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Genetics of Human Lupus Nephritis

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Abstract

Systemic lupus erythematosus (SLE) is an inflammatory autoimmune disease characterized by immune complex formation with multi-organ manifestations. Lupus nephritis (LN) is one of the most severe types of organ damage in SLE, and it clearly contributes to increased morbidity and mortality due to SLE. LN occurs more frequently and is more severe in non-European ancestral backgrounds, although the cause of this disparity remains largely unknown. Genetic factors play an important role in the pathogenesis of SLE. Although many SLE susceptibility genes have been identified, the genetic basis of LN is not as well understood. While some of the established general SLE susceptibility genes are associated with LN, recent discoveries highlight a number of genes with renal functions that are specifically associated with LN. Some of these genes associated with LN help to explain the disparity in the prevalence of nephritis between individuals with SLE, and also partially explain differences in LN between ancestral backgrounds. Moreover, not only the gene mutations, but also post-translational modifications seem to play important roles in the pathogenesis of LN. Overall it seems likely that a combination of general SLE susceptibility genes will outline the genetic contribution to LN and describe possible roles of LN susceptibility genes.

Keywords

Systemic lupus erythematosus; Lupus nephritis; Genetics; APOL1; PDGFR; HAS2

1. Introduction

Systemic lupus erythematosus (SLE) is an inflammatory autoimmune disease characterized by multi-organ system involvement. It affects women 9 times more often than men, and often is diagnosed in early adulthood, frequently during the childbearing years.

Genetic and environmental factors play important roles in the pathogenesis of SLE [1]. The incidence and prevalence of SLE is higher in non-European ancestry, especially in African ancestry. The severity of SLE also varies among the ethnic groups, being more severe in

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non-European populations [2–4]. Supporting a genetic contribution to disease, monozygotic twins are much more likely to be concordant for SLE than dizygotic twins (concordance rate 24% and 2%, respectively) [5]. Familial aggregation of SLE has also been clearly documented, and most pedigrees support a non-Mendelian complex inheritance [6]. These facts strongly support notion of a polygenic genetic contribution to SLE pathogenesis.

Among the various organ manifestations of SLE, lupus nephritis (LN) is one of the most feared, potentially resulting in organ damage and renal insufficiency that results in poor clinical outcomes despite recent improvements in SLE treatment.

Genome-Wide-Association Studies (GWAS) and candidate gene association studies have revealed numerous risk genes for SLE, including loci which contain genes that function in the innate and adaptive immune system [7, 8]. Some of these genes are also closely associated with LN. However, most of these previous studies were not primarily focused on the nephritis phenotype, and less is known about which genes predispose to LN. Some recent studies which have focused on identifying the genes specifically responsible for LN have identified intrarenal genes that are associated with LN, but not associated with the susceptibility of SLE in general [9, 10]. While the exact functional mechanisms of these renal-related candidate genes remains unclear, it seems that the genetic basis of LN involves a combination of general SLE susceptibility genes which function in the immune system and genes which are more renal-specific that predispose specifically to LN (Figure 1). In this review, we outline the genetic contribution to LN and also discuss possible roles of each candidate genes in the pathogenesis of LN.

2. Difference in incidence rate and severity of LN between ethnicities

Among various organ manifestations of SLE, LN is one of the most severe, and can progress to end-stage renal disease (ESRD) leading to increased morbidity and mortality. LN affects about 40% of SLE patients throughout their lifetime [11]. Despite recent advances in treatment, patients with LN still have higher morbidity and mortality compared with those without LN [11-13]. It is well known that African, Asian, and Hispanic populations are more likely to develop LN as compared to European ancestry [3, 11, 14–16]. There are also differences in severity of LN among the ethnicities, with LN being more severe in non-European populations [3, 11]. This disparity between ancestral backgrounds could be related to genetic or environmental factors [17]. To investigate the importance of genetic factors, Sanchez et al. conducted a study evaluating the genetic impact of the proportion of Amerindian vs. European genetic ancestry in admixed populations living in South America. This is an informative way to study the contribution of genetics to LN with some control over environment, as different individuals living in the same population and same location will have different proportional genetic ancestry. This study revealed that an increased proportion of Amerindian genetic ancestry correlated with increased risk of developing LN [2]. Another study demonstrated familial clustering of ESRD African ancestry SLE patients with LN, suggesting shared genetic factors contributing to LN in these families [18]. These studies support the idea that genetic factors contribute to the pathogenesis of LN.

