

AMPK-autophagy inhibition sensitizes icaritin-induced anti-colorectal cancer cell activity

Chunxian Zhou^{1,*}, Jun Gu^{2,*}, Gang Zhang¹, Da Dong¹, Qunying Yang¹, Min-Bin Chen³, Dongfeng Xu¹

¹Department of Interventional Radiology, Wujiang Hospital Affiliated to Nantong University, Wujiang, Suzhou, China

²The Department of Orthopedics, The Second Affiliated Hospital of Soochow University, Suzhou, China

³Department of Oncology, Kunshan First People's Hospital Affiliated to Jiangsu University, Kunshan, 215300, China

* Co-first authors

Correspondence to: Dongfeng Xu, **email:** drxudongfeng6@126.com

Min-Bin Chen, **email:** cmb1981@163.com

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ABSTRACT

The current research studied the potential effect of autophagy on icaritin-induced anti-colorectal cancer (CRC) cell activity. Treatment of icaritin in both primary and established (HT-29) CRC cells induced feedback activation of autophagy, evidenced by p62 degradation, Beclin-1 and autophagy-related gene-5 (ATG-5) upregulation, as well as light chain 3B (LC3B)-GFP puncta formation. Pharmacological inhibiting of autophagy dramatically potentiated icaritin-induced CRC cell death and apoptosis. Meanwhile, shRNA-mediated knockdown of Beclin-1 or ATG-5 also sensitized icaritin-induced CRC cell death and apoptosis. Icaritin activated AMP-activated protein kinase (AMPK) signaling in CRC cells, functioning as the upstream signaling for autophagy activation. shRNA/siRNA-mediated knockdown of AMPK α 1 inhibited icaritin-induced autophagy activation, but exacerbated CRC cell death. On the other hand, the AMPK activator compound 13 (C13) or the autophagy activator MHY1485 attenuated icaritin-induced cytotoxicity. In nude mice, icaritin (oral administration)-induced HT-29 tumor growth inhibition was potentiated when combined with AMPK α 1 shRNA knockdown in tumors. We conclude that feedback activation of AMPK-autophagy pathway could be a primary resistance factor of icaritin.

INTRODUCTION

The colorectal cancer (CRC) has long been a major health problem in both Eastern and Western countries [1–3]. Hence, our group [10] and others [1, 8] have been focusing on exploring novel anti-CRC agents. Icaritin, the hydrolytic product of Icarin, is the main active ingredient of *Epimedium*, which is a known Traditional Chinese Medicine [11]. Studies have tested its biological functions in different experimental settings, including its anti-cancer activity [12–14]. For instance, icaritin was shown to inhibit proliferation of prostate cancer cells [12], breast cancer cells [13], as well as hepatocellular carcinoma (HCC) cells [15] and endometrial cancer cells [16]. Our recent study has demonstrated that icaritin could also inhibit CRC cells *in vitro* and *in vivo* [10]. At the molecular level, we

found that icaritin activated JNK-dependent mitochondrial permeability transition pore (mPTP) necrosis pathway to kill CRC cells [10].

One important aim of the current study is to identify possible icaritin's resistance factor. We here focused on the potential involvement of autophagy in the process. Existing studies have displayed feedback activation of autophagy in many cancer cells following treatment of a variety of anti-cancer drugs [17–21], which could be a key resistance factor to inhibit cancer cell death and apoptosis [17, 21, 22]. Reversely, genetic or pharmacological inactivation of autophagy could then sensitize the anti-cancer activity by these anti-cancer drugs [17–22]. In the current study, we showed that autophagy inhibition dramatically sensitizes icaritin-induced anti-CRC cell activity.

RESULTS

Icaritin activates autophagy in human CRC cells

In order to test the potential effect of icaritin on autophagy, Western blot assay was performed to test expression of autophagy-associated proteins in icaritin-treated cells. As demonstrated, treatment of icaritin in HT-29 cells dose-dependently upregulated Beclin-1, autophagy-related gene-5 (ATG-5) and light chain 3B-II (LC3B-II), but downregulated p62 (Figure 1A). Meanwhile, the percentage of LC3B-GFP puncta positive cells, or autophagic cells, was also significantly increased following icaritin (5–25 μ M) treatment (Figure 1B). These results suggested autophagy activation in HT-29 cells after icaritin treatment [23–26]. Similarly, in two lines of primary colon cancer cells (patient-derived), icaritin (10 μ M) treatment induced Beclin-1, ATG-5 and LC3B-II upregulation but p62 degradation (Figure 1C). Further, the number of autophagic cells was also significantly increased after icaritin treatment in the primary colon cancer cells (Figure 1D). Thus, these results indicate that icaritin induces autophagy activation in established and primary CRC cells.

Autophagy inhibitors potentate icaritin-induced CRC cell death and apoptosis

To study the potential effect of autophagy in icaritin-mediated anti-CRC cell activity, various autophagy inhibitors were applied, including chloroquine (Cq), ammonium chloride (NH_4Cl) and 3-methyladenine (3-MA). MTT assay results showed that, in the presence of these autophagy inhibitors, icaritin-induced viability reduction was significantly potentiated (Figure 2A). The icaritin's IC₅₀, the concentration that inhibits 50% of cell viability, decreased from over 20 μ M to less than 5 μ M with co-treatment of the autophagy inhibitors (Figure 2A). Icaritin-induced HT-29 cell death, tested by the lactate dehydrogenase (LDH) release, was also significantly augmented with the autophagy inhibitors (Figure 2B). In line with our previous findings [10], treatment with icaritin (10 μ M) alone failed to induce significant apoptosis activation in HT-29 cells (Figure 2C). Remarkably, when combined with the autophagy inhibitors, icaritin provoked dramatic apoptosis (Figure 2C), which was tested by the TUNEL staining assay (Figure 2C). In the primary colon cancer cells, the above autophagy inhibitors similarly potentiated icaritin-induced cell viability reduction (Figure 2D). Thus, pharmacological inhibition of autophagy potentates icaritin's cytotoxicity in CRC cells. On the other hand, in the primary human colon epithelial cells, treatment with icaritin or together with these autophagy inhibitors failed to induce significant cell viability reduction (Figure 2E) and apoptosis (Figure 2F). Thus, icaritin combination

with the autophagy inhibitor was only cytotoxic to cancerous cells.

Beclin-1 or ATG-5 shRNA knockdown sensitizes icaritin-induced cytotoxicity against CRC cells

The above pharmacological evidences suggest that autophagy activation possibly serves as a potential resistance factor of icaritin. To exclude the possible off-target effect of the applied autophagy inhibitors, shRNA method was utilized to silence key autophagy-associated proteins, including Beclin-1 [27] and ATG-5 [28]. As demonstrated in Figure 3A, the two different Beclin-1 shRNAs ("a/b") both efficiently and stably downregulated Beclin-1 in HT-29 cells. Similarly, the two specific shRNAs ("a/b") [29] also silenced ATG-5 in HT-29 cells (Figure 3A). Remarkably, icaritin-induced HT-29 cell viability reduction (Figure 3B), cell death (Figure 3C) and apoptosis (Figure 3D) were significantly intensified with Beclin-1 or ATG-5 shRNA knockdown. Yet, knockdown of these proteins alone showed no significant effect on cell survival and apoptosis (Figure 3B and 3D). These genetic evidences further suggest that autophagy inhibition could sensitize icaritin's activity in CRC cells.

