

ORIGINAL RESEARCH

Immunosurveillance and immunoediting in MMTV-PyMT-induced mammary oncogenesis

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ABSTRACT

Evidence of cancer immunosurveillance and immunoediting processes has been primarily demonstrated in mouse models of chemically induced oncogenesis. Although these models are very tractable, they are characterized by high mutational loads that represent a minority of human cancers. In this study, we sought to determine whether cancer immunosurveillance and immunoediting could be demonstrated in a more clinically relevant oncogene-induced model of carcinogenesis, the MMTV-PyMT (PyMT) mammary carcinoma model. This model system in the FVB/NJ strain background was previously used to demonstrate that adaptive immunity had no role in limiting primary cancer formation and in fact promoted metastasis, thus calling into question whether cancer immunosurveillance operated in preventing the development of breast cancer. Our current study in the C57BL/6 strain backgrounds provides a different conclusion, as we report here the existence of an adaptive immunosurveillance of PyMT mammary carcinomas using two independent models of immune deficiency. PyMT mice bred onto a *Rag1*^{-/-} background or immune suppressed by chronic tacrolimus therapy both demonstrated accelerated development of mammary carcinomas. By generating a bank of cell lines from these animals, we further show that a subset of PyMT cell lines had delayed growth after transplantation into wild-type (WT) syngeneic, but not immune-deficient hosts. This reduced growth rate in immunocompetent animals was characterized by an increase in immune cell infiltration and tissue differentiation. Furthermore, loss of the immune cell infiltration that characterized immunoediting of slow growing cell lines, changed them into fast growing variants capable of progressing in the immunocompetent model. In conclusion, our study provides evidence that immunosurveillance and immunoediting of PyMT-derived cell lines modulate tumor progression in this oncogene-induced model of cancer.

Abbreviations: BFA, brefeldin A; HPF, high-power field; MCA, methylchloranthrene; MMTV-PyMT, mouse mammary tumor virus-polyoma virus middle T antigen; *vs.*, *versus*; WT, wild type; d, days

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Introduction

Evidence for cancer immunosurveillance comes from studies monitoring differences in cancer incidence in immunocompetent *versus* (*vs.*) immunodeficient animals. These studies utilize chemical carcinogenesis models, spontaneous models, and genetic cancer models. Using the well-studied 3-methylcholanthrene (MCA) chemically induced tumorigenesis model, several groups showed a requirement for adaptive immunity, natural killer (NK) cells, and NK-T cells in the surveillance of murine sarcomas.¹⁻⁷ These studies have shown that immunosurveillance of cancer cells occurs as part of an elimination step in the cancer immunoediting process.⁸ Animals that fail to completely eradicate MCA-induced sarcoma cells can undergo an equilibrium phase,¹ in which a small number of persisting tumor cells is held in check by an active antitumor immune response. This stage can be followed by the emergence of immune “edited,” escaped cancers that have acquired or abolished the expression of certain genes to avoid immune recognition.^{3,9-12} In addition to the MCA model, other studies using

spontaneous or genetic models of cancer have found a role for perforin, NKT cells, NKG2D, and CD226 in the surveillance of hematologic malignancies.^{7,13-17} Furthermore, genetic models showed that hepatocarcinoma and prostate cancer are surveilled by adaptive immune responses.¹⁸⁻²⁰

Although considerable support for cancer immunoediting has emerged, there is a paucity of studies and lack of consensus on the role of immunity in oncogene-induced non-hematologic cancers. In fact, recent studies using oncogene-induced sarcomas have shown that there is no cancer surveillance and immunoediting unless specific antigens were introduced into these cancers,²¹ in contrast to the robust cancer surveillance and immunoediting seen with MCA-induced sarcoma models.^{2,12,22} The basis for this difference could lie in the scarcity of mutations in genetic *vs.* carcinogen-induced models of cancer.¹²

Breast cancer has a large inflammatory component that typically promotes cancer cell growth and metastasis.²³⁻²⁶ Mouse models of breast cancer are abundant,²⁷ but perhaps the most studied model is the MMTV-PyMT (PyMT) model,²⁸ in which

the middle T antigen from the polyoma virus is expressed as an oncogene in mammary tissues to drive tumor formation. Studies using this model have come to differing conclusions on the role of immunity in tumor formation and progression. One group found that genetic depletion of the main immune components of the adaptive immune system performed by generating PyMT mice with a homozygous null mutation in *Rag1*, or mice more specifically lacking CD4⁺, CD8⁺, or B cells had no impact on primary tumor latency or progression, but were resistant to metastasis.²⁵ In contrast, recent studies have found that deficiency in IL-15 led to an increase in cancer development in PyMT animals, suggesting that innate lymphocytes could inhibit breast cancer formation in this model.²⁹

Earlier studies investigating the effect of immunity on PyMT tumor formation had been performed in the FVB/NJ strain background, which has a tumor latency 6 weeks shorter than the C57BL/6J strain.^{30,25} We sought to determine if this discrepancy was due to the extremely short tumor latency and rapid tumor progression observed in FVB/NJ PyMT mice. Indeed, we hypothesized that immunosurveillance would be insufficient to control the rapid and aggressive oncogene-driven tumor progression in the FVB/NJ background.

In the current study, we show that the adaptive immune system can delay cancer formation in PyMT mice. In addition, by generating a bank of cell lines with varying growth kinetics that depend on the presence of adaptive immune cells, we further demonstrate the occurrence of immunosurveillance and immunoediting in breast cancer. This study provides much-needed evidence that immunosurveillance and immunoediting of cancer cells can occur outside of carcinogen-induced cancer models and moreover suggest that breast cancer patients may benefit from cancer immune therapy.

Materials and methods

Experimental procedures

All experiments involving mice were conducted under an animal protocol approved by the University of California, San Diego Institutional Animal Care and Use Committee (IACUC protocol #S06201).

Mice

C57BL/6 (PyMT) MMTV-PyMT³⁰ and FVB/NJ (PyMT) MMTV-PyMT²⁸ mice were used in these studies. C57BL/6 PyMT mice were bred with recombinase active gene 1 (*Rag1*)^{-/-} mice to generate PyMT/*Rag1*^{-/-}. C57BL/6 WT, C57BL/6 *Rag1*^{-/-}, and *Rag2*^{-/-} x γ c^{-/-} mice were used for tumor transplantation experiments. All mice were bred in-house and therefore exposed to similar microbiota. In syngeneic model systems, tumor cell lines were transplanted into sex-matched animals. In spontaneous tumor formation experiments, tumor sites were palpated biweekly to monitor tumor latency and progression.

