RESEARCH

The role of near-patient coeliac serology testing in the follow-up of patients with coeliac disease

D A George,¹ L L Hui,² D Rattehalli,¹ T Lovatt,¹ I Perry,¹ M Green,¹ K Robinson,¹ J R F Walters,² M J Brookes¹

ABSTRACT

Objective This pilot study was undertaken to assess the validity and effectiveness of near-patient coeliac immunological testing, compared to standard laboratory immunological techniques, used in the context of dietician-led coeliac disease follow-up clinics.

Design The study was designed in two phases, each assessing the near-patient test and standard laboratory immunological techniques. Phase 1 analysed stored serum samples; Phase 2 analysed whole blood from patients attending the dietician-led coeliac disease clinics.

Setting Patients were recruited from New Cross Hospital, Wolverhampton (n=50), and Imperial College London (n=30), between March 2010 and February 2011.

Patients Those with a diagnosis of coeliac disease for greater than 12 months attending dietician-led coeliac disease clinics.

Interventions In addition to whole blood taken for routine analysis, patients required a capillary finger-prick blood sample.

Main outcome measure To determine if the whole blood and serum near-patient test results were in correlation with outcomes of standard laboratory evaluation.

Results Phase 1 demonstrated that the nearpatient serum test had a sensitivity of 93.5% (95% CI 0.79% to 0.98%), specificity of 94.9% (0.83% to 0.99%), when compared to standard laboratory ELISA. Phase 2, involving patients whole blood, had a sensitivity of 77.8% (0.45% to 0.93%), and specificity of 100% (0.94% to 1%).

Conclusions This pilot study has demonstrated that there appears to be a role for near-patient testing in coeliac disease, but further studies are recommended.

INTRODUCTION

Coeliac disease is a common autoimmune disorder precipitated by the ingestion of

wheat gluten and similar proteins in barley and rye. Population studies suggest that this condition affects around 1% of Europeans.¹ The disease is commonly associated with a number of autoimmune disorders through its genetic linkage with the HLA-DQ2 and -DQ8 haplotypes.² The diagnosis can be made clinically where patients may present with a plethora of features, including lethargy, tiredness, abdominal bloating, or diarrhoea. Only a minority of patients however, will present with classical malabsorptive problems, including steatorrhoea and/or weight loss.³

The investigation of coeliac disease has been aided by the increasing use and reliability of laboratory-based serological autoantibody testing. In 1984, IgA Endomysial antibodies (EmAs) were first described directed against the intermyofibril substance of smooth muscle, suggesting a high specificity for patients with coeliac disease, and a correlation between EmAs titre and severity of small bowel histology.¹ Further investigations have confirmed that EmAs were superior to serum antigiadin antibody and antireticulin antibody, previously used for diagnostic assessment of patients with symptoms consistent with coeliac disease.^{2–4} In 1997, Dieterich *et al*⁵ identified tissue transglutaminase (tTGA) as the antigen against which EmA was directed, and since this, tTGA testing has followed.

Near-patient testing has recently been introduced to allow for bedside testing for tTGA, using immunochromatographic assays, and these have demonstrated good efficacy when compared with serum antibody testing in the diagnosis of coeliac disease.^{6 7} The sensitivities of near-patient testing kits compared with laboratory

¹Department of Gastroenterology, New Cross Hospital, Wolverhampton, UK ²Department of Gastroenterology, Imperial College London, London, UK

Correspondence to

Dr Matthew Brookes, Department of Gastroenterology, New Cross Hospital, Wolverhampton Road, Wolverhampton WV10 0QP, UK; m.j.brookes@bham.ac.uk

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To cite: George DA, Hui LL, Rattehalli D, *et al. Frontline Gastroenterology* 2014;**5**: 20–25. ELISAs, have been reported to be between 96% and 100% with specificities of between 95% and 100%.⁶ When these were tested in 51 patients with untreated known coeliac disease and 36 controls they demonstrated a similar efficacy to the laboratory serum EmA and tTGA with all tests having 100% specificity.⁸

The ease of use of these near-patient blood droplet kits has led to an enthusiastic response for investigating potential cases of coeliac disease by paramedical teams including district nurses.⁹ However, further data is required to confirm its full efficacy in this setting, and there are no studies to look at the role of near-patient testing in the follow-up of treated cases of coeliac disease.