3. LN risk genes and possible functions

Although GWAS studies have identified numerous SLE risk genes, thus far fewer genes have been identified as predisposing to the LN phenotype. Most of the reported risk genes for LN have been studied in candidate gene studies, after being identified as SLE risk factors in GWAS studies [7, 8], with the hypothesis that SLE susceptibility genes may also be LN risk genes. More recently, studies have examined the LN phenotype directly, comparing patients that have developed LN to those patients who have not developed LN.

3.1. SLE susceptibility genes associated with LN

3.1.1. HLA-DR—The major histocompatibility complex (MHC) region which contains human leukocyte antigen (HLA) genes is located on human chromosome 6. This locus contains more than 200 genes, many of which function in the immune system. The HLA Class II region contains HLA-DR, -DQ, and –DP genes. They are highly expressed on antigen-presenting cells and are important for the activation of CD4+ T cells and other immune responses. Polymorphisms in the HLA region were among the first to be discovered as SLE risk factors, and this locus remains the strongest common genetic risk factor for SLE [19].

Among the HLA alleles, HLA-DR2 and HLA-DR3 are the most established SLE susceptibility loci proven in previous studies [7, 20]. Large phenotypic association studies also revealed HLA-DR3 [9, 21, 22] as a susceptibility gene for LN in European ancestry. Additionally, meta-analysis of multiethnic case-control studies identified 7 HLA Class II candidate alleles in the association of LN. HLA-DR15 (part of the older HLA-DR2 serotype group) and -DR3 were found as risk alleles for LN, and interestingly -DR4 and -DR11 were associated with protection from LN [23]. Despite the strong association of HLA alleles with both SLE and LN, the actual mechanism of how these HLA-DR haplotypes predispose to LN has not been elucidated. Some studies support the idea that particular MHC molecules may permit presentation of self peptides in an abnormal way. One study identified a dominant HLA-DR3 restricted T cell epitope on the SmD autoantigen, and those mimicry peptides were able to activate T cells that were reactive to this epitope [24]. This HLA restricted autoreactive T cell activation may then lead to stimulation of autoreactive B cells, resulting in the production of autoantibodies. Another group showed the association of HLA-DR3 and the presence of anti-Ro autoantibodies in SLE patients, which may also support the above mechanism [20].

3.1.2. ITGAM—The ITGAM gene is located on human chromosome 16. It encodes CD11b-integrin (alpha M) which is a subunit composing alpha M beta-2 integrin (also called complement receptor 3 or Mac-1). Mac-1 is highly expressed in granulocytes, macrophages and dendritic cells. Complement (e.g. iC3b) is one of the ligands of Mac-1, and iC3b-coated particles such as apoptotic cells will induce phagocytosis by phagocytes via engagement to Mac-1. Mac-1 also controls leukocyte migration to inflammation site and facilitate adherence to vascular endothelium[25].

A non-synonymous SNP (rs1143679) in the ITGAM gene results in a change of arginine (R) to histidine (H) at position 77 (R77H). Human primary monocytes carrying 77H allele