Tumor transplantation

Orthotopic engraftment of mammary tumor cell lines was performed by injections of 20 μ L of 10⁵ PyMT cells in PBS mixed

1:1 with matrigel (#E1270, Sigma) into the #4 and #9 fat pads of C57BL/6 WT, C57BL/6 *Rag1*^{-/-}, and *Rag2*^{-/-} x γ c^{-/-} mice anesthetized by a combination of ketamine (Ketaset, Zoetis) and xylazine (AnaSed, Lloyd) delivered intraperitoneally (i.p.). Tumor sites were palpated biweekly to determine the initiation of tumor formation.

Tacrolimus-induced immune suppression

13 mg biodegradable 90 d-releasable tacrolimus pellets or placebo-containing pellets (Innovative Research of America, FL, USA and Tacropellet, MD, USA) were implanted subcutaneously in the interscapular region of the back in C57BL/6 PyMT and FVB/NJ PyMT mice anesthetized by a combination of ketamine and xylazine, as described previously.³¹ C57BL/6 WT control animals were implanted with 13 mg biodegradable 90 d-releasable tacrolimus pellets or placebo-containing pellets.

Generation of PyMT cell lines

C57BL/6 PyMT and C57BL/6 PyMT/*Rag1*^{-/-} mammary tumor cell lines were established from spontaneous mammary tumors arising in C57BL/6 PyMT females as described previously.³² Tumors were minced and incubated at 37°C with 250 rpm agitation for at least an hour in 4 mL of Ham's F12K medium containing 1 mg/mL collagenase type II (#LS004174, Worthington Biochemicals, Lakewood, NJ, USA), 2 mg/mL soybean trypsin inhibitor (#T6522, Sigma-Aldrich, St. Louis, MO, USA), and 2% BSA (Sigma-Aldrich, St. Louis, MO, USA). Minced and digested tissues were sequentially re-suspended in red blood cell lysis solution (Sigma), then 0.25% trypsin/EDTA solution (Gibco) and dispase (#07923, Stem Cell technologies).

After multiple washes with a FCS-containing medium, the suspension was passed through a 70 μ m nylon filter (Fisher Scientific, Pittsburg, PA, USA) and the single cells were pelleted by centrifugation and cultured in Ham's F12K medium (Gibco) containing 5% FCS, 2.5 μ g/mL fungizone (#FG-70, Omega Scientific), 50 μ g/mL gentamycin (Gibco), and MITO+ (#355006, Corning). The cells were grown in flasks for several weeks. Typically, after the first week, a small population of immortalized cancer cells emerges. These are expanded and frozen. All cell lines were derived in an identical manner and are considered representative of subpopulations of tumor cells within the original tumor mass.

Histology and immunostaining

Tumors were fixed for 24 h in 10% neutral buffered formalin (BDH) and stored in 70% ethanol before embedding in paraffin. Sections were stained with H&E for pathological analysis and adjacent sections were evaluated by immunohistochemistry (IHC) with an anti-CD45 antibody (#ab10558, Abcam). For IHC, the slides were de-paraffinized and blocked for endogenous peroxidase and endogenous biotin and overlaid with the antibody at 5 μ g/mL in a humidified chamber overnight at 4°C. Slides were washed with tris buffered saline with 0.1% tween 20 (TBST) after every incubation. Separate slides received irrelevant IgG control. Bound antibody was detected using biotinylated anti-mouse antibody, followed by HRP

labeled Streptavidin and substrate color development used AEC (Amino-ethyl carbazole, Vector Labs Burlingame CA), following the manufacturer's recommendations. Nuclei were counter-stained using Mayer's hematoxylin and the slides were mounted in aqueous mounting media for visualization using an Olympus BH2 light microscope with digital photomicrography using Olympus Magnafire software. The number of CD45⁺ events per 40× high-power field were counted using a Leica DM2500 microscope.

Splenocyte isolation

Spleens were dissected from placebo or tacrolimus pellet implanted animals and homogenized by mechanical disruption. To obtain a single-cell suspension, the homogenized tissues were first subjected to red blood cell lysis, washed with PBS and then filtered through a 70 μ m nylon filter (Fisher Scientific, Pittsburgh, PA, USA).

Antibodies and flow cytometry

Splenocytes were vigorously re-suspended and washed in FACS buffer (PBS + 1% FCS + 0.05% NaN₃; Sigma-Aldrich). The following anti-mouse antibodies were used anti-: CD45 (#103116), CD4⁺ (GK1.5, #103116) (Biolegend) and CD8⁺ (53-6.7, 11-0081-85) (eBioscience). Cell surface staining was conducted for 15–20 min at 4°C in FACS tubes containing 1–2 × 10⁶ total cells, 0.1–0.2 μ g of antibody, 1 μ g of Fc blocking anti-CD16/32 antibody (for tumor transplant analysis), and 100 μ L of FACS buffer. 7AAD (EMD Millipore) was added at 1 μ g/mL immediately before FACS analysis.

For intracellular staining, stimulated and unstimulated splenocytes were incubated with or without 1 μ g/mL Brefeldin A (BFA, BD Biosciences) for 5 h and then prepared for intracellular FACS staining. Briefly, the cells were washed with PBS, incubated with Cytofix for 15 min at 4°C and washed twice with Perm/Wash solution (BD biosciences). Fixed cells were then incubated for 30 min in the dark with antibodies against the intracellular proteins IFN γ and IL-2 (eBioscience) diluted in the Perm/Wash solution. Cells were analyzed on a BD FACSCanto.

T cell stimulation

Splenocytes were cultured in 48-well plates pre-coated with 1 μ g/mL anti-CD3 (Clone: 17A2, Catalog#: BE0002, BioXcell) and anti-CD28 antibodies (Clone: 37.51 Catalog#: BE0015-1, BioXcell) for either 5 h with BFA or 3 d with 50 U/mL of IL-2 (#589102, Biolegend). Stimulated splenocytes were then processed for FACS staining.

Statistical analysis

Statistical significance between two groups at defined time points was determined by the Student's *t*-test using two-tailed analysis to obtain *p*-values. The Log-Rank (Mantel-Cox) test was used to compare the survival of mice across tumor transplantation or primary oncogenesis and * reflects the *p* values obtained in between groups at a given time point calculated by the Student's *t*-test. Error bars are depicted using the SEM, mean is represented as center values and **p* < 0.05, ***p* < 0.01, ****p* < 0.001. All experiments were done at least twice and representative data are shown. In primary transplant experiments, non-growing tumors in the *Rag2*^{-/-} × *γ c*^{-/-} environment were excluded, as the fat pad morphology in this immune background did not allow a 100% rate of success for the mammary fat pad injections in contrast to WT mice.