Dietician-led follow-up of patients with coeliac disease now involves a full dietary history and serological assessment to determine a patient's compliance with a gluten-free diet (GFD). The subsequent laboratory-based immunological testing can then take up to 1 week to be made available (any elevation in serum antibody levels signifies a patient's noncompliance). This delay in the availability of laboratorybased assays often means that a patient's dietary assessment in clinic is 'blind' to their immunology, which is inefficient and can waste the patient's and the dietician's time. In the current clinical setting, if the laboratory immunology is positive, the dietician often has to make a further follow-up appointment to reassess the GFD.

If the dietician had access to a simple, efficient and reliable near-patient immunological test, the assessment of the patient's GFD could be more thorough, and the patient could be informed of their results at the same clinic. Although this technology is available and has been investigated in the diagnosis of coeliac disease, it has not yet been evaluated in this setting.

The aim of this pilot study is to assess the validity and effectiveness of near-patient immunological testing used in the context of dietician-led coeliac disease clinics, compared to standard laboratory immunological evaluation. Our hypothesis is that the results of the near-patient testing will be similar to the outcomes of standard laboratory evaluation, in the follow-up of patients with coeliac disease.

METHODS

The study was undertaken in two phases: (1) laboratory stored serum samples and (2) patients who were recruited from the dietician-led coeliac disease outpatient clinics.

Patients

Patients were identified at Imperial College London, between 2 March and 11 May 2010, and at New Cross Hospital, Wolverhampton, between 14 September 2010 and 1 February 2011. Patients were screened and recruited from the dietician-led coeliac disease outpatient clinics. The group involved patients over the age of 16 years with an established histological diagnosis of coeliac disease, based on the modified Marsh criteria for the grading of villus atrophy,¹⁰ who had received treatment for at least 12 months. Those patients fulfilling the inclusion and exclusion criteria (age <16 years, pregnant, known existing liver disease (liver fibrosis or above), rheumatoid arthritis, psoriatic arthritis, ankylosing spondylitis, end-stage heart failure) were included.

In this prospective evaluation, 80 consecutive patients (median age 58 years, range 18–89) with previously diagnosed biopsy-proven coeliac disease and known serum total IgA levels took part. The time since diagnosis, and presumed GFD was from 12 months to 50 years (median: 2.17 years).

Clinical data collection

Data obtained during the consultation was a dietary evaluation, including whether a patient's GFD is assumed compliant or non-compliant (if patient openly/ obviously not compliant, has symptoms, or is known from past medical history), current and previous symptoms, known complications of coeliac disease, and body mass index. As per normal protocol, patients who were deemed symptomatic, non-compliant or had positive laboratory serology were referred for further endoscopic and histological assessment.

Near-patient testing

Medical personnel trained in performing the nearpatient test included research nurses, dieticians and a medical student. After receiving the patient's consent, a capillary finger prick (whole) blood sample was used to test for tTGA (IgG and IgA) using the 'Biohit Coeliac Quick Test' near-patient kit as per manufacturer's instructions. This test detects IgA, IgG and IgM classes of antibodies against tTGA, and is specific for whole blood requiring no laboratory expertise in its execution. All reagents were available within the single-use kit. The result of this test was read on site and available within 10 min. A positive test led to further dietary assessments. The positivity can be further divided into a positive or strong positive result, depending upon the intensity of near-patient test indicator. In order to minimise interobserver variation, two independent researchers, in a blinded fashion, read all near-patient testing samples from the patient cohort.

Blood samples

During Phase 1, stored serum samples were analysed using the near-patient test, and analysed for antibody levels using standard ELISA techniques.

In addition to the capillary finger prick, blood samples taken from patients in Phase 2, 20 mL of whole blood was drawn and analysed for full blood count, serum liver function, calcium profile, immunoglobulins vitamin B12, folate and ferritin, using standard laboratory techniques.

