



Published in final edited form as:

Curr Neurovasc Res. 2017 ; 14(1): 82–88. doi:10.2174/1567202613666161129112822.

Harnessing the Power of SIRT1 and Non-coding RNAs in Vascular Disease

Kenneth Maiese^{1,*}

¹Cellular and Molecular Signaling, Newark, New Jersey 07101

Abstract

Noncommunicable diseases (NCDs) contribute to a significant amount of disability and death in the world. Of these disorders, vascular disease is ranked high, falls within the five leading causes of death, and impacts multiple other disease entities such as those of the cardiac system, nervous system, and metabolic disease. Targeting the silent mating type information regulation 2 homolog 1 (*Saccharomyces cerevisiae*) (SIRT1) pathway and the modulation of micro ribonucleic acids (miRNAs) may hold great promise for the development of novel strategies for the treatment of vascular disease since each of these pathways are highly relevant to cardiac and nervous system disorders as well as to metabolic dysfunction. SIRT1 is vital in determining the course of stem cell development and the survival, metabolism, and life span of differentiated cells that are overseen by both autophagy and apoptosis. SIRT1 interfaces with a number of pathways that involve forkhead transcription factors, mechanistic of rapamycin (mTOR), AMP activated protein kinase (AMPK) and Wnt1 inducible signaling pathway protein 1 (WISP1) such that the level of activity of SIRT1 can become a critical determinant for biological and clinical outcomes. The essential fine control of SIRT1 is directly tied to the world of non-coding RNAs that ultimately oversee SIRT1 activity to either extend or end cellular survival. Future studies that can further elucidate the crosstalk between SIRT1 and non-coding RNAs should serve well our ability to harness the power of SIRT1 and non-coding RNAs for the treatment of vascular disorders.

Keywords

aging; apoptosis; autophagy; biomarker; cardiovascular disease; CCN4; cyclin-dependent kinase 2 (CDK2); cyclin-dependent kinase inhibitor 1 (p21); cell cycle; circular RNA; diabetes mellitus; endothelial cells; forkhead transcription factors; FoxO; metabolism; microRNA; non-coding RNA; oxidative stress; programmed cell death; senescence; SIRT1; stem cells; transcription factors; WISP1; Wnt signaling

The Global Presence of Vascular Disease

Noncommunicable diseases (NCDs) contribute to a significant amount of deaths across the globe. At least sixty percent of the fifty-seven million global deaths result from NCDs and almost eighty percent of these NCDs occur in low and middle-income countries (1). In

*Correspondence to: Kenneth Maiese, MD, Cellular and Molecular Signaling, USA. wntin75@yahoo.com.

Competing Interests: There are no conflicts of interest to declare.

regards to the leading causes of NCDs, vascular disorders rank high and fall within the five leading causes of death that include cardiac disease, cancer, chronic lower respiratory disease, stroke, and traumatic accidents (2). Interestingly, vascular disorders not only affect cardiac disease, but also disorders of the nervous system. Hypertension and elevated serum lipids are significant risk factors for stroke, a disorder that affects approximately fifteen million individuals every year and leads to an annual cost of seventy-five billion dollars in the United States (3–7). Other NCDs such as diabetes mellitus (DM) also can lead to neurodegenerative disease (8–11). DM is increasing in incidence throughout the world. Approximately 350 million individuals currently have DM and an additional eight million individuals are believed to suffer from metabolic disorders and are currently undiagnosed (12, 13). Although DM can affect any system of the body, DM significantly impacts vascular disease resulting in platelet dysfunction (14, 15), atherosclerosis (9, 16, 17), endothelial cell senescence (18, 19), injury to endothelial cells (20–27), endothelial progenitor cell dysfunction (27–30), impaired angiogenesis (31–33), and cardiovascular disease (24, 34–38).

SIRT1 as a Target for Vascular Disease

The extent of vascular disease in the population is immense. Coupled to the progression of vascular disease in the world is the increasing age of the global population. Life expectancy is approaching eighty years of age for all individuals and is accompanied by a one percent decrease in the age-adjusted death rate from the years 2000 through 2011 (39). In addition, the number of individuals over the age of sixty-five also has doubled during the past fifty years (40). As a result, new avenues of investigation to develop novel therapies against vascular disease are required to stem this tide of vascular disease. One such avenue involves the silent mating type information regulation 2 homolog 1 (*Saccharomyces cerevisiae*) (SIRT1) that is under such consideration for the development of new treatments against vascular disease (29, 35, 41–47).

SIRT1, a member of the sirtuin family, is a histone deacetylase that transfers acetyl groups from ϵ -N-acetyl lysine amino acids on the histones of DNA to control transcription (29, 48–51). Seven identified mammalian homologues of Sir2 exist that include SIRT1 through SIRT7 that control post-translational changes of proteins and oversee cellular proliferation, survival, and senescence. In regards to SIRT1, it is dependent upon nicotinamide adenine dinucleotide (NAD⁺) as substrate (48, 52–54). Nicotinamide phosphoribosyltransferase (NAMPT) catalyzes the conversion of nicotinamide to nicotinamide mononucleotide through the salvage pathway of NAD⁺ synthesis (55–57). Nicotinamide mononucleotide is then converted to NAD⁺ by enzymes in the nicotinamide/nicotinic acid mononucleotide adenylyltransferase (NMNAT) family.

The level of SIRT1 activity is believed to be an important factor in determining cell survival and protection against toxic insults. NAMPT is a rate-limiting enzyme in mammalian NAD⁺ biosynthesis pathway such that elevated levels of NAMPT activity increase cellular NAD⁺ levels and the activity of SIRT1 transcription. Other factors also can influence SIRT1 activity. NMNAT can modulate the deacetylating activity of SIRT1. Mammalian forkhead transcription factors (58, 59) also can bind to the SIRT1 promoter region that contains a

cluster of five insulin-responsive core repeat motifs (IRS-1) and a forkhead-like consensus-binding site (FKHD-L). This allows forkhead transcription factors, such as FoxO1, to increase SIRT1 expression (60). AMP activated protein kinase (AMPK) is another mechanism for the control of SIRT1 activity (61–67). AMPK is a member of the mechanistic of rapamycin (mTOR) pathway that phosphorylates tuberous sclerosis protein 2 (TSC2) and inhibits the activity of mTORC1 (67, 68). AMPK increases the activity of NAMPT (51). This process catalyzes the conversion of nicotinamide to nicotinamide mononucleotide (69), increases NAD⁺ levels (69), decreases levels of nicotinamide (70), an inhibitor of SIRT1 (69, 71–73), and leads to SIRT1 transcription (13, 48, 74). As a result of increasing the intracellular NAD⁺/NADH ratio, AMPK leads to deacetylation of the SIRT1 targets peroxisome proliferator-activated receptor-gamma coactivator 1 (PGC-1 α) and forkhead transcription factors that include FoxO1 (37) and FoxO3a (75). Crosstalk between SIRT1 and AMPK also occurs (74). Resveratrol, a SIRT1 activator, has been demonstrated to activate AMPK (76).