Results

Early immune suppression and adaptive immunodeficiency accelerates mammary tumor latency in C57BL/6 PyMT mice

To study the impact of acute immunosuppression on tumor latency, 6- or 11-week-old PyMT C57BL/6 mice were implanted with a 13 mg slow-release tacrolimus pellet in the interscapular region of the back (Figs. 1A and B).³¹ CD4⁺ and CD8⁺ T cells are activated by IL-2 produced by a calcium-dependent *Il-2* gene transcription that is blocked by tacrolimus.³³ We confirmed that the slow release of tacrolimus led to sustained immune suppression as CD8⁺ mediated-IL-2 production under basal conditions as well as after CD3/CD28 stimulation was blocked (Fig. S1A). Moreover, IL-2 production by CD4⁺ cells after CD3/CD28 stimulation (Fig. S1B) as well as

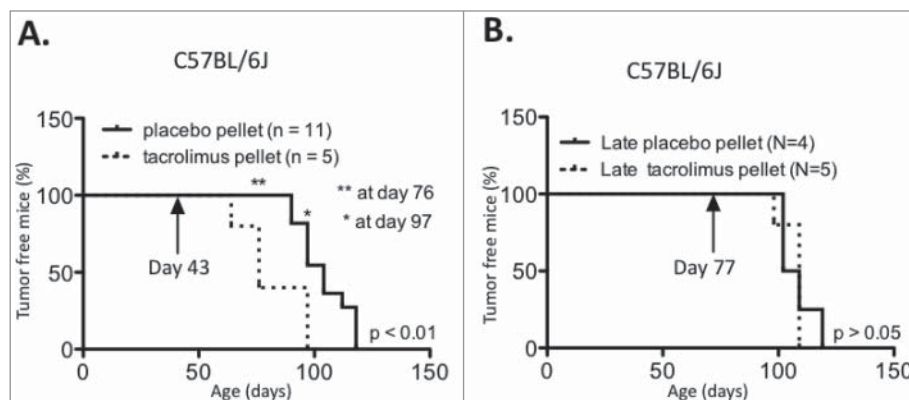


Figure 1. Tacrolimus immune suppression can impact PyMT tumor latency in the C56BL/6 background. PyMT C56BL/6 mice were implanted at age (A) 43 d or (B) 77 d with a 13 mg biodegradable 90 d-releasable tacrolimus or placebo pellet in the interscapular region of the back, and tumor onset was recorded over time.

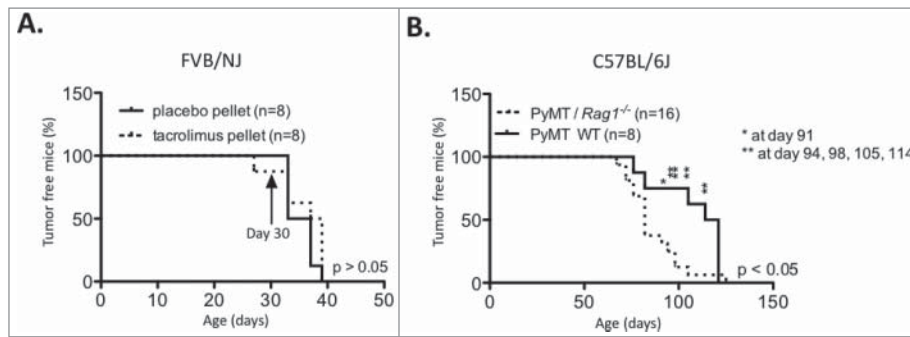


Figure 2. Impact of adaptive immune deficiency in the C57BL/6 PyMT background and strain specific tumor latency in the PyMT FVB/NJ background. (A) PyMT FVB/NJ mice were implanted at 30 d of age with a 13 mg biodegradable 90 d-releasable tacrolimus or placebo pellet in the interscapular region of the back. (B) Tumor onset was compared between C57BL/6 PyMT/*Rag1*^{-/-} and PyMT WT mice.

the number and percentages of CD4⁺ cells with or without CD3/CD28/IL-2 stimulation (Figs. S1F and H) was diminished. NK cells, however, were not impacted by tacrolimus-mediated immune suppression (data not shown).

Having confirmed that tacrolimus caused immune suppression, we compared tumor formation in cohorts of C57BL/6J PyMT mice implanted with tacrolimus *vs.* placebo pellets. The results showed that in mice implanted at 6 weeks, tumor latency was accelerated in the presence of tacrolimus, with an average tumor onset of 82 d *vs.* 104 d for mice implanted with the placebo (Fig. 1A, $p < 0.01$). In concordance, the number of tumor-bearing mice was significantly increased with tacrolimus treatment at days 76 and 97. However, when the pellet was inserted at 11 weeks of age, tacrolimus-mediated immune suppression had no impact on tumor latency (Fig. 1B).

We tested our hypothesis that strain background had a significant effect on the efficacy of immunosurveillance by implanting 13 mg biodegradable 90 d-releasable tacrolimus pellets at age 30 d in the interscapular region of the backs of FVB/NJ PyMT mice. Immune suppression had no impact on tumor latency in the FVB/NJ background (Fig. 2A), suggesting a strain-specific immunosurveillance of tumorigenesis that could

be attributed to the early and aggressive tumor progression that characterizes PyMT mice generated in the FVB/NJ background.

We next sought to confirm the tacrolimus results using a genetic approach similar to that used in previous studies.²⁵ Fig. 2B shows that PyMT/*Rag1*^{-/-} mice in the C57BL/6J background displayed a significantly earlier tumor onset than PyMT WT mice (86.5 *vs.* 107.62 d, $p < 0.05$). These data contradict previously published results obtained on the FVB/NJ background^{25,34} and provide further support for our hypothesis.

Immune-dependent growth kinetics of PyMT cell lines

We next hypothesized that the earlier cancer formation occurring in PyMT/*Rag1*^{-/-} mice resulted in unedited cancers. To test this hypothesis, 12 cell lines were generated from primary mammary tumors arising in either PyMT/*Rag1*^{-/-} hosts ($n = 6$) or PyMT WT hosts ($n = 6$) by producing a single-cell suspension from the tumor masses and cultivating the cells *in vitro* for a week (Fig. 3A). The immunogenicity of each cell line was determined by injecting 10⁵ cells into the mammary fat pads of severely immunodeficient *Rag2*^{-/-} x γ c^{-/-} or WT