showed reduced ability of phagocytosis to iC3b-opsonized particles [26]. Those monocytes with 77H allele also failed to repress the production of inflammatory cytokines such as interleukin (IL)-1b, IL-6 and tumor necrosis factor alpha, when stimulated with Toll-like receptor (TLR) 7/8 ligands. Association of the ITGAM gene with SLE susceptibility has been well established in several studies [7, 8, 27, 28]. Associations of ITGAM variants with renal involvement in SLE patients has also been shown in several studies [9, 21, 29–31]. Yang et al. reported higher prevalence of LN in SLE patients carrying the R77H variant in Hong Kong Chinese SLE patients, as compared to SLE patients without LN (OR = 3.35, p =0.029). Kim-Howard et al also demonstrated association of the R77H variant with LN in large European ancestry SLE cohort (OR = 2.15, p = 4.69×10^{-22}). Recently, two large studies, primarily designed to identify the association between genetic variants and LN, have also confirmed ITGAM as a susceptibility gene for LN [9, 31]. Collectively, defective function of the complement receptor due to these variants may result in incomplete clearance of glomerular deposits, and facilitate a pro-inflammatory environment in the kidney which leads to LN. Interestingly, while many of the other SLE susceptibility genes are shared with other autoimmune diseases, ITGAM seems to be an SLE-specific genetic risk factor (with the possible exception of systemic sclerosis)[32]. This could indicate a specific role for ITGAM in the pathogenesis of SLE and LN, rather than a more general proinflammatory role or global alteration in phagocytic function, but further investigation is needed.

3.1.3. FCGR—The FCGR gene locus is located on human chromosome 1 and encodes Fc gamma receptors (FcgRs). One key role of FcgRs is to remove immune complexes (ICs) [33], and FcgR2A and FcgR3A are expressed on antigen presenting cells (e.g. macrophage and dendritic cells). Defective clearance of apoptotic cells is identified in SLE patients, and is thought to contribute to disease pathogenesis [34, 35]. Failure of appropriate phagocytosis will allow apoptotic cells to proceed to secondary necrosis resulting in the release of nuclear self-antigens, which should lead to greater IC formation when combined with SLE-associated anti-nuclear antibodies. There is one high-affinity receptor (e.g. FcgR1) and two low-affinity receptors (e.g. FcgR2A and FcgR3) which are activating-type FcgRs, and there is one inhibitory-type FcgR known as FcgR2b. Each of these receptors has a different binding affinity to IgG subclasses and are present in most of myeloid cells [36]. Genetic associations between variants of low-affinity FcgR (i.e. FcgR2A and FcgR3A) and SLE have been reported [7, 37]. FcgR2A and FcgR3A are also associated with LN which have been proven in several individual studies among multiple ethnicities [38–43].

A non-synonymous SNP (rs1801274) of FCGR2A encoding a G-to-A allele change at position 131 results in amino-acid change from arginine (R) to histidine (H). These alleles, 131R and 131H, have different binding capacity for IgG2. Cells which are homozygous for the histidine allele (131H/H) strongly bind IgG2, and cells that are homozygous for the arginine allele (131R/R) show a reduction in binding capacity for IgG2 opsonized ICs (131H/R has intermediate binding capacity) [44]. In a study of anti-C1q antibody positive SLE patients, presence of FCGR2A 131R was associated in the increased risk for LN [45]. Zuniga et al. identified that the frequency of FCGR2A 131R allele was significantly higher in LN patients with intense IgG2 deposition in the kidney [46]. Also, the frequency of

homozygous FCGR2A 131R/R was increased in the LN population as compared to nonproliferative idiopathic glomerulonephritis[39]. These data suggest that the FCGR2A 131R allele results in dysfunctional clearance of IgG2-opsonized ICs, which could allow greater IC deposition in the kidney and increased inflammation.

Also, a non-synonymous SNP (rs396991) of FCGR3A encoding T-to-G allele change at nucleotide 559 results in an amino-acid conversion from phenylalanine (F) to valine (V) [33]. FcgR3A-158V allele binds to IgG1, IgG3 and IgG4 in higher affinity compared to the 158F allele [33]. Several reports including meta-analysis article revealed that FCGR3A 158F allele is strongly associated with LN [40–43]. Although, in the recent large multiethnic lupus cohort, patients with SLE who were homozygous for FCGR3A 158V/V progressed to ESRD more frequently than compared to those with the 158F/V or 158F/F genotypes (21.9% versus 7.5%, p=0.0175) [47]. Low IgG binding capacity FCGR3A (i.e. 158V/V) may be needed to initiate LN, while high IgG binding FCGR3A (i.e. 158V/V) may play an important role in accelerating the inflammation of LN towards ESRD to explain the above discrepancy.

