

# Specific Roles of NMDA Receptor Subunits in Mental Disorders

H. Yamamoto, Y. Hagino, S. Kasai and K. Ikeda\*

Addictive Substance Project, Tokyo Metropolitan Institute of Medical Science, 2-1-6 Kamikitazawa, Setagaya-ku, Tokyo 156-8506, Japan



K. Ikeda

**Abstract:** N-methyl-D-aspartate (NMDA) receptor plays important roles in learning and memory. NMDA receptors are a tetramer that consists of two glycine-binding subunits GluN1, two glutamate-binding subunits (i.e., GluN2A, GluN2B, GluN2C, and GluN2D), a combination of a GluN2 subunit and glycine-binding GluN3 subunit (i.e., GluN3A or GluN3B), or two GluN3 subunits. Recent studies revealed that the specific expression and distribution of each subunit are deeply involved in neural excitability, plasticity, and synaptic deficits. The present article summarizes reports on the dysfunction of NMDA receptors and responsible subunits in various neurological and psychiatric disorders, including schizophrenia, autoimmune-induced glutamatergic receptor dysfunction, mood disorders, and autism. A key role for the GluN2D subunit in NMDA receptor antagonist-induced psychosis has been recently revealed.

**Keywords:** GluN1, GluN2D, knockout mice, NMDA receptor subtype, phencyclidine, psychiatric disorders.

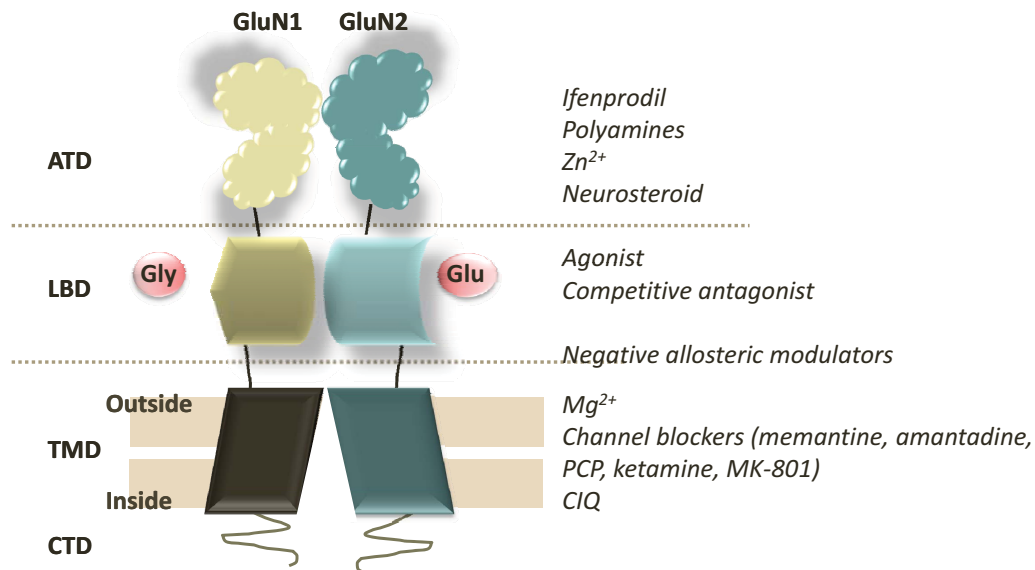
## INTRODUCTION

Postsynaptic N-Methyl-D-aspartate (NMDA) receptor is ionotropic glutamate receptor that mediates excitatory signaling in the presence of glycine and glutamate. NMDA receptors are a tetramer that consists of two glycine-binding subunits GluN1, two glutamate-binding subunits (i.e., GluN2A, GluN2B, GluN2C, and GluN2D), a combination of a GluN2 subunit and glycine-binding GluN3 subunit (i.e., GluN3A or GluN3B), or two GluN3 subunits. Moriyoshi *et al.* first isolated the cDNA of the rat NMDA receptor [1]. Subsequently, each subunit of the NMDA receptor was successfully cloned, including GluN1 [2], GluN2A-2C [3, 4], GluN2D [5], splice variants of GluN1 [2, 6-9], and GluN3A-3B [10-15]. GluN2 subunits are differentially expressed throughout the central nervous system [16-18]. NMDA receptor ion channels allow Na<sup>+</sup> and Ca<sup>2+</sup> ionic flows into the cell and K<sup>+</sup> ions from the cell to outside in a voltage-dependent manner. Therefore, single-channel conductance is calculated from the summation of these three ionic flows. Subunit composition influences single-channel conductance, Mg<sup>2+</sup> blockade, and Ca<sup>2+</sup> permeability [19-22]. The NMDA receptor channel containing the GluN1 subunit and a GluN2A or GluN2B subunit yields a single larger conductance level associated with higher permeability of Ca<sup>2+</sup> ions and higher sensitivity to extracellular Mg<sup>2+</sup> blockade. In contrast, the NMDA receptor channel containing GluN1 and the glutamate-binding subunit GluN2C shows two lower conductance sublevels with frequent direct transitions, reduced permeability of Ca<sup>2+</sup> ions, and less sensitivity to extracellular Mg<sup>2+</sup> blockade

[19]. Receptor channels containing GluN1 and the glutamate-binding subunit GluN2D also have a low open probability, two conductance levels, reduced sensitivity to Mg<sup>2+</sup> blockade, minimal desensitization, and a markedly slow deactivation time course [17, 23-29]. The deactivation time of the NMDA receptor channel composed of the GluN1 and GluN2D subunits is also affected by its splice variants GluN1-1a and GluN1-1b [30].