Serum antibody measurements

The IgA class serum antibodies against TG2 were measured with human recombinant TG2 using the Celikey (Pharmacia Diagnostics, Freiburg, Germany) ELISA, according to the manufacturer's instructions. Cut-off for positivity was 5 U/mL. EMA was determined on monkey oesophagus sections by indirect immunofluorescence as described elsewhere.¹¹ Samples reactive at a serum dilution of 1:2.5 were considered positive.

Statistics

Data from each phase was analysed to assess the sensitivity and specificity (Phase 1 stored serum samples; Phase 2 whole blood samples) between the laboratorybased ELISA techniques and the near-patient test using Fishers Exact test using SPSS (V16) for Windows. Positive and negative predictive values were determined for the respective assays.

RESULTS

Serum ELISA and near-patient testing (Phase 1)

Phase 1 of the study analysed a total of 70 serum samples. The established laboratory ELISA techniques identified 31 samples (44.3%) to be positive for IgA tTGA, IgG tTGA, EmA or immunoglobulin levels. The near-patient test similarly identified 31 samples to be positive for elevated levels of tTGA (table 1). Two false negative near-patient tests were seen in patients with low tTGA titres (15 and 8 U/mL). These cases were not shown to have potential confounding diagnoses, such as raised IgA, liver disease or abnormal biopsy findings.

Whole blood ELISA and near-patient testing (Phase 2)

Phase 2 prospectively identified 50 patients at the New Cross Hospital and 30 patients from Imperial College London (n=80). The majority of the patients were women (72.5%), the mean age of 57.4 years (SD 14.52) with a mean age at diagnosis of 42.9 years (SD 19.3).

Of the near-patient test true positive results (n=7), stronger positives accounted for higher levels of

laboratory tTGA levels (53, 59 U/mL, and three >80 U/mL), whereas, weaker positives accounted for lower levels (20 U/mL, and 50 U/mL). However, false near-patient test negative results (n=2) were found in patients whose laboratory levels were 20 U/mL and 35 U/mL.

Further details on patients recruited from New Cross Hospital only

The majority of patients were women (74.0%), the mean age was 55 years, a mean age at diagnosis of 46.4 years with the mean time since diagnosis was 9.17 years (median 4.5 years). Prior to the blood test results, only 35 patients (70%) were assumed compliant with their GFD, whereas the laboratory findings confirmed that 46 patients (92.0) were compliant.

A single false negative result was seen in a patient with a high tTGA titre of 79.0 U/mL. Three patients were true positives with tTGA titres of 50.0 U/mL and >80.0 U/mL in two patients. One of these cases was assumed to have been compliant with a GFD at the clinic assessment. The mean age of these true positive patients was 44 years and they had been diagnosed with coeliac disease for an average of 16 months. At their most recent consultation, the symptoms were mainly of fatigue (66.6%) and bone/ joint pains (66.6%). Blood levels showed a low level of haemoglobin in one patient (9.6 g/dL), and ferritin in two (3.9 ng/mL and 14.3 ng/mL), with normal ranges for platelets, White Cell Count (WCC), Mean Corpuscular Volume (MCV), vitamin B12 and folate levels.

Forty-six patients (92%) were true negatives with tTGA levels ranging from 0.1 U/mL to 30.0 U/mL (table 2). The mean age was 57 years, mean age at diagnosis 47.3 years, and have had coeliac disease for an average of 9.6 years.

The majority of patients were assumed compliant (34; 68%), eight assumed non-compliant, four however, were unknown. The main prediagnostic symptoms of this group were abdominal bloating, abdominal pain and fatigue (table 3).

Table 1	Statistical significance of nea	r-patient and corresponding	laboratory test results,	for stored serum and whole blood samples	

	Laboratory testing					
	Stored serum samples	s (Phase 1)	Whole blood samples (Phase 2)			
				ELISA tTG		
	ELISA tTGA (n=70)	lgA tTGA (n=50)	EMA (n=50)	Both sites (n=80)	New Cross Hospital (n=50)	
Sensitivity (%)	93.5	92.6	93.5	77.8	75.0	
Specificity (%)	94.9	95.7	94.7	100	100	
PPV (%)	93.5	96.2	96.7	100	100	
NPV (%)	94.9	91.7	90	97.3	97.9	
p Value	<0.05	<0.05	<0.05	<0.05	<0.01	

p Value: statistically significant if p<0.05.