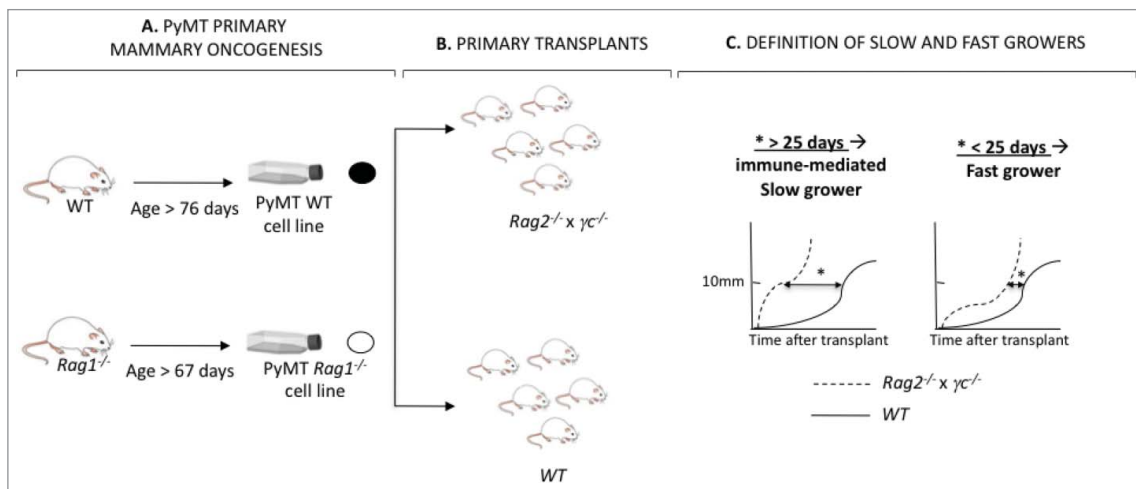


Figure 3. Generation of PyMT cell lines from *Rag1*^{-/-} and WT hosts. (A) PyMT mice in the *Rag1*^{-/-} or WT immune background developed mammary tumors over time. Tumors from each animal were harvested into a single-cell suspension and a cell line was generated. (B) Each cell line was transplanted into five *Rag2*^{-/-} x γ c^{-/-} and five WT mice. (C) The kinetic of each cell line in immune deficient *Rag2*^{-/-} x γ c^{-/-} *vs.* immune competent WT mice was compared by determining the difference in time to reach 10 × 10 mm in size in *Rag2*^{-/-} x γ c^{-/-} *vs.* WT mice. When this value (*) was inferior to 25 d, the cell lines were identified as immune-mediated slow growers. Cell lines for which this value was superior to 25 d, the cell lines were identified as fast growers.

Table 1. Characteristics of the PyMT cell lines generated from tumors arising in a WT or a *Rag1*^{-/-} background.

#	Genotype	PRIMARY ONCOGENESIS		PRIMARY TRANSPLANT		Time ¹ - Time ²	Growth phenotype
		Age onset (days)	Age at tumor harvest (days)	Average time: 10×10 mm in WT host ¹ (days)	Average time: 10×10 mm in <i>Rag2</i> ^{-/-} x <i>γc</i> ^{-/-} host ² (days)		
53	PyMT/ <i>Rag1</i> ^{-/-}	79	32	32	33	-1	FAST
29	PyMT/ <i>Rag1</i> ^{-/-}	69	40	40	35	5	FAST
31	PyMT/ <i>Rag1</i> ^{-/-}	97	62	62	56	6	FAST
32	PyMT/ <i>Rag1</i> ^{-/-}	97	47	47	26	21	FAST
55	PyMT/ <i>Rag1</i> ^{-/-}	91	90	90	50	40	SLOW
69	PyMT/ <i>Rag1</i> ^{-/-}	80	92	92	25	67	SLOW
252	PyMT WT	74	26	26	29	-3	FAST
234	PyMT WT	117	37	37	33	4	FAST
256	PyMT WT	115	74	74	74	0	FAST
235	PyMT WT	93	36	36	14	22	FAST
276	PyMT WT	78	96	96	59	37	SLOW
238	PyMT WT	113	108	108	53	55	SLOW

mice (Fig. 3B), as performed previously to define regressor and progressor cells in the MCA-sarcoma system.^{2,3}

The influence of the immune system on tumor growth was examined by comparing the growth of transplanted cell lines in severely immunodeficient *Rag2*^{-/-} x *γc*^{-/-} hosts vs. immunocompetent WT host (Fig. 3C). Classic regressor growth was not observed in this model system. Rather, we observed that some cell lines exhibited delayed growth in WT but not *Rag2*^{-/-} x *γc*^{-/-} hosts. To demonstrate the

growth delay caused by host immunity, we calculated the average time to reach a tumor size of 10×10 mm in WT vs. *Rag2*^{-/-} x *γc*^{-/-} for all the cell lines (Table 1: Time^{10×10(WT)} - Time^{10×10(Rag2^{-/-} x γc^{-/-})}). Based on this measure, cell lines were subsequently assigned either a slow grower (Time^{10×10(WT)} - Time^{10×10(Rag2^{-/-} x γc^{-/-})} < 25 d) or fast grower (Time^{10×10(WT)} - Time^{10×10(Rag2^{-/-} x γc^{-/-})} > 25 d) phenotype (Figs. 3C, 4A and B, Table 1). Surprisingly, the number of slow growers and fast growers were almost

