

Role of the Cytokine-like Hormone Leptin in Muscle-bone Crosstalk with Aging

Mark W. Hamrick

Department of Cellular Biology and Anatomy, Medical College of Georgia, Augusta University, Augusta, GA, USA

Corresponding author

Mark W. Hamrick
Department of Cellular Biology and Anatomy,
Medical College of Georgia, Augusta
University, Laney Walker Blvd. CB2915,
Augusta, GA 30912, USA
Tel: +1-706-721-1958
Fax: +1-706-721-6120
E-mail: mhamrick@augusta.edu

Received: December 9, 2016

Accepted: December 20, 2016

No potential conflict of interest relevant to this article was reported.

The cytokine-like hormone leptin is a classic adipokine that is secreted by adipocytes, increases with weight gain, and decreases with weight loss. Additional studies have, however, shown that leptin is also produced by skeletal muscle, and leptin receptors are abundant in both skeletal muscle and bone-derived mesenchymal (stromal) stem cells. These findings suggest that leptin may play an important role in muscle-bone crosstalk. Leptin treatment *in vitro* increases the expression of myogenic genes in primary myoblasts, and leptin treatment *in vivo* increases the expression of microRNAs involved in myogenesis. Bone marrow adipogenesis is associated with low bone mass in humans and rodents, and leptin can reduce marrow adipogenesis centrally through its receptors in the hypothalamus as well as directly via its receptors in bone marrow stem cells. Yet, central leptin resistance can increase with age, and low circulating levels of leptin have been observed among the frail elderly. Thus, aging appears to significantly alter leptin-mediated crosstalk among various organs and tissues. Aging is associated with bone loss and muscle atrophy, contributing to frailty, postural instability, and the incidence of falls. Therapeutic interventions such as protein and amino acid supplementation that can increase muscle mass and muscle-derived leptin may have multiple benefits for the elderly that can potentially reduce the incidence of falls and fractures.

Key Words: Insulin-like growth factor I, Leptin, Mesenchymal stromal cells, Osteoporosis, Sarcopenia

INTRODUCTION

The incidence of debilitating bone fractures increases with age, and as the aging population expands in nations around the world the public health burden of these age-related bone fractures continues to grow. The risk of hip fracture doubles every 5 to 6 years after age 60, underscoring the dramatic increase in fracture risk that accompanies older age.[1] Older individuals who are at greatest risk of fracture frequently show a spectrum of features broadly categorized as “frailty”, defined by unintentional weight loss, self-reported exhaustion, muscle weakness, slow walking speed, and low physical activity.[2] The functional and behavioral aspects of frailty such as muscle weakness and reduced walking speed are, not surprisingly, also associated with loss of lean mass in the form of sarcopenia.[3] The muscle weakness and reduced muscle mass that characterize sarcopenia are associated with an increased risk of falling, which is in turn a primary cause of bone frac-

Copyright © 2017 The Korean Society for Bone and Mineral Research

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

tures.[4] Thus, while middle-age is often associated with increased body weight in many developed countries, later life is frequently characterized by loss of body mass, loss muscle mass, bone loss, and ultimately bone fractures.

The constellation of factors that characterizes the frailty phenotype suggests that some of these features may be physiologically related through endocrine or paracrine pathways. The integrative physiology of exercise provides a very clear example of the extensive crosstalk that can exist among different organs and tissues.[5] For example, muscle is now recognized as a source of circulating myokines that can be impacted by resistance training and physical activity,[6,7] fat is a source of secreted adipokines that can be modulated by weight loss and may effect a number of different organs and tissues,[8] and bone is now acknowledged to produce a circulating osteokine osteocalcin that can alter insulin sensitivity and exercise adaptation.[9] It is also becoming better appreciated that some of these pathways are significantly altered with aging, modifying the network for tissue crosstalk that exists in younger individuals.[10]

The cytokine-like hormone leptin is a classic adipokine that is secreted by adipocytes, increases with weight gain, and decreases with weight loss.[8] Additional studies have, however, shown that leptin is also produced by skeletal muscle [11-14] as well as bone cells,[15] and leptin receptors are abundant in these musculoskeletal tissues.[16,17] The leptin receptor is highly conserved in vertebrates, and leptin functions as a growth factor for both muscle and bone early in life.[18] In addition, altered leptin signaling may play a key role in the aging process, as leptin resistance is known to occur in the brain of aging rodents.[19] Leptin levels are positively associated with longevity in centenarians,[20] and the ratio of leptin to adiponectin is positively correlated with muscle strength in older adults.[21] In addition, higher leptin levels are associated with a reduced risk of dementia in elderly people.[22,23] Leptin signaling and changes in leptin sensitivity with age are therefore likely to contribute to age-related degeneration of multiple organs and tissues. This review summarizes the evidence for leptin's role in the maintenance of muscle and bone mass with aging, and suggests future directions for research aimed at defining the basic mechanisms of organ crosstalk linking age-related changes in musculoskeletal tissues.