At times, reduced SIRT1 activity may be required to promote cellular survival involving trophic factors such as insulin growth factor-1 (77). However, insufficient SIRT1 activity may have a detrimental affect upon vascular cell survival (23, 78, 79) and block protection against cardiovascular disease (53, 80–82). Overall, SIRT1 controls multiple cellular functions that are involved with modulation of vascular survival and senescence (18, 83, 84), control of vascular tone through the transient receptor potential cation channel A1 (TRPA1) (85), development of atherosclerosis (80, 86–89), extension of lifespan (42, 73, 90, 91), alterations in cellular metabolism (10, 31, 57, 92–96), enhancement of neuronal survival and cognition (46, 59, 92, 97), and protection against oxidative stress (58, 81, 94, 98–103).

SIRT1, Stem Cells, Autophagy, and Apoptosis

SIRT1 can have a protective effective over stem cell development and maturation. Through pathways of autophagy (104, 105), SIRT1 can control stem cell survival by modulating autophagic flux (106). SIRT1 has a role in autophagic flux by promoting autophagy in mitochondria (107) that may be required to maintain a healthy pool of mitochondria (108). SIRT1 is required to initiate autophagy and transition muscle stem cells from a quiescence state to an active state (109). SIRT1 also inhibits apoptotic cell injury in endothelial progenitor cells during oxidative stress through the induction of autophagy (110). In endothelial cells exposed to oxidized low density lipoproteins that can lead to atherosclerosis, SIRT1 activity is increased in conjunction with AMPK that leads to autophagy and promotes cellular protection (86). SIRT1 activation of autophagy also involves the inhibition of other pathways such as the mechanistic target of rapamycin (mTOR) (33, 101, 111, 112). SIRT1 can have an inverse relationship with mTOR in embryonic stem cells (57, 113). SIRT1 inhibits mTOR pathways to promote autophagy and preserve the integrity of embryonic stem cells during oxidant stress (101) as well as to promote neuronal growth (114).

Apoptotic pathways with SIRT1 also play a role with stem cell survival (23, 42, 78, 103, 115, 116). For example, SIRT1 activation prevents external membrane phosphatidylserine (PS) exposure during the early phases of apoptosis in mature cells (23, 78, 117, 118). SIRT1

activity can counteract apoptosis initiated by tumor necrosis factor- α (TNF- α) in endothelial progenitor cells (84). During exposure to TNF- α , SIRT1 also has been shown to prevent skeletal myoblast injury (119). In the presence of decreased SIRT1 activity, human mesenchymal stem cells have been shown not to proliferate and succumb to apoptosis (83). Loss of SIRT1 expression in endothelial progenitor cells leads to apoptotic cell death that can occur in smokers and chronic obstructive disease patients (120).

During apoptosis, SIRT1 activity can be reduced by the degradation of caspases. Caspase degradation of the SIRT1 protein (121) also can accelerate further activation of caspases (121, 122). Other pathways such as p38 (123) and JNK1 (124) can degrade SIRT1 as well. Some pathways may protect SIRT1 against caspase degradation. The CCN family (defined by the first three members of the family that include Cysteine-rich protein 61, Connective tissue growth factor, and Nephroblastoma over-expressed gene) (125) member Wnt1 inducible signaling pathway protein 1 (WISP1) increases SIRT1 activity to protect cells from oxidative stress and apoptotic injury (126). WISP1 prevents SIRT1 caspase degradation and modulates forkhead transcription activity with SIRT1 to block FoxO3a activity and prevent caspase activation that would otherwise lead to the degradation of SIRT1 (126–129).

SIRT1, Non-Coding RNAs, and Stem Cell Proliferation

SIRT1 modulation of vascular disease as well as stem cell proliferation may be closely tied to the pathways of small non-coding ribonucleic acids (RNAs) that are termed microRNAs (miRNAs) (43, 49, 130, 131). MiRNAs oversee gene expression by silencing targeted messenger RNAs (mRNAs) translated by specific genes and are composed of 19–25 nucleotides. These small non-coding ribonucleic acids have an important role with SIRT1 to control stem cell development and differentiated cell survival. Under some conditions, increased SIRT1 activity is beneficial. Silencing of miR-195 in old mesenchymal stem cells promotes stem cell proliferation by increasing SIRT1 activity to restore anti-aging factors expression that include telomerase reverse transcriptase, the forkhead transcription factor FOXO1 (50), and protein kinase B (Akt) (131). Increased SIRT1 activity with loss of miR-204 promotes the proliferation of spermatogonial stem cells (132). Proliferation of stem cells also may require increased SIRT1 activity in combination with the inhibition or dysfunction of mTOR signaling that is controlled by miRNAs (133). However, it should be noted that a reduction in SIRT1 activity controlled by miRNAs may at times offer a benefit to stem cell populations. Neuronal differentiation can occur through miR-34a that leads to decreased SIRT1 expression and DNA-binding of p53 in mouse neural stem cells (134). Furthermore, miR-34a under other conditions that increase the SIRT1 and FoxO3a pathways can result in mitochondrial dysfunction and activation of intrinsic apoptotic pathways that are detrimental to mesenchymal stem cells (135).

Other non-coding RNAs also may have a critical role in vascular disease and be related to SIRT1 activity. For example, circular ribonucleic acids (circRNAs) are non-coding RNAs of approximately 100 nucleotides in length that were initially identified as being circular in nature (136–138). These non-coding RNAs have covalent bonds that maintain their circular structure. CircRNAs have both *cis* and *trans* regulation, regulate gene expression through the sponging of microRNAs (miRNAs) (139), can function as biomarkers, and oversee cellular

survival through programmed cell death involving apoptosis (104, 105). Circular antisense non-coding RNA in the INK4 locus (circANRIL) in vascular smooth muscle cells and macrophages can prevent exonuclease-mediated pre-ribosomal RNA processing, ribosome biogenesis, and proliferation of cells that may lead to atherosclerosis through the induction of apoptosis (140). CircRNA can function as an endogenous miR-223 sponge to inhibit cardiac hypertrophy and heart failure (141). Yet, circRNAs may not always be protective against apoptosis. Up-regulation of specific circRNAs may foster apoptotic cell injury during cell models of ischemia-reperfusion injury (142). In experimental models of myocardial infarction, the circRNA Cdr1as can increase cardiac infarct size and function as a sponge for “cytoprotective” miR-7a (143).

In the cardiovascular system, circRNA generated from Foxo3 (circ-Foxo3) has been found to be expressed in aged patients that may be linked to processes of aging. Silencing circ-Foxo3 inhibits senescence in mouse embryonic fibroblasts and over-expression of circ-Foxo3 results in cell senescence (144). Circ-Foxo3 appears to block cell cycle progression by binding to the cell cycle proteins cyclin-dependent kinase 2 (CDK2) and cyclin-dependent kinase inhibitor 1 (p21) to prevent cellular proliferation (145). With advanced age, increased expression of circRNAs also has been demonstrated in the skeletal muscles of monkeys (146). Given the intimate association among SIRT1, forkhead transcription factors, lifespan extension, and aging, it is conceivable that SIRT1 may have a significant role in the vascular system with aging and circRNAs.