3.1.4. IRF5—The IRF5 gene is located on human chromosome 7 and encodes interferon regulatory factor 5 (IRF5), which is a transcription factor which induces the expression of inflammatory genes and type I interferon (IFN)-related genes downstream of TLR engagement. The genetic association between IRF5 and SLE was first reported by Sigurdsson et al. in 2005 [48]. Further studies demonstrated that a haplotype which contains multiple functional genetic elements was strongly associated with SLE [49, 50]. Type I interferon is known to play a critical role in the pathogenesis of SLE [51] and risk haplotype of IRF5 was associated with higher IFN alpha activity in the presence with auto-antibodies (i.e. anti-RNA binding protein or anti-double-stranded DNA autoantibodies) [52]. SLE patients carrying the risk haplotype of IRF5 also have elevated expression of IRF5 protein [53]. Moreover, the IRF5 gene has been associated with autoantibody formation in SLE patients and healthy individuals [54, 55]. This same risk haplotype was also identified as a LN susceptibility locus strongly associated with proliferative nephritis and severe renal insufficiency in patients with SLE [21]. While it is not known exactly how IRF5 genotype contributes to nephritis, this gene impacts on both autoantibody formation and type I IFN, and these factors may contribute.

3.1.5. TNIP1—The TNIP1 gene is located on human chromosome 5 and encodes the protein A20 binding and inhibitor of NF-kappaB 1 (ABIN1). The ubiquitin-binding protein ABIN1 is an adaptor protein of the ABIN family that cooperates with A20/TNFAIP3 to limit inflammatory responses. A20 is a ubiquitin-editing protein that inhibits transcription factor NF-kappaB [56, 57] which is a key player of inflammation in immune system.

In mice, Caster et al. reported that the knockin mouse expressing ABIN1[D485N], an inactive form of ABIN1, develops glomerulonephritis similar to class III and IV in human LN with enhanced activation of NF-kappaB and MAPK (mitogen-activated protein kinase) [58].

In humans, variants in TNIP1 gene were detected as susceptibility loci for SLE in multiethnic populations (rs7708392 and rs10036748) [59–61]. TNIP1 variants were also identified as LN susceptibility loci in Swedish Caucasian (rs7708392) [21], European American population (rs7708392) [58] and African American population (rs4958881) [58], supporting the important role of ABIN1 in the pathogenesis of LN observed in mice experiment. Another study from Japan also revealed the association with LN in Japanese population (rs7708392) [62]. Two partial loss-of-function risk haplotypes of TNIP1, accompanied with reduced ABIN1 protein expression was identified in SLE patients among European population [63]. Therefore, TNIP1 variants may partly be involved in the pathogenesis of LN by reducing ABIN1 protein expression and causing the dysregulation of NF-kappaB pathways.

3.1.6. STAT4—The STAT4 gene is located on human chromosome 2 and encodes signal transducer and activators of transcription-4 (STAT4). STAT4 is known as a susceptibility gene for SLE [64]. In 2008, large study conducted by Taylor et al. showed the associations between STAT4 (rs7574865) and the presence of LN and anti-dsDNA antibody in European ancestry SLE cohorts [65]. Additionally, a recent study in 2013 further confirmed the association of STAT4 with the development of proliferative nephritis, a severe subtype of LN [21]. A report from Japan confirmed the association of STAT4 (rs7574865) in both SLE patients with LN and in those without in this population [66]. The association was stronger in the SLE patients with LN and anti-dsDNA antibody, but small sample size precluded strong statistical significance for this comparison [66].

STAT4 is a transcriptional factor which is important for T cell signaling. STAT4 is activated by interleukin (IL)-12, leading naïve CD4+ T cells to differentiate to Th1 and Th17 cells which then play an important roles in inflammation and immune responses. IL-17 producing T cells are present in the kidney of LN patients, suggesting that these T cells play a role in the pathogenesis of LN [67]. STAT4 is also activated by type I IFN [68], and SLE patients carrying the STAT4 risk allele rs7574865 have an increased sensitivity to type I IFN supported by the increased expression of IFN inducible genes [69]. The functions of STAT4 in T cell differentiation, the type I IFN pathway, and the association of this risk allele with anti-dsDNA antibody formation could all play a role in considering how this risk allele predisposes to LN.