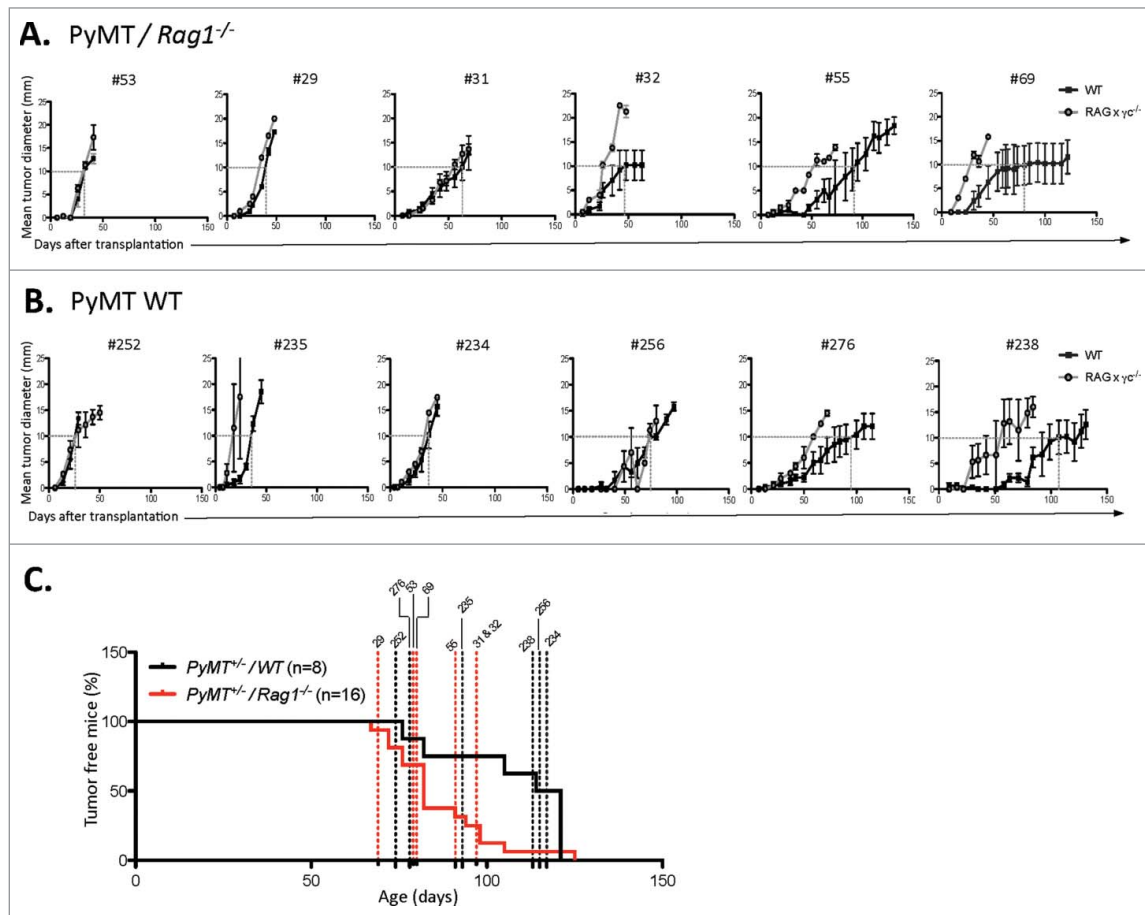


Figure 4. Immune-dependent growth kinetics of PyMT cell lines. PyMT cell lines were generated from primary tumors from PyMT mice (A) *Rag1*^{-/-} or (B) WT mice. Each cell line was subsequently passaged in either severely immune deficient *Rag2*^{-/-} x *γc*^{-/-} (open circle) or WT mice (closed square). For each cell line, a dotted line shows the age at which tumor size reached a 10×10 mm mean tumor diameter. (C) The primary tumor latency of each cell lines was reported on the graph showing the primary tumor onset of PyMT/*Rag1*^{-/-} mice vs. PyMT WT mice.

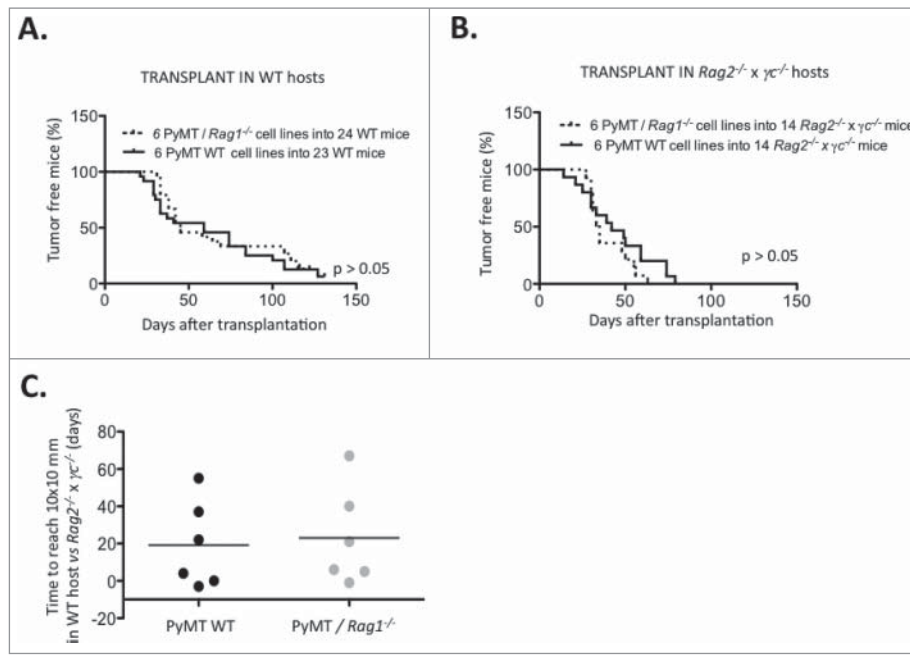


Figure 5. Immune-dependent growth kinetics of PyMT cell lines. Using a cut-off of 10×10 mm mean tumor diameter, survival of (A) 24 WT mice and (B) 14 $Rag2^{-/-} \times \gamma C^{-/-}$ mice transplanted with either six PyMT/ $Rag1^{-/-}$ cell lines or six PyMT WT cell lines was analyzed. (C) The time difference to reach 10×10 mm in WT vs. $Rag2^{-/-} \times \gamma C^{-/-}$ hosts is plotted.

identical for both genotypes from which the cell lines originated. Four out of the six PyMT/ $Rag1^{-/-}$ cell lines (#53, #29, #31, #32) and PyMT WT cell lines (#252, #234, #256, #235) showed a fast grower phenotype.

Two cell lines from PyMT/ $Rag1^{-/-}$ hosts (#69 and #55) and two cell lines from PyMT WT hosts (#276 and #238) displayed a slow grower phenotype in the presence of complete immunity, taking more than 90 d to form tumors in WT hosts. No correlations were found regarding age of tumor onset, age of tumor harvest, and tumor burden at end point between slow and fast growers (Fig. 4C, Table 1).

In accordance with the identical number of slow and fast growers in each of the genotypes tested, no significant differences were found when comparing the genotype of the cell lines for their survival after primary transplants in immunocompetent (Fig. 5A) vs. immunodeficient hosts (Fig. 5B). Similarly, there were no significant differences in duration to reach 10×10 mm of diameter in WT vs. $Rag2^{-/-} \times \gamma C^{-/-}$ hosts (Fig. 5C).

Nevertheless, several cell lines (#55, #69, #238, #276) displayed a clear delay in growth after transplantation into WT vs. immune deficient $Rag2^{-/-} \times \gamma C^{-/-}$. These cell lines were defined as immunogenic and selected for further study.

