



Genome Analysis of the Carbapenemand Colistin-Resistant *Escherichia coli* Isolate NRZ14408 Reveals Horizontal Gene Transfer Pathways towards Panresistance and Enhanced Virulence

Linda Falgenhauer,^a Hiren Ghosh,^a Swapnil Doijad,^a Yancheng Yao,^a Boyke Bunk,^b Cathrin Spröer,^b Martin Kaase,^{c*} Rolf Hilker,^{d*} Jörg Overmann,^b Can Imirzalioglu,^a Trinad Chakraborty^a

Institute of Medical Microbiology, Justus Liebig University Giessen, and German Center for Infection Research (DZIF), Partner Site Giessen-Marburg-Langen, Giessen, Germanya; Leibniz Institute DSMZ-German Collection of Microorganisms and Cell Cultures, and German Center for Infection Research (DZIF), Partner site Hannover-Braunschweig, Braunschweig, Germanyb; Department of Medical Microbiology, Ruhr University, Bochum, Germanyc; Institute of Bioinformatics and Systems Biology, Justus Liebig University Giessen, Giessen, Germanyd

KEYWORDS *Escherichia coli*, carbapenem resistance, colistin resistance, *mcr-1*, multidrug resistance, plasmid-mediated resistance

colistin is used as a last-resort antibiotic for treating infections caused by multidrugresistant members of the family *Enterobacteriaceae*, in particular for carbapenemresistant isolates. Resistance to colistin in *Enterobacteriaceae* species has been attributed to chromosomal mutations (1), but recently a plasmid-encoded colistin resistance gene, *mcr-1*, was described in isolates from different sources and in many countries worldwide (2, 3). The emergence of plasmid-encoded colistin resistance is critical, as its transfer could render carbapenem-resistant bacteria pandrug resistant, resulting in virtually untreatable pathogens.

We recently reported the first *mcr-1/bla*_{KPC-2}-encoding carbapenem-resistant *Escherichia coli* isolate (NRZ14408) from a patient with a wound infection (4). In order to characterize the isolate, we sequenced the whole genome to completion (see the supplemental material).

E. coli NRZ14408 harbored a chromosome of 5,344,876 bp with a GC content of 50.65%, four plasmids of about 12 to 238 kbp (p14408_M, p14408_1, p14408_2, and p14408_3; see Table S1 in the supplemental material), and two phages of 41 and 19 kbp (14408_1, 14408_2; see Table S1 in the supplemental material). *E. coli* NRZ14408 is an O7:H6 isolate of multilocus sequence type 362 (ST362) and phylogenetic group D, which is associated with extraintestinal infections (5).

Plasmid p14408_3 harbored no antibiotic resistance genes but did have colicin R genes (*cra*, *cri*, and *crl*) and the *mobC* gene for mobilization of the plasmid. The *mcr-1* gene was flanked by two ISApl1 elements and located on an IncHI2 plasmid (p14408_M) with high similarity to other *mcr-1*-encoding IncHI2 plasmids (pHNSHP45-2 [accession no. KU341381.1], pSA26-MCR-1 [KU743384.1], and pS38 [KX129782]). p14408_M differed from these plasmids by the presence of five additional insertion sequences (ISKpn11, ISKpn12, ISAba14, and two copies of IS629; Fig. 1) and a different set of antibiotic resistance genes (see Table S2 in the supplemental material). Plasmids p14408_1 and p14408_2 harbored the same 36,152-bp region, including nine antibiotic resistance genes (see Table S1 in the supplemental material) and a large portion of the

Accepted manuscript posted online 17 January 2017

Citation Falgenhauer L, Ghosh H, Doijad S, Yao Y, Bunk B, Spröer C, Kaase M, Hilker R, Overmann J, Imirzalioglu C, Chakraborty T. 2017. Genome analysis of the carbapenemand colistin-resistant *Escherichia coli* isolate NRZ14408 reveals horizontal gene transfer pathways towards panresistance and enhanced virulence. Antimicrob Agents Chemother 61:e02359-16. https://doi.org/10.1128/AAC.02359-16.

Copyright © 2017 American Society for Microbiology. All Rights Reserved.

Address correspondence to Trinad Chakraborty, Trinad.chakraborty@mikrobio.med.uni-giessen.de.

* Present address: Martin Kaase, Department for Infection Control and Infectious Diseases, University Medical Center, Göttingen, Germany; Rolf Hilker, BioNTech, Mainz, Germany.

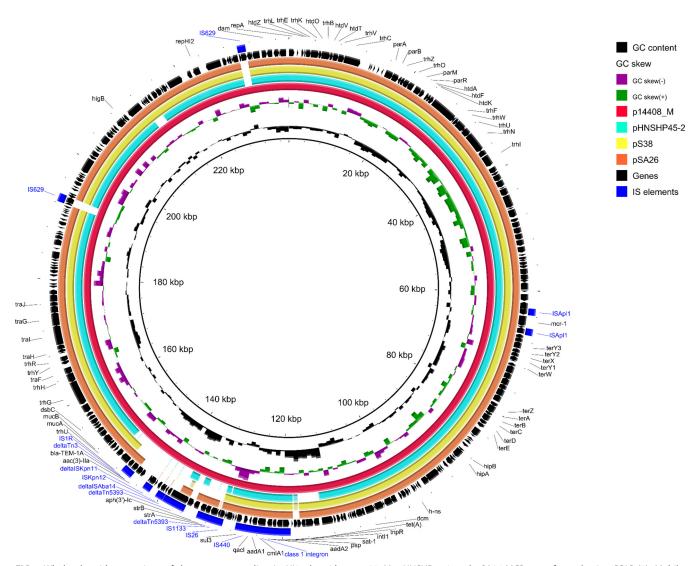


FIG 1 Whole plasmid comparison of three *mcr-1*-encoding IncHI2 plasmids, p14408_M, pHNSHP45-2 and pSA26-MCR-1, performed using BRIG (9). Mobile elements are marked in blue color; genes are marked with black arrows.