3.1.7. TNFSF4—The TNFSF4 gene is located on human chromosome 1 and encodes the TNF Super Family 4 (TNFSF4) protein. TNFSF4, also known as OX40L or CD252 is expressed on antigen presenting cells (APC) (e.g. monocytes, dendritic cells and B cells) and is a co-stimulatory factor for T cell activation. A genetic variant in TNFSF4 (rs2205960) was associated with LN in a European ancestry SLE cohort (OR = 1.14, p = 0.0013) [31], and this association was also observed in a Chinese population (rs2205960 and rs10489265) [70]. Serum levels of TNFSF4 are significantly higher among SLE patients with LN than those without LN [71]. Also, the expression level of TNFSF4 receptor on CD4 T cells has been associated with nephritis and disease activity in patients with SLE [72].

3.2.1. APOL1—As shown in Figure 2, while some general SLE susceptibility genes predispose to LN, there are also some examples of genes which are specifically related to LN and not general SLE susceptibility. APOL1 gene is one example of an LN specific gene. The APOL genes located on human chromosome 22 and encodes apolipoprotein L-1 (APOL1). It is a component of circulating HDL (high density lipoprotein) and also exists abundantly in many organs including the kidney [73]. APOL1 gene variants were known to have a protective effect against African *Trypanosoma brucei rhodesiense* (*T. b. rhodesiense*) infection [74]. Serum apolipoprotein L-1 from individuals with the APOL1 variant lyses Tripanosomes more efficiently than that from individuals without the APOL1 variant [74]. Thus, the high prevalence of APOL1 variants in individuals of African descent seems likely to have been driven by evolutionary pressure from *T. b. rhodesiense* infection [75], and this variant is almost absent in European ancestry. Interestingly, this APOL1 gene variant in African ancestry populations has also been associated with multiple kidney diseases, such as focal segmental glomerulonephritis, ESRD in non-diabetic nephropathy and human immunodeficiency virus-associated nephropathy [74, 76–78].

More recently, Freedman et al. reported the association of risk variants of APOL1 with ESRD in African Americans with LN [10]. SLE patients with LN having APOL1 risk alleles G1/G2 were much more likely to progress to ESRD (OR = 2.72, $P = 6.23 \times 10^{-6}$) as compared to those patients without risk alleles [10]. However, Lin et al. reported that APOL1 risk alleles were not associated with LN in European American population and also did not show significant relations with mild LN even in African American population [79]. Thus the association between APOL1 and LN could be limited to severe LN which is likely to progress to ESRD. From the immunological aspect, APOL1 may also have roles in innate immunity and anti-viral activities. Nichols et al. have shown that APOL1 is induced by TLR3 agonists and interferons (interferon-alpha, -beta and –gamma) [80]. Additionally, APOL1 is involved in autophagy pathway [81]. Of note, APOL1 has not been identified as a general SLE susceptibility gene, so it appears to be a modifier gene, important to nephritis risk and progression in African ancestry populations. Thus, this gene may partially explain the increased incidence and severity of LN in African ancestry SLE patients.

3.2.2. PDGFRA—In a recent study, a variant in the PDGFRA gene (rs1364989) was found to be strongly associated with LN in the European population [9]. The PDGFRA gene is located on human chromosome 4 and encodes platelet-derived growth factor receptor alpha (PDGFRA). PDGF and PDGFR (e.g. PDGFRA) are constitutively expressed in most renal cells (e.g. mesangial cells, fibroblasts and vascular smooth muscle cells), and are involved in normal kidney development. Alteration of the PDGF system is common in human kidney diseases [82]. PDGF-BB (one of the isoforms of PDGF), which is a ligand for PDGFRA, is an important factor that promotes mesangioproliferative disease and renal interstitial fibrosis [83]. In LN, increased PDGFRA mRNA expression was observed in the kidney (glomeruli and tubulointerstitial compartment) of LN patients [9].