Icaritin activates AMPK as upstream of autophagy in CRC cells

Next, we explored the potential upstream signaling responsible for autophagy activation by icaritin. Growing evidences have suggested that AMP-activated protein kinase (AMPK) activation could provoke autophagy under a number of stimuli [30–32]. Here, we showed that icaritin treatment in HT-29 cells dose-dependently induced AMPK activation, which was tested by phosphorylation ("p") of AMPK α (Thr-172) and its major downstream kinase ACC (acetyl-CoA carboxylase, Ser-79) (Figure 4A). Expression of regular AMPK α 1 and ACC was unchanged following icaritin treatment (Figure 4A). To study the activity of AMPK in icaritin-induced autophagy activation, we again utilized shRNA method to stably knockdown AMPK α 1 in HT-29 cells. As demonstrated, the three targeted-shRNAs ("a/b/c") dramatically downregulated AMPK α 1 in HT-29 cells (Figure 4B). Consequently, icaritin-induced AMPK activation, or AMPK α -ACC phosphorylation, was also dramatically inhibited (Figure 4B). Remarkably, AMPK α 1 knockdown almost blocked icaritin-induced autophagy activation in HT-29 cells (Figure 4C), indicating that AMPK activation is required for icaritin-induced autophagy activation.

If AMPK activation is the upstream signaling for icaritin-mediated autophagy activation, blockage of AMPK should also potentate icaritin's cytotoxicity in CRC cells. Indeed, we showed that icaritin-induced viability reduction (Figure 4D), cell death (Figure 4E) and apoptosis (Figure 4F)

were remarkably augmented in AMPK α 1-shRNA-expressing HT-29 cells. In another word, AMPK α 1 silence could significantly facilitate icaritin-induced lethality against HT-29 cells (Figure 4D–4F). In the primary colon cancer cells, siRNA was applied to temporary knockdown AMPK α 1. The targeted siRNA decreased AMPK α 1 expression and inhibited icaritin-induced AMPK activation (Figure 4G). Significantly, icaritin-induced cytotoxicity, or viability reduction, was also exacerbated with AMPK α 1 siRNA knockdown in the primary cancer cells (Figure 4H). Collectively, these results indicate that icaritin-induced AMPK activation serves as the upstream signaling for subsequent autophagy activation. Reversely, AMPK inactivation by siRNA/shRNA knockdown of AMPK α 1 significantly potentiates icaritin-induced cytotoxicity in CRC cells.

AMPK-autophagy activators attenuate icaritin's cytotoxicity in CRC cells

Based on the results above, we would speculate that forced-activation of AMPK-autophagy pathway should attenuate icaritin's cytotoxicity. In the current study, an α 1 specific AMPK activator Compound 13 (C13) [33, 34] and an autophagy activator MHY1485 [35] were utilized. Results showed that pre-treatment of the two activators indeed inhibited icaritin-induced HT-29 cell viability reduction (Figure 5A) and cell death (Figure 5B). Similarly in the primary colon cancer cells, Compound 13 and MHY1485 attenuated icaritin's lethality (Figure 5C and 5D). Therefore, pharmacologic forced-activation of AMPK-autophagy pathway attenuates icaritin's cytotoxicity in CRC cells.

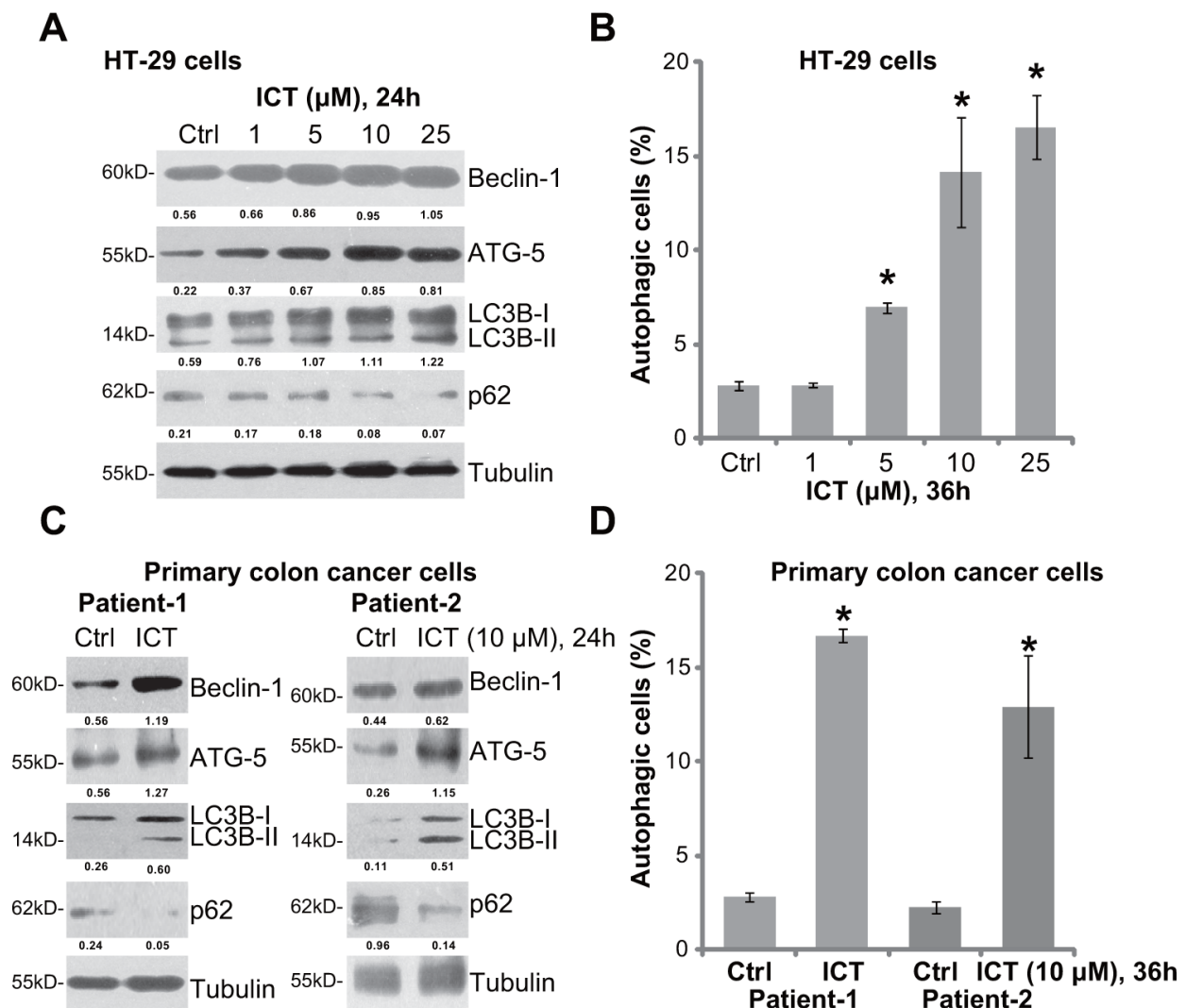


Figure 1: Icaritin activates autophagy in human CRC cells. HT-29 cells (A and B) or the primary colon cancer cells (two lines, "Patient-1/-2") (C and D), were either left untreated ("Ctrl", same for all figures), or treated with applied concentration of icaritin (ICT, same for all figures) for indicated time; Expression of listed proteins was shown (A and C); Percentage of LC3B-GFP puncta positive cells, or autophagic cells, was also recorded (B and D). Expression of listed proteins was quantified and normalized to Tubulin (A and C). Data were expressed as mean \pm standard deviation (SD), experiments were repeated five times. $n = 5$ for each assay. * $p < 0.05$ vs. "Ctrl" group.