THE ANABOLIC EFFECTS OF LEPTIN ON SKELETAL MUSCLE

Leptin levels normalized for total protein are actually higher in mouse skeletal muscle than in mouse adipose tissue (Fig. 1A).[24] Muscle can accumulate fat, but fat does not accumulate substantially in normal mouse skeletal muscle, suggesting that the high concentrations of leptin in muscle are derived from muscle cells themselves.[25] This is further indicated by the observation that myoblasts secrete leptin *in vitro*, [12] and that leptin is released from skeletal muscle *in vivo*. [13,14] Importantly, leptin release from skeletal muscle is only slightly less than release from adipose tissue (per unit tissue mass), yet muscle comprises a much greater percentage of body composition than fat, revealing that muscle is an important source of circulating leptin.[14] As noted above, leptin receptors are abundant in skeletal muscle (Fig. 1B), [17] their expression in skeletal muscle is altered with changes in physical activity, [17,26] and the absence of functional leptin receptors in skeletal muscle impairs the capacity for myoblast proliferation and differentiation.[16] On the other hand, treatment of isolated primary myoblasts with leptin increases proliferation and the expression of myogenic genes (Fig. 1C). [16] These findings point to an autocrine function of muscle-derived leptin, but circulating leptin is also likely to have important anabolic effects on skeletal muscle. Leptin treatment increases muscle mass and decreases the expression of atrophy-related factors such as myostatin, muscle RING-finger protein-1 (MuRF1), and muscle atrophy F-box (MAFbx) in muscle (Fig. 1C). [16,27] Circulating and muscle-derived insulin-like growth factor 1 (IGF-1) are both increased with leptin treatment in aged mice as well as in leptin-deficient mice, [24,28] indicating that many of the anabolic effects of leptin on skeletal muscle are likely to be mediated by IGF-1. Finally, recent data suggest that leptin may also stimulate follistatin production in a circadian manner, and follistatin is a potent antagonist of myostatin.[29]

We have not detected a significant change in muscle-derived leptin with age (Fig. 1D), nor have we identified any changes in leptin receptor expression in skeletal muscle with age.[16] These observations are consistent with the finding that exogenous leptin can increase the expression of myogenic genes in primary myoblasts from aged mice,[16] and leptin treatment can increase muscle mass in aged mice and alter the expression of myogenic microR-