Slow growers are characterized by increased immune cell infiltration and a more differentiated tumor histology

To determine the impact of infiltrating immune cells on the immunogenicity of cancer cell lines subsequently derived from a primary tumor, sections from primary PyMT tumors (Fig. 3A) were analyzed for their CD45 immune infiltration content using IHC. Primary tumors from which slow growing cell lines (#69, #238, #55) were derived showed massive

CD45⁺ cell infiltration within the tumor mass. In contrast, primary tumors from which fast growers (#252, #234, #29) were derived showed the presence of CD45⁺ cells confined to the periphery of the tumor tissue (Fig. 6A). Quantitative analysis of immune cell infiltration within the H&E positive tumor tissues showed a significant increase in immune cell infiltration within the primary slow grower tumor mass (Fig. 6B). Interestingly, the number of CD45⁺ infiltrating cells correlated with the genotype of the mouse from which the tumor originated: PyMT/ $Rag1^{-/-}$ were more infiltrated with immune cells compared with PyMT WT (Fig. 6C). Histological analysis of primary tumors for dedifferentiation, necrosis, and cystic changes revealed a significant increase in cystic changes in slow growers, indicative of tissue differentiation (Fig. 6D).³⁵ The amount of necrosis and the area of poorly differentiated tissue were not significantly altered between slow growers and fast growers (Fig. S2).

Immune pressure in WT hosts converts slow grower tumors into fast growers—evidence of immunoediting in the PyMT model

To determine if cell lines were edited during their growth in WT recipients, passaged cell lines were generated after transplantation of the primary cell lines and tested for immunogenicity (Figs. 7A and B). Cell lines #69 (slow grower) and #252 (fast grower) (Fig. 7A) were transplanted into either WT or $Rag2^{-/-} \times \gamma C^{-/-}$ hosts. Cell lines generated from the resulting tumor masses (Fig. 7B) were then re-transplanted into syngeneic WT or $Rag2^{-/-} \times \gamma C^{-/-}$ hosts (Fig. 7C). The tumorigenicity of the WT- or $Rag2^{-/-} \times \gamma C^{-/-}$ -passaged-cell lines after secondary transplantations into immune deficient or WT hosts is shown in Fig. 8. Notably, the slow growing cell line #69,

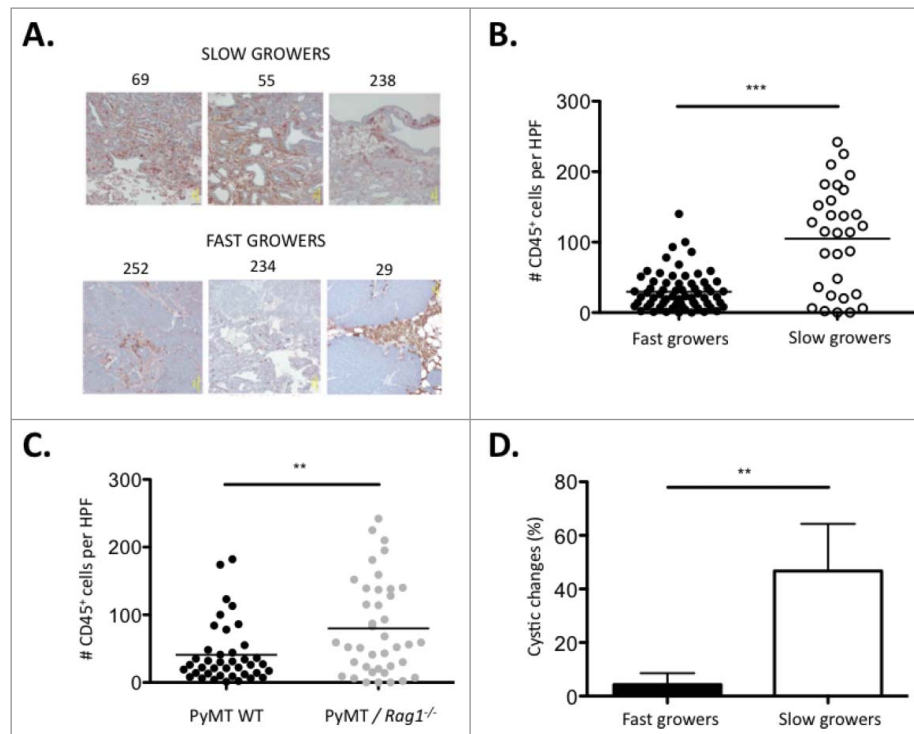


Figure 6. Slow growing tumors are characterized by increased immune cell infiltration and a more differentiated tumor histology. (A) Representative micrographs of primary tumors from three immune-mediated slow growers (#69, #55, #238) and fast growers (#252, #234, #29) sectioned and stained with H&E and anti-CD45 antibody. (B) Number of tumor-infiltrating CD45⁺ cells per high-power field (HPF) in primary tumors from three immune-mediated slow growers (#69, #55, #238) vs. five fast growers (#252, #234, #29, #235, #256). Ten HPF were counted per tumor sample. (C) Number of tumor infiltrating CD45⁺ cells per HPF in primary tumors from four PyMT/*Rag1*^{-/-} (#69, #55, #29, #31) vs. four PyMT WT (#252, #234, #256, #238) cell lines. Ten HPF were counted per tumor sample. The percent of cystic changes as a morphological indication of tumor differentiation was examined on H&E stained sections in the primary tumor of three immune-mediated slow growers (#69, #55, #238) vs. five fast growers (#252, #234, #29, #235, #256).

when passaged through WT hosts, was converted from a slow grower to a fast grower (Figs. 7B, C and 8B), whereas passaging through severely immunodeficient *Rag2*^{-/-} x *γc*^{-/-} hosts did not change the slow growing profile (Figs. 7B, C and 8A). Indeed, the passaged cell line failed to grow in three out of four WT mice transplanted (Fig. 8A). Fig. 8C shows the combined survival curves of the secondary-transplanted WT- and *Rag2*^{-/-} x *γc*^{-/-}-passaged #69 cell lines in WT recipients, demonstrating a significant difference ($p < 0.01$) in tumorigenicity of cell lines passaged through WT vs *Rag2*^{-/-} x *γc*^{-/-} mice. In contrast, the growth kinetics of the fast grower #252 cell line in secondary transplants remained unchanged after passage in WT hosts compared its *Rag2*^{-/-} x *γc*^{-/-}-passaged counterpart (Figs. 7C, 8E and F).

The difference in tumorigenicity between the secondary *Rag2*^{-/-} x *γc*^{-/-}-passaged vs. the WT-passaged #69 cell line could be due to intrinsic differences in growth rate. To eliminate this possibility, the growth of the primary-passaged #69 cell lines was compared *in vitro* and after transplantation into immune deficient animals. Fig. S3 shows that the passaged cell lines had equivalent growth *in vitro* and Figs. 8D and 8G show that the passaged cell lines also had equivalent growth in immune deficient *Rag1*^{-/-} and *Rag2*^{-/-} x *γc*^{-/-} mice. Interestingly, despite having identical growth kinetics, there was still a heavy infiltration of CD45⁺ cells in the *Rag2*^{-/-} x *γc*^{-/-}-passaged vs. the WT-passaged #69 cell line after secondary transplant into *Rag1*^{-/-} mice (Fig. 8H).