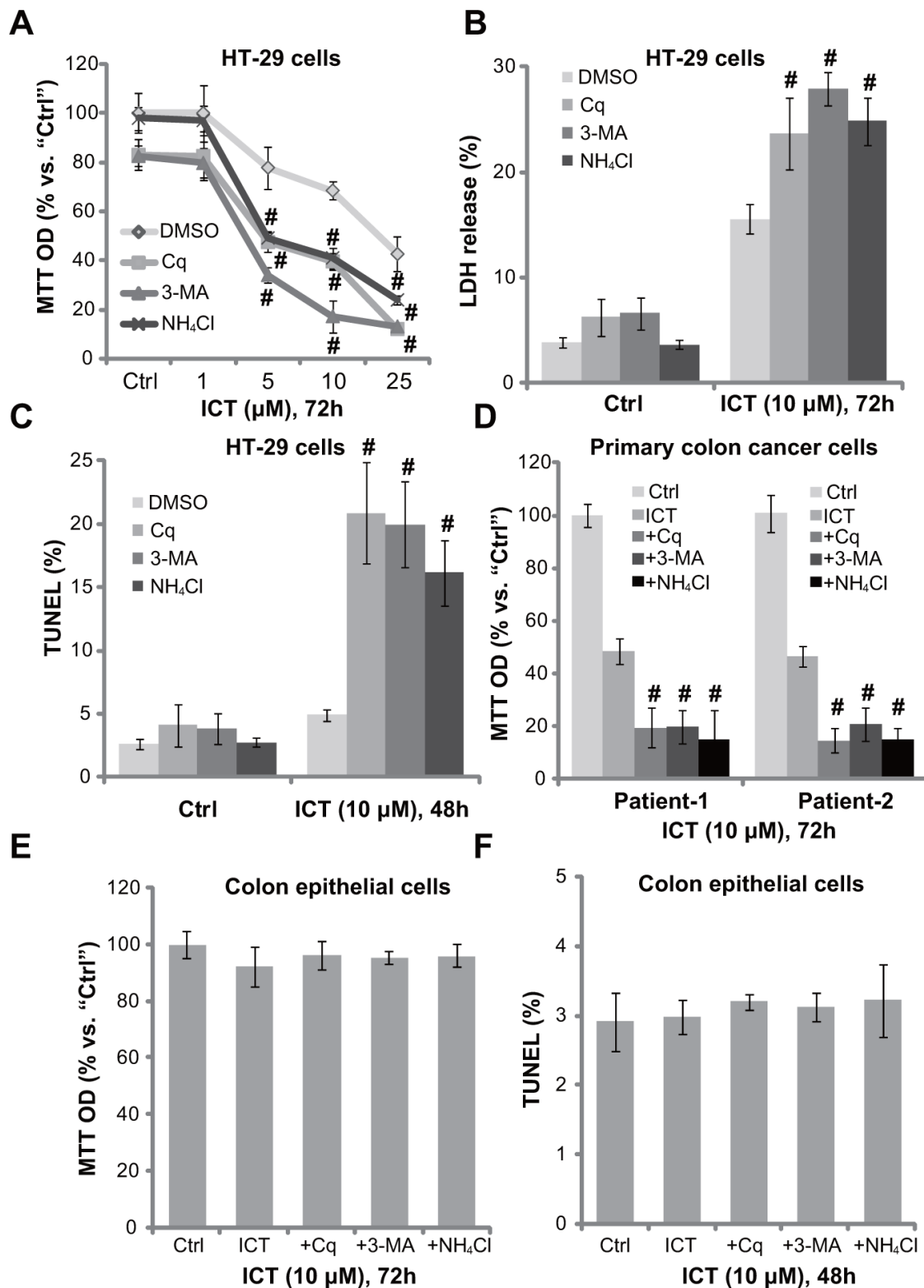


Figure 2: Autophagy inhibitors potentate icaritin-induced CRC cell death and apoptosis. HT-29 cells (A–C), primary colon cancer cells (two lines, “Patient-1/-2”) (D), or the primary colon epithelial cells (E and F), pre-treated for 30 min with applied autophagy inhibitors: 3-methyladenine (3-MA, 1.0 mM), chloroquine (Cq, 5 μM) or ammonium chloride (NH_4Cl , 2.5 mM), were subsequently treated with ICT (10 μM) for designated time; Cell viability, cell death and apoptosis were tested by MTT assay (A, D and E), LDH release assay (B) and TUNEL staining assay (C and F), respectively. Experiments in this figure were repeated three times, and similar results were obtained. “DMSO” stands for 0.1% DMSO of vehicle control. Data were expressed as mean \pm SD, experiments were repeated five times. $n = 5$ for each assay. $\#p < 0.05$ vs. “DMSO” group.

AMPK α 1 silence sensitizes icaritin-induced anti-HT-29 tumor activity *in vivo*

Above *in vitro* studies imply that AMPK-autophagy inhibition could sensitize icaritin's anti-CRC cell activity. We next tested this hypothesis *in vivo*, using the HT-29 xenograft model [10]. Tumor growth curve results in Figure 6A demonstrated that oral administration of icaritin (10 mg/kg, daily, for 21 days) in the nude mice inhibited growth of HT-29 xenografts with scramble non-sense control shRNA ("Ctl shRNA"). Remarkably, icaritin-induced anti-tumor activity was dramatically potentiated in HT-29 tumors with AMPK α 1 shRNA-a (Figure 6A). Daily tumor growth results further confirmed that AMPK α 1 silence sensitized icaritin-mediated anti-

HT-29 tumor activity (Figure 6B). Notably, there was no significant difference in the growth of HT-29 tumors expressing AMPK α 1 shRNA or Ctl shRNA (Figure 6A and 6B). Intriguingly, the mice body weight was not significantly different among the four groups (Figure 6C), neither did we notice any apparent toxicities. These results imply that these nude mice were well-tolerated to the above treatment regimens. When analyzing tumor tissue lysate samples, we showed that AMPK α 1 was indeed silenced in HT-29 tumors expressing AMPK α 1 shRNA-a (Figure 6D, quantified data of three repeats). Meanwhile, icaritin-induced autophagy activation, evidenced by strong Beclin-1 expression, was also significantly attenuated with AMPK α 1 shRNA knockdown (Figure 6D, quantified data of three repeats). These results indicate that activation of