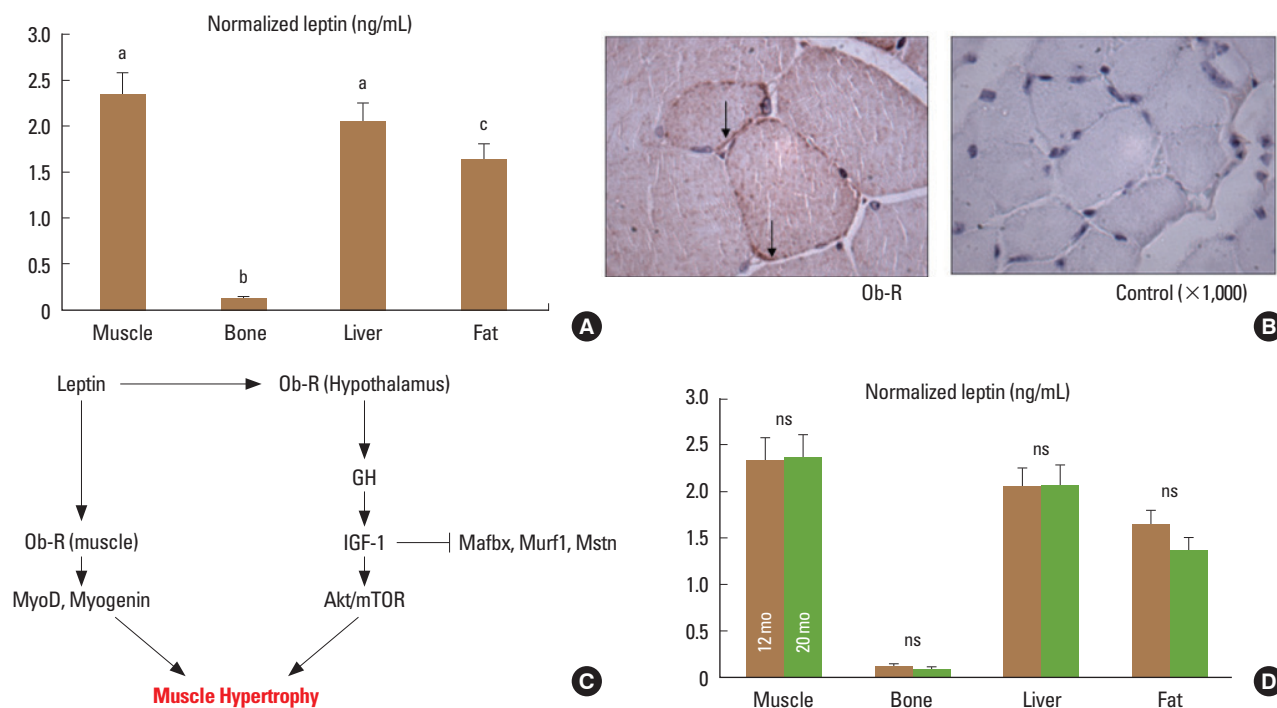


Fig. 1. (A) Leptin levels measured in homogenized mouse tissue normalized by total protein. Means with different superscripts differ significantly from one another ($P < 0.05$). (B) Immunostaining for the long form of the leptin receptor (Ob-R) in cross-sections of mouse skeletal muscle, showing positive staining relative to control samples (no primary antibody). (C) Peripheral, direct (muscle) and central (hypothalamic) pathways through which leptin alters pathways regulating skeletal muscle hypertrophy. (D) Leptin levels measured in homogenized mouse tissue normalized by total protein in young adult (12 months) and aged (12 months) mouse samples. ns, not statistically significant ($P < 0.05$) for age-related differences; GH, growth hormone; IGF, insulin-like growth factor; mTOR, mechanistic target of rapamycin.

NAs such as miR-31 and miR-223.[30] In addition, leptin treatment produces a marked decrease in miR-489 expression in aged mice,[30] and miR-489 is known to maintain muscle satellite cells in a quiescent state.[31] Thus, leptin-induced suppression of miR-489 activity would be expected to enhance the capacity for muscle regeneration and repair in older animals. Although bone-derived mesenchymal stem cells appear to lose their responsiveness to leptin with age (see below), this does not seem to be the case in skeletal muscle. Age-related differences in the response of muscle and bone progenitor cells to leptin with age may also result from intrinsic differences in the progenitor cells themselves, as bone-derived stem cells appear to exhibit greater impairments with age compared to stem cells isolated from muscle or adipose tissue.[32]

LEPTIN AND AGE-RELATED CHANGES IN BONE MARROW CELL POPULATIONS

The central effects of leptin on bone, mediated by hypo-

thalamic leptin receptors and the beta-adrenergic signaling network, were initially thought to suppress bone formation producing a low bone mass phenotype.[33] More recent studies show that the effects of leptin on the skeleton are quite complex, and that leptin deficiency is associated with low bone mass primarily due to reduced cortical bone.[34-36] Central infusions of leptin in leptin-deficient ob/ob mice actually increase cortical bone formation and total bone mass,[28] but leptin also has important peripheral, direct effects on osteoblasts and bone-derived mesenchymal stem (stromal) cells (BMSCs). In fact, the leptin receptor is now regarded a marker of BMSCs[37] that mediates the switch between osteogenesis and adipogenesis.[38,39] Leptin receptors are also abundant in the periosteum surrounding cortical bone (Fig. 2A). Leptin treatment of leptin-sensitive BMSCs increases the expression of bone morphogenetic protein 2 (BMP-2) as well as the secretion of the chemokine stromal cell-derived factor 1 (SDF-1) (CXCL12).[40] Other studies indicate that replacement of bone marrow cell populations with stem cells lacking the leptin re-