Each NMDA receptor subunit is composed of four discrete semiautonomous domains, including the extracellular amino-terminal domain (ATD), extracellular ligand-binding domain (LBD), transmembrane domain (TMD), and intracellular carboxyl-terminal domain (CTD; Fig. 1) [19]. The GluN2 ATD regulates agonist potency, the deactivation time course, the open probability, and the mean open/closed duration of different GluN2 subunits [25, 26]. Ryan *et al.* (2013) [31] found unique functions of the GluN2A CTD (i.e., the regulation of locomotor activity and impulsivity) and GluN2B CTD (i.e., the regulation of perceptual learning, anxiety, impulsivity, and motor coordination). In contrast, the GluN2A and GluN2B CTDs had similar functions in the regulation of reversal learning, associative learning, and motor learning. Corticostriatal or striatal GluN2B deletion and GluN2B antagonism in dorsal striatum impair choice learning, whereas cortical GluN2B deletion and GluN2B antagonism in orbitofrontal cortex impair shifting [32]. Although the GluN2B subunit has been implicated in both the acquisition and extinction of conditioned fear, GluN2C subunits in the amygdala are involved in the consolidation of learned fear responses. D-cycloserine selectively enhances the activity of NMDA receptors containing GluN2C subunit. The increased activity of GluN2C receptors may underlie the enhancement of fear extinction by D-cycloserine [33].

\*Address correspondence to this author at the Addictive Substance Project, Tokyo Metropolitan Institute of Medical Science, 2-1-6 Kamikitazawa, Setagaya-ku, Tokyo 156-8506, Japan; Tel: +81-3- 6834-2379; Fax: +81-3-6834-2390; E-mail: [ikeda-kz@igakuken.or.jp](mailto:ikeda-kz@igakuken.or.jp)



**Fig. (1). Model of the structure of NMDA receptors.** NMDA receptors are glutamate-activated ion channels in the presence of glycine expressed throughout the central nervous system. The full NMDA receptor is a tetramer, but only a GluN1/GluN2 dimer is shown. These receptors are mainly composed of two glycine (Gly)-binding GluN1 subunits and two glutamate (Glu)-binding GluN2 subunits. Each subunit is organized into four distinct domains: an extracellular N-terminal domain (ATD), a ligand-binding domain (LBD), a transmembrane pore-forming domain (TMD), and an intracellular C-terminal domain (CTD) [19].

The expression of GluN3A protein is observed throughout the central nervous system in multiple neuronal cell types. GluN3A subunit expression is high in the postnatal week and then decreases in adult animals [34, 35]. The GluN3A subunit makes NMDA receptor ion channels less Ca<sup>2+</sup>-permeable and Mg<sup>2+</sup>-insensitive [36]. In the absence of GluN3A, the expression of markers of synaptic maturation is accelerated [37]. Enhanced responses of NMDA receptors and increased number of dendritic spines are observed in early stage of postnatal cerebrocortical neurons [38], and inducing long-term potentiation (LTP) at young synapses is easier [39]. GluN3A knockout (KO) mice showed impaired locomotor activity, increased sensitivity to acute and subacute inflammatory pain, and enhanced recognition, spatial learning, and memory function [40]. The overexpression of GluN3A retards synaptic maturation and attenuates LTP at adult synapses [39]. These data indicate that GluN3A acts as a molecular brake to limit the plasticity and maturation of excitatory synapses [34, 37] and may have a profound impact on several functional/behavioral activities in adult animals [40] (Table 1).

## NMDA RECEPTOR DYSFUNCTION IS INVOLVED IN VARIOUS DISORDERS

NMDA receptors are implicated in neuronal development, synaptic plasticity, and learning and memory [19, 41]. NMDA receptor dysfunction is also involved in various psychiatric disorders, including schizophrenia, autoimmune-induced glutamatergic receptor dysfunction, mood disorders, autism, and drug-induced psychosis (Table 2).

## Schizophrenia (NMDA Receptor Hypofunction)

Schizophrenia has been treated as a disease with hyperdopaminergic function, in which dopamine receptor antagonists effectively treat positive symptoms. However, dopamine antagonist treatment has not improved negative symptoms and/or cognitive deficits. Presently, the hypofunction theory of glutamatergic neurons in schizophrenia is also widely accepted. Furthermore, environmental factors and multiple genes that enhanced the risk for the onset of schizophrenia are being investigated [79]. As an example of research on environmental factors, a nationwide study of 2,486,646 Danish people reported familial and environmental risk factors for schizophrenia, rate ratios, population-attributable risks, and sex-specific cumulative incidences of several risk factors [80]. In parallel, multiple genetic loci have been reported by genetic linkage studies [81] and association studies [43, 44]. Genome-wide association studies (GWASs) investigate candidate genes, including voltage-gated calcium channel genes, miR-137 targets,  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor genes (*GRIA1*, *GRIA4*), an NMDA receptor gene (*GRIN2B*), a metabotropic glutamate receptor gene (*GRM5*), an enzyme involved in glutamate metabolism *GAD1*, and a glutamate transporter *SLC1A2*. Gene expression data using human postmortem brain and human blood and relevant animal model data to identify candidate genes involved in schizophrenia integrate such studies [44-46]. In addition to the aforementioned candidate genes, genes with lower scores have also been reported, including an NMDA receptor gene (*GRIN2A*), a high-affinity sodium-coupled glutamate aspartate transporter *GLAST-1* gene (*SLC1A3*), an AMPA

Table 1. NEW IUPHAR nomenclature for glutamate ionotropic receptor subunits [<http://www.iuphar-db.org/LGICNomenclature.jsp>].