SIRT1, miRNAs, and Vascular Disease

In a number of recent experimental paradigms, miRNAs appear to significantly impact the ability of SIRT1 to control cell survival in the vascular and nervous system. In studies involving aging and the apoptotic loss of cochlear hair cells, high expression of miR-29b in aged C57BL/6 mice results in decreased expression of SIRT1 and mitochondrial dysfunction, suggesting a protective role for SIRT1 (147). Diabetic endothelial vascular dysfunction that occurs during hyperglycemia elevated free fatty acids can result from the up-regulation of miR-34a that depresses the expression of SIRT1. In rat glomerular mesangial cells, elevated glucose increases miR-217 that decreases SIRT1 expression as well to result in inflammation and fibrosis (148). During periods of hyperglycemia, angiogenesis is impaired as a result of suppressed SIRT1 expression and the up-regulation of miR-34a expression (31). The ill effects that can occur during DM also extend to the retinal microvasculature. Studies in rats demonstrate that an up-regulation of senescence-associated markers that include miR-34a depresses SIRT1 expression and accelerates aging and oxidative stress injury in the retinal vasculature (94). In additional studies, endothelial vascular dysfunction and oxidative stress are prevented with the maintenance of SIRT1 and the inhibition of miR-34a (95). Vascular miR-204 controlled by gut microbiome also impairs endothelial function by targeting SIRT1 and depressing its function (149).

Future Perspectives

Targeting the SIRT1 pathway and the modulation of miRNAs may hold great promise for the development of novel strategies for the treatment of vascular disease. Clearly, as a NCD,

cardiovascular disorders affect a large proportion of the global population as a leading cause of death and disability (29, 42, 150). In addition, vascular disorders not only impact the cardiac system, but also play a significant role in disease progression for other disease entities such as those that involve the nervous system and metabolic disease (36, 151–153). In its self, SIRT1 is critical in controlling cell development, survival, metabolism, life span, and programmed cell death with apoptosis and autophagy. SIRT1 also interfaces with a number of vital pathways that involve forkhead transcription factors, mTOR, AMPK and WISPI. Yet, it appears that the level of SIRT1 activity can be a vital determinant in both biological and clinical outcomes that requires careful analysis and targeting. The level of activity of SIRT1 can affect the balance of the programmed death pathways of apoptosis and autophagy that ultimately determine cell survival as well as alter the course of stem cell development, maintenance, and differentiation. For example, SIRT1 activity is required for neuronal cell protection (126, 154, 155) and vascular cell survival (23, 78, 79, 156). This necessary fine control of SIRT1 activity also translates into the world of miRNAs such that in several circumstances loss of SIRT1 activity that is controlled by high miRNA expression can be toxic to cell survival and stem cell maintenance. However, evidence exists that under other conditions, a reduction in SIRT1 activity by miRNAs is required in cases that can promote stem cell differentiation and stem cell survival (134, 135). Future work that can foster new insight into these challenges will be extremely conducive for the development of harnessing the power of SIRT1 and non-coding RNAs for the treatment of vascular disorders.

Acknowledgments

This research was supported by the following grants to Kenneth Maiese: American Diabetes Association, American Heart Association, NIH NIEHS, NIH NIA, NIH NINDS, and NIH ARRA.

References

1. Organization WH. Global status report on noncommunicable diseases 2010. 2011 Apr. Description of the global burden of NCDs, their risk factors and determinants; p. 1-176.
2. Minino AM, Murphy SL. Death in the United States, 2010. NCHS data brief. 2012; (99):1–8.
3. Kim JY, Park J, Chang JY, Kim SH, Lee JE. Inflammation after Ischemic Stroke: The Role of Leukocytes and Glial Cells. *Experimental neurobiology*. 2016; 25(5):241–51. [PubMed: 27790058]
4. Maiese K. Cutting through the Complexities of mTOR for the Treatment of Stroke. *Curr Neurovasc Res*. 2014; 11(2):177–86. [PubMed: 24712647]
5. Maiese K. Driving neural regeneration through the mammalian target of rapamycin. *Neural regeneration research*. 2014; 9(15):1413–7. [PubMed: 25317149]
6. Shahjouei S, Ansari S, Pourmotabbed T, Zand R. Potential Roles of Adropin in Central Nervous System: Review of Current Literature. *Frontiers in molecular biosciences*. 2016; 3:25. [PubMed: 27446928]
7. Zhao EY, Efendizade A, Cai L, Ding Y. The role of Akt (protein kinase B) and protein kinase C in ischemia-reperfusion injury. *Neurol Res*. 2016:1–8.
8. Albiero M, Poncina N, Tjwa M, Ciciliot S, Menegazzo L, Ceolotto G, et al. Diabetes causes bone marrow autonomic neuropathy and impairs stem cell mobilization via dysregulated p66Shc and Sirt1. *Diabetes*. 2014; 63(4):1353–65. [PubMed: 24270983]
9. Maiese K. mTOR: Driving apoptosis and autophagy for neurocardiac complications of diabetes mellitus. *World J Diabetes*. 2015; 6(2):217–24. [PubMed: 25789103]

