to decrease pathogenic bacteria in vaginal environments of women diagnosed with BV (2, 3).

In this study, the strain *Lactobacillus jensenii* SNUV360, isolated from the vaginas of healthy women, shows functional properties in the treatment of vaginal infection caused by lactobacilli deficiency. Here, we present the complete genome sequence of this probiotic strain.

In order to perform the complete genome sequencing of the strain *L. jensenii* SNUV360, high throughput sequencing technology was implemented using the PacBio platform (Pacific Biosciences, Menlo Park, CA). A 20-Kb library was constructed with purified DNA affixed to single-molecule real-time (SMRT) cell and was sequenced using P6-C4 chemistry with a data collection time of 4 h. The sequencing run provided a total of 143,207 reads with a quality score of Q20. The number of bases was 964,237,020 bp. *De novo* assembly employed the default parameters of the Hierarchical Genome Assembly Process approach version 3 (HGAP3) (4). The *L. jensenii* SNUV360 genome consisted of a 1,672,949 bp single-chromosome contig, with coverage of 420× and G+C content of 34.4%. No plasmids were detected.

The assembled genome sequences were annotated using the Prokka annotation pipeline, version 1.11 (5), which predicted tRNA, rRNA, and mRNA genes. Putative gene products were then assigned to protein-coding genes (CDSs) based on their similarity to the sequences in the respective database. Curated virulence factors and antibiotic resistance genes were estimated by using IslandViewer3 (6) against the Virulence Factor Database (VFDB) (7) of virulence factors, and Comprehensive Antibiotic Resistance Database (CARD) (8) of antibiotic resistance genes.

The genome contains 1,595 CDSs, 58 tRNA genes, and 12 rRNA genes. The *L. jensenii* SNUV360 genome was compared with the reference strain *L. jensenii* TL2937 (genome accession number NZ_MDTN01000000) with the Rapid Annotations using the Subsys-

Received 29 December 2016 Accepted 11 January 2017 Published 9 March 2017

Citation Lee S, You HJ, Kwon B, Ko G. 2017. Complete genome sequence of *Lactobacillus jensenii* strain SNUV360, a probiotic for treatment of bacterial vaginosis isolated from the vagina of a healthy Korean woman. Genome Announc 5:e01757-16. <https://doi.org/10.1128/genomeA.01757-16>.

Copyright © 2017 Lee et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to GwangPyo Ko, gko@snu.ac.kr.

tems Technology (RAST) server (9). In this comparison, we detected 42 elements absent in the published strain *L. jensenii* TL2937. No remarkable antibiotic resistance or virulence-associated genes were found. The analysis of the complete genome of *L. jensenii* SNUV360 may assist in understanding the mechanisms involved in its effect against bacterial vaginosis.

Accession number(s). The results of this whole-genome project have been deposited at GenBank under accession no. [CP018809](https://www.ncbi.nlm.nih.gov/nuclseq/CP018809).

ACKNOWLEDGMENT

This project was supported by the National Research Foundation of Korea (NRF) (Grant No. NRF-2015R1A2A1A10054078).

REFERENCES

1. Koumans EH, Sternberg M, Bruce C, McQuillan G, Kendrick J, Sutton M, Markowitz LE. 2007. The prevalence of bacterial vaginosis in the United States, 2001–2004; associations with symptoms, sexual behaviors, and reproductive health. *Sex Transm Dis* 34:864–869. <https://doi.org/10.1097/OLQ.0b013e318074e565>.
2. Anukam K, Osazuwa E, Ahonkhai I, Ngwu M, Osemene G, Bruce AW, Reid G. 2006. Augmentation of antimicrobial metronidazole therapy of bacterial vaginosis with oral probiotic *Lactobacillus rhamnosus* GR-1 and *Lactobacillus reuteri* RC-14: randomized, double-blind, placebo controlled trial. *Microbes Infect* 8:1450–1454. <https://doi.org/10.1016/j.micinf.2006.01.003>.
3. Reid G, Charbonneau D, Erb J, Kochanowski B, Beuerman D, Poehner R, Bruce AW. 2003. Oral use of *Lactobacillus rhamnosus* GR-1 and *L. fermentum* RC-14 significantly alters vaginal flora: randomized, placebo-controlled trial in 64 healthy women. *FEMS Immunol Med Microbiol* 35:131–134. [https://doi.org/10.1016/S0928-8244\(02\)00465-0](https://doi.org/10.1016/S0928-8244(02)00465-0).
4. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
5. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
6. Dhillon BK, Laird MR, Shay JA, Winsor GL, Lo R, Nizam F, Pereira SK, Waglechner N, McArthur AG, Langille MG, Brinkman FS. 2015. IslandViewer 3: more flexible, interactive genomic island discovery, visualization and analysis. *Nucleic Acids Res* 43:W104–W108. <https://doi.org/10.1093/nar/gkv401>.
7. Chen L, Xiong Z, Sun L, Yang J, Jin Q. 2012. VFDB 2012 update: toward the genetic diversity and molecular evolution of bacterial virulence factors. *Nucleic Acids Res* 40:D641–645. <https://doi.org/10.1093/nar/gkr989>.
8. McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yan M, Yu T, Wright GD. 2013. The comprehensive antibiotic resistance database. *Antimicrob Agents Chemother* 57:3348–3357. <https://doi.org/10.1128/AAC.00419-13>.
9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-1189-1175>.