NC-IUPHAR NMDA Receptor Subunit Nomenclature	Gene Name		Previous Nomenclatures	
	Human	Rat and Mouse	Human	Rat and Mouse
GluN1	<i>GRIN1</i>	<i>Grin1</i>	RP11-350O14.1, GluN1, MRD8, NMDA1, NMDAR1, NR1	GluN1, NMDAR1, NR1 RP23-132N23.20-010, GluRε1, M100174, Nmdar, Rgsc174
GluN2A	<i>GRIN2A</i>	<i>Grin2a</i>	EPND, FESD, GluN2A, LKS, NMDAR2A, NR2A, RP11-297M9.2	GluN2A, NMDAR2A, NR2A, GluRε1
GluN2B	<i>GRIN2B</i>	<i>Grin2b</i>	GluN2B, MRD6, NMDAR2B, NR2B, hNR3	AW490526, GluN2B, NR2B, Nmdar2b, GluRε2
GluN2C	<i>GRIN2C</i>	<i>Grin2c</i>	GluN2C, NMDAR2C, NR2C	RP23-117K15.2, GluN2C, NMDAR2C, NR2C, GluRε3
GluN2D	<i>GRIN2D</i>	<i>Grin2d</i>	EB11, GluN2D, NMDAR2D, NR2D	GluN2D, NMDAR2D, NR2D, GluRε4
GluN3A	<i>GRIN3A</i>	<i>Grin3a</i>	GluN3A, NMDAR-L, NR3A	GluN3A, NR3, chi-1, mCG_120729, 6430537F04, A830097C19Rik, NMDAR-L, NR3A, mKIAA1973
GluN3B	<i>GRIN3B</i>	<i>Grin3b</i>	GluN3B, NR3B	GluN3B, NMDAR3B, NR3B
AmpA Receptor Subunit Nomenclature	Human	Rat and Mouse	Human	Rat and Mouse
GluA1	<i>GRIA1</i>	<i>Gria1</i>	GLUH1, GLUR1, GLURA, HBGR1	RP23-102H8.1, 2900051M01Rik, Glr-1, Glr1, GluR-A, GluRA, Glur-1, Glur1, HIPA1, gluR-K1
GluA2	<i>GRIA2</i>	<i>Gria2</i>	GLUR2, GLURB, GluR-K2, HBGR2	GluR-K2, GluR2, gluR-B GluR-B, Glur-2, Glur2
GluA3	<i>GRIA3</i>	<i>Gria3</i>	RP11-349N19.3, GLUR-C, GLUR-K3, GLUR3, GLURC, GluA3, MRX94	GLUR3, GluR-3, GluR-C, GluR-K3 RP23-471M13.1, 2900064119Rik, Glur-3, Glur3, Gluralpha3
GluA4	<i>GRIA4</i>	<i>Gria4</i>	GLUR4, GLUR4C, GLURD	GluR-D, GluR4, Glur-4, Glur4, Gluralpha4, spkw1

NC-IUPHAR (the International Union of Basic and Clinical Pharmacology Committee on Receptor Nomenclature and Drug Classification). Greek symbols in NMDA receptor subunit names were applied to the mouse orthologue only. The protein name mirrors the gene name, with just the two letter code difference (i.e., *GRIN1* translates to GluN1, *Gria1* translates to GluA1).

receptor gene (*GRIA3*), a kainate receptor gene (*GRIK4*), and metabotropic glutamate receptor genes (*GRM1*, *GRM4*, and *GRM7*) [45]. Two *de novo* mutations in the *GRIN2A* gene, which encodes the NMDA receptor GluN2A subunit, have been reported in patients with sporadic schizophrenia [47]. In the Japanese population, the GluN2D subunit gene is also reported as a possible genome that is involved in schizophrenia susceptibility [82]. More precisely, the transcripts that encode the ionotropic glutamate receptor subunits GluN2D, GluA3, GluK2, and GluK3 and intracellular proteins GRIP1 and SynGAP1 are reduced in relay neurons in the medial dorsal thalamus in schizophrenia [83]. The susceptibility to schizophrenia is known to be increased by a small number of rare, recurrent genomic copy number variants (CNVs). Many small *de novo* mutations are found in the glutamatergic postsynaptic proteins, including NMDA receptor complex [84, 85]. These mutation-induced defects in glutamatergic transmission, especially *via* NMDA receptors, are consistent with the glutamatergic neuron hypofunction theory of schizophrenia.

Changes in binding of glutamate receptor, gene expression, and subunit protein expression, especially

decreased GluN1 expression, in the prefrontal cortex [86-90], thalamus [91-94], hippocampus [95-99], and cerebellum [89] are shown using postmortem brain of patients with schizophrenia. The transcription of the GluN1 and GluN2B subunits of the NMDA receptor are regulated by nuclear respiratory factors 1 and 2 (NRF-1 and NRF-2). Specificity protein4 functionally regulates GluN1, GluN2A, and GluN2B in a complementary and concurrent/parallel manner with nuclear respiratory factors 1 and 2 (NRF-1 and NRF-2) [100]. One research group reported an increase in the expression of one spliced isoform, GluN1<sup>c2</sup>, that was significantly increased in the anterior cingulate cortex in postmortem samples from aged schizophrenia patients [87]. With regard to the GluN1 subunit, *GRIN1* (which encodes human GluN1) is not included in the candidate genes for schizophrenia, and complicated defects in glutamatergic postsynaptic signaling complexes, including NMDA receptors, AMPA receptors, and ARC, may induce schizophrenia symptoms.