10. Maiese K. Novel nervous and multi-system regenerative therapeutic strategies for diabetes mellitus with mTOR. *Neural regeneration research*. 2016; 11(3):372–85. [PubMed: 27127460]
11. White MF. IRS2 integrates insulin/IGF1 signalling with metabolism, neurodegeneration and longevity. *Diabetes Obes Metab*. 2014; 16(Suppl 1):4–15. [PubMed: 25200290]
12. Haldar SR, Chakrabarty A, Chowdhury S, Haldar A, Sengupta S, Bhattacharyya M. Oxidative stress-related genes in type 2 diabetes: association analysis and their clinical impact. *Biochemical genetics*. 2015; 53(4–6):93–119. [PubMed: 25991559]
13. Maiese K. New Insights for Oxidative Stress and Diabetes Mellitus. *Oxid Med Cell Longev*. 2015; 2015:875961. [PubMed: 26064426]
14. Alexandru N, Popov D, Georgescu A. Platelet dysfunction in vascular pathologies and how can it be treated. *Thromb Res*. 2012; 129(2):116–26. [PubMed: 22035630]
15. Balestrieri ML, Servillo L, Esposito A, D’Onofrio N, Giovane A, Casale R, et al. Poor glycaemic control in type 2 diabetes patients reduces endothelial progenitor cell number by influencing SIRT1 signalling via platelet-activating factor receptor activation. *Diabetologia*. 2013; 56(1):162–72. [PubMed: 23070058]
16. Di Rosa M, Malaguarnera L. Chitotriosidase: A New Inflammatory Marker in Diabetic Complications. *Pathobiology*. 2016; 83(4):211–9. [PubMed: 27116685]
17. Mikhed Y, Daiber A, Steven S. Mitochondrial Oxidative Stress, Mitochondrial DNA Damage and Their Role in Age-Related Vascular Dysfunction. *International journal of molecular sciences*. 2015; 16(7):15918–53. [PubMed: 26184181]
18. Arunachalam G, Samuel SM, Marei I, Ding H, Triggler CR. Metformin modulates hyperglycaemia-induced endothelial senescence and apoptosis through SIRT1. *Br J Pharmacol*. 2014; 171(2):523–35. [PubMed: 24372553]
19. Orimo M, Minamino T, Miyauchi H, Tateno K, Okada S, Moriya J, et al. Protective role of SIRT1 in diabetic vascular dysfunction. *Arterioscler Thromb Vasc Biol*. 2009; 29(6):889–94. [PubMed: 19286634]
20. Chong ZZ, Hou J, Shang YC, Wang S, Maiese K. EPO Relies upon Novel Signaling of Wnt1 that Requires Akt1, FoxO3a, GSK-3beta, and beta-Catenin to Foster Vascular Integrity During Experimental Diabetes. *Curr Neurovasc Res*. 2011; 8(2):103–20. [PubMed: 21443457]
21. Chong ZZ, Shang YC, Maiese K. Vascular injury during elevated glucose can be mitigated by erythropoietin and Wnt signaling. *Curr Neurovasc Res*. 2007; 4(3):194–204. [PubMed: 17691973]
22. Hou J, Chong ZZ, Shang YC, Maiese K. FoxO3a governs early and late apoptotic endothelial programs during elevated glucose through mitochondrial and caspase signaling. *Mol Cell Endocrinol*. 2010; 321(2):194–206. [PubMed: 20211690]
23. Hou J, Chong ZZ, Shang YC, Maiese K. Early apoptotic vascular signaling is determined by Sirt1 through nuclear shuttling, forkhead trafficking, bad, and mitochondrial caspase activation. *Curr Neurovasc Res*. 2010; 7(2):95–112. [PubMed: 20370652]
24. Kandula V, Kosuru R, Li H, Yan D, Zhu Q, Lian Q, et al. Forkhead box transcription factor 1: role in the pathogenesis of diabetic cardiomyopathy. *Cardiovasc Diabetol*. 2016; 15(1):44. [PubMed: 26956801]
25. Li FQ, Zeng DK, Jia CL, Zhou P, Yin L, Zhang B, et al. The effects of sodium tanshinone IIA sulfonate pretreatment on high glucose-induced expression of fractalkine and apoptosis in human umbilical vein endothelial cells. *International journal of clinical and experimental medicine*. 2015; 8(4):5279–86. [PubMed: 26131102]
26. Weikel KA, Cacicedo JM, Ruderman NB, Ido Y. Knockdown of GSK3beta Increases Basal Autophagy and AMPK Signaling in Nutrient-laden Human Aortic Endothelial Cells. *Bioscience reports*. 2016
27. Wu H, Li R, Wei ZH, Zhang XL, Chen JZ, Dai Q, et al. Diabetes-Induced Oxidative Stress in Endothelial Progenitor Cells May Be Sustained by a Positive Feedback Loop Involving High Mobility Group Box-1. *Oxid Med Cell Longev*. 2016; 2016:1943918. [PubMed: 26798412]
28. Barthelmes D, Zhu L, Shen W, Gillies MC, Irhimeh MR. Differential gene expression in Lin-/VEGF-R2+ bone marrow-derived endothelial progenitor cells isolated from diabetic mice. *Cardiovasc Diabetol*. 2014; 13(1):42. [PubMed: 24521356]

29. D'Onofrio N, Vitiello M, Casale R, Servillo L, Giovane A, Balestrieri ML. Sirtuins in vascular diseases: Emerging roles and therapeutic potential. *Biochim Biophys Acta*. 2015; 1852(7):1311–22. [PubMed: 25766107]
30. Kim KA, Shin YJ, Akram M, Kim ES, Choi KW, Suh H, et al. High glucose condition induces autophagy in endothelial progenitor cells contributing to angiogenic impairment. *Biol Pharm Bull*. 2014; 37(7):1248–52. [PubMed: 24989016]
31. Arunachalam G, Lakshmanan AP, Samuel SM, Triggle CR, Ding H. Molecular Interplay between microRNA-34a and Sirtuin1 in Hyperglycemia-Mediated Impaired Angiogenesis in Endothelial Cells: Effects of Metformin. *J Pharmacol Exp Ther*. 2016; 356(2):314–23. [PubMed: 26582729]
32. Maiese K. Novel applications of trophic factors, Wnt and WISP for neuronal repair and regeneration in metabolic disease. *Neural regeneration research*. 2015; 10(4):518–28. [PubMed: 26170801]
33. Maiese K. Programming apoptosis and autophagy with novel approaches for diabetes mellitus. *Curr Neurovasc Res*. 2015; 12(2):173–88. [PubMed: 25742566]
34. Chong ZZ, Maiese K. Mammalian Target of Rapamycin Signaling in Diabetic Cardiovascular Disease. *Cardiovasc Diabetol*. 2012; 11(1):45. [PubMed: 22545721]
35. Esser N, Paquot N, Scheen AJ. Anti-inflammatory agents to treat or prevent type 2 diabetes, metabolic syndrome and cardiovascular disease. *Expert opinion on investigational drugs*. 2015; 24(3):283–307. [PubMed: 25345753]
36. Lawler JM, Rodriguez DA, Hord JM. Mitochondria in the middle: Exercise preconditioning protection of striated muscle. *J Physiol*. 2016
37. Maiese K, Chong ZZ, Shang YC, Hou J. FoxO proteins: cunning concepts and considerations for the cardiovascular system. *Clin Sci (Lond)*. 2009; 116(3):191–203. [PubMed: 19118491]
38. Perez-Hernandez N, Vargas-Alarcon G, Posadas-Sanchez R, Martinez-Rodriguez N, Tovilla-Zarate CA, Rodriguez-Cortes AA, et al. PHACTR1 Gene Polymorphism Is Associated with Increased Risk of Developing Premature Coronary Artery Disease in Mexican Population. *International journal of environmental research and public health*. 2016; 13(8)
39. Minino AM. Death in the United States, 2011. *NCHS data brief*. 2013; (115):1–8.
40. Hayutin A. Global demographic shifts create challenges and opportunities. *PREA Quarterly*. 2007 Fall;:46–53.
41. Favero G, Franceschetti L, Rodella LF, Rezzani R. Sirtuins, aging, and cardiovascular risks. *Age (Dordr)*. 2015; 37(4):9804. [PubMed: 26099749]
42. Maiese K. SIRT1 and stem cells: In the forefront with cardiovascular disease, neurodegeneration and cancer. *World J Stem Cells*. 2015; 7(2):235–42. [PubMed: 25815111]
43. Maiese K. MicroRNAs and SIRT1: A Strategy for Stem Cell Renewal and Clinical Development? *J Transl Sci*. 2015; 1(3):55–7. [PubMed: 26561536]
44. Maiese K. Regeneration in the nervous system with erythropoietin. *Frontiers in bioscience (Landmark edition)*. 2016; 21:561–96.
45. Maiese K. The bright side of reactive oxygen species: lifespan extension without cellular demise. *J Transl Sci*. 2016; 2(3):185–7. [PubMed: 27200181]
46. Martin A, Tegla CA, Cudrici CD, Kruszewski AM, Azimzadeh P, Boodhoo D, et al. Role of SIRT1 in autoimmune demyelination and neurodegeneration. *Immunologic research*. 2015; 61(3):187–97. [PubMed: 25281273]
47. Poulouse N, Raju R. Sirtuin regulation in aging and injury. *Biochim Biophys Acta*. 2015; 1852(11):2442–55. [PubMed: 26303641]
48. Chong ZZ, Shang YC, Wang S, Maiese K. SIRT1: New avenues of discovery for disorders of oxidative stress. *Expert opinion on therapeutic targets*. 2012; 16(2):167–78. [PubMed: 22233091]
49. Gabay O, Clouse KA. Epigenetics of cartilage diseases. *Joint, bone, spine : revue du rhumatisme*. 2015
50. Maiese K. FoxO Proteins in the Nervous System. *Anal Cell Pathol (Amst)*. 2015; 2015:569392. [PubMed: 26171319]
51. Maiese K. Targeting molecules to medicine with mTOR, autophagy, and neurodegenerative disorders. *Br J Clin Pharmacol*. 2015