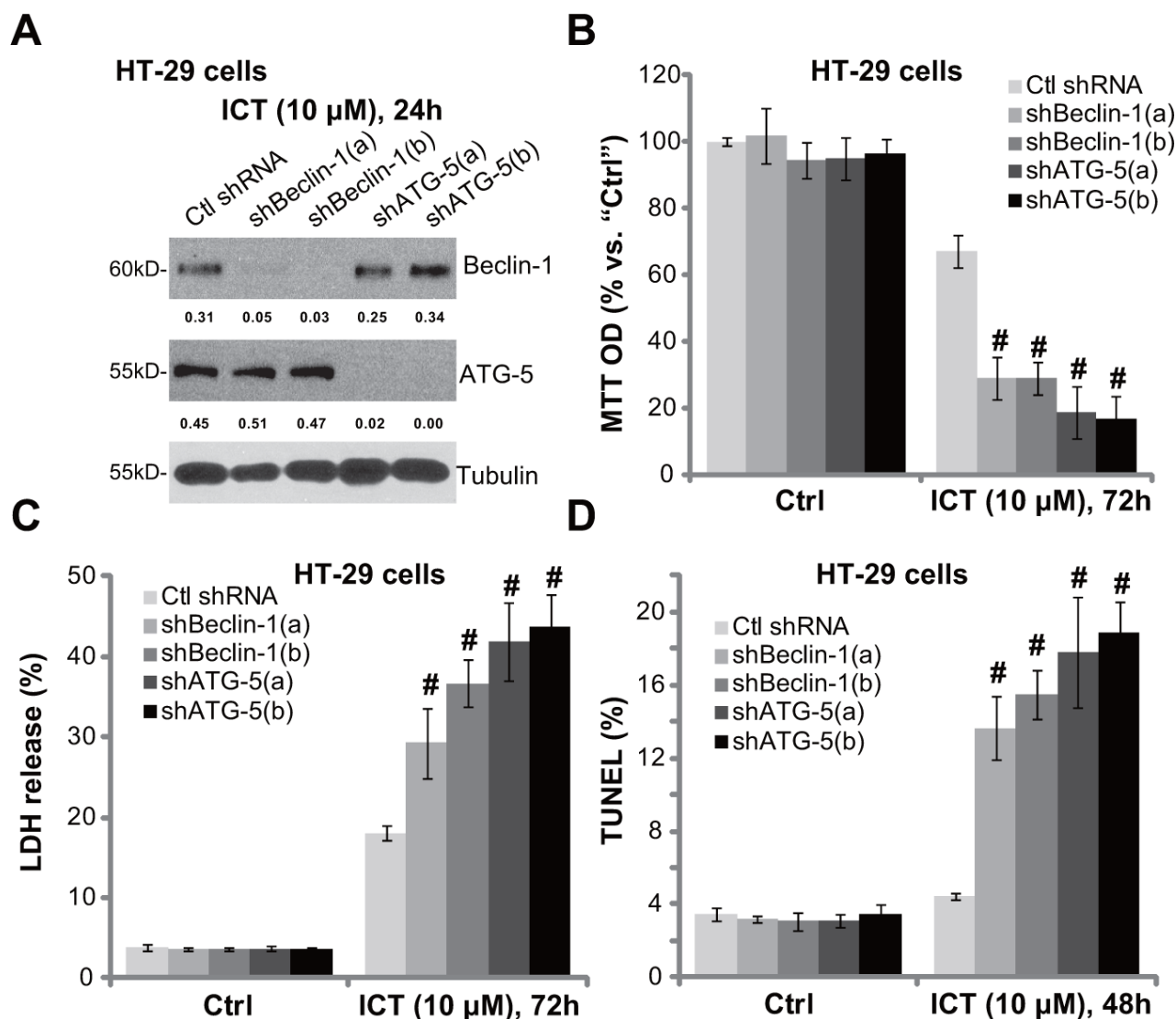


Figure 3: Beclin-1 or ATG-5 shRNA knockdown sensitizes icaritin-induced cytotoxicity against CRC cells. The stably HT-29 cells with scramble non-sense control shRNA ("Ctl shRNA"), Beclin-1 shRNA ("shBeclin-1-a/b") or ATG-5 shRNA ("sh ATG-5-a/b") were treated with ICT (10 μ M) for designated time; Expression of listed proteins was tested by Western blot assay (A); Cell viability (B), cell death (C) and apoptosis (D) were also tested. Expression of Beclin-1 and ATG-5 was quantified and normalized to Tubulin (A). Data were expressed as mean \pm SD, experiments were repeated three times. $n = 5$ for each assay. $^{\#}p < 0.05$ vs. "Ctl shRNA" group.

AMPK is possibly also important for autophagy activation and icaritin resistance *in vivo*.

DISCUSSION

Several inhibitors were applied here. Chloroquine (Cq) and ammonium chloride (NH₄Cl) both increase

intra-lysosomal pH to block degradation of autophagic proteins [36]. On the other hand, 3-methyladenine (3-MA) inhibits LC3B-I to LC3B-II conversion, and inhibits autophagosome initiation [37]. Here, treatment of these autophagy inhibitors dramatically potentiated icaritin-induced lethality against primary and established CRC cells. More intriguingly, pharmacological autophagy