Alterations in cortical inhibitory  $\gamma$ -aminobutyric acid (GABA) neurons have been demonstrated in many postmortem studies. For example, a subset of these neurons expressing the calcium-binding protein parvalbumin (PV) appear to have lower mRNA and

**Table 2. Dysfunction of NMDA receptors and its experimental therapeutic treatments.**

Disorders	NMDAR-Subunit Related Alterations	Experimental Therapeutic Treatments
Schizophrenia	<ul style="list-style-type: none"> <li>Hypofunction of NMDA receptor on GABAergic neurons induce an imbalance in neural network activity [42].</li> <li>A majority of candidate genes associated with increasing risk for schizophrenia can modulate glutamate receptor functions or receptor-interacting proteins, and then affect signal transduction pathways [43-46].</li> <li>Two <i>de novo</i> mutations in <i>GluN2A</i> gene are reported in patients with sporadic schizophrenia [47].</li> <li>Expression of NMDA receptor subunit (GluN2A subunit in particular) is reduced in postmortem brain with schizophrenia [48].</li> </ul>	<ul style="list-style-type: none"> <li>Agonists for glycine site (D-serine, glycine) or glycine transporter 1 inhibitors [49, 50].</li> <li>GluN2A-selective potentiators are of potential interest [51].</li> </ul>
Anti-NMDAR encephalitis	<ul style="list-style-type: none"> <li>Autoimmune-induced glutamatergic receptor dysfunctions. Anti-NMDA receptor antibodies reduced density of NMDA receptor and induced severe neurological symptoms including hallucinations, psychosis, and seizures [52].</li> </ul>	<ul style="list-style-type: none"> <li>Immunotherapy is effective for most patients with anti-NMDA receptor encephalitis [53].</li> </ul>
Depression	<ul style="list-style-type: none"> <li>Inhibitors of NMDA receptors, in particular GluN2B-containing receptors, result in fast and sustained therapeutic effects in depressive symptoms [54, 55].</li> </ul>	<ul style="list-style-type: none"> <li>Ketamine and GluN2B-selective antagonists [54, 55].</li> <li>NMDA receptor functional glycine site partial agonist [56].</li> </ul>
Autism spectrum disorders	<ul style="list-style-type: none"> <li>Alteration of GluN2B and SHANK3 are genetic risk factors. Reduced or enhanced functions of NMDA receptor are involved. Mechanisms are obscure [22, 47, 57-62].</li> </ul>	<ul style="list-style-type: none"> <li>Potential interest for NMDA receptor partial agonist [63].</li> </ul>
NMDA receptor antagonist-induced psychosis	<ul style="list-style-type: none"> <li>Administration of NMDA receptor antagonists induces abnormal behaviors in rodents and psychosis in humans [64-66].</li> <li>NMDA receptor antagonists induce increases in the cortical high frequency or gamma oscillations [67, 68].</li> <li>Effect of PCP is attenuated in GluN2D KO mice [69, 70].</li> </ul>	<ul style="list-style-type: none"> <li>Potential interest for NMDA receptor co-agonists, glycine type I transport inhibitors, mGluR2/3 agonist or NMDA receptor potentiators [70-77].</li> <li>Prior embryonic medial ganglionic eminence cell transplantation into the medial prefrontal cortex [78].</li> </ul>

protein levels of the glutamic acid decarboxylase (GAD67; 67kDa isozyme for GABA synthesis) and PV [101], indicating abnormal GABA neurotransmission by PV neurons in schizophrenia. Altered PV neuron function appears to underlie cognitive deficits in schizophrenia by disturbing the generation of cortical gamma-oscillations [101]. Interestingly, GluN2A mRNA expression is reduced to undetectable levels in approximately 50% of PV neurons in the prefrontal cortex in subjects with schizophrenia [48]. Genetically engineered mice, in which the GluN1 gene is eliminated during early postnatal development but not adulthood, replicated the reduced expression of both GAD67 and PV in cortical PV neurons [102]. These findings indicate that reduced transmission through NMDA receptors could be an upstream mechanism of altered PV neuron function in schizophrenia.

Numerous studies provided genetic, pharmacological, and behavioral data to indicate that a reduction of glutamatergic function, especially NMDA receptors on inhibitory GABAergic interneurons, induces an imbalance between excitatory input and inhibitory input. These neural circuitry perturbations that underlie cognitive and executive dysfunctions lead to eventually psychosis [42].