52. Duan W. Sirtuins: from metabolic regulation to brain aging. *Frontiers in aging neuroscience*. 2013; 5:36. [PubMed: 23888142]
53. Maiese K, Chong ZZ, Shang YC, Wang S. Translating cell survival and cell longevity into treatment strategies with SIRT1. *Rom J Morphol Embryol*. 2011; 52(4):1173–85. [PubMed: 22203920]
54. Srivastava S, Haigis MC. Role of sirtuins and calorie restriction in neuroprotection: implications in Alzheimer's and Parkinson's diseases. *Curr Pharm Des*. 2011; 17(31):3418–33. [PubMed: 21902666]
55. Chong ZZ, Wang S, Shang YC, Maiese K. Targeting cardiovascular disease with novel SIRT1 pathways. *Future Cardiol*. 2012; 8(1):89–100. [PubMed: 22185448]
56. Maiese K. Picking a bone with WISP1 (CCN4): new strategies against degenerative joint disease. *J Transl Sci*. 2016; 1(3):83–5. [PubMed: 26893943]
57. Maiese K, Chong ZZ, Shang YC, Wang S. Novel directions for diabetes mellitus drug discovery. *Expert opinion on drug discovery*. 2013; 8(1):35–48. [PubMed: 23092114]
58. Maiese K. FoxO Transcription Factors and Regenerative Pathways in Diabetes Mellitus. *Curr Neurovasc Res*. 2015; 12(4):404–13. [PubMed: 26256004]
59. Maiese K. Forkhead transcription factors: new considerations for alzheimer's disease and dementia. *J Transl Sci*. 2016; 2(4):241–7. [PubMed: 27390624]
60. Xiong S, Salazar G, Patrushev N, Alexander RW. FoxO1 Mediates an Autofeedback Loop Regulating SIRT1 Expression. *J Biol Chem*. 2011; 286(7):5289–99. [PubMed: 21149440]
61. Cai Z, Chen G, He W, Xiao M, Yan LJ. Activation of mTOR: a culprit of Alzheimer's disease? *Neuropsychiatric disease and treatment*. 2015; 11:1015–30. [PubMed: 25914534]
62. Chong ZZ, Shang YC, Wang S, Maiese K. Shedding new light on neurodegenerative diseases through the mammalian target of rapamycin. *Prog Neurobiol*. 2012; 99(2):128–48. [PubMed: 22980037]
63. Citraro R, Leo A, Constanti A, Russo E, De Sarro G. mTOR pathway inhibition as a new therapeutic strategy in epilepsy and epileptogenesis. *Pharmacol Res*. 2016; 107:333–43. [PubMed: 27049136]
64. Johnson SC, Sangesland M, Kaerberlein M, Rabinovitch PS. Modulating mTOR in aging and health. *Interdisciplinary topics in gerontology*. 2015; 40:107–27. [PubMed: 25341517]
65. Lan AP, Chen J, Zhao Y, Chai Z, Hu Y. mTOR Signaling in Parkinson's Disease. *Neuromolecular Med*. 2016
66. Maiese K. Taking aim at Alzheimer's disease through the mammalian target of rapamycin. *Ann Med*. 2014; 46(8):587–96. [PubMed: 25105207]
67. Maiese K, Chong ZZ, Shang YC, Wang S. mTOR: on target for novel therapeutic strategies in the nervous system. *Trends Mol Med*. 2013; 19(1):51–60. [PubMed: 23265840]
68. Shang YC, Chong ZZ, Wang S, Maiese K. Tuberous sclerosis protein 2 (TSC2) modulates CCN4 cytoprotection during apoptotic amyloid toxicity in microglia. *Curr Neurovasc Res*. 2013; 10(1): 29–38. [PubMed: 23244622]
69. Maiese K, Chong ZZ, Hou J, Shang YC. The vitamin nicotinamide: translating nutrition into clinical care. *Molecules*. 2009; 14(9):3446–85. [PubMed: 19783937]
70. Fulco M, Cen Y, Zhao P, Hoffman EP, McBurney MW, Sauve AA, et al. Glucose restriction inhibits skeletal myoblast differentiation by activating SIRT1 through AMPK-mediated regulation of Nampt. *Dev Cell*. 2008; 14(5):661–73. [PubMed: 18477450]
71. Marshall CA, Borbon IA, Erickson RP. Relative efficacy of nicotinamide treatment of a mouse model of infantile Niemann-Pick C1 disease. *Journal of applied genetics*. 2016
72. Naia L, Rosenstock TR, Oliveira AM, Oliveira-Sousa SI, Caldeira GL, Carmo C, et al. Comparative Mitochondrial-Based Protective Effects of Resveratrol and Nicotinamide in Huntington's Disease Models. *Mol Neurobiol*. 2016
73. Poljsak B, Milisav I. The NAD(+)-depletion theory of ageing: NAD(+) as the link between oxidative stress, inflammation, caloric restriction, exercise, DNA repair, longevity and health span. *Rejuvenation Res*. 2016