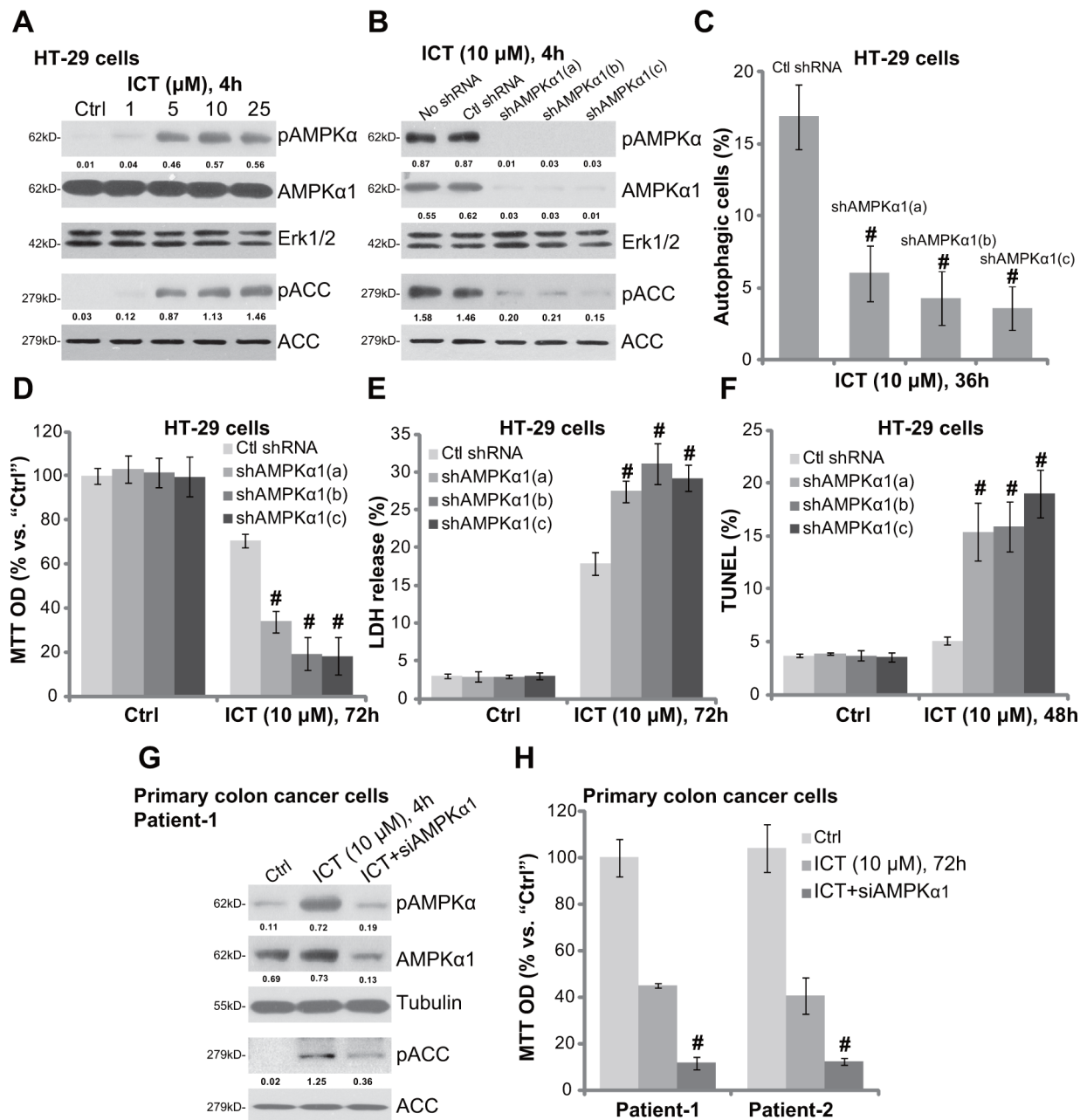


Figure 4: Icaritin activates AMPK as upstream of autophagy in CRC cells. HT-29 cells were treated with applied concentration of ICT for 4 hours, expression of listed proteins was shown (A); The stable HT-29 cells, expressing scramble non-sense control shRNA ("Ctl shRNA") or indicated AMPKα1 shRNA ("a/b/c"), were treated with/out ICT (10 μM) for designated time; Expression of listed proteins was shown (B); Percentage of LC3B-GFP puncta cells ("autophagic cells") was recorded (C); Cell viability (D), cell death (E) and apoptosis (F) were also tested. Primary colon cancer cells with/out AMPKα1 siRNA (siAMPKα1, 200 nM, 48 hours) were treated with ICT (10 μM) for designated time; Expression of listed proteins was shown (G); Cell viability was also tested (H). Phosphorylation ("p") of AMPKα (vs. loading Erk1/2 or Tubulin) and ACC (vs. regular ACC) was quantified. Data were expressed as mean ± SD, experiments were repeated three times. *n* = 5 for each assay. #*p* < 0.05 vs. "ICT" only group.

inhibition could restore CRC cell apoptosis following icaritin treatment. Therefore, feedback autophagy activation could be a primary reason of non-apoptosis induction in icaritin-treated CRC cells, as reported in our previous study [10]. Inhibition of autophagy then reprovokes apoptosis with icaritin treatment in CRC cells.

In the process of autophagy, autophagosome, the double-membrane structure, will enclose the cellular components [17, 21, 22]. These autophagosomes will

then be fused within lysosome to digest the enclosed components, and to supply nutrients for cell survival and apoptosis prevention [17, 21, 22]. Both *ATG-5* and *Beclin-1* [27] are indispensable for autophagosome formation [17, 21, 22]. In the current study, we showed that shRNA stable knockdown of *ATG-5* or *Beclin-1*, which presumably blocked autophagy progression, dramatically potentiated icaritin-induced CRC cell death and apoptosis. These genetic evidences further confirm

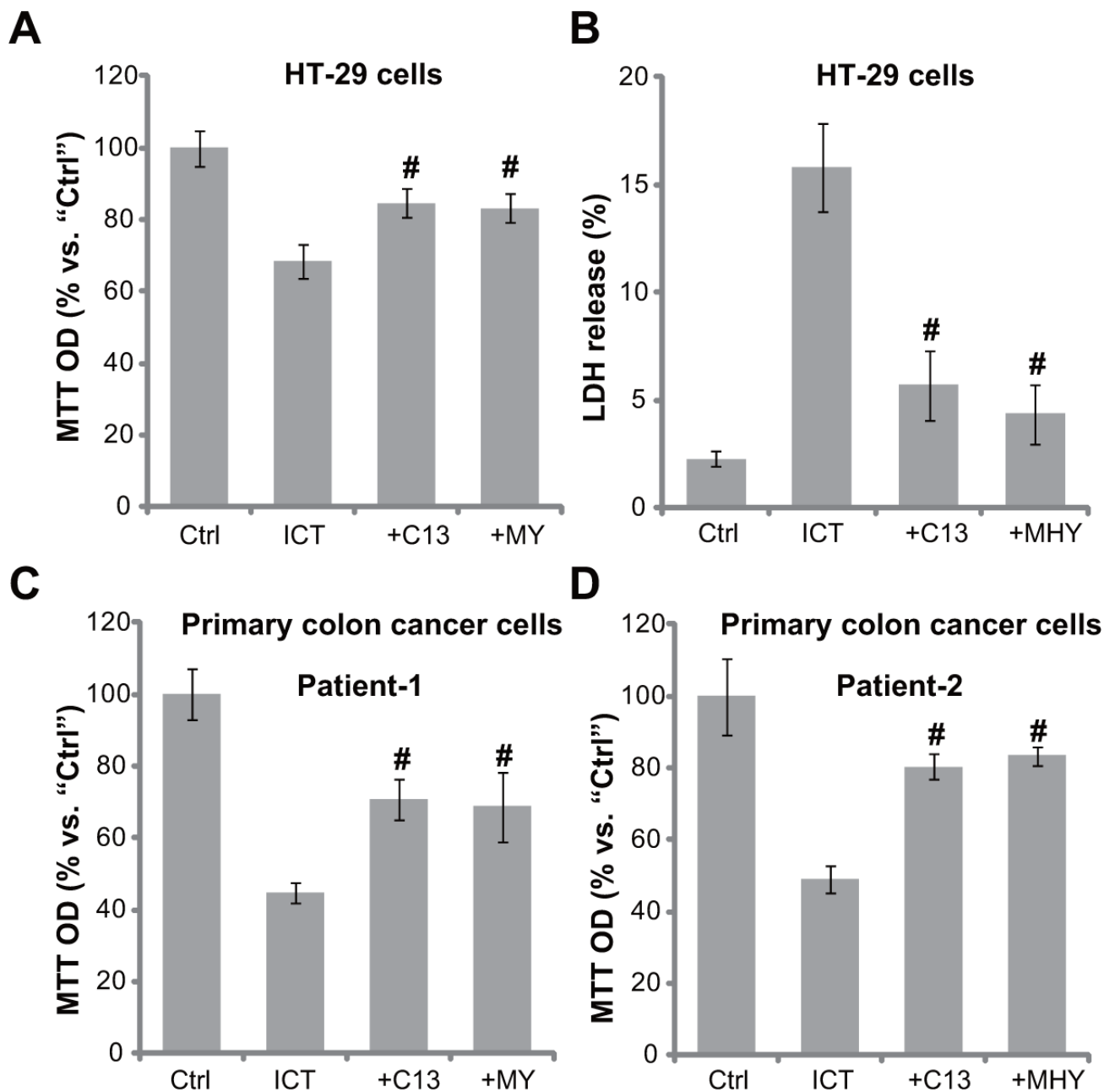


Figure 5: AMPK-autophagy activators attenuate icaritin's cytotoxicity in CRC cells. HT-29 cells (A–B) or the primary colon cancer cells (two lines, "Patient-1/-2") (C and D) were pre-treated for 30 min with the AMPK activator Compound 13 ("C13", 10 μ M) or the autophagy activator MHY1485 ("MHY", 10 μ M), followed by ICT (10 μ M) treatment for 72 hours; Cell viability and cell death were tested by MTT assay (A, C and D) and LDH release assay (B), respectively. Data were expressed as mean \pm SD, experiments were repeated five times. $n = 5$ for each assay. [#] $p < 0.05$ vs. "ICT" only group.

that feedback autophagy activation serves as a primary resistance factor of icaritin in CRC cells.