### Autoimmune-Induced Glutamatergic Receptor Dysfunction

Recently, autoimmune synaptic encephalitis are defined brain diseases in human that cause severe neurological symptoms, including hallucinations, psychosis, and seizures, by autoantibody reactions with brain tissue. Among autoimmune encephalitis patients, autoantibodies against the extracellular domains of the NMDA receptor are frequently detected and the concentrations of autoantibodies are relevant to the developing psychotic and neurological symptoms [52]. Notably, in addition to having autoimmune encephalitis, some patients also have anti-NMDA receptor antibodies. Using serum from 459 patients admitted with acute schizophrenia, major depression (MD), and borderline personality disorder (BLPD) and matched controls, diverse NMDA receptor antibodies are detected in 15 subjects [103]. They were classified primarily as an initial schizophrenia (9.9%), MD (2.8%) or BLPD (0%) and controls (0.4%). Among these patients, two young females with acute disorganized behavior or catatonia were diagnosed with NMDA receptor encephalitis according to specific immunoglobulin G (IgG) antibodies against GluN1a, which were also increased in cerebrospinal fluid (CSF)

[103]. The other two patients had a different epitope of IgG NMDA receptor antibodies against GluN1a/GluN2b but not against GluN1a alone, and they had no immunoreactive antibodies in CSF [103]. IgA and IgM anti-NMDA receptor antibodies were present in all of the other seropositive cases. The presence of IgG GluN1 antibodies in CSF is important for the diagnosis of NMDA receptor encephalitis because antibodies in some patients are detected only in CSF [104, 105]. Kayser *et al.* (2013) reported the frequency, symptoms types, and outcome in patients with anti-NMDA receptor encephalitis and isolated psychiatric manifestations in 571 patients with IgG antibodies against the GluN1 subunit of the NMDA receptor [53]. Of these 571 patients, 23 (4%) developed isolated psychiatric episodes. Interestingly, for all 23 patients, age (median, 20 years), sex (91% female), and tumor association (43%; ovarian teratoma in all cases) were similar to the population at large. Predominant symptoms included delusional thinking (74%), mood disturbances (70%, usually manic), and aggression (57%). Brain magnetic resonance imaging findings were abnormal in 10 of 22 patients (45%), and CSF analysis showed pleocytosis in 17 of 22 patients (77%). Gresa-Arribas *et al.* (2014) [105] recently studied 250 patients with anti-NMDA receptor encephalitis and reported antibodies that targeted a main epitope region at GluN1 amino acid 369. The epitope repertoire did not differ between patients with different outcomes and did not change during relapses. Although autoimmune encephalitis is a rare disease, systematic checks of NMDA receptor antibodies are desired for patients with acute psychiatric symptoms, especially for young females. Patients with anti-NMDA receptor encephalitis mostly respond to immunotherapy [53]. When first-line treatments (e.g., steroids, intravenous immunoglobulin, and plasmapheresis) fail, second-line immunotherapy (e.g., rituximab and cyclophosphamide) is usually effective [104].

### Mood Disorders and NMDA Receptor GluN2B Subunit

Major depressive disorder is considered a mood disorder caused by a malfunction of the monoaminergic systems, but the possibility of the involvement of the glutamatergic system has also been suggested [106]. Gene linkage analysis confirmed a role specifically for the GluN2B subunit of the NMDA receptor in bipolar disorder [107]. A recent study reported statistically significant differences in allele and genotype frequencies between treatment-resistant depression (TRD) and non-TRD groups for the rs1805502 polymorphism within the *GRIN2B* gene [108]. Interestingly, the nonselective NMDA receptor antagonist ketamine produces a fast and sustained reduction of depressive symptoms in patients with TRD, supporting this hypothesis [54, 55]. The antidepressant-like effect of ketamine is estimated to occur through the following pathways. The activity of cortical GABA interneurons is regulated by NMDA receptors. The basal activity of pyramidal neurons, in contrast, is not directly regulated by NMDA receptors

[109]. Ketamine blocks NMDA receptors, and the subsequent suppression of tonic glutamate input to GABAergic interneurons results in the disinhibition of glutamatergic transmission to pyramidal neurons in the prefrontal cortex.

In animals, low doses of ketamine induce an increase in spine density, enhance mammalian target of rapamycin (mTOR) signaling, and increase protein synthesis in the prefrontal cortex, accompanied by an antidepressant effect [110]. This is consistent with reports of the reduced expression of mTOR and its downstream signaling targets in postmortem brain samples from depressed patients [111]. The GluN2B-specific antagonist Ro 25-6981 (Table 3) induced robust antidepressant-like effects [112]. GluN2B antagonists trigger their antidepressant effects by altering the activity of mTOR [110, 113], suggesting that inhibiting the activity of GluN2B subunit-containing NMDA receptors accounts for the majority of the antidepressant effects of ketamine. A recent study by Wang *et al.* [114] reported that replacing GluN2B with GluN2A in genetically modified mice enhanced the expression of synaptic AMPA receptors by activating mTOR signaling, which resembles ketamine-induced changes.

GLYX-13 is an amidated tetrapeptide (threonine-proline-proline-threonine) and glycine-site modulator at the NMDA receptor. It also preferentially modulates GluN2B subunit-containing NMDA receptors [115]. GLYX-13 and ketamine increased both GluN2B and GluR1 protein levels, but no changes in mRNA expression level [56]. GLYX-13 induces an antidepressant-like effect in the absence of the usual side effects associated with ketamine, at least partially by directly modulating GluN2B subunit-containing NMDA receptors in the medial prefrontal cortex [56]. Altogether, a direct or indirect antagonist that is selective for the GluN2B subunit would be a new-generation antidepressant candidate for the treatment of TRD.