EMA, endomysial antibody; IgA tTG, tissue transglutaminase antibody; NPV, negative predictive value; PPV, positive predictive value.

Table 2	Levels of	tTGA in	true	negative	results	(n=46)

tTGA (U/mL)	Frequency (n)	Percentage
<1.0	17	37.0
≥1.0-<2.0	6	13.0
≥2.0-<4.0	7	15.2
≥4.0-<8.0	9	19.6
≥8.0	7	15.2

Blood results for each patient presenting to New Cross Hospital were available within 1 week of the consultation. The majority of the blood levels were within the normal range, however, 26.1% of true negatives had deficiencies in ferritin and 2.2% of folate (table 4).

All available data on patients recruited from Imperial College London are shown in table 1.

DISCUSSION

We have demonstrated the high sensitivity and specificity of a near-patient test with an efficacy that is comparable to established laboratory immunology, using the patients' whole blood and stored serum in the follow-up of patients with coeliac disease.

Current management of patients with established diagnosis of coeliac disease, highlights the importance of adherence to the GFD and of regular dietetics assessment. Compliance with the GFD involves a dietary history and the measurement of serum laboratory antibodies, but a repeat duodenal biopsy is not routinely advocated.¹² ¹³ Indeed, it is now commonplace for coeliac disease patients on a GFD with a complete resolution of symptoms and negative follow-up laboratory tTGA to not require any further histological assessment, though practice is varied due to a lack of evidence.

Given that histological surveillance is becoming less commonplace, this will emphasise the importance of accurate dietary and serological assessment follow-up of coeliac disease.¹⁴ Laboratory serology used to monitor dietary compliance is of no use in assessing a patient's compliance in the immediate clinical setting as results are often delayed.

Use of the near-patient test in the dietician-led clinics was neither invasive nor inconvenient for the patient, allowing real-time assessment of IgA, IgG and IgM classes of antibodies against tTG. The 'Biohit Coeliac Quick Test' near-patient kit is simple to use, requiring minimal training in its use and recognition of a positive or negative reading. During the study, a second opinion was always sought if a practitioner was unclear of the reading. This can be translated to the established dietician clinic, as a practitioner taking bloods is present for confirmation. By comparison, laboratory readings are calibrated to factory settings, the near-patient test is dependent upon the practitioner's eyesight and recognition of a positive reading.

A patient was assumed compliant to their GFD by the dietician, based upon their clinical experience, patient's current symptoms and previous encounters with the patient. The dieticians state that the immediate availability of the test helped to direct their consultation in only 20% of the time. However, our findings have shown that many patients thought to have been non-compliant were in fact confirmed compliant by the near-patient test and laboratory findings. Therefore, the availability of immediate tTGA levels, and therefore confirmation of compliance, would prevent unnecessary discussions relating to diet and possible further interventions that may be needed.

The near-patient test was further analysed against the stored serum of patients with known liver cirrhosis and myeloma, without a history of coeliac disease. The IgA levels raised in both of these conditions did not lead to (false) positive results. These findings suggest that the specificity of the test is not affected by polyclonally raised IgA. Further analysis would be needed to evaluate the sensitivity of this test in coeliac patients with IgA deficiency.

A strong, positive, near-patient test result is related to higher levels of tTGA, as well as higher rates of

Table 3 Prediagnostic and current symptoms in true negative patients presenting at New Cross Hospital (n=46)

Symptoms	Prediagnostic	(n, %)	Current symptoms (n, %)		
Abdominal bloating	27	58.7	13	28.3	
Abdominal pain	25	54.3	8	17.4	
Chronic diarrhoea	20	43.5	3	6.5	
Vomiting	2	4.3	1	2.2	
Constipation	12	26.1	11	23.9	
Pale, foul-smelling, or fatty stool	21	45.7	6	13.0	
Weight loss	19	41.3	4	8.7	
Fatigue	29	63.0	14	30.4	
Bone or joint pain	9	19.6	18	39.1	
Arthritis	4	8.7	5	10.9	
Depression or anxiety	14	30.4	7	15.2	
Other	6	13.0	0	_	