Discussion

In this study, we show that adaptive immunosurveillance occurs in the oncogene-induced PyMT C57BL/6 mammary carcinoma model, where genetic ablation of adaptive immune responses and early acute immune suppression with tacrolimus accelerated tumor latency. We also generated a bank of PyMT tumor cell lines, with differing degrees of growth capacity in immune competent vs. immune deficient hosts. Using this novel set of cell lines, we show that immune infiltration into the primary tumor mass correlated with tumor cystic changes and could impact the immunogenicity of cell lines subsequently generated from the tumor. Overall, these results extend the cancer immunoeediting paradigm to breast cancer and an oncogene-induced model system, while providing a useful set of reagents to model immune cell infiltration into breast cancer.

Our results are consistent with recent reports demonstrating that immunosurveillance of PyMT tumors was mediated in part by tissue resident innate lymphocytes that depended on IL-15 and perforin for their development and activity.²⁹ Among these novel actors of immunosurveillance, a population of T cell receptor-expressing innate lymphoid cells, termed ILTC1, was identified. Since all T cells require RAG to develop, our mice also lacked this population of cells; however, it is not known whether the lack of immunosurveillance we observed is due to lack of ILTC1 cells or another cell type that requires

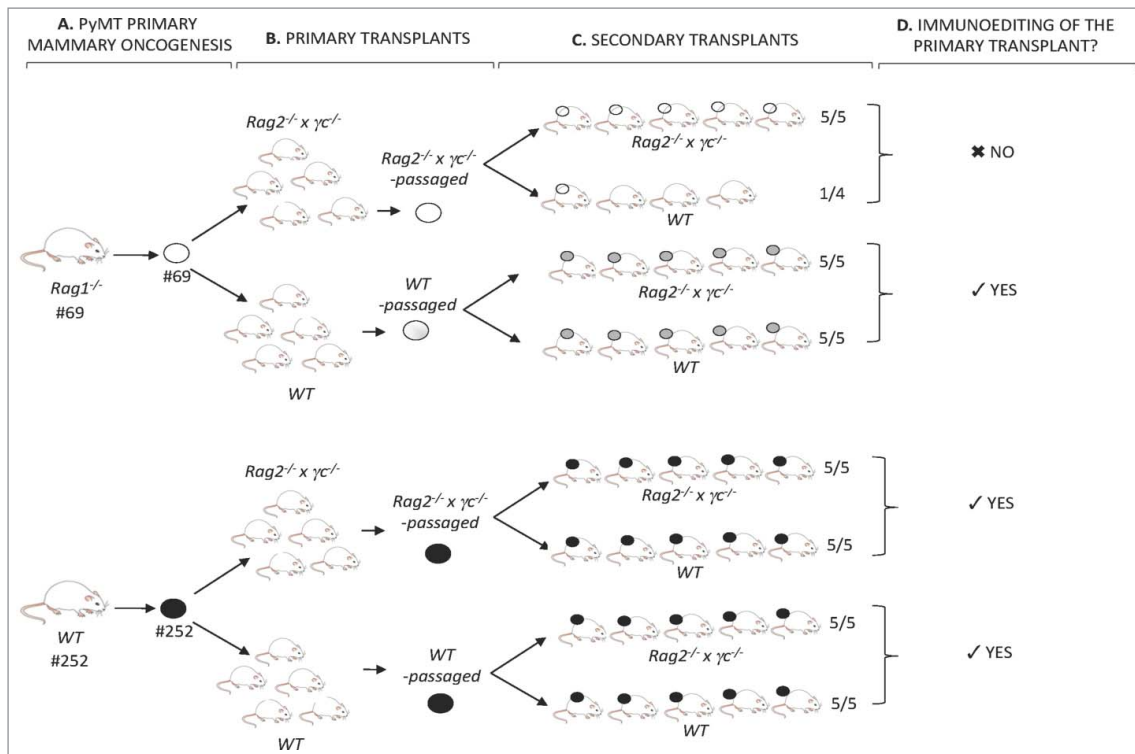


Figure 7. Identification of immunoediting in the PyMT model. (A) PyMT mice #69 and #252, respectively, of the *Rag1*^{-/-} or WT immune background, developed mammary tumors over time. Tumors from each animal were harvested into a single-cell suspension and a cell line was generated. (B) Each cell line was transplanted into five *Rag2*^{-/-} *x* *γc*^{-/-} and five WT mice. Tumors were harvested into a single-cell suspension and *Rag2*^{-/-} *x* *γc*^{-/-}-passaged and WT-passaged cell lines of both #69 and #252 primary cell lines were generated. (C) *Rag2*^{-/-} *x* *γc*^{-/-} and WT-passaged primary transplants of #69 and #252 were transplanted as secondary transplants in either *Rag2*^{-/-} *x* *γc*^{-/-} or WT mice. (D) Secondary transplant kinetics in immune deficient *Rag2*^{-/-} *x* *γc*^{-/-} and immune competent WT mice revealed the occurrence of immunoediting of the primary-passaged transplant.

RAG for its development. Our possession of cell lines derived from these mice will allow us to determine whether the slow growing cell lines are enriched in genes that stimulate ILTC1 cells or other cell types that could mediate cancer immunoediting.

We found that PyMT/*Rag1*^{-/-} mice had significantly reduced tumor latency compared with PyMT WT mice in the C57BL/6 background, a result that was not seen when similar animals were studied in the FVB/NJ background.²⁵ This strain specificity is likely to be due to the rapid oncogenesis occurring in the FVB/NJ strain where the oncogene-driven rapid primary tumor progression does not allow for an environmental immune cell influence. Alternatively, it is possible that the Th1 tendencies in C57BL/6 animals^{36,37} promotes better antitumor immunity than the Th2 bias in FVB/NJ animals.^{38,39} Other groups have also observed a strain specificity in tumor progression in this animal model. In studying the role of nitric oxide in PyMT tumor progression, Davie *et al.* found that PyMT/*iNOS*^{-/-} mice had delayed tumor formation in the C57BL/6 but not in the FVB/NJ background.³⁰