### Autism and Animal Models

In the *Diagnostic and Statistical Manual of Mental Disorders*, 5th edition (<http://www.dsm5.org/proposedrevision/Pages/NeurodevelopmentalDisorders.aspx>; accessed April 11, 2014), social communication impairments and restricted repetitive patterns of behavior are described as core symptoms of autism spectrum disorder (ASD). Many changes in ASD are intratelencephalic (IT)-related (for review, see [118]). The pathological changes in ASD patient are increased thickness of cortical tissue, abnormal interhemispheric and long-range cortico-cortical synchrony, resulting in a relative disconnection of IT neurons in the contralateral cortex, and reduced thickness of the corpus callosum. Causal genetic alterations have not yet been determined, but several genetic risk factors contribute to idiopathic ASD. Genetic variants of GluN2A and GluN2B in human have reportedly been associated with mental retardation, epilepsy, and autism [22, 57, 58]. One *de novo* mutation in GluN2B in a patient with

**Table 3. Affinities of compounds for recombinant NMDA receptors [116, 117].**

<b>Agonist and co-agonist</b>	<b>GluN2A</b>	<b>GluN2B</b>	<b>GluN2C</b>	<b>GluN2D</b>	<b>GluN3A</b>	<b>GluN3B</b>
<b>EC<sub>50</sub> (μM)</b>						
Glutamate	7.7	2.3	1	0.39		
Glycine	1.2	0.38	0.32	0.12	57	95
<b>Competitive antagonists</b>						
<b>Ki (μM)</b>						
(R)-AP5	0.28	0.46	1.6	3.7		
UBP141	14	19	4.2	2.8		
<b>Channel blockers</b>						
<b>IC<sub>50</sub> (μM)</b>						
(+)MK801	0.015	0.0099	0.024	0.038		
Ketamine	5.4	5.1	1.2	2.9		
Phencyclidine	0.82	0.16	0.16	0.22		
<b>Noncompetitive antagonists</b>						
<b>IC<sub>50</sub> (μM)</b>						
Ifenprodil	39	0.15	29	76		
Ro25-6981	52	0.009				
<b>Allosteric potentiators</b>						
<b>EC<sub>50</sub> (μM)</b>						
CIQ	NE	NE	2.8	3.0		

NE, no detectable effect.

ASD has been reported [47]. Using 151 Korean trios, a family-based association test (FBAT) provides a statistically significant association between ASD and *GRIN2B* haplotype [59]. *GRIN2B* gene alterations, including mutations and gene disruption by apparently balanced chromosomal rearrangements, have been described in patients with intellectual disability and ASD [119].

Rare mutations in *SHANK3* have been reported in patients with idiopathic ASD [60, 61]. Shank family proteins are scaffolding proteins that organize a cytoskeleton-associated signaling complex at the postsynaptic density of excitatory synapses, including NMDA receptors. Mutation in *SHANK3* induces ASD, however the population of *SHANK3* mutations is small in ASD [62]. Similarly, mutations in *SHANK2* and *SHANK1* associate with idiopathic ASD [61]. A *de novo* deletion in *SHANK1* is detected in an unrelated male individual with ASD with higher functioning [121]. A rare autosomal *SHANK1* deletion that is limited to males provides a possible contributory model for elucidating the male gender bias in autism [120].

Behavioral analyses of *Shank1* mutant mice have shown impairments in social interaction and communication, increased self-grooming, and repetitive behaviors [121, 122] but enhanced spatial learning and memory [123]. *Shank2* mutant mice carry a mutation that is identical to the ASD-associated microdeletion of *SHANK2* gene (exons 6 and 7) [124]. This mutation results in a markedly decrease in NMDA receptor

complex function and ASD-like behavior, containing reduced social interaction and communication (reflected by ultrasonic vocalizations, USVs), and repetitive jumping [124]. The NMDA receptor-AMPA receptor ratio in the synapse of *Shank2* mutant mice is reduced, and GluN2A and GluN2B subunit-containing NMDA receptors are equally affected in *Shank2* mutant mice [124]. *Shank2* mutant mice with exon 7 deletion showed similar behavioral abnormalities and NMDA receptor hyperfunction [125]. Several lines of *Shank3* mutant mice shows reduced social interaction and affiliation behaviors [126-129]. These mice exhibited alterations in the levels of synaptic glutamate receptors, that is, GluA1 (AMPA receptor subunit) in the hippocampus, GluA2 (AMPA receptor subunit), GluN2A and GluN2B in the striatum. *Shank3* knockdown with a small-interfering RNA (siRNA) caused significant reductions of ionic or synaptic currents *via* NMDA receptors and reduced surface GluR1 expressions in rat cortical cultures [130].

Neuroligins are neuronal postsynaptic cell adhesion molecules. Neuroligin-1 (NL-1) is preferentially distributed at excitatory synapses [132]. NL-1 KO mice are also known as an animal model of ASD that exhibits a marked increase in repetitive grooming behavior similar to increased repetitive behavior observed in autism. This repetitive grooming abnormality in NL1 KO mice is associated with decrease in the ratio of the NMDA to the AMPA at synapses in corticostriatal pathway [132].