AMPK is a metabolic switcher that senses change in the intracellular AMP/ATP ratio [38]. Recent studies have proposed a key function of AMPK in the activation of autophagy [31]. There are at least two ways for AMPK to activate autophagy. AMPK phosphorylates and activates its downstream Ulk1, the latter is able to initiate cell autophagy [39, 40]. Secondly, AMPK phosphorylates

and in-activates TSC2, which then leads to mTORC1 (mTOR complex 1) inhibition [41, 42]. Activation of mTORC1 will silence autophagy, and AMPK-induced mTORC1 inactivation could then provoke autophagy [39, 40]. Here, we found that icaritin activated AMPK cascade in CRC cells, which served as the upstream signaling for autophagy. Blockage of AMPK activation by AMPK α 1 shRNA/siRNA largely inhibited icaritin-induced autophagy activation, yet exacerbating CRC

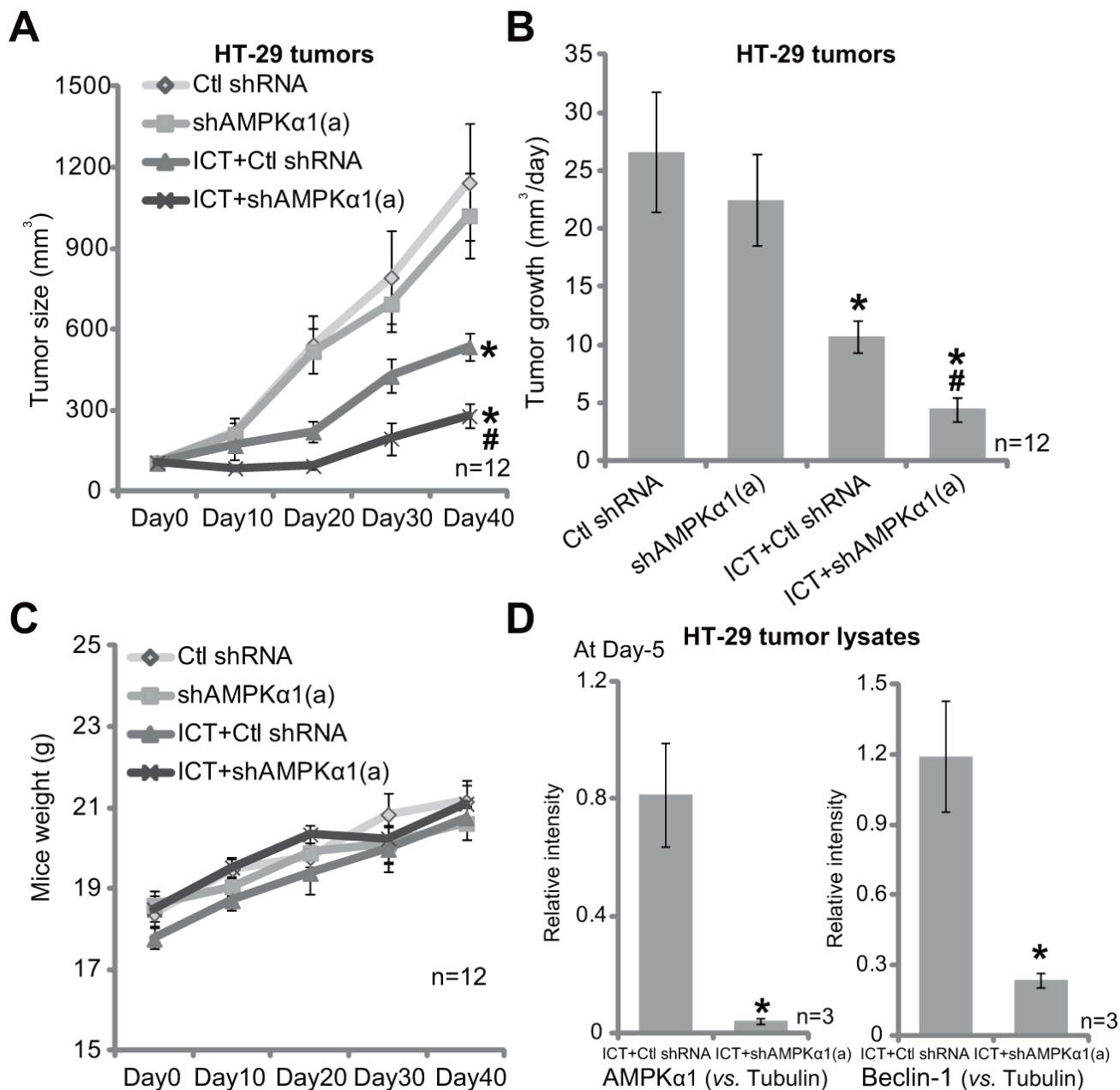


Figure 6: AMPK α 1 silence sensitizes icaritin-induced anti-HT-29 tumor activity *in vivo*. Nude mice bearing HT-29 tumors, expressing non-sense control shRNA (“Ctl shRNA”) or AMPK α 1 shRNA (“a”), were administrated daily with ICT (10 mg/kg, oral gavage) for 21 consecutive days; Tumor volume (A) and mice body weight (C) were recorded every 10 days for a total of 40 days; Tumor daily growth was also calculated (B). Day-5 after initial ICT administration, HT-29 tumors of three mice of ICT treatment group were isolated, expression of listed proteins in fresh tumor lysates was tested, and results were quantified (D, three repeats). Data were expressed as mean \pm SD. * $p < 0.05$ vs. “Ctl shRNA” only group. # $p < 0.05$ vs. “ICT” treatment of “Ctl shRNA” group.

cell death and apoptosis. In nude mice, AMPK α 1 shRNA knockdown in HT-29 tumors facilitated icaritin-induced inhibition on tumor growth. Thus, AMPK activation mediates autophagy induction and icaritin resistance in CRC cells. Further studies will be needed to explore the underlying mechanisms of AMPK activation by icaritin, and how activated-AMPK provokes autophagy in icaritin-treated cells.

In summary, our results suggest that feedback activation of AMPK-autophagy cascade could be a major resistance factor of icaritin in CRC cells. Blockage of this signaling dramatically sensitizes icaritin-induced anti-CRC cell activity *in vitro* and *in vivo*.

MATERIALS AND METHODS

Chemicals and reagents

Icaritin with a purity of 99.5% was obtained from Shenogen Pharma Co (Beijing, China). Icaritin preparation *in vitro* and *in vivo* was described previously [10]. Autophagy inhibitors chloroquine (Cq), ammonium chloride (NH₄Cl) and 3-methyladenine (3-MA) were obtained from Sigma-Aldrich (St. Louis, MO). Compound 13 was from Dr. Zhao's group [43]. MHY1485 was purchased from Biotools (Shanghai, China). Antibodies utilized in this study were all obtained from Cell Signaling Tech (Santa Cruz, CA).

HT-29 cell culture

HT-29 cells were maintained in DMEM medium with 10% FBS, in a CO₂ incubator.

Primary colon cancer and epithelial cell culture

Colon cancer tissues from written-informed patients (two patients, Patient-1/-2, both male, 57/48 years old) were washed, and digested as described [24]. Afterwards, individual cells were pelleted and cultured [44]. The primary human colon epithelial cells were provided by Dr. Wei-Hao Sun at the First Affiliated Hospital of Nanjing Medical University (Nanjing, China) [45]. The primary colon epithelial cells were cultured as described [46]. The study was approved by the institutional review board of all authors' institutions, and was in accordance with the principles expressed in the Declaration of Helsinki. Enrolled patients didn't receive chemotherapy or radiation prior to the surgery.

Detection of autophagic cells

The light chain 3 (LC3)-GFP-pcDNA3-puromycin, a gift from Dr. Shen [47], was transfected to HT-29 cells or primary colon cancer cells via Lipofectamine 2000 protocol. Afterwards, puromycin (5.0 μ g/mL) was added for 6–8 days to select stable cells. Cells were

then seeded onto confocal cover-slips. Following the applied treatment, GFP-LC3 accumulation was tested by fluorescence microscopy. Autophagic cells were tested by counting the percentage of cells with intense GFP-LC3 puncta, analyzing at least 200 cells per preparation in five independent experiments [24].