Genetic models of oncogenesis have been used extensively to document immune-mediated tumor promotion in solid cancers.^{25,30,40-42} However, evidence of immunosurveillance and immunoediting in genetic models of “solid,” non-lymphoid, oncogenesis have been sparse and have yielded conflicting results.^{34,43} For example, in the *p53*^{-/+} model of oncogenesis, knockout of *Ifngr1* accelerated tumor onset and the spectrum of tumors of non-lymphoid origins.³⁴ In the TRAMP model, deficiencies in NKG2D or TCRδ resulted in higher incidence,

higher grade, or extensive prostate carcinomas.^{17,44} Conversely, deficiencies in TRAIL and TRAIL receptor DR5 did not affect tumor onset in colon adenocarcinoma and mammary tumors.^{45,46} In genetic models of sarcoma and lung cancer, immunosurveillance and immunoediting were shown to be mediated through the lentiviral-enforced expression of exogenous antigens by the primary tumor, therefore positing the expression of tumor-specific antigen as key for the occurrence of immunoediting.^{21,47} In addition, naturally occurring immunosurveillance against a self-antigen was documented in a genetic model of prostate cancer.²⁰ Relevant to breast cancer, using the Her2/Neu model of murine breast cancer Street *et al.* showed that perforin could diminish mammary cancer progression but had no effect on incidence.⁴⁸ *Stat1*^{-/-} mice also had a higher incidence of breast cancer, although the effect was attributed to a role for STAT1 as a cell intrinsic tumor suppressor rather than an immune deficiency.⁴⁹ Our results provide evidence that adaptive immunity is required for optimal breast cancer surveillance.

In addition, we show here that innate immune cells, found in *Rag1*^{-/-} mice, could be recruited to slow growing tumors and correlated with the cystic changes in the tumor histology, as reported in an oncogene-induced model of carcinogenesis in a perforin-deficient model.⁵⁰ We speculate that the ability to recruit innate immune cells is a pre-requisite to create an environment that will in turn recruit adaptive immune cells. Additionally, this increase in immune cell infiltration in *Rag1*^{-/-} primary and transplant recipients suggests that this infiltration

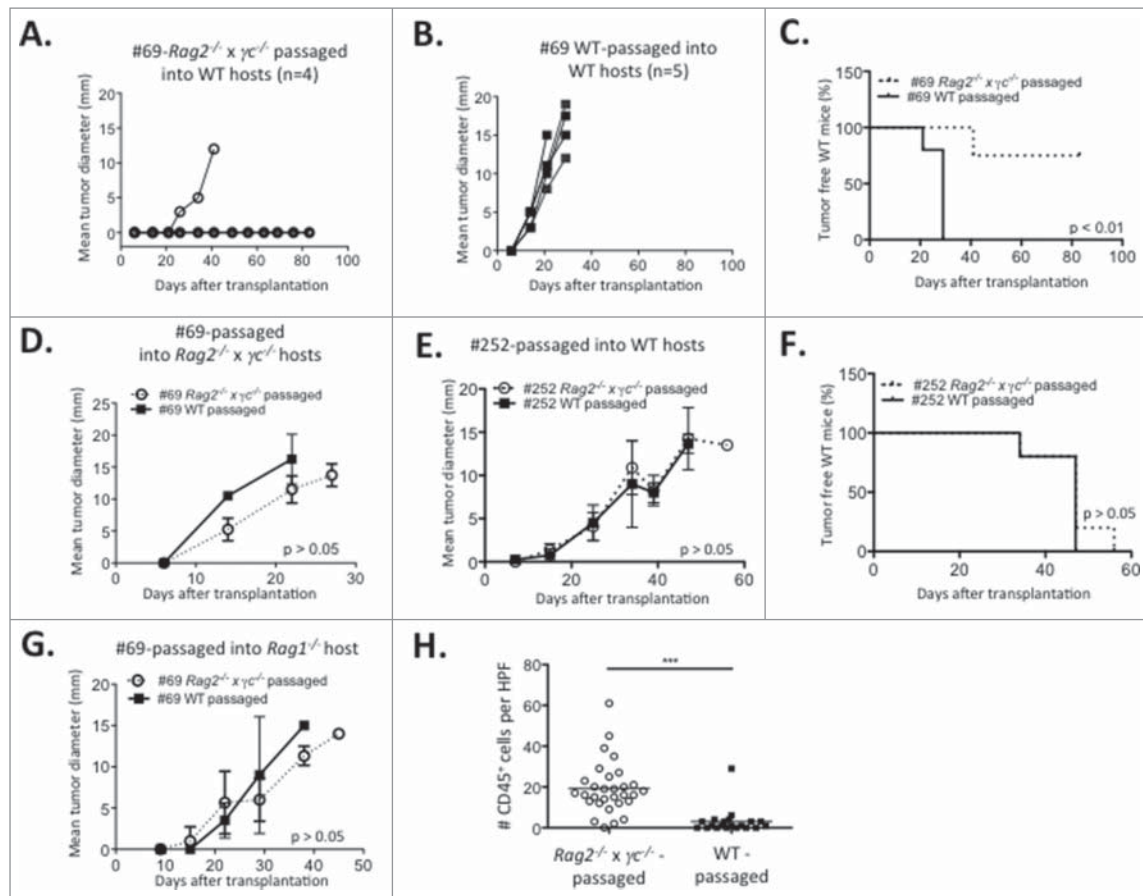


Figure 8. Immunoeediting in the PyMT model. Tumor progression over time of (A) *Rag2*^{-/-} x *γc*^{-/-} and (B) WT- passaged slow growing cell line #69 transplanted in WT recipients. *Rag2*^{-/-} x *γc*^{-/-} or WT-passaged slow growing cell line #69 were transplanted into (C) WT or (D) *Rag2*^{-/-} x *γc*^{-/-} recipients and (C) survival or (D) tumor progression was assessed. (E) Tumor progression of the *Rag2*^{-/-} x *γc*^{-/-} and WT-passaged cell lines originating from the fast grower #252 in WT recipients. (F) Survival of WT mice transplanted with *Rag2*^{-/-} x *γc*^{-/-} and WT-passaged cell lines originating from the fast grower #252. (G) Tumor progression of *Rag1*^{-/-} mice transplanted with either *Rag2*^{-/-} x *γc*^{-/-} or WT-passaged slow growing cell line #69. (H) Number of tumor-infiltrating CD45⁺ cells per high-power field (HPF) in the *Rag2*^{-/-} x *γc*^{-/-} and WT-passaged #69 cell lines secondarily transplanted in *Rag1*^{-/-} host. Experiments were repeated twice.

is independent of the antigenicity of the tumor, as adaptive immune cells are absent. Whether this increased immune recruitment is due to a chemokine or cytokine secreted by immunogenic tumor cells³ is not known but will be studied in the future. Importantly, identification of these key molecular mediators of innate cell recruitment, as we have done previously, may open new avenues for cancer therapy of tumors with a low level of antigenicity such as breast cancer.^{3,51}

Disclosure of potential conflicts of interest

No potential conflicts of interest were disclosed.

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