Autism has been reliably associated with electrophysiological endophenotypes that may be caused by NMDA receptor disruption on parvalbumin (PV)-containing interneurons [133]. In human, M1 (N1 event-related potential as measured by magnetoencephalography) latency is shifted by approximately 10% in ASD patients [134]. The N1 event-related potential is a negative peak approximately 100 ms after sensory stimulation that is linked to early attention [135]. In PV containing cell-type selective GluN1 KO mice, delayed N1 event-related potential latency, reduced sociability, and impairment of mating-related USVs are observed [133]. In mice, social behavior is known to be expressed as social investigation, intermale aggression, sexual behavior, and parental behavior. When aggressive behavior or sexual motivation of the test mouse toward the stimulus mouse is low in the social interaction test, a significant correlation is found between delayed N1 latency and reduced sociability (i.e., social approach with same-sex gonadectomized mouse) but not between N1 latency and pre-mating USV power (i.e., social approach with female mouse) or T-maze performance [133]. Reduced sociability is one of core symptoms in the patients of ASD [134]. Poor USV emission mimics the social communication impairments in ASD. T-maze learning involves finding a food reward in one of two available locations at opposite ends of a T-shaped apparatus. Reversal requires the mouse to extinguish the location of the reinforcer in one arm and learn a new location of the reinforcer in the other arm. Failure to switch to the new position may be analogous to the inflexibility of routines that is characteristic of autism [136]. Therefore, the increased N1 latency, impairments of sociability and USVs in PV-selective GluR1 KO mice have some analogy with characteristic of autism [133]. Treatment with the NMDA receptor functional glycine site partial agonist GLYX-13 rescued the USV deficits [63]. Considering that GLYX-13 rescued the pro-social USV deficits, the reduction of NMDA receptor function may be an important therapeutic target for autism. Further investigation of the precise mechanisms that modulate the reduction of sociability, poor USV, and inflexible reversal learning is required.

Genetic alterations of GluN2A or 2B subunits are reported in humans with autism. As animal models of ASD, Shank mutations or NL1 KO alter NMDA receptor function and induce ASD-like behavior, including reduced sociability and stereotyped grooming behavior. PV-selective GluN1 KO mice exhibited delayed N1 latency, reduced sociability, impaired mating-related USVs, and inflexible T-maze learning. Various animal studies of ASD revealed that reduced NMDA receptor function in corticostriatal PV-containing neurons is involved in ASD-like behavior. The improvement of USV deficits by the partial agonist GLYX-13 in animal models might be extended to the treatment of autism in humans.

### NMDA Receptor Antagonist-Induced Psychosis

NMDA receptor antagonists, including PCP, ketamine, and MK-801, induce experimentally abnormal behavior in rodents and transient psychosis in humans. NMDA receptor antagonists overstimulate neurons, but locally administered PCP does not induce excitation. The mechanism of NMDA receptor antagonist-induced excitation has been hypothesized to involve the NMDA receptor-modulated inhibition of GABA interneurons and disinhibition of pyramidal neurons, consistent with reductions of extracellular GABA levels and elevations of extracellular glutamate levels [64-66].

NMDA receptor antagonists result in an increase of cortical high-frequency or gamma oscillations in animals, and these phenomena might be involved in psychosis [67, 68]. Systemic administration of an NMDA receptor antagonist increased the power of high-frequency oscillations. Tetrodotoxin infusion directly into the nucleus accumbens immediately and markedly reduced the power of accumbal high-frequency oscillations, that is correlated with changes in high-frequency oscillations recorded in distant cortical sites, suggesting that the nucleus accumbens is an important neural generator of oscillations [137].

Ketamine induces negative symptoms which are related to the degree of its occupancy of NMDA receptors [138]. Neuroimaging data suggest that the production of schizophrenia-like symptoms by ketamine is associated with frontal cortical activation, reflected by increases in frontal cortical perfusion [139-141], cortical glucose metabolism [142, 143], and glutamate levels [144]. Acute or chronic administration of ketamine both differentially and brain region-specifically modulates acetylcholine, dopamine, serotonin, and norepinephrine levels. Additionally, chronic administration of ketamine markedly reduces the glycine levels and induced gene expression changes of important neurotransmitter receptor systems, e.g. some members of the dopamine and serotonin receptor families [145]. Chronic administration of ketamine but not MK-801 elicited a reduction of the peak oscillatory frequency of kainic acid-elicited oscillations in *ex vivo* slices [146]. de Bartolomeisa *et al.* [147] show that ketamine and MK-801 induce the expression of Homer1a and Arc early genes in cortical regions, whereas ketamine and MK-801 reduce Homer1b and PSD-95 expression in cortical and striatal regions. These findings suggest that ketamine and MK-801 might have slightly different mechanisms of action in the induction of experimental psychotic-like behavior in rodents.

Systemic PCP also produces long-lasting excitation in the prefrontal cortex along with the enhanced locomotor activity and behavioral stereotypies in rats [148]. Comparisons of the effects of acute ketamine and PCP administration generally show that PCP increases impulsive responding. Ketamine does not

have the same effect like PCP on impulsive responding and consequently produces more subtle cognitive deficits in attentional set-shifting [149]. Interestingly, ketamine, which is more frequently used in clinical settings, does not result in extensive cognitive deficits induced by using PCP administration [149]. As previously mentioned, inhibition of the activity of GluN2B-containing NMDA receptors can be responsible for the majority of the antidepressant effects of ketamine. Although ketamine can act on the GluN2B subunit, the influence of GluN2B antagonism by ketamine may differ between wildtype and disease-model mice. Acute application of Ro 25-6981, a GluN2B-selective antagonist, rescues LTP and gamma oscillation deficits in slices derived from Ts65Dn mice (i.e., a Down syndrome model mice associated with cognitive impairment), whereas prolonged treatment induces the persistent rescue of LTP [150]. In contrast, Ro 25-6981 has no effect on LTP in wildtype mice but reduces gamma oscillations both acutely and following prolonged treatment [150].