Cell viability assay

Cell viability was tested by the MTT assay as previously described [10].

TUNEL staining assay

Following treatment of cells, cell apoptosis was tested by the TUNEL staining assay as described [10]. Percentage of TUNEL positive nuclei was recorded, analyzing at least 200 cells per preparation in five independent experiments.

LDH assay of cell death

Cell death was tested by lactate dehydrogenase (LDH) release assay, the detailed protocol is described in our previous study [10].

Western blot assay

Aliquots of 30 μ g of lysed proteins were separated by 10% SDS polyacrylamide gel and transferred onto PVDF membranes (Millipore, Bedford, MA). After blocking, the membranes were incubated with specific primary and secondary antibodies. The targeted protein band was visualized via enhanced chemiluminescence (ECL) reagents. The intensity of each blot was quantified using ImageJ software after normalization to the corresponding loading control.

siRNA and transfection

The AMPK α 1 siRNA and the negative control scramble siRNA were purchased from Dharmacon Research Inc. (Lafayette, CO). siRNA transfection (200 nM, 48 hours) was performed using Lipofectamine plus reagents based on the previous protocol [48]. The efficiency of siRNA was determined by Western blots.

shRNA knockdown and stable cell selection

The two different lentiviral Beclin-1 shRNAs (“-a/-b”) were purchased from Santa Cruz Biotech (sc-29797-V, “Beclin-1 shRNA-a”) and Genepharma (Shanghai, China, “Beclin-1 shRNA-b”), respectively. The two different lentiviral ATG-5 shRNAs were gifts from Dr. Qin Jiang's lab [24]. The three non-overlapping AMPK α 1 shRNAs were designed and synthesized by Genepharma. The lentiviral shRNA (20 μ L/mL medium) was added directly

to the cultured HT-29 cells for 24 hours. Afterwards, cells were cultured in fresh medium for additional 24 hours. Stable cells were selected by puromycin (5.0 $\mu\text{g}/\text{mL}$) for 10–12 days. Control cells were infected with same amount of lentiviral scramble control shRNA (sc-108080-V, Santa Cruz, Beijing, China). Expression of targeted protein was verified by Western blot assay.

HT-29 xenograft

The *in vivo* experimental protocols were approved by the Institutional Animal Care and Use Committee (IACUC) and Ethics Review Board of authors' institutions. Male nude mice (7–8 week age, 18–20 g weight) were injected subcutaneously with HT-29 cells (4×10^6 cells/mice) expressing AMPK α 1 shRNA or scramble non-sense control shRNA. The treatment was initiated 14 days post tumor cell inoculation, when established tumors were around 100 mm^3 in volume. Treatment protocol was described. Tumor volumes were measured every 10 days by the modified ellipsoid formula: $(\pi / 6) \times AB^2$, and A is the longest and B is the shortest perpendicular axis of a tumor mass [10].

Statistical analysis

Data were expressed as mean \pm standard deviation (SD). Data were analyzed by one-way ANOVA followed by a Scheffe's f -test by using SPSS 18.0 software (SPSS Inc., Chicago, IL). Significance was chosen as $p < 0.05$. IC-50 was also calculated by SPSS 18.0.

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CONFLICTS OF INTEREST

The authors have no conflicts of interests.

Authors' contributions

All authors conceived the idea and designed the work, contributed to acquisition of data. CZ, JG, MC and DX wrote the paper; All authors performed the experiments of the paper, CZ, JG MC and DX made the support for all the instruments and reagents.

REFERENCES

1. Schmoll HJ, Stein A. Colorectal cancer in 2013: Towards improved drugs, combinations and patient selection. *Nat Rev Clin Oncol*. 2014; 11:79–80.

2. Kuipers EJ, Rosch T, Bretthauer M. Colorectal cancer screening—optimizing current strategies and new directions. *Nat Rev Clin Oncol*. 2013; 10:130–142.
3. Brouquet A, Nordlinger B. Metastatic colorectal cancer outcome and fatty liver disease. *Nat Rev Gastroenterol Hepatol*. 2013; 10:266–267.
4. Kim JH. Chemotherapy for colorectal cancer in the elderly. *World J Gastroenterol*. 2015; 21:5158–5166.
5. Hubbard JM, Grothey A. Colorectal cancer in 2014: Progress in defining first-line and maintenance therapies. *Nat Rev Clin Oncol*. 2015; 12:73–74.
6. Gonzalez-Pons M, Cruz-Correa M. Colorectal Cancer Biomarkers: Where Are We Now? *Biomed Res Int*. 2015; 2015:149014.
7. Sugarbaker PH. Colorectal Cancer: Prevention and Management of Metastatic Disease. *Biomed Res Int*. 2014; 2014:782890.
8. Palta M, Czito BG, Willett CG. Colorectal cancer: adjuvant chemotherapy for rectal cancer—an unresolved issue. *Nat Rev Clin Oncol*. 2014; 11:182–184.
9. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin*. 2014; 64:9–29.
10. Zhou C, Chen Z, Lu X, Wu H, Yang Q, Xu D. Icaritin activates JNK-dependent mPTP necrosis pathway in colorectal cancer cells. *Tumour Biol*. 2016; 37:3135–3144.
11. Li C, Li Q, Mei Q, Lu T. Pharmacological effects and pharmacokinetic properties of icaritin, the major bioactive component in *Herba Epimedii*. *Life Sci*. 2015; 126:57–68.
12. Huang X, Zhu D, Lou Y. A novel anticancer agent, icaritin, induced cell growth inhibition, G1 arrest and mitochondrial transmembrane potential drop in human prostate carcinoma PC-3 cells. *Eur J Pharmacol*. 2007; 564:26–36.
13. Guo Y, Zhang X, Meng J, Wang ZY. An anticancer agent icaritin induces sustained activation of the extracellular signal-regulated kinase (ERK) pathway and inhibits growth of breast cancer cells. *Eur J Pharmacol*. 2011; 658:114–122.
14. Sun L, Chen W, Qu L, Wu J, Si J. Icaritin reverses multidrug resistance of HepG2/ADR human hepatoma cells via downregulation of MDR1 and Pglycoprotein expression. *Mol Med Rep*. 2013; 8:1883–1887.
15. Zhao H, Guo Y, Li S, Han R, Ying J, Zhu H, Wang Y, Yin L, Han Y, Sun L, Wang Z, Lin Q, Bi X, et al. A novel anticancer agent Icaritin suppresses hepatocellular carcinoma initiation and malignant growth through the IL-6/Jak2/Stat3 pathway. *Oncotarget*. 2015; 6:31927–31943. doi: 10.18632/oncotarget.5578.
16. Tong JS, Zhang QH, Huang X, Fu XQ, Qi ST, Wang YP, Hou Y, Sheng J, Sun QY. Icaritin causes sustained ERK1/2 activation and induces apoptosis in human endometrial cancer cells. *PLoS One*. 2011; 6:e16781.
17. Janku F, McConkey DJ, Hong DS, Kurzrock R. Autophagy as a target for anticancer therapy. *Nat Rev Clin Oncol*. 2011; 8:528–539.
18. Reyjal J, Cormier K, Turcotte S. Autophagy and cell death to target cancer cells: exploiting synthetic lethality as cancer therapies. *Adv Exp Med Biol*. 2014; 772:167–188.