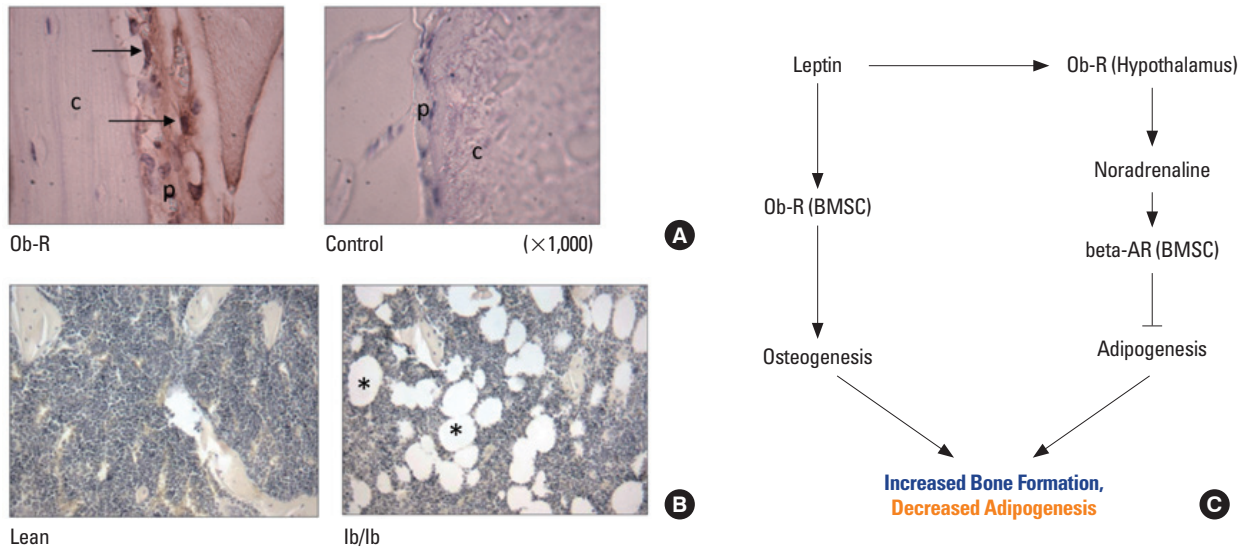


Fig. 2. (A) Immunostaining for the long form of the leptin receptor (Ob-R) in cross-sections of mouse cortical bone (c), showing positive staining in periosteal cells (p) relative to control samples (no primary antibody). (B) Increased number of bone marrow adipocytes (asterisks) in bone cross-sections from POUND mice lacking both forms of the leptin receptor (lb/lb) compared to marrow from normal lean mice. (C) Peripheral, direct (bone-derived mesenchymal stromal cells [BMSC]) and central (hypothalamic) pathways through which leptin alters pathways regulating osteogenesis and adipogenesis in bone marrow. beta-AR, beta-adrenergic receptor.

ceptor impairs bone formation, whereas restoring direct leptin signaling by transplanting cells expressing the leptin receptor enhances bone formation.[41] Moreover, as noted above, leptin can stimulate the growth hormone (GH)/IGF-1 axis, and IGF-1 is a potent osteogenic factor. Thus, leptin can effectively couple food intake and energy reserves with bone formation through both central and peripheral (direct) actions.

Aging appears to impact leptin's effects on bone metabolism in several ways. First, central leptin resistance, which is known to increase with aging,[19] is likely to attenuate leptin's central effects mediated by the hypothalamus. This may have a particularly important influence on bone marrow adipogenesis and the accumulation of bone marrow fat with age. Intrahypothalamic injection of leptin reduces marrow adipocytes in both mice[42] and rats[43], which appears to be mediated by beta-adrenergic signaling.[42,44] Moreover, leptin deficiency due to either absence of leptin or calorie restriction increases marrow adipocytes (Fig. 2B).[45] These findings suggest that reduced hypothalamic sensitivity to leptin would be expected to support an overall increase in marrow adipocytes with aging (Fig. 2C). An increase in marrow fat with age-associated leptin resistance would also be consistent with the well-known phenomenon of increased marrow fat seen with advanced age in humans.