74. Wang Y, Liang Y, Vanhoutte PM. SIRT1 and AMPK in regulating mammalian senescence: a critical review and a working model. *FEBS Lett.* 2011; 585(7):986–94. [PubMed: 21130086]
75. Canto C, Auwerx J. Caloric restriction, SIRT1 and longevity. *Trends Endocrinol Metab.* 2009; 20(7):325–31. [PubMed: 19713122]
76. Chen S, Xiao X, Feng X, Li W, Zhou N, Zheng L, et al. Resveratrol induces Sirt1-dependent apoptosis in 3T3-L1 preadipocytes by activating AMPK and suppressing AKT activity and survivin expression. *The Journal of nutritional biochemistry.* 2012; 23(9):1100–12. [PubMed: 22137261]
77. Sansone L, Reali V, Pellegrini L, Villanova L, Aventaggiato M, Marfe G, et al. SIRT1 silencing confers neuroprotection through IGF-1 pathway activation. *J Cell Physiol.* 2013; 228(8):1754–61. [PubMed: 23359486]
78. Hou J, Wang S, Shang YC, Chong ZZ, Maiese K. Erythropoietin Employs Cell Longevity Pathways of SIRT1 to Foster Endothelial Vascular Integrity During Oxidant Stress. *Curr Neurovasc Res.* 2011; 8(3):220–35. [PubMed: 21722091]
79. Marampon F, Gravina GL, Scarsella L, Festuccia C, Lovat F, Ciccarelli C, et al. Angiotensin-converting-enzyme inhibition counteracts angiotensin II-mediated endothelial cell dysfunction by modulating the p38/SirT1 axis. *Journal of hypertension.* 2013; 31(10):1972–83. [PubMed: 23868084]
80. Hung CH, Chan SH, Chu PM, Tsai KL. Quercetin is a potent anti-atherosclerotic compound by activation of SIRT1 signaling under oxLDL stimulation. *Molecular nutrition & food research.* 2015
81. Saboori S, Koohdani F, Nematipour E, Yousefi Rad E, Saboor-Yaraghi AA, Javanbakht MH, et al. Beneficial effects of omega-3 and vitamin E coadministration on gene expression of SIRT1 and PGC1alpha and serum antioxidant enzymes in patients with coronary artery disease. *Nutrition, metabolism, and cardiovascular diseases : NMCD.* 2016; 26(6):489–94.
82. Yu L, Liang H, Dong X, Zhao G, Jin Z, Zhai M, et al. Reduced silent information regulator 1 signaling exacerbates myocardial ischemia-reperfusion injury in type 2 diabetic rats and the protective effect of melatonin. *J Pineal Res.* 2015; 59(3):376–90. [PubMed: 26327197]
83. Chiara B, Ilaria C, Antonietta C, Francesca C, Marco M, Lucia A, et al. SIRT1 Inhibition Affects Angiogenic Properties of Human MSCs. *BioMed research international.* 2014; 2014:783459. [PubMed: 25243179]
84. Du G, Song Y, Zhang T, Ma L, Bian N, Chen X, et al. Simvastatin attenuates TNFalpha-induced apoptosis in endothelial progenitor cells via the upregulation of SIRT1. *Int J Mol Med.* 2014; 34(1):177–82. [PubMed: 24718722]
85. Paziienza V, Pomara C, Cappello F, Calogero R, Carrara M, Mazzoccoli G, et al. The TRPA1 channel is a cardiac target of mIGF-1/SIRT1 signaling. *Am J Physiol Heart Circ Physiol.* 2014; 307(7):H939–44. [PubMed: 25108014]
86. Jin X, Chen M, Yi L, Chang H, Zhang T, Wang L, et al. Delphinidin-3-glucoside protects human umbilical vein endothelial cells against oxidized low-density lipoprotein-induced injury by autophagy upregulation via the AMPK/SIRT1 signaling pathway. *Molecular nutrition & food research.* 2014; 58(10):1941–51. [PubMed: 25047736]
87. Kedenko L, Lamina C, Kedenko I, Kollerits B, Kiesslich T, Iglseder B, et al. Genetic polymorphisms at SIRT1 and FOXO1 are associated with carotid atherosclerosis in the SAPHIR cohort. *BMC Med Genet.* 2014; 15(1):112. [PubMed: 25273948]
88. Luo XY, Qu SL, Tang ZH, Zhang Y, Liu MH, Peng J, et al. SIRT1 in cardiovascular aging. *Clin Chim Acta.* 2014; 437:106–14. [PubMed: 25063737]
89. Xu S, Bai P, Little PJ, Liu P. Poly(ADP-ribose) polymerase 1 (PARP1) in atherosclerosis: from molecular mechanisms to therapeutic implications. *Med Res Rev.* 2014; 34(3):644–75. [PubMed: 24002940]
90. Balan V, Miller GS, Kaplun L, Balan K, Chong ZZ, Li F, et al. Life span extension and neuronal cell protection by Drosophila nicotinamidase. *J Biol Chem.* 2008; 283(41):27810–9. [PubMed: 18678867]

91. Moroz N, Carmona JJ, Anderson E, Hart AC, Sinclair DA, Blackwell TK. Dietary restriction involves NAD⁻dependent mechanisms and a shift toward oxidative metabolism. *Aging Cell*. 2014; 13(6):1075–85. [PubMed: 25257342]
92. Du LL, Chai DM, Zhao LN, Li XH, Zhang FC, Zhang HB, et al. AMPK Activation Ameliorates Alzheimer's Disease-Like Pathology and Spatial Memory Impairment in a Streptozotocin-Induced Alzheimer's Disease Model in Rats. *J Alzheimers Dis*. 2014
93. Hardeland R. Melatonin and the pathologies of weakened or dysregulated circadian oscillators. *J Pineal Res*. 2016
94. Lamoke F, Shaw S, Yuan J, Ananth S, Duncan M, Martin P, et al. Increased Oxidative and Nitritative Stress Accelerates Aging of the Retinal Vasculature in the Diabetic Retina. *PLoS One*. 2015; 10(10):e0139664. [PubMed: 26466127]
95. Li Q, Kim YR, Vikram A, Kumar S, Kassan M, Gabani M, et al. P66Shc-Induced MicroRNA-34a Causes Diabetic Endothelial Dysfunction by Downregulating Sirtuin1. *Arterioscler Thromb Vasc Biol*. 2016
96. Tulsulkar J, Nada SE, Slotterbeck BD, McInerney MF, Shah ZA. Obesity and hyperglycemia lead to impaired post-ischemic recovery after permanent ischemia in mice. *Obesity (Silver Spring, Md)*. 2015
97. Min JJ, Huo XL, Xiang LY, Qin YQ, Chai KQ, Wu B, et al. Protective effect of DI-3n-butylphthalide on learning and memory impairment induced by chronic intermittent hypoxia-hypercapnia exposure. *Scientific reports*. 2014; 4:5555. [PubMed: 24990154]
98. Akasaki Y, Alvarez-Garcia O, Saito M, Carames B, Iwamoto Y, Lotz MK. FOXO transcription factors support oxidative stress resistance in human chondrocytes. *Arthritis & rheumatology (Hoboken, NJ)*. 2014; 66(12):3349–58.
99. Chong ZZ, Maiese K. Enhanced Tolerance against Early and Late Apoptotic Oxidative Stress in Mammalian Neurons through Nicotinamidase and Sirtuin Mediated Pathways. *Curr Neurovasc Res*. 2008; 5(3):159–70. [PubMed: 18691073]
100. Colak Y, Yesil A, Mutlu HH, Caklili OT, Ulasoglu C, Senates E, et al. A potential treatment of non-alcoholic fatty liver disease with SIRT1 activators. *Journal of gastrointestinal and liver diseases : JGLD*. 2014; 23(3):311–9. [PubMed: 25267960]
101. Ou X, Lee MR, Huang X, Messina-Graham S, Broxmeyer HE. SIRT1 positively regulates autophagy and mitochondria function in embryonic stem cells under oxidative stress. *Stem Cells*. 2014; 32(5):1183–94. [PubMed: 24449278]
102. Zhang F, Hu Y, Xu X, Zhai X, Wang G, Ning S, et al. Icarin protects against intestinal ischemia-reperfusion injury. *J Surg Res*. 2015; 194(1):127–38. [PubMed: 25472572]
103. Zhang XS, Wu Q, Wu LY, Ye ZN, Jiang TW, Li W, et al. Sirtuin 1 activation protects against early brain injury after experimental subarachnoid hemorrhage in rats. *Cell death & disease*. 2016; 7(10):e2416. [PubMed: 27735947]
104. Klionsky DJ, Abdelmohsen K, Abe A, Abedin MJ, Abeliovich H, Acevedo Arozena A, et al. Guidelines for the use and interpretation of assays for monitoring autophagy (3rd edition). *Autophagy*. 2016; 12(1):1–222. [PubMed: 26799652]
105. Maiese K, Chong ZZ, Shang YC, Wang S. Targeting disease through novel pathways of apoptosis and autophagy. *Expert opinion on therapeutic targets*. 2012; 16(12):1203–14. [PubMed: 22924465]
106. Mazzoccoli G, Tevy MF, Borghesan M, Delle Vergini MR, Vinciguerra M. Caloric restriction and aging stem cells: the stick and the carrot? *Exp Gerontol*. 2014; 50:137–48. [PubMed: 24211426]
107. Jang SY, Kang HT, Hwang ES. Nicotinamide-induced mitophagy: event mediated by high NAD⁺/NADH ratio and SIRT1 protein activation. *J Biol Chem*. 2012; 287(23):19304–14. [PubMed: 22493485]
108. Fang EF, Scheibye-Knudsen M, Brace LE, Kassahun H, SenGupta T, Nilsen H, et al. Defective mitophagy in XPA via PARP-1 hyperactivation and NAD⁽⁺⁾/SIRT1 reduction. *Cell*. 2014; 157(4):882–96. [PubMed: 24813611]
109. Tang AH, Rando TA. Induction of autophagy supports the bioenergetic demands of quiescent muscle stem cell activation. *EMBO J*. 2014; 33(23):2782–97. [PubMed: 25316028]