Imatinib is a tyrosine-kinase inhibitor that has been used in the treatment of Philadelphia chromosome-positive chronic myelogenous leukemia, KIT-positive gastrointestinal stromal

tumors, and other diseases. Imatinib is also known to inhibit PDGFR signaling [82]. Administration of Imatinib in several experimental studies in lupus mouse models showed a significant decrease in LN progression confirmed by histological findings[84, 85], and reduced PDGFR expression [84]. However, some animal experiments using trapidil, a broad PDGF antagonist, have failed to treat nephritis [86]. These data could support the idea of targeting PDGFR in LN, although it seems that the type of inhibitor may be important, as the murine studies are not consistent across all types of PDGFR blockade.

3.2.3. HAS2—The HAS2 gene is located on human chromosome 8 that encodes Hyaluronan synthase 2 (HAS2); an enzyme responsible for hyaluronan (HA) synthesis. HA plays a crucial role in fibrosis in various organs. Renal fibrosis is the principle pathologic process that moves LN toward chronic kidney disease and to ESRD. Several reports provide evidence that intra- and extraglomerular mesangial cells respond to various cytokines and growth factors (e.g. IL-1 beta, PDGF) and result in induction of HA, accompanied by upregulation of HAS2 [87–90]. Yung et al. have shown that increased expression of HA in the mesangium and tubular portion of the kidney in renal biopsy specimens from LN patients [91]. Serum HA concentration was higher in the active phase of LN than in inactive LN. They also identified that anti-DNA antibody induced IL-1 beta, which then generates HA via overexpression of HAS2 in human mesangial cells [91]. Recent genetic association studies have demonstrated association of a variant in the HAS2 gene (rs7834765) with LN (OR = 3.15) [9]. Again, HAS2 has not been shown to be associated with SLE susceptibility in general, but seems to play a role in LN pathogenesis, possibly via HA production and a contribution to fibrosis.

4. Epigenetics in LN

Not only genetic changes, but epigenetic changes (i.e. post-translational modifications) also play an important role in the pathogenesis of SLE [92]. DNA methylation is one of the important post-translational regulatory modifications, typically occuring at CG dinucleotides. DNA methylation results in gene silencing by tightening the chromatin structure and limiting the access of transcriptional factors, while DNA hypomethylation induce transcription of genes.

Impaired DNA methylation status in CD4+ T cells of SLE patients was reported more than 20 years ago [93]. As next-generation sequencing technology has advanced, genome-wide methylation studies have demonstrated the differences in methylation profiles of CD4 T cells in SLE patients compared to those of healthy controls. Some studies have shown a difference in methylation profiles between different groups of SLE patients [94, 95]. Of note, hypomethylation of type I IFN-regulated genes known to play important roles in the pathogenesis of SLE are reported in SLE patients [96, 97]. More recently, Coit et al. identified that there are more robust differences in methylation status of type I IFN-regulated genes when compared between SLE patients with LN and SLE patients without LN [98].

These studies shed light to another aspect of genetic involvement in the pathogenesis of SLE and LN, although there is still much work to be done to clarify their specific role to LN, and take advantage of this knowledge to design treatments.

5. Conclusions

Genetic factors play an important role in predisposing SLE patients to LN. The genetic factors discussed above are summarized in Table 1. The prevalence and severity of LN differs among individuals and ethnicities, and there are some examples of differences in genetic factors predisposing to LN in different populations. Despite the identification of numerous general SLE susceptibility genes, only a small number of these general SLE genes are associated with LN. Of note, the magnitude of the OR for general SLE susceptibility genes as risk factors for LN was lower than their impact upon SLE in general. This supports the idea that these general SLE susceptibility genes impact SLE more broadly than their impact upon LN alone. The more recent discovery of genes with renal function that are associated specifically with LN is fascinating and provides additional insight into the pathogenesis of LN. It seems that the predisposition to LN involves a combination of general SLE susceptibility genes and genes with renal function that are associated specifically with LN. Figure 3 shows some potential functional relationships between the loci discussed in this review. Moreover, new generation sequencing technique has revealed not only the genome mutation, but the post-translational modification may contribute in the pathogenesis of SLE and LN. Further epidemiologic and functional studies will better define the relationships of these genes with LN pathogenesis, and likely will identify additional gene loci involved in this serious disease manifestation.