Second, evidence indicates that aging has a significant effect on BMSCs and their response to leptin. Individuals with osteoporosis have reduced levels of leptin in the bone marrow microenvironment,[46] BMSCs isolated from osteoporotic donors show lower leptin binding capacity than BMSCs from normal donors,[47] and leptin can suppress the adipogenic differentiation of BMSCs from healthy donors but not in BMSCs from osteoporotic donors.[48] The study by Astudillo and colleagues[47] suggests that leptin insensitivity due to decreased leptin binding occurs with age, but the molecular mechanisms underlying this phenomenon are not well understood. It is also unclear how leptin receptor expression, or the expression of microRNAs targeting the leptin receptor, are altered with age in BMSCs. BMSCs isolated from older individuals are observed to have impaired proliferation, increased senescence, and reduced potential for osteoblastic differentiation.[32,49-51] It is certainly possible that some of these age-associated changes in BMSC function documented in older donors may be associated with alterations in factors associated with leptin signaling, such as microRNAs targeting the leptin receptor or molecules such as suppressor of cytokine signaling 3 (SOCS3) and protein tyrosine phosphatase 1B (PT-1B) that inhibit leptin signal transduction.[52,53]

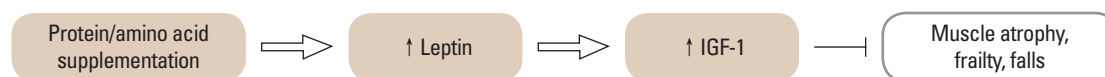


Fig. 3. Proposed interactions among dietary protein, muscle-derived leptin, insulin-like growth factor 1 (IGF-1) and muscle mass. Protein and amino acid supplementation is suggested to increase muscle-derived leptin and IGF-1, increasing muscle mass and strength and reducing the risk of falls and fractures.

DISCUSSION

Leptin secretion links food intake and energy reserves with energy expenditure, growth, and reproduction.[54] The role of altered leptin signaling in aging has, however, received less attention. Aging is associated with overall reduced calorie intake as well as reduced protein intake,[55-57] which would be expected to lower circulating levels of leptin. Not surprisingly malnutrition in the elderly is in many cases associated with reduced leptin,[58,59] as well as with reduced subcutaneous fat and reduced lean mass.[60] Many of the physical changes observed with older age such as reduced lean mass, bone loss, and cognitive decline may be related. For example, frailty and muscle weakness are associated with dementia,[61,62] and aging and leptin resistance have been linked with the development of Alzheimer's disease.[63] These observations then raise the question of what interventions might increase leptin production and lean mass in older patients. We have found that the amino acid tryptophan increases muscle-derived leptin, IGF-1, and follistatin in mice on a low protein diet,[64] and others have found that amino acid supplements plus exercise increase both serum leptin and musculoskeletal function in older adults.[65] These studies suggest that dietary interventions may provide one approach for reducing loss of lean mass with aging,[66] perhaps through a leptin-mediated pathway (Fig. 3). Recombinant human leptin (rhLep) has shown potential for increasing IGF-1 and markers of bone formation in women with hypothalamic amenorrhea,[67] although other studies indicate that the effects of rhLep on lean mass may be more modest.[68]

Resistance exercise can improve muscle strength and power in the elderly[69,70] whereas bone is much less responsive to mechanical stimuli with age.[10] This may indicate that targeting muscle through nutritional supplementation (Fig. 3) as well as with resistance exercise is likely to have a greater impact on reducing the risk of falls and fractures than targeting bone itself; however, given that leptin can mediate the differentiation of bone marrow stromal

cells directly and reduce bone marrow adipogenesis via its receptors in the hypothalamus, increasing leptin levels with increasing muscle mass may have some positive effects on the skeleton. These changes, in addition to the fact that leptin may prevent some neurodegenerative decline with aging, suggest that increasing muscle mass and strength in the elderly may have multiple, positive effects on the brain and the skeleton.

ACKNOWLEDGMENTS

Funding for this research was provided by the National Institute on Aging (P01 AG036675). This work was presented at the Korean Bone and Mineral Society and the Asan Medical Center in November, 2016. I am grateful to Drs. J.M. Koh, B.J. Kim and the Korean Bone and Mineral Society for providing this opportunity, and to the late Clifton Baile for his encouragement and collaboration on our many leptin-related studies.

REFERENCES

1. Kim SH, Meehan JP, Blumenfeld T, et al. Hip fractures in the United States: 2008 nationwide emergency department sample. *Arthritis Care Res (Hoboken)* 2012;64:751-7.
2. Fried LP, Tangen CM, Walston J, et al. Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci* 2001;56:M146-56.
3. Dawson-Hughes B, Bischoff-Ferrari H. Considerations concerning the definition of sarcopenia. *Osteoporos Int* 2016; 27:3139-44.
4. Järvinen TL, Sievanen H, Khan KM, et al. Shifting the focus in fracture prevention from osteoporosis to falls. *BMJ* 2008; 336:124-6.
5. Fiuzza-Luces C, Garatachea N, Berger NA, et al. Exercise is the real polypill. *Physiology (Bethesda)* 2013;28:330-58.
6. Pedersen BK. Muscles and their myokines. *J Exp Biol* 2011; 214:337-46.
7. Hamrick MW. A role for myokines in muscle-bone interac-