110. Chen J, Xavier S, Moskowitz-Kassai E, Chen R, Lu CY, Sanduski K, et al. Cathepsin cleavage of sirtuin 1 in endothelial progenitor cells mediates stress-induced premature senescence. *Am J Pathol.* 2012; 180(3):973–83. [PubMed: 22234173]
111. Ma L, Dong W, Wang R, Li Y, Xu B, Zhang J, et al. Effect of caloric restriction on the SIRT1/mTOR signaling pathways in senile mice. *Brain Res Bull.* 2015; 116:67–72. [PubMed: 26135885]
112. Maiese K. Stem cell guidance through the mechanistic target of rapamycin. *World J Stem Cells.* 2015; 7(7):999–1009. [PubMed: 26328016]
113. Maiese K. Erythropoietin and mTOR: A “One-Two Punch” for Aging-Related Disorders Accompanied by Enhanced Life Expectancy. *Curr Neurovasc Res.* 2016; 13(4):329–40. [PubMed: 27488211]
114. Guo W, Qian L, Zhang J, Zhang W, Morrison A, Hayes P, et al. Sirt1 overexpression in neurons promotes neurite outgrowth and cell survival through inhibition of the mTOR signaling. *J Neurosci Res.* 2011; 89(11):1723–36. [PubMed: 21826702]
115. Lin CL, Huang WN, Li HH, Huang CN, Hsieh S, Lai C, et al. Hydrogen-rich water attenuates amyloid beta-induced cytotoxicity through upregulation of Sirt1-FoxO3a by stimulation of AMP-activated protein kinase in SK-N-MC cells. *Chem Biol Interact.* 2015; 240:12–21. [PubMed: 26271894]
116. Wu H, Wang H, Zhang W, Wei X, Zhao J, Yan P, et al. rhEPO affects apoptosis in hippocampus of aging rats by upregulating SIRT1. *Int J Clin Exp Pathol.* 2015; 8(6):6870–80. [PubMed: 26261574]
117. Fong Y, Lin YC, Wu CY, Wang HM, Lin LL, Chou HL, et al. The antiproliferative and apoptotic effects of sirtinol, a sirtuin inhibitor on human lung cancer cells by modulating Akt/beta-catenin-Foxo3a axis. *Scientific World Journal.* 2014; 2014:937051. [PubMed: 25184156]
118. Wang T, Cui H, Ma N, Jiang Y. Nicotinamide-mediated inhibition of SIRT1 deacetylase is associated with the viability of cancer cells exposed to antitumor agents and apoptosis. *Oncology letters.* 2013; 6(2):600–4. [PubMed: 24137378]
119. Saini A, Al-Shanti N, Sharples AP, Stewart CE. Sirtuin 1 regulates skeletal myoblast survival and enhances differentiation in the presence of resveratrol. *Experimental physiology.* 2012; 97(3):400–18. [PubMed: 22125309]
120. Paschalaki KE, Starke RD, Hu Y, Mercado N, Margariti A, Gorgoulis VG, et al. Dysfunction of endothelial progenitor cells from smokers and chronic obstructive pulmonary disease patients due to increased DNA damage and senescence. *Stem Cells.* 2013; 31(12):2813–26. [PubMed: 23897750]
121. Kozako T, Aikawa A, Shoji T, Fujimoto T, Yoshimitsu M, Shirasawa S, et al. High expression of the longevity gene product SIRT1 and apoptosis induction by sirtinol in adult T-cell leukemia cells. *Int J Cancer.* 2012; 131(9):2044–55. [PubMed: 22322739]
122. Balaiya S, Ferguson LR, Chalam KV. Evaluation of sirtuin role in neuroprotection of retinal ganglion cells in hypoxia. *Invest Ophthalmol Vis Sci.* 2012; 53(7):4315–22. [PubMed: 22669716]
123. Hong EH, Lee SJ, Kim JS, Lee KH, Um HD, Kim JH, et al. Ionizing radiation induces cellular senescence of articular chondrocytes via negative regulation of SIRT1 by p38 kinase. *J Biol Chem.* 2010; 285(2):1283–95. [PubMed: 19887452]
124. Gao Z, Zhang J, Kheterpal I, Kennedy N, Davis RJ, Ye J. Sirtuin 1 (SIRT1) protein degradation in response to persistent c-Jun N-terminal kinase 1 (JNK1) activation contributes to hepatic steatosis in obesity. *J Biol Chem.* 2011; 286(25):22227–34. [PubMed: 21540183]
125. Maiese K. WISP1: Clinical Insights for a Proliferative and Restorative Member of the CCN Family. *Curr Neurovasc Res.* 2014; 11(4):378–89. [PubMed: 25219658]
126. Wang S, Chong ZZ, Shang YC, Maiese K. WISP1 neuroprotection requires FoxO3a post-translational modulation with autoregulatory control of SIRT1. *Curr Neurovasc Res.* 2013; 10(1):54–60. [PubMed: 23151077]
127. Dong L, Zhou S, Yang X, Chen Q, He Y, Huang W. Magnolol protects against oxidative stress-mediated neural cell damage by modulating mitochondrial dysfunction and PI3K/Akt signaling. *J Mol Neurosci.* 2013; 50(3):469–81. [PubMed: 23404573]