IncN plasmid conjugation machinery (see Fig. S2 in the supplemental material). It displayed a unique $bla_{\rm KPC-2}$ cassette [IS26, aac(3)-Ild, ISCfr1, $bla_{\rm KPC-2}$, $bla_{\rm TEM-1B}$, and ISKpn16], present only in the IncN plasmid pCF8698_KPC2, which was isolated during an Enterobacteriaceae $bla_{\rm KPC-2}$ outbreak in southern Hesse in Germany (6). The region was flanked by two IS26 insertion sequences in p14408_1, indicating an IS26-mediated transfer (7, 8).

The genome of *E. coli* NRZ14408 displayed a distinct distribution of antibiotic resistance and virulence genes. Antibiotic resistance genes were located exclusively on plasmids (see Table S1 in the supplemental material), whereas virulence genes were located solely on the chromosome (see Table S3 in the supplemental material). Analysis of other *E. coli* ST362 isolates from 1952 to 2014 (see Table S4 in the supplemental material) harboring virulence determinants or antibiotic resistance genes provided further evidence supporting this observation (see Fig. S2 in the supplemental material). In addition, the total number of virulence genes was generally higher in ST362 isolates harboring antibiotic resistance genes than in antibiotic-susceptible isolates (see Fig. S3 in the supplemental material).

As ST362 is associated with extraintestinal infections, it may be a representative of those *E. coli* clones where increased antibiotic resistance together with enhanced virulence is being selected through the use of injudicious antibiotic regimens.

Accession number(s). The closed genome and plasmid sequences obtained in this study were deposited in the European Nucleotide Archive under accession numbers LT599825 to LT599831.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at https://doi.org/10.1128/AAC.02359-16.

TEXT S1, PDF file, 0.5 MB.

ACKNOWLEDGMENTS

We thank Christina Gerstmann, Natalia Lest, Simone Severitt, and Nicole Heyer for excellent technical assistance.

This study was supported by grants within the framework of the German Center of Infection Research (DZIF) through the German Federal Ministry of Education and Research (BMBF) to T.C. and C.I. (grant 8000 701–3 [HZI]) and to T.C. (grant Tl06.001, 8032808811).

REFERENCES

- Olaitan AO, Morand S, Rolain J-M. 2014. Mechanisms of polymyxin resistance: acquired and intrinsic resistance in bacteria. Front Microbiol 5:643. https://doi.org/10.3389/fmicb.2014.00643.
- Liu Y-Y, Wang Y, Walsh TR, Yi L-X, Zhang R, Spencer J, Doi Y, Tian G, Dong B, Huang X, Yu L-F, Gu D, Ren H, Chen X, Lv L, He D, Zhou H, Liang Z, Liu J-H, Shen J. 2016. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis 16:161–168. https://doi.org/10.1016/S1473-3099(15)00424-7.
- Schwarz S, Johnson AP. 2016. Transferable resistance to colistin: a new but old threat. J Antimicrob Chemother 71:2066–2070. https://doi.org/ 10.1093/jac/dkw274.
- Falgenhauer L, Waezsada S-E, Yao Y, Imirzalioglu C, Käsbohrer A, Roesler U, Michael GB, Schwarz S, Werner G, Kreienbrock L, Chakraborty T. 2016. Colistin resistance gene mcr-1 in extended-spectrum β-lactamase-producing and carbapenemase-producing Gram-negative bacteria in Germany. Lancet Infect Dis 16:282–283. https://doi.org/10.1016/S1473-3099(16)00009-8.
- 5. Clermont O, Bonacorsi S, Bingen E, Bonacorsi P. 2000. Rapid and simple determination of the Escherichia coli phylogenetic group. Appl Environ

- Microbiol 66:4555–4558. https://doi.org/10.1128/AEM.66.10.4555-4558.2000.
- Yao Y, Imirzalioglu C, Hain T, Kaase M, Gatermann S, Exner M, Mielke M, Hauri A, Dragneva Y, Bill R, Wendt C, Wirtz A, Domann E, Chakraborty T. 2014. Complete nucleotide sequence of a Citrobacter freundii plasmid carrying KPC-2 in a unique genetic environment. Genome Announc 2:e01157-14. https://doi.org/10.1128/genomeA.01157-14.
- Harmer CJ, Moran RA, Hall RM. 2014. Movement of IS26-associated antibiotic resistance genes occurs via a translocatable unit that includes a single IS26 and preferentially inserts adjacent to another IS26. mBio 5:e01801-14. https://doi.org/10.1128/mBio.01801-14.
- Ghosh H, Doijad S, Bunk B, Falgenhauer L, Yao Y, Spröer C, Gentil K, Schmiedel J, Imirzalioglu C, Overmann J, Chakraborty T. 2016. Detection of translocatable units in a bla_{CTX-M-15} extended-spectrum β-lactamase-producing ST131 Escherichia coli isolate using a hybrid sequencing approach. Int J Antimicrob Agents 47:245–247. https://doi.org/10.1016/j.ijantimicag.2016.01.003.
- Alikhan N-F, Petty NK, Ben Zakour NL, Beatson SA. 2011. BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. BMC Genomics 12:402. https://doi.org/10.1186/1471-2164-12-402.