- tions. *Exerc Sport Sci Rev* 2011;39:43-7.
8. Spiegelman BM, Flier JS. Obesity and the regulation of energy balance. *Cell* 2001;104:531-43.
 9. Mera P, Laue K, Ferron M, et al. Osteocalcin signaling in myofibers is necessary and sufficient for optimum adaptation to exercise. *Cell Metab* 2016;23:1078-92.
 10. Novotny SA, Warren GL, Hamrick MW. Aging and the muscle-bone relationship. *Physiology (Bethesda)* 2015;30:8-16.
 11. Fernández-Real JM, Vayreda M, Casamitjana R, et al. The fat-free mass compartment influences serum leptin in men. *Eur J Endocrinol* 2000;142:25-9.
 12. Solberg R, Aas V, Thoresen GH, et al. Leptin expression in human primary skeletal muscle cells is reduced during differentiation. *J Cell Biochem* 2005;96:89-96.
 13. Wang J, Liu R, Hawkins M, et al. A nutrient-sensing pathway regulates leptin gene expression in muscle and fat. *Nature* 1998;393:684-8.
 14. Wolsk E, Mygind H, Grondahl TS, et al. Human skeletal muscle releases leptin in vivo. *Cytokine* 2012;60:667-73.
 15. Thomas T. The complex effects of leptin on bone metabolism through multiple pathways. *Curr Opin Pharmacol* 2004;4:295-300.
 16. Arounleut P, Bowser M, Upadhyay S, et al. Absence of functional leptin receptor isoforms in the POUND (*Lepr(db/lb)*) mouse is associated with muscle atrophy and altered myoblast proliferation and differentiation. *PLoS One* 2013;8:e72330.
 17. Guerra B, Santana A, Fuentes T, et al. Leptin receptors in human skeletal muscle. *J Appl Physiol (1985)* 2007;102:1786-92.
 18. Crespi EJ, Denver RJ. Leptin (*ob* gene) of the South African clawed frog *Xenopus laevis*. *Proc Natl Acad Sci U S A* 2006;103:10092-7.
 19. Balaskó M, Soós S, Székely M, et al. Leptin and aging: Review and questions with particular emphasis on its role in the central regulation of energy balance. *J Chem Neuroanat* 2014;61-62:248-55.
 20. Pareja-Galeano H, Santos-Lozano A, Sanchis-Gomar F, et al. Circulating leptin and adiponectin concentrations in healthy exceptional longevity. *Mech Ageing Dev* 2016.
 21. Bucci L, Yani SL, Fabbri C, et al. Circulating levels of adipokines and IGF-1 are associated with skeletal muscle strength of young and old healthy subjects. *Biogerontology* 2013;14:261-72.
 22. Holden KF, Lindquist K, Tylavsky FA, et al. Serum leptin level and cognition in the elderly: Findings from the Health ABC Study. *Neurobiol Aging* 2009;30:1483-9.
 23. Zeki Al Hazzouri A, Stone KL, Haan MN, et al. Leptin, mild cognitive impairment, and dementia among elderly women. *J Gerontol A Biol Sci Med Sci* 2013;68:175-80.
 24. Hamrick MW, Dukes A, Arounleut P, et al. The adipokine leptin mediates muscle- and liver-derived IGF-1 in aged mice. *Exp Gerontol* 2015;70:92-6.
 25. Hamrick MW, McGee-Lawrence ME, Frechette DM. Fatty infiltration of skeletal muscle: mechanisms and comparisons with bone marrow adiposity. *Front Endocrinol (Lausanne)* 2016;7:69.
 26. Chen YW, Gregory CM, Scarborough MT, et al. Transcriptional pathways associated with skeletal muscle disuse atrophy in humans. *Physiol Genomics* 2007;31:510-20.
 27. Sáinz N, Rodríguez A, Catalán V, et al. Leptin administration favors muscle mass accretion by decreasing FoxO3a and increasing PGC-1alpha in *ob/ob* mice. *PLoS One* 2009;4:e6808.
 28. Bartell SM, Rayalam S, Ambati S, et al. Central (ICV) leptin injection increases bone formation, bone mineral density, muscle mass, serum IGF-1, and the expression of osteogenic genes in leptin-deficient *ob/ob* mice. *J Bone Miner Res* 2011;26:1710-20.
 29. Anastasilakis AD, Polyzos SA, Skouvaklidou EC, et al. Circulating follistatin displays a day-night rhythm and is associated with muscle mass and circulating leptin levels in healthy, young humans. *Metabolism* 2016;65:1459-65.
 30. Hamrick MW, Herberg S, Arounleut P, et al. The adipokine leptin increases skeletal muscle mass and significantly alters skeletal muscle miRNA expression profile in aged mice. *Biochem Biophys Res Commun* 2010;400:379-83.
 31. Cheung TH, Quach NL, Charville GW, et al. Maintenance of muscle stem-cell quiescence by microRNA-489. *Nature* 2012;482:524-8.
 32. Beane OS, Fonseca VC, Cooper LL, et al. Impact of aging on the regenerative properties of bone marrow-, muscle-, and adipose-derived mesenchymal stem/stromal cells. *PLoS One* 2014;9:e115963.
 33. Ducey P, Amling M, Takeda S, et al. Leptin inhibits bone formation through a hypothalamic relay: a central control of bone mass. *Cell* 2000;100:197-207.
 34. Hamrick MW, Ding KH, Ponnala S, et al. Caloric restriction decreases cortical bone mass but spares trabecular bone in the mouse skeleton: implications for the regulation of