128. Qi XF, Li YJ, Chen ZY, Kim SK, Lee KJ, Cai DQ. Involvement of the FoxO3a pathway in the ischemia/reperfusion injury of cardiac microvascular endothelial cells. *Experimental and molecular pathology*. 2013; 95(2):242–7. [PubMed: 23948278]
129. Yang Y, Su Y, Wang D, Chen Y, Wu T, Li G, et al. Tanshinol attenuates the deleterious effects of oxidative stress on osteoblastic differentiation via Wnt/FoxO3a signaling. *Oxid Med Cell Longev*. 2013; 2013:351895. [PubMed: 24489983]
130. Chen H, Lu Q, Fei X, Shen L, Jiang D, Dai D. miR-22 inhibits the proliferation, motility, and invasion of human glioblastoma cells by directly targeting SIRT1. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015
131. Okada M, Kim HW, Matsu-Ura K, Wang YG, Xu M, Ashraf M. Abrogation of Age-Induced MicroRNA-195 Rejuvenates the Senescent Mesenchymal Stem Cells by Reactivating Telomerase. *Stem Cells*. 2015
132. Hua J. miR-204 regulated the proliferation of dairy goat spermatogonial stem cells via targeting to Sirt1. *Rejuvenation Res*. 2015
133. Jung CJ, Iyengar S, Blahnik KR, Jiang JX, Tahimic C, Torok NJ, et al. Human ESC self-renewal promoting microRNAs induce epithelial-mesenchymal transition in hepatocytes by controlling the PTEN and TGFbeta tumor suppressor signaling pathways. *Mol Cancer Res*. 2012; 10(7):979–91. [PubMed: 22622027]
134. Aranha MM, Santos DM, Sola S, Steer CJ, Rodrigues CM. miR-34a regulates mouse neural stem cell differentiation. *PLoS One*. 2011; 6(8):e21396. [PubMed: 21857907]
135. Zhang F, Cui J, Liu X, Lv B, Liu X, Xie Z, et al. Roles of microRNA-34a targeting SIRT1 in mesenchymal stem cells. *Stem cell research & therapy*. 2015; 6(1):195. [PubMed: 26446137]
136. Cocquerelle C, Mascrez B, Hetuin D, Bailleul B. Mis-splicing yields circular RNA molecules. *FASEB J*. 1993; 7(1):155–60. [PubMed: 7678559]
137. Hsu MT, Coca-Prados M. Electron microscopic evidence for the circular form of RNA in the cytoplasm of eukaryotic cells. *Nature*. 1979; 280(5720):339–40. [PubMed: 460409]
138. Maiese K. Disease onset and aging in the world of circular RNAs. *J Transl Sci*. 2016; 2(6):327–9. [PubMed: 27642518]
139. Zheng Q, Bao C, Guo W, Li S, Chen J, Chen B, et al. Circular RNA profiling reveals an abundant circHIPK3 that regulates cell growth by sponging multiple miRNAs. *Nature communications*. 2016; 7:11215.
140. Holdt LM, Stahlinger A, Sass K, Pichler G, Kulak NA, Wilfert W, et al. Circular non-coding RNA ANRIL modulates ribosomal RNA maturation and atherosclerosis in humans. *Nature communications*. 2016; 7:12429.
141. Wang K, Long B, Liu F, Wang JX, Liu CY, Zhao B, et al. A circular RNA protects the heart from pathological hypertrophy and heart failure by targeting miR-223. *Eur Heart J*. 2016
142. Lin SP, Ye S, Long Y, Fan Y, Mao HF, Chen MT, et al. Circular RNA expression alterations are involved in OGD/R-induced neuron injury. *Biochem Biophys Res Commun*. 2016; 471(1):52–6. [PubMed: 26845359]
143. Geng HH, Li R, Su YM, Xiao J, Pan M, Cai XX, et al. The Circular RNA Cdr1as Promotes Myocardial Infarction by Mediating the Regulation of miR-7a on Its Target Genes Expression. *PLoS One*. 2016; 11(3):e0151753. [PubMed: 26998750]
144. Du WW, Yang W, Chen Y, Wu ZK, Foster FS, Yang Z, et al. Foxo3 circular RNA promotes cardiac senescence by modulating multiple factors associated with stress and senescence responses. *Eur Heart J*. 2016
145. Du WW, Yang W, Liu E, Yang Z, Dhaliwal P, Yang BB. Foxo3 circular RNA retards cell cycle progression via forming ternary complexes with p21 and CDK2. *Nucleic Acids Res*. 2016; 44(6):2846–58. [PubMed: 26861625]
146. Abdelmohsen K, Panda AC, De S, Grammatikakis I, Kim J, Ding J, et al. Circular RNAs in monkey muscle: age-dependent changes. *Aging (Albany NY)*. 2015; 7(11):903–10. [PubMed: 26546448]
147. Xue T, Wei L, Zha DJ, Qiu JH, Chen FQ, Qiao L, et al. miR-29b overexpression induces cochlear hair cell apoptosis through the regulation of SIRT1/PGC-1alpha signaling: Implications for age-related hearing loss. *Int J Mol Med*. 2016

148. Shao Y, Lv C, Wu C, Zhou Y, Wang Q. Mir-217 promotes inflammation and fibrosis in high glucose cultured rat glomerular mesangial cells via Sirt1/HIF-1 α signaling pathway. *Diabetes Metab Res Rev.* 2016; 32(6):534–43. [PubMed: 26891083]
149. Vikram A, Kim YR, Kumar S, Li Q, Kassan M, Jacobs JS, et al. Vascular microRNA-204 is remotely governed by the microbiome and impairs endothelium-dependent vasorelaxation by downregulating Sirtuin1. *Nature communications.* 2016; 7:12565.
150. Maiese K. *Molecules to Medicine with mTOR: Translating Critical Pathways into Novel Therapeutic Strategies.* Elsevier and Academic Press; 2016.
151. Chong ZZ, Li F, Maiese K. Oxidative stress in the brain: Novel cellular targets that govern survival during neurodegenerative disease. *Prog Neurobiol.* 2005; 75(3):207–46. [PubMed: 15882775]
152. Ferro A. Mechanistic target of rapamycin modulation: an emerging therapeutic approach in a wide variety of disease processes. *Br J Clin Pharmacol.* 2016; 82(5):1156–7. [PubMed: 27734581]
153. Maiese K, Li F, Chong ZZ. New avenues of exploration for erythropoietin. *Jama.* 2005; 293(1): 90–5. [PubMed: 15632341]
154. Kim DW, Kim YM, Kang SD, Han YM, Pae HO. Effects of Resveratrol and trans-3,5,4'-Trimethoxystilbene on Glutamate-Induced Cytotoxicity, Heme Oxygenase-1, and Sirtuin 1 in HT22 Neuronal Cells. *Biomolecules & therapeutics.* 2012; 20(3):306–12. [PubMed: 24130928]
155. Zhang J, Feng X, Wu J, Xu H, Li G, Zhu D, et al. Neuroprotective effects of resveratrol on damages of mouse cortical neurons induced by beta-amyloid through activation of SIRT1/Akt1 pathway. *BioFactors (Oxford, England).* 2013; 40(2):258–67.
156. Kilic U, Gok O, Bacaksiz A, Izmirli M, Elibol-Can B, Uysal O. SIRT1 Gene Polymorphisms Affect the Protein Expression in Cardiovascular Diseases. *PLoS One.* 2014; 9(2):e90428. [PubMed: 24587358]