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Highlights

- Some general SLE susceptibility genes predispose to LN
- APOL1, PDGFR and HAS2 are novel LN susceptibility genes
- Combination of general SLE and LN specific genes contribute to LN

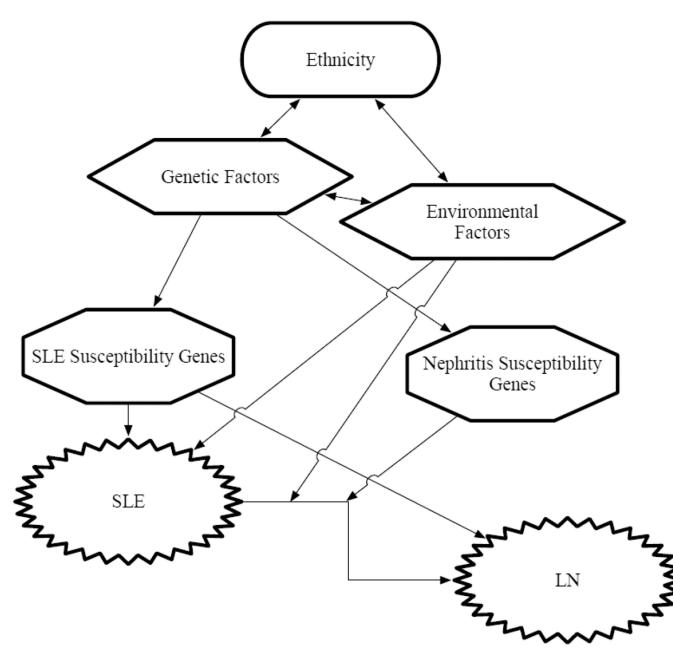


Figure 1. Interactions between multiple factors predisposing to LN Interactions between genetic factors and environmental factors predispose to SLE and LN. LN, lupus nephritis; SLE, systemic lupus erythematosus

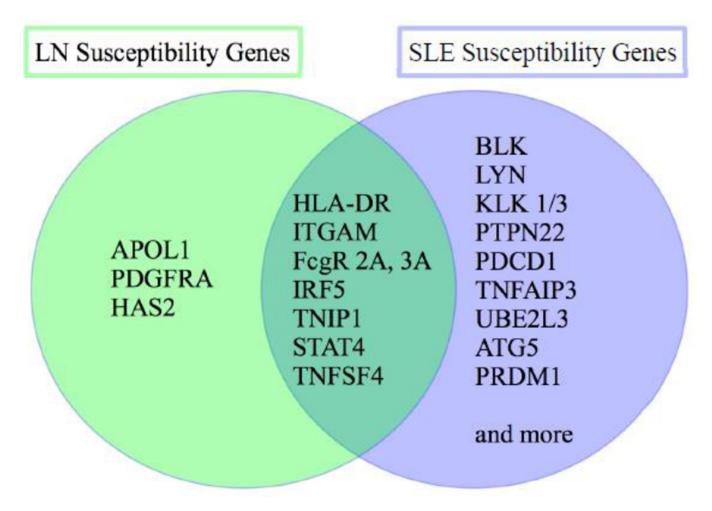


Figure 2. SLE and LN susceptibility genes Venn diagram of SLE and LN susceptibility genes. LN, lupus nephritis; SLE, systemic lupus erythematosus