19. Marino G, Niso-Santano M, Baehrecke EH, Kroemer G. Self-consumption: the interplay of autophagy and apoptosis. *Nat Rev Mol Cell Biol.* 2014; 15:81–94.
20. Rubinsztein DC, Codogno P, Levine B. Autophagy modulation as a potential therapeutic target for diverse diseases. *Nat Rev Drug Discov.* 2012; 11:709–730.
21. Li Y, Zhang J, Chen X, Liu T, He W, Chen Y, Zeng X. Molecular machinery of autophagy and its implication in cancer. *Am J Med Sci.* 2012; 343:155–161.
22. Kondo Y, Kanzawa T, Sawaya R, Kondo S. The role of autophagy in cancer development and response to therapy. *Nat Rev Cancer.* 2005; 5:726–734.
23. Zheng B, Mao JH, Qian L, Zhu H, Gu DH, Pan XD, Yi F, Ji DM. Pre-clinical evaluation of AZD-2014, a novel mTORC1/2 dual inhibitor, against renal cell carcinoma. *Cancer Lett.* 2015; 357:468–475.
24. Zhang Q, Yang M, Qu Z, Zhou J, Jiang Q. Autophagy prevention sensitizes AKTi-1/2-induced anti-hepatocellular carcinoma cell activity *in vitro* and *in vivo*. *Biochem Biophys Res Commun.* 2016.
25. Tong Y, Huang H, Pan H. Inhibition of MEK/ERK activation attenuates autophagy and potentiates pemetrexed-induced activity against HepG2 hepatocellular carcinoma cells. *Biochem Biophys Res Commun.* 2015; 456:86–91.
26. Tang JY, Dai T, Zhang H, Xiong WJ, Xu MZ, Wang XJ, Tang QH, Chen B, Xu M. GDC-0980-induced apoptosis is enhanced by autophagy inhibition in human pancreatic cancer cells. *Biochem Biophys Res Commun.* 2014; 453:533–538.
27. Fu LL, Cheng Y, Liu B. Beclin-1: autophagic regulator and therapeutic target in cancer. *Int J Biochem Cell Biol.* 2013; 45:921–924.
28. Zheng B, Zhu H, Gu D, Pan X, Qian L, Xue B, Yang D, Zhou J, Shan Y. MiRNA-30a-mediated autophagy inhibition sensitizes renal cell carcinoma cells to sorafenib. *Biochem Biophys Res Commun.* 2015.
29. Zhen YF, Li ST, Zhu YR, Wang XD, Zhou XZ, Zhu LQ. Identification of DNA-PKcs as a primary resistance factor of salinomycin in osteosarcoma cells. *Oncotarget.* 2016; 7:79417–79427. doi: 10.18632/oncotarget.12712.
30. Papandreou I, Lim AL, Laderoute K, Denko NC. Hypoxia signals autophagy in tumor cells via AMPK activity, independent of HIF-1, BNIP3, and BNIP3L. *Cell Death Differ.* 2008; 15:1572–1581.
31. Mihaylova MM, Shaw RJ. The AMPK signalling pathway coordinates cell growth, autophagy and metabolism. *Nat Cell Biol.* 2011; 13:1016–1023.
32. Khan SH, Kumar R. Role of an intrinsically disordered conformation in AMPK-mediated phosphorylation of ULK1 and regulation of autophagy. *Mol Biosyst.* 2012; 8:91–96.
33. Hu X, Jiang F, Bao Q, Qian H, Fang Q, Shao Z. Compound 13, an alpha1-selective small molecule activator of AMPK, potently inhibits melanoma cell proliferation. *Tumour Biol.* 2016; 37:1071–1078.
34. Guo S, Mao L, Ji F, Wang S, Xie Y, Fei H, Wang XD. Activating AMP-activated protein kinase by an alpha1 selective activator compound 13 attenuates dexamethasone-induced osteoblast cell death. *Biochem Biophys Res Commun.* 2016; 471:545–552.
35. Choi YJ, Park YJ, Park JY, Jeong HO, Kim DH, Ha YM, Kim JM, Song YM, Heo HS, Yu BP, Chun P, Moon HR, Chung HY. Inhibitory effect of mTOR activator MHY1485 on autophagy: suppression of lysosomal fusion. *PLoS One.* 2012; 7:e43418.
36. Boya P, Gonzalez-Polo RA, Casares N, Perfettini JL, Dessen P, Larochette N, Metivier D, Meley D, Souquere S, Yoshimori T, Pierron G, Codogno P, Kroemer G. Inhibition of macroautophagy triggers apoptosis. *Mol Cell Biol.* 2005; 25:1025–1040.
37. Seglen PO, Gordon PB. 3-Methyladenine: specific inhibitor of autophagic/lysosomal protein degradation in isolated rat hepatocytes. *Proc Natl Acad Sci USA.* 1982; 79:1889–1892.
38. Hardie DG, Schaffer BE, Brunet A. AMPK: An Energy-Sensing Pathway with Multiple Inputs and Outputs. *Trends Cell Biol.* 2016; 26:190–201.
39. Egan DF, Shackelford DB, Mihaylova MM, Gelino S, Kohnz RA, Mair W, Vasquez DS, Joshi A, Gwinn DM, Taylor R, Asara JM, Fitzpatrick J, Dillin A, et al. Phosphorylation of ULK1 (hATG1) by AMP-activated protein kinase connects energy sensing to mitophagy. *Science.* 2011; 331:456–461.
40. Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol.* 2011; 13:132–141.
41. Inoki K, Ouyang H, Zhu T, Lindvall C, Wang Y, Zhang X, Yang Q, Bennett C, Harada Y, Stankunas K, Wang CY, He X, MacDougald OA, et al. TSC2 integrates Wnt and energy signals via a coordinated phosphorylation by AMPK and GSK3 to regulate cell growth. *Cell.* 2006; 126:955–968.
42. Inoki K, Zhu T, Guan KL. TSC2 mediates cellular energy response to control cell growth and survival. *Cell.* 2003; 115:577–590.
43. Zhao H, Zhu H, Lin Z, Lin G, Lv G. Compound 13, an alpha1-selective small molecule activator of AMPK, inhibits *Helicobacter pylori*-induced oxidative stresses and gastric epithelial cell apoptosis. *Biochem Biophys Res Commun.* 2015; 463:510–517.
44. Li C, Cui JF, Chen MB, Liu CY, Liu F, Zhang QD, Zou J, Lu PH. The preclinical evaluation of the dual mTORC1/2 inhibitor INK-128 as a potential anti-colorectal cancer agent. *Cancer Biol Ther.* 2015; 16:34–42.
45. Li JP, Huang ZJ, Lu XS, Zhou YC, Shao Y, He XP, Chen SR, Wang DD, Qin LS, Sun WH. Pre-clinical characterization of PKC412, a multi-kinase inhibitor, against colorectal cancer cells. *Oncotarget.* 2016; 7:77815–77824. doi: 10.18632/oncotarget.12802.
46. Zhijun H, Shusheng W, Han M, Jianping L, Li-Sen Q, Dechun L. Pre-clinical characterization of 4SC-202, a novel

- class I HDAC inhibitor, against colorectal cancer cells. *Tumour Biol.* 2016.
47. Yu R, Zhang ZQ, Wang B, Jiang HX, Cheng L, Shen LM. Berberine-induced apoptotic and autophagic death of HepG2 cells requires AMPK activation. *Cancer Cell Int.* 2014; 14:49.
48. Qiu Y, Yu T, Wang W, Pan K, Shi D, Sun H. Curcumin-induced melanoma cell death is associated with mitochondrial permeability transition pore (mPTP) opening. *Biochem Biophys Res Commun.* 2014; 448:15–21.