- bone mass by body weight. *J Bone Miner Res* 2008;23:870-8.
35. Hamrick MW, Ferrari SL. Leptin and the sympathetic connection of fat to bone. *Osteoporos Int* 2008;19:905-12.
 36. Hamrick MW, Della-Fera MA, Choi YH, et al. Leptin treatment induces loss of bone marrow adipocytes and increases bone formation in leptin-deficient ob/ob mice. *J Bone Miner Res* 2005;20:994-1001.
 37. Zhou BO, Yue R, Murphy MM, et al. Leptin-receptor-expressing mesenchymal stromal cells represent the main source of bone formed by adult bone marrow. *Cell Stem Cell* 2014;15:154-68.
 38. Thomas T, Gori F, Khosla S, et al. Leptin acts on human marrow stromal cells to enhance differentiation to osteoblasts and to inhibit differentiation to adipocytes. *Endocrinology* 1999;140:1630-8.
 39. Yue R, Zhou BO, Shimada IS, et al. Leptin receptor promotes adipogenesis and reduces osteogenesis by regulating mesenchymal stromal cells in adult bone marrow. *Cell Stem Cell* 2016;18:782-96.
 40. Periyasamy-Thandavan S, Herberg S, Arounleut P, et al. Caloric restriction and the adipokine leptin alter the SDF-1 signaling axis in bone marrow and in bone marrow derived mesenchymal stem cells. *Mol Cell Endocrinol* 2015;410:64-72.
 41. Turner RT, Kalra SP, Wong CP, et al. Peripheral leptin regulates bone formation. *J Bone Miner Res* 2013;28:22-34.
 42. Hamrick MW, Della Fera MA, Choi YH, et al. Injections of leptin into rat ventromedial hypothalamus increase adipocyte apoptosis in peripheral fat and in bone marrow. *Cell Tissue Res* 2007;327:133-41.
 43. Lindenmaier LB, Philbrick KA, Branscum AJ, et al. Hypothalamic leptin gene therapy reduces bone marrow adiposity in ob/ob mice fed regular and high-fat diets. *Front Endocrinol (Lausanne)* 2016;7:110.
 44. Li H, Fong C, Chen Y, et al. Beta-adrenergic signals regulate adipogenesis of mouse mesenchymal stem cells via cAMP/PKA pathway. *Mol Cell Endocrinol* 2010;323:201-7.
 45. Devlin MJ, Cloutier AM, Thomas NA, et al. Caloric restriction leads to high marrow adiposity and low bone mass in growing mice. *J Bone Miner Res* 2010;25:2078-88.
 46. Pino AM, Ríos S, Astudillo P, et al. Concentration of adipogenic and proinflammatory cytokines in the bone marrow supernatant fluid of osteoporotic women. *J Bone Miner Res* 2010;25:492-8.
 47. Astudillo P, Ríos S, Pastenes L, et al. Increased adipogenesis of osteoporotic human-mesenchymal stem cells (MSCs) characterizes by impaired leptin action. *J Cell Biochem* 2008;103:1054-65.
 48. Hess R, Pino AM, Ríos S, et al. High affinity leptin receptors are present in human mesenchymal stem cells (MSCs) derived from control and osteoporotic donors. *J Cell Biochem* 2005;94:50-7.
 49. Fafián-Labora J, Fernández-Pernas P, Fuentes I, et al. Influence of age on rat bone-marrow mesenchymal stem cells potential. *Sci Rep* 2015;5:16765.
 50. Sui BD, Hu CH, Zheng CX, et al. Microenvironmental views on mesenchymal stem cell differentiation in aging. *J Dent Res* 2016;95:1333-40.
 51. Zhang W, Ou G, Hamrick M, et al. Age-related changes in the osteogenic differentiation potential of mouse bone marrow stromal cells. *J Bone Miner Res* 2008;23:1118-28.
 52. Howard JK, Flier JS. Attenuation of leptin and insulin signaling by SOCS proteins. *Trends Endocrinol Metab* 2006;17:365-71.
 53. Guadalupe-Grau A, Larsen S, Guerra B, et al. Influence of age on leptin induced skeletal muscle signalling. *Acta Physiol (Oxf)* 2014;211:214-28.
 54. Hamrick MW. Leptin, bone mass, and the thrifty phenotype. *J Bone Miner Res* 2004;19:1607-11.
 55. Chapman IM. Endocrinology of anorexia of ageing. *Best Pract Res Clin Endocrinol Metab* 2004;18:437-52.
 56. Fulgoni VL 3rd. Current protein intake in America: analysis of the National Health and Nutrition Examination Survey, 2003-2004. *Am J Clin Nutr* 2008;87:1554s-7s.
 57. Wengreen HJ, Munger RG, West NA, et al. Dietary protein intake and risk of osteoporotic hip fracture in elderly residents of Utah. *J Bone Miner Res* 2004;19:537-45.
 58. Hubbard RE, O'Mahony MS, Calver BL, et al. Nutrition, inflammation, and leptin levels in aging and frailty. *J Am Geriatr Soc* 2008;56:279-84.
 59. Hamrick MW, Ding KH, Pennington C, et al. Age-related loss of muscle mass and bone strength in mice is associated with a decline in physical activity and serum leptin. *Bone* 2006;39:845-53.
 60. Morley JE. Anorexia, sarcopenia, and aging. *Nutrition* 2001;17:660-3.
 61. Demontis F, Piccirillo R, Goldberg AL, et al. The influence of skeletal muscle on systemic aging and lifespan. *Aging Cell* 2013;12:943-9.
 62. Gray SL, Anderson ML, Hubbard RA, et al. Frailty and inci-