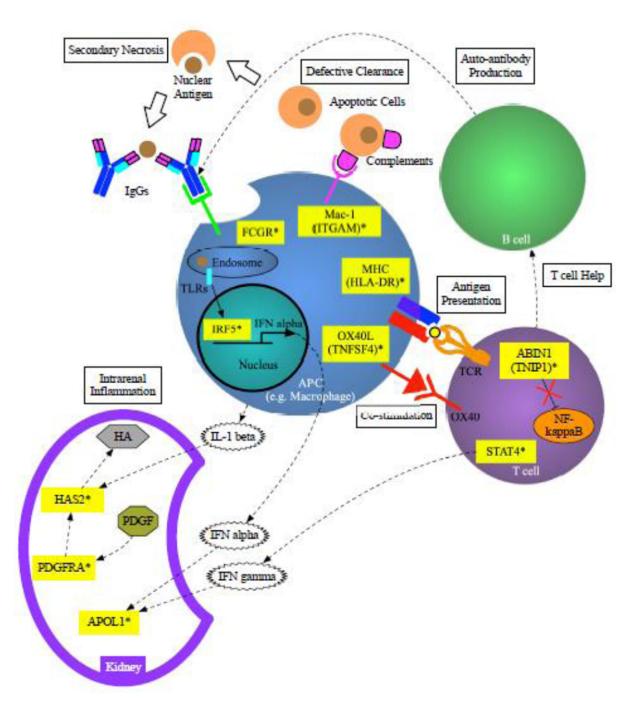


Figure 3. Genetic contribution to immune system in the pathogenesis of LN

Altered functions of complement receptors (Mac-1) and FcgR will result in defective clearance of apoptotic cells and move towards secondary necrosis. Phagocyted nuclear antigens will activate TLRs and altered IRF5 leads to increased production of IFN alpha. Co-stimulatory factors (i.e. HLA and OX40L) activate T cells and result in IFN gamma production via STAT4 activation. Altered ABIN1 function, caused by TNIP1 variant leads to dysregulation of NF-kappaB activation. Auto-antibodies will be produced by B cells with the help of activated T cells. IFNs will induce APOL1 expression in the kidney. IL-1 beta,

secreted by activated APC (e.g. macrophages) and the signal of PDGF via PDGFR will eventually lead to increased expression of HA via HAS2. ABIN1, A20 binding and inhibitor of NF-kappaB 1; APC, antigen presenting cell; APOL1, apolipoprotein L-1; FCGR, Fc gamma receptor; HA, hyaluronan; HAS2, hyaluronan synthase 2; HLA, human leukocyte antigen; IFN, interferon; IL, interleukin; IRF5,

interferon regulatory factor 5; ITGAM, integrin alpha M; OX40L, OX40 ligand; PDGFRA, platelet-derived growth factor receptor alpha; STAT4, signal transducers and activators of transcription 4; TLRs, Toll-like receptors; TNFSF4, tumor necrosis factor super family 4; * Susceptible genes.

Table 1

Confirmed Susceptibility Loci Associated with lupus Nephritis

Genes associated with SLE and LN

Gene name	Chromosome	SNP associated with LN	Known fuction	Reference
HLA-DR	6	-	Antigen presentation	[9, 21–23]
ITGAM	16	rs1143679 rs9888739	Clearance of immune complex	[9, 21, 29–31]
FcgR 2A FcgR 3A	1	rs1801274 (H131/R131 polymorphism) rs396991 (F158/V158 polymorphism)	Clearance of immune complex	[38, 39, 46] [40–43, 47]
IRF5	7	rs10488631 rs2070197	Innate immunity, type I IFN pathway	[21]
TNIP1	5	rs7708392 rs4958881 rs6889239	Inhibition of NF-kappaB pathway	[21, 58, 62]
STAT4	2	rs7574865 rs7568275 rs7582694 rs11889341	T cell signaling	[21, 65]
TNFSF4	1	rs 22D5960 rs10489265	Costimulatory factor for T cell activation	[9, 31, 70]
Genes not as	sociated with SL	E, but associated with LN		
Gene name	Chromosome	SNP associated with LN	Known function	Reference
APOL1	22	Gl/G2 haplotype	Innate immunity autophagy	[10]
PDGFRA	4	rs1364989	Kidney development	[9]
HAS2	8	rs7834765	Organ fibrosis	[9]

APOL1, apolipoprotein L-1; FCGR, Fc gamma receptor; F158, phenylalanine residue at position 158; H131, histidine residue at position 131; HAS2, hyaluronan synthase 2; ITGAM, integrin alpha M; LN, lupus nephritis; PDGFRA, platelet-derived growth factor receptor alpha; R131, arginine residue at position 131; SLE, systemic lupus erythematosus; SNP, single nucleotide polymorphism; STAT4, signal transducers and activators of transcription 4; TNFSF4, tumor necrosis factor super family 4; TNIP1, TNFAIP3-interacting protein 1; V158, valine residue at position 158