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Table 4	Blood levels for true negat	ives and true positives	presenting to New	Cross Hospital

	Normal range	True ne	gatives (n=46)		True positives (n=3)		
Values		Below range (n, %)		Mean	Below range (n, %)		Mean
Hb (g/dL)	13–18 (male) 11.5–16 (female)	0	-	13.8	1	33.3	13.26
Plt (×10 ⁹ /L)	150-450	0	_	294	0	_	338
WCC (×10 ⁹ /L)	4–11	2	4.3	5.9	0	_	6.77
MCV (fL)	80–100	1	2.2	89.35	0	_	79.3
Vitamin B12 (ng/L)	>150	0	_	499.5	0	_	599.3
Folate (µg/L)	3–30	1	2.2	9.03	0	_	13.28
Ferritin (µg/L)	28–365	12	26.1	65.1	2	66.7	17.17

Hb, haemoglobin; Plt, platelet; WCC, White Cell Count; MCV, Mean Corpuscular Volume.

symptoms and deficiencies, such as haemoglobin and ferritin (66.7%). However, it was shown that a large proportion of true negative patients, despite having been compliant with their GFD, were still deficient in ferritin (26.1%) and remained symptomatic of coeliac disease (although the frequency of symptoms had halved since their initial diagnosis). This may reflect an irregularity in the correlation between IgA levels, compliance with GFD, and subsequent blood deficiencies.

Unfortunately in this study, we did not examine patient serum with the near-patient test in Phase 2. It is likely that the small differences in the sensitivity and specificity seen between Phases 1 and 2 are occurring as a result of the higher concentrations of antibody present in serum rather than whole blood. In this study, it does obviously lead to differences in the statistical results, however, in wider practice, this difference is likely to be irrelevant, as the near-patient testing will only be based on whole blood sampling taken within the clinic environment. Furthermore, this project is a small pilot study and, as such, the low number of tTG positivity may have significantly influenced the statistical outcomes of this work.

Despite these shortcomings, we have demonstrated encouraging findings, and would use the results to inform the design of large controlled trials to determine the cost effectiveness of replacing current dietician-led clinics and laboratory immunological assessments with a 'one-stop' clinic assessment using the near-patient test. This will allow for a more streamlined, efficient and cost-effective clinic to be run. It would also improve our service to patients, allowing them to receive their immunology results immediately, and preventing those with positive results having to be brought back to the clinic another time to re-evaluate their diet. As this is such a common disorder, it will have a large impact on practice, reducing both the need for further clinic assessment, and reducing the need for laboratory blood samples, thus, improving the patient experience and reducing clinical costs as well.

What is already known on this topic

Near-patient immunological testing has recently been introduced to allow for bedside testing for tTGA levels in the diagnosis of coeliac disease. Results have shown a high sensitivity (96-100%) and specificity (95-100%) compared to standard laboratory immunological evaluation.

What this study adds

This study assesses the validity and effectiveness of near-patient testing, in the context of a dietician-led follow up clinic of treated cases of coeliac disease. Utilising the near-patient test on stored serum, there was a sensitivity of 93.5% and specificity of 94.9%, when compared to standard laboratory immunological evaluation. In the clinic, patient's whole blood was shown to have a sensitivity of 77.8%, and specificity of 100%.

How might it impact on clinical practice in the foreseeable future

We would utilise these encouraging results to inform the design of large controlled trials, to determine the cost-effectiveness of replacing current dietician-led clinics and laboratory immunological assessments, with a "one-stop" clinic assessment. The ultimate aim would be to enable a streamlined, efficient and cost-effective clinic to be run, and improve our service to patients by allowing them to receive their immunology results immediately, and reduce the need for laboratory blood samples. **Acknowledgements** We would like to thank Core and Dr Falk Pharma for their kind bursary, which helped fund the study.

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