- dent dementia. *J Gerontol A Biol Sci Med Sci* 2013;68:1083-90.
63. Maioli S, Lodeiro M, Merino-Serrais P, et al. Alterations in brain leptin signalling in spite of unchanged CSF leptin levels in Alzheimer's disease. *Aging Cell* 2015;14:122-9.
 64. Dukes A, Davis C, El Refaey M, et al. The aromatic amino acid tryptophan stimulates skeletal muscle IGF1/p70s6k/mTor signaling in vivo and the expression of myogenic genes in vitro. *Nutrition* 2015;31:1018-24.
 65. Kim H, Kim M, Kojima N, et al. Exercise and nutritional supplementation on community-dwelling elderly Japanese women with sarcopenic obesity: a randomized controlled trial. *J Am Med Dir Assoc* 2016;17:1011-9.
 66. Isanejad M, Mursu J, Sirola J, et al. Association of protein intake with the change of lean mass among elderly women: The Osteoporosis Risk Factor and Prevention - Fracture Prevention Study (OSTPRE-FPS). *J Nutr Sci* 2015;4:e41.
 67. Welt CK, Chan JL, Bullen J, et al. Recombinant human leptin in women with hypothalamic amenorrhea. *N Engl J Med* 2004;351:987-97.
 68. Brinkoetter M, Magkos F, Vamvini M, et al. Leptin treatment reduces body fat but does not affect lean body mass or the myostatin-follistatin-activin axis in lean hypoleptinemic women. *Am J Physiol Endocrinol Metab* 2011;301:E99-104.
 69. Leenders M, Verdijk LB, van der Hoeven L, et al. Elderly men and women benefit equally from prolonged resistance-type exercise training. *J Gerontol A Biol Sci Med Sci* 2013;68:769-79.
 70. Fielding RA, LeBrasseur NK, Cuoco A, et al. High-velocity resistance training increases skeletal muscle peak power in older women. *J Am Geriatr Soc* 2002;50:655-62.