PCP is not known to have antidepressive actions. Chronic PCP exposure intensely affects the relative immunoreactivity of GluN2 subunits in the frontal cortex. PCP induces significant increases in GluN2D immunoreactivity and protein expression, and a shift to a predominance of the GluN2D subunit in the frontal cortex [151]. Recently, we showed that the action of PCP involves the GluN2D subunit. In wildtype and GluN2A KO mice, PCP increases locomotor activity and extracellular dopamine levels in the striatum and prefrontal cortex, but not in GluN2D KO mice [69] (Table 4). Acute or repeated administration of PCP is not able to increase locomotor activity in GluN2D KO mice [69] (Table 4). These results indicate that acting of PCP at the GluN2D subunit induces enhanced dopaminergic transmission and increases in locomotor activity. To determine the precise mechanism of action of PCP, the effects of a GluN2C/2D antagonist and GluN2C/2D potentiator were investigated in wildtype and GluN2D KO mice. PCP and UBP141 (a GluN2C/2D antagonist) induced potent motor impairment in wildtype mice but not in GluN2D KO

mice [70] (Table 4). In contrast, CIQ, a GluN2C/2D potentiator, induced severe motor impairment in GluN2D KO mice but not in wildtype mice [70] (Table 4), suggesting that the GluN2D subunit plays an essential role in the effects of PCP and UBP141, and an appropriate balance between the GluN2C and GluN2D subunits might be needed for appropriate motor performance. GluN2D subunits mainly exist in brainstem structures, the globus pallidus, the thalamus, and the subthalamic nucleus. The *c-fos* gene is differentially and PCP-dependently expressed in wildtype and GluN2D KO mice, and the number of Fos-positive cells increases after PCP administration in the basal ganglia motor circuit in wildtype mice but not in GluN2D KO mice [70] (Table 4). These results suggest that the GluN2D subunit within motor circuitry is a key subunit for PCP-induced motor impairment, which requires an intricate balance between GluN2C- and GluN2D-mediated excitatory outputs.

PCP and ketamine are used as experimental psychosis-inducing drugs in rodents. Prenatal treatment with PCP causes cognitive impairment of memory, PCP sensitization, and deficits of sensorimotor gating [152, 153]. These behavioral deficits may be caused by impairments in the neuronal progenitor proliferations and decreased densities of glutamatergic neurons in the prefrontal cortex following prenatal PCP treatment [154]. Prenatally PCP-administered mice display behavioral deficits in cognitive memory and sensorimotor gating until adulthood, and they are used as an experimental model of schizophrenia. The administration of D-serine improved the PCP-induced reduction of prepulse inhibition and cognitive deficits [71, 72]. Glycine type I (GlyT1) transport inhibitors (GTIs) are applied for PCP-induced deficits and have been proven effective (for review, see [73]). Furthermore, the cognitive impairment and behavioral effects caused by NMDA receptor antagonists may be reduced by facilitating GABAergic neurotransmission [155], administering metabotropic glutamate 2/3 receptor agonists [74-76], or prior embryonic medial ganglionic eminence cell transplantation in the medial prefrontal cortex that

**Table 4. Differential effects of antagonists or potentiators between GluN2 subunit genotypes.**

	Treatment	WT	GluN2A KO	GluN2D KO	Ref. No.
<b>Dopamine release (<i>in vivo</i> brain dialysis)</b>	PCP (3 mg/kg, s.c.)	↑↑	↑↑	→	[69]
<b>Locomotor activity</b>	PCP (3 mg/kg, s.c.)	↑↑	↑↑	→	[69]
<b>Sensitization</b>	Subchronic PCP (3 mg/kg, s.c.)	↑↑	→	→	[69]
<b>Motor performance (rotarod test)</b>	PCP (3 or 5 mg/kg, s.c.)	↓↓	n.d.	↓	[70]
	UBP141 (3 mM, 20 μl, i.c.)	↓↓	n.d.	→	[70]
	CIQ (20 mg/kg, i.p.)	→	n.d.	↓↓	[70]
<b><i>c-fos</i> expression</b>	PCP (10 mg/kg, s.c.)	↑↑↑	n.d.	↑ or →	[70]
	UBP141 (3 mM, 20 μl, i.c.)	↑	n.d.	→	[70]
	CIQ (20 mg/kg, i.p.)	↑↑↑	n.d.	↑↑↑	[70]

n.d., not determined.



many of these cells differentiate into cortical GABAergic interneurons [78]. Systemic administration of CIQ (a positive allosteric modulator selective for GluN2C/GluN2D-containing NMDA receptors) reversed deficits induced by MK-801 in prepulse inhibition in mice [77].

## CONCLUDING REMARKS

NMDA receptors have been implicated in physiological processes, and the dysfunction of NMDA receptors is known to be involved in various psychiatric disorders. The hypofunction of NMDA receptors might be a common mechanism that underlies schizophrenia and anti-NMDA receptor encephalitis. Unique findings related to the physiological functions of the GluN2B subunit in mood disorders provide important information that may have wide clinical applications. Our finding that PCP preferentially acts on the GluN2D subunit *in vivo* suggests the possibility that the GluN2D molecule might be a main target related to PCP-induced psychosis. Subunit-selective treatments may have important clinical